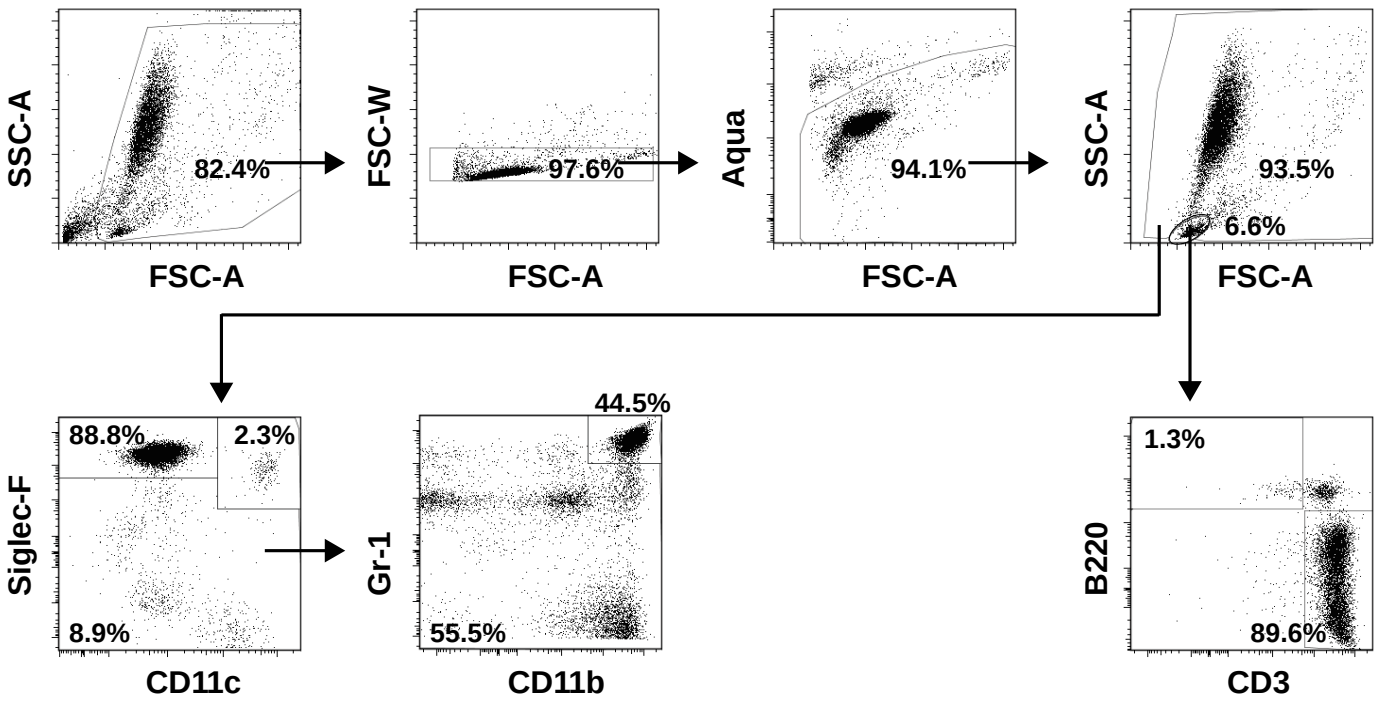
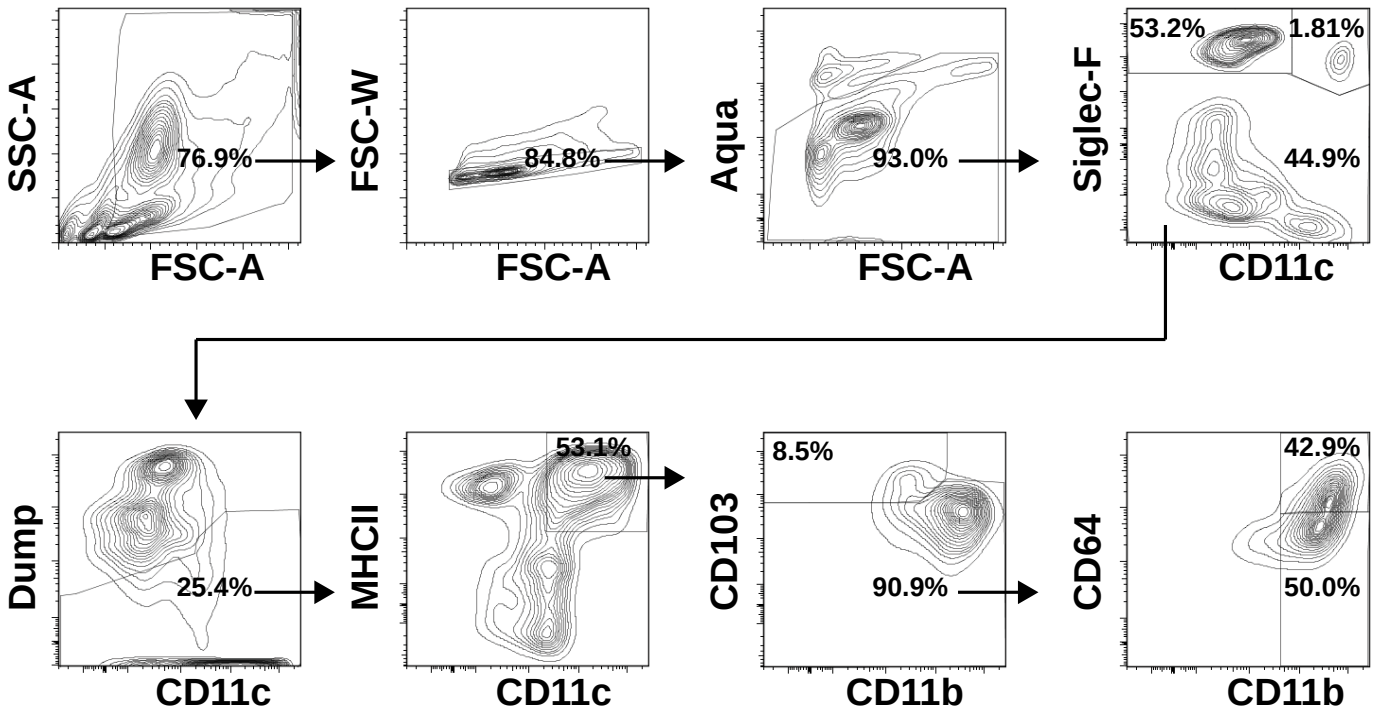


Supplemental figure 1. Gating strategy for flow cytometry analysis of cells in the BAL fluid. Cells were Aqua stained and formaldehyde fixed. Single, alive ($Aqua^-$) cells were separated into lymphocytes and non-lymphocytes based on the FSC/SSC of the different populations. In the non-lymphocyte gate, eosinophilic granulocytes ($CD11c^-Siglec\text{-}F^+$), alveolar macrophages ($CD11c^+Siglec\text{-}F^+$), and neutrophilic granulocytes ($CD11b^+Gr\text{-}1^+$) were distinguished. In the lymphocyte gate, T cells ($CD3^+$) and B cells ($B220^+$) were analysed.



Supplemental figure 2. Gating strategy for flow cytometry analysis of DC subtypes in lung tissue. Cells were Aqua stained and formaldehyde fixed. From single, alive (Aqua$^-$) cells the eosinophils (CD11c$^-$ Siglec-F$^+$) and alveolar macrophages (CD11c$^+$Siglec-F$^+$) were distinguished. A dump channel made it possible to delete T cells (CD3$^+$), B cells (B220$^+$), neutrophils (Gr-1$^+$) and NK cells (NK1.1$^+$) from the analysis. Dendritic cells (CD11c$^+$MHCII$^+$) were separated in CD103$^+$ and CD103$^-$ cells. From the CD103$^-$ DC a distinction was made in CD64$^+$ (monocyte derived, pro-inflammatory DCs) and CD64$^-$ DCs.