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IL-1RL2 and Its Ligands Contribute to the Cytokine Network in Psoriasis

Hal Blumberg,*† Huyen Dinh,* Charles Dean, Jr.,* Esther S. Trueblood,* Keith Bailey,* Donna Shows,* Narasimharao Bhagavathula,† Muhammad Nadeem Aslam,† James Varani,† Jennifer E. Towne,* and John E. Sims*

Psoriasis is a chronic immune-mediated disease in European populations; it is characterized by inflammation and altered epidermal differentiation leading to redness and scaling. T cells are thought to be the main driver, but there is also evidence for an epidermal contribution. In this article, we show that treatment of mouse skin overexpressing the IL-1 family member, IL-1F6, with phorbol ester leads to an inflammatory condition with macroscopic and histological similarities to human psoriasis. Inflammatory cytokines thought to be important in psoriasis, such as TNF-α, IL-17A, and IL-23, are upregulated in the mouse skin. These cytokines are induced by and can induce IL-1F6 and related IL-1 family cytokines. Inhibition of TNF or IL-23 inhibits the increased epidermal thickness, inflammation, and cytokine production. Blockade of IL-1F6 receptor also resolves the inflammatory changes in human psoriatic lesional skin transplanted onto immunodeficient mice. These data suggest a role for IL-1F family members in psoriasis. The Journal of Immunology, 2010, 185: 000–000.

Psoriasis appears to involve a cytokine network centered around IL-17, IL-22, IL-23, and TNF, all of which are elevated in lesional skin (14–17). IL-22 (along with its related cytokines IL-19, IL-20, and IL-24) was shown to cause epidermal hyperplasia, primarily by downregulation of genes involved in terminal keratinocyte differentiation (18, 19). IL-17 acts on keratinocytes to induce chemokines that lead to neutrophil, inflammatory dendritic cell (TNF/iNOS-producing dendritic cells, or TIP-DC) (20), and Th17 cell influx into the skin (21). IL-17 and IL-22 induce keratinocytes to produce antimicrobial peptides, such as S100A8, S100A9, β-defensin 2, and LL-37, known to be elevated in lesional skin (22–24). These protect against infection and act as endogenous ligands for TLRs, such as TLR4 and TLR9 expressed on keratinocytes and dendritic cells (DCs) (25, 26). Human β-defensin 2 is also chemotactic for CCR6+ cells (27), which include neutrophils, DCs, and Th17 cells, and for CCR2+ cells (28), which include TIP-DCs. IL-23 enhances the production of IL-17 and IL-22 from Th17 and other cells (29, 30). TIP-DCs make inducible NO synthase (leading to NO, which can dilate dermal blood vessels) and secrete TNF, IL-20, and IL-23 (31–35). TNF amplifies many of these responses, further activating DC populations in the skin and inducing cytokines, such as IL-1, IL-6, and IL-8, from keratinocytes and fibroblasts to promote continued Th17 differentiation and neutrophil recruitment. Neutrophils, in turn, can cause tissue damage via reactive oxygen species and proteases, which exposes self-Ags and generates endogenous TLR ligands. TLR stimulation of skin-resident DCs causes IL-12 synthesis, which can promote Th1 development, IFN-γ production, and generation of CXCR3 chemokines that recruit more T cells to the lesion (36). There may be a role for IFN-α in maturation of DCs, particularly at plaque initiation (6). Known triggers for psoriasis include trauma and infection, resulting in generation of exogenous or endogenous TLR ligands and other pathogen-associated and damage-associated molecular patterns, which may provide entry points into the cycle of mutually reinforcing gene expression and cell-recruitment loops described above.

The IL-1 family contains 11 members (37, 38). IL-1α, IL-1β, IL-18 and IL-33 all have known roles as agonists affecting inflammation and/or adaptive immunity, whereas IL-1ra acts to inhibit...
IL-1α and IL-1β action. IL-1F6, -1F8, and -1F9 are agonists of the IL-1R family member IL-1R1L2 (also known as IL-1R rp2) (39, 40). IL-1F5 serves to antagonize these responses in a manner parallel to that used by IL-1ra for IL-1 responses (39) (J.E. Towne, B.R. Renshaw, J. Douangpanya, B.P. Lipsky, M. Shen, C.A. Gabel, J.E. Sims, unpublished data). Global-expression analysis of IL-1 family members demonstrated that IL-1F6, -1F8, and -1F9 were highly expressed in only a few tissues (39, 41–43) (data not shown). Among these, the most abundant expression was in skin. We surveyed a variety of human inflammatory skin conditions and found that IL-1F6, -1F8, and -1F9 were upregulated in psoriatic lesions (44) (H. Blumberg, H. Dinh, D. Shows, and J.E. Sims, unpublished data). Microarray studies of psoriatic skin also found upregulation of IL-1F9 (45).

We previously reported that transgenic overexpression of IL-1F6 under control of the keratin-14 promoter leads to an inflammatory skin phenotype at birth, which resolves after 2–3 wk of age. A number of inflammatory cytokines and chemokines are upregulated in the inflamed skin, and TNF inhibition leads to a decrease in epidermal thickness. In this article, we demonstrate that application of an irritant can rapidly induce psoriatic-like skin inflammation in phentypically normal skin from 2–3-mo-old transgenic mice. We characterize the skin changes and cytokine involvement in much more detail than in our previous publication. We show that cytokines, such as IL-17, IL-22, and IL-23, known to be involved in human psoriasis, are overexpressed in this model and that they can induce IL-1F6, which, in turn, can induce IL-17, IL-22, and IL-23, thus establishing a self-amplifying gene-expression loop. We also provide a direct link to human psoriasis by demonstrating that agents approved for clinical treatment of psoriasis are beneficial in this group) received 12.5 μg TPA (Sigma-Aldrich, St. Louis MO) in 200 μl acetone by topical administration to the shaved back. At 4 or 48 h (day 6) after the second TPA injection, photos were taken for gross observation, and full-thickness back skin was excised and divided into three sections: lower back skin was saved in formalin for histopathology, midback skin was processed for RNA, and upper back skin (collected using a 8-mm biopsy punch to obtain a fixed area) was snap-frozen in liquid nitrogen for protein analysis. Abs and soluble TNFRp75Fc (500 μg each) were injected i.p. on days −1 and 3.

Intradermal injections

Eight- to nine-week-old male FVB mice (Tacronic Farms) were injected intradermally into back skin on 2 consecutive days (two injections; short protocol) or every other day for 12 d (six injections; long protocol) with 500 ng each cytokine or PBS. Four hours after the final injection, full-thickness skin around the injection site was harvested for RNA isolation.

RNA isolation, cDNA synthesis, and quantitative PCR

Total skin RNA was extracted using the RNeasy mini kit (Qiagen, Valencia, CA), and first-strand cDNA synthesis was accomplished using High-Capacity cDNA Reverse Transcription Kits (Applied Biosystems, Foster City, CA), according to the manufacturers’ directions. Quantitative RT-PCR was done using a customized TaqMan low density array (TLDA) plate with 62 query genes and 2 control genes (HPRT1 and 18S rRNA). Genes not on the TLDA plate were analyzed using Assay-on-Demand primer and probe sets (Applied Biosystems). Assay numbers (Mm00439307_m1, Mm00426321_m1, Mm00321041_g1, Mm00569256_m1, Mm00438270_m1, Mm004238_m1, Mm009990114_s1, Mm00438285_m1, Mm00731768_m1, Mm00806979_m1, Mm00454341_m1, Mm00519250_m1, Mm01291775_m1, Mm0043919_1_m1, and Mm01281447_m1) used were as follows: HB-EGF, TGF-α, S100A8, S100A9, CCR2, CCL20, CCR6, CAMP, BDE4F, BDEF14, IL-20, IL-1RL2, STAT3, and APEX, respectively.

Multianalyte profiling-analysis protocol and details

Full-thickness skin samples from each group were collected with 8-mm biopsy punches and homogenized in 1.6 ml lysis buffer (Cell Signaling Technology, Beverly, MA), containing one protease inhibitor tablet EDTA-free (Roche, Indianapolis, IN). Aliquots of the lysates (250 μl) were sent to Rules-Based Medicine (Austin, TX) for multianalyte profiling. Additional ELISAs were run for IL-17E, IL-20, and IL-22 using specific duo kit ELISAs (R&D Systems).

Skin transplantation and treatment protocol

Replicate 6-mm punch biopsies of full-thickness plaque skin were obtained from human skin donors with psoriasis. Sun-protected skin from nonpsoriatic donors was obtained as control. This project was approved by the University of Michigan Institutional Review Board. All subjects provided written informed consent prior to biopsy. SCID mice (CB-17 strain; Tacronic Farms) were used as tissue recipients. One piece of tissue from each normal or psoriatic volunteer was transplanted onto the dorsal surface of a recipient mouse as follows. After the animal was anesthetized, the dorsal surface of the mouse was shaved. Mouse skin was surgically removed to size and replaced with the human tissue. This tissue was secured to the back of the mouse with absorbable sutures (4-0 Dexon “S”, Davis-Geck, Manati, Puerto Rico). The transplant was bandaged with Xeroform petrolatum dressing (Kendall, Mansfield, MA) for 5 d. The animals were maintained in a pathogen-free environment throughout the preparation and treatment phases. Treatment was initiated 1–2 wk posttransplantation, depending on the healing rate of the transplanted tissue. Animals with the human skin transplants were divided into treatment groups (isotype control Ab, anti-IL-1RL2 Ab, or etanercept as a positive control). Animals were treated with seven injections each of 150 μg anti-human IL-1RL2, isotype matched control Ab, or etanercept i.p. on alternate days. All procedures involving animals were approved by the University of Michigan Committee on Use and Care of Animals. At the end of the treatment phase, animals were photographed and then euthanized. The transplanted human tissue along with the surrounding mouse skin was surgically removed and fixed in 10% formalin. After embedding the tissue in paraffin, multiple 5-μm sections were cut from each tissue piece (~50 μm between sections), mounted on microscope slides, and stained with H&E.

Immunohistochemistry

For cytokeratin 6 and CD3 immunohistochemistry (IHC), skin samples were fixed in 10% neutral buffered formalin and embedded in paraffin. Deparaffinized tissue sections were subjected to Ag retrieval using Citra solution (no. HK086-9K; BioGenex, Sun Ramon, CA) in a Decloaking Chamber (Biocare, Concord, CA). Tissue sections were incubated with a 1:500 dilution of the cytokeratin 6 Ab (no. PRB-169P; Covance, Berkeley, CA) or with a 1:200 dilution of the CD3 Ab (no CP215C; Biocare) at room temperature for
Detection was performed with an anti-rabbit Mach 3 Rabbit AP Polymer Kit (nos. RP531L & RAP533L; Biocare), followed by Permanent Red chromogen solution (no. K0640; Dako North America, Carpinteria, CA). For the CD11c IHC, skin samples were frozen in OCT Compound (no. 4583; Sakura, Torrance, CA). Tissue sections were incubated with a 1:100 dilution of a biotinylated CD11c Ab (no. 553800; BD Biosciences, Pharmingen, San Diego, CA) at room temperature for 60 min. Detection was performed with streptavidin-alkaline phosphatase (no. NEL750; PerkinElmer, Waltham, MA) with tyramide signal amplification (no. SAT700B; PerkinElmer), followed by Permanent Red solution. For the CD31 IHC, skin samples were fixed in IHC zinc fixative (no. 550523; BD Biosciences, Pharmingen) and embedded in paraffin. Deparaffinized tissue sections were incubated with a 1:30 dilution of CD31 Ab (no. 533370; BD Biosciences Pharmingen) for 60 min at room temperature. Detection was performed with a Vectastain ABC-AP kit (no. AK-5000; Vector Laboratories, Burlingame, CA), followed by Permanent Red solution. Following the IHC-staining procedure, slides were counterstained with hematoxylin (no. S3309; Dako North America), dehydrated, cleared, and coverslipped. To quantify changes in the number of CD3+ and CD11c+ cells and the area of CD31+ vessels in response to TPA treatment, five representative digital images of each IHC assay from each individual animal were analyzed using MetaVue morphometry software (version 6.2r6; Universal Imaging, Downingtown, PA). The CD3 and CD11c images were taken with an ×20 microscope objective (∼200 magnification) and are expressed as the number of positive cells per mm². The CD31 images were taken with an ×40 microscope objective (∼400 magnification) and are expressed as the CD31+ area in μm²/mm² of dermis evaluated.

Results

We previously reported that transgenic overexpression of IL-1F6 in mouse skin results in a hyperproliferative, inflammatory skin condition in newborn animals that resolves by 3 wk of age only to reappear at ~6 mo (44). Resolution could be prevented and the phenotype exacerbated by eliminating one copy of the IL-1F5 an-
agonist gene (44). We have now found that treatment of transgenic mouse skin with TPA at a time when it is phenotypically normal (≈2–3 mo of age) elicits skin inflammation to a much greater extent than TPA treatment of nontransgenic skin (Fig. 1A, 1B, Supplemental Fig. 1A). The morphology of TPA-treated transgenic skin is similar to lesional skin of human psoriasis. Macroscopically, the skin appears reddened, thickened, scaly, and crusted (Fig 1A). Histologically, there is epidermal hyperplasia (acanthosis); a thickened stratum corneum (hyperkeratosis) containing nucleated cells (parakeratosis); neutrophilic microabscesses in the stratum spinosum and the stratum corneum; a mixed dermal infiltrate containing macrophages/DCs, neutrophils, and lymphocytes; and an increase in and dilution of superficial dermal blood vessels (as evidenced by CD31 staining, which is increased 6-fold in TPA-treated transgenic mice compared with 1.8-fold in TPA-treated wild-type mice) (Fig. 1B, 1C, Supplemental Fig. 1A, 1B, 1D). A granular layer (normally difficult to see in mouse skin) becomes prominent in the hyperplastic skin of TPA-treated wild-type mice but is mostly lost in TPA-treated transgenic mice, particularly in areas of more severe lesion. Although the most obvious epidermal penetrations into the dermis are associated with hair follicles, there is more unevenness to the lower margin of the epidermis than is typical for mouse, suggestive of rete ridges found in psoriatic skin. Cytokeratin 6, indicative of proliferating keratinocytes, is expressed throughout the epidermis (Fig. 1D) in wild-type and transgenic mice in response to TPA. However, it is noteworthy that even in apparently normal untreated transgenic mice, cytokeratin 6 staining reveals focal patches of mildly proliferative skin (Fig. 1D, Supplemental Fig. 1C), suggesting that the transgenic skin is poised on the brink of abnormal proliferative changes. These focal proliferative patches are not seen in wild-type mice. CD3+ T cells are also increased in the epidermis after TPA treatment in transgenic and wild-type mice (2.3-fold and 1.8-fold, respectively) (Fig. 1E, Supplemental Fig. 1D). There is a significant increase in CD11c+ DCs, especially in the upper layers of the dermis (9.7-fold in transgenic mice, 2.7-fold in wild-type mice after TPA treatment) (Fig. 1F, Supplemental Fig. 1D). An excess of DCs over T cells, as seen in the transgenic mice after treatment with TPA, is typical of psoriasis (46). Although the general presentation is similar to that of human psoriatic skin, there are also points of difference. The most obvious of these is the role of T lymphocytes. Disease in transgenic mice occurs on a lymphocyte-deficient rag2−/− background (Fig. 1G), suggesting that T cells are not required, whereas T lymphocytes are believed to play a key role in human psoriasis. Other differences include the variable presence or loss of a granular-appearing layer beneath the stratum corneum, the relative paucity of lymphocytes in the infiltrate in the mouse disease, and the uncertain presence of rete ridges. Elicitation of skin hyperplasia and inflammation by TPA is dependent on the IL-1F6 transgene, because it can be prevented by injection of an anti–IL-1RL2 neutralizing mAb (Fig. 1H). These findings made us wonder whether IL-1F6, -1F8, and/or -1F9 might account for part of the epidermal contribution to human psoriasis.

**Gene-expression patterns**

To probe the similarity between human psoriasis and skin inflammation caused by overexpression of IL-1F6, we examined the expression of a number of genes relevant to skin or inflammation by quantitative PCR. Skin samples were taken from nontransgenic and K14/F6 transgenic mice, either untreated or treated with TPA. Genes substantially upregulated in transgenic animals treated with TPA are indicated in Fig. 2. TaqMan quantitative PCR was performed, and the cycle threshold (CT) value for each gene was normalized to that for HPRT. The normalized values for selected genes are shown. Each data point represents one mouse. The figure is representative of two experiments.
TPA, but not in other groups, included cytokines, chemokines, and antimicrobial peptides that are known to be upregulated in psoriasis (Fig. 2, Supplemental Table 1). These include IL-17A, IL-22, IL-23 (both p19 and p40 subunits), and the antimicrobial peptides S100A8, S100A9, and β-defensin 4 (the ortholog of human β-defensin 2). IL-1β, a regulator of Th17 differentiation, was also upregulated, as were IL-19, IL-20, and IL-24, cytokines related to IL-22 and possessing similar activities. TNF-α was upregulated by TPA in transgenic and nontransgenic mice.

**Mutual induction of inflammatory cytokines and IL-1 family proteins**

To establish directly whether IL-1F6 is capable of inducing genes implicated in psoriasis pathology, we injected IL-1F6 intradermally into wild-type mice and analyzed transcript levels in skin for selected genes (Fig. 3A, Supplemental Table 2). IL-1F6 led to substantial increases in IL-17A, IL-23, TNF, and IFN-γ mRNA. IL-1F6 also strongly induced itself, as well as other IL-1 family members, chemokines, growth factors, and antimicrobial proteins.

We next asked whether cytokines known to be elevated in psoriatic skin could induce IL-1F6 and other family members. TNF, IL-17A, IL-23, and IFN-γ were able to induce IL-1F6 mRNA following intradermal injection of cytokines into wild-type mice (Fig. 3B, Supplemental Table 3, data not shown). Induction was usually stronger when combinations of these cytokines, as well as IL-22, were used (Supplemental Table 3). Some of these cytokines also induced IL-1F8 and IL-1F9 (Supplemental Table 3). Thus, a positive feedback loop may exist in psoriatic skin, with cytokines, such as IL-17, IL-22, IL-23, and TNF, inducing IL-1F6 and IL-1F9, which, in turn, amplify expression of the previously implicated inflammatory cytokines as well as themselves.

**Response to therapeutics**

TNF inhibitors are effective in treating psoriasis (47) and are approved for this purpose in many countries. Abs that block the action of IL-12 and IL-23 have proven successful in clinical trials (48), and one is approved for treating psoriasis in the European Union, Canada, and the United States. Human-expression data and genetic-susceptibility studies suggest that Abs inhibiting the action of IL-23 alone would be effective as well (16, 49, 50). Because the morphologic appearance of and gene-expression patterns in the skin of K14/F6 transgenic mice treated with TPA were similar to those of human psoriatic skin, we asked whether agents known or suspected to be efficacious in treating psoriasis would have therapeutic effects in the mice. K14/F6 transgenic mice were treated with candidate agents on days −1 and 3 and with TPA at days 0 and 4. Mouse skin was evaluated macroscopically and histologically at day 6, and samples were taken for mRNA and protein analysis at 48 h after the second TPA application. Therapeutic agents examined were soluble TNFRp75-Fc, an anti–IL-12/23p40 Ab, and an anti–IL-23p19 Ab. Mice treated with any of these agents showed a markedly improved appearance of the skin (Fig. 4). The skin was also improved histologically, with reduced acanthosis, hyperkeratosis, parakeratosis, inflammatory cell infiltrate, dilation of dermal capillaries, and loss of granular layer compared with skin from mice treated with isotype-matched control Abs (Fig. 4). Moreover, gene-expression studies (Fig. 5, Supplemental Table 4) and protein analyses (Supplemental Table 5) showed reductions in mRNA for IL-17A and -17F, IL-22, IL-1β, chemokines, and antimicrobial peptides after treatment, as well as reductions in protein levels for most of these as well (IL-17A protein was below the level of detection for the assay used, and protein levels were not analyzed for antimicrobial proteins). For all of these features, inhibition of IL-12/23p40 or IL-23p19 was more effective than TNF inhibition, although the
effectiveness of TNF inhibition varied from modest to excellent in different experiments. These results indicate that the psoriasis-like pathology in the skin of TPA-treated transgenic mice is responsive to the same therapeutic approaches used effectively for human psoriasis.

Mouse and human skin are quite different. Therefore, many investigators have studied psoriasis preclinically by transplanting skin from psoriasis patients onto immunodeficient mice (5, 6). Technical limitations, especially the small amount of tissue available, make this approach challenging. However, the results obtained provide important insights into the pathogenesis of psoriasis and the potential for developing new therapies.

**FIGURE 4.** Skin pathology in K14/F6 mice treated with TPA is ameliorated by inhibitors of TNF, IL-12/23p40, and IL-23p19. Transgenic mice were treated with TPA to induce skin inflammation and were given antagonists of TNF (TNFRp75-Fc), IL-12/23p40, or IL-23p19 or isotype control Ab, as described in Materials and Methods. The photographs show the macroscopic appearance of the skin, as well as skin sections stained with H&E, taken 2 d after the second TPA application. Treatment with anti–IL-12/23p40 or anti–IL-23p19 led to reduction in acanthosis, hyperkeratosis, and parakeratosis, as well as to loss of the granular cell layer, inflammatory infiltrate, and dermal blood vessel prominence. Rare intracorneal pustules are still present in mice treated with anti–IL-23p19 or anti–IL-12/23p40 (data not shown) Ab. Treatment with TNFRp75-Fc led overall to less severe disease than in the control mice, but parakeratosis, loss of the granular cell layer, and intracorneal and intraepidermal pustules persist in some areas. The figure is representative of three experiments. Scale bar, 0.02 mm. Original magnification ×200.

**FIGURE 5.** Inhibition of TNF, IL-12/23p40, or IL-23p19 results in decreased expression of cytokines, antimicrobial peptides, and chemokines in K14/F6 mouse skin treated with TPA. Transgenic mice were treated with TPA to induce skin inflammation and were given antagonists of TNF (TNFRp75-Fc), IL-12/23p40, or IL-23p19 or isotype-matched control Ab, as described in Materials and Methods. RNA was prepared from full-thickness skin sections harvested 48 h after the second TPA treatment. TaqMan quantitative PCR was performed, and the CT value for each gene was normalized to that for HPRT. The normalized values for selected genes are shown. Each data point represents one mouse. The figure is representative of two experiments for anti-TNF and anti-p40 treatment and of six experiments for anti–IL-23p19 treatment.
mouse skin inflammation, we have not investigated whether the T cells play important roles in disease. Although T cells are not required for the mice for each patient. Original magnification 200×.

Representative histology images of H&E stained skin sections are shown from four patients transplanted onto at least two mice in the experiment shown. Thus, forced IL-1F6 expression would simply be a different entry point to that provided by activated T cells for the upregulation of a cytokine network similar to that found in psoriasis. In the absence of T cells, it is not known whether other cytokines, such as TNF, IL-20, and IL-24, and IL-1F6 proteins can substitute for the key functions of IL-17 and IL-22 or whether IL-17 and IL-22 might still be present but made by other, non-rag2–dependent cell types. It is notable that mast cells can make IL-17 (53), that neutrophils contribute IL-17 during *Leishmania* infection (54), and that NK cells and DCs can express IL-22 (55, 56). In addition, IL-17 is induced, and IL-23 is critical for disease, in two T cell-independent colitis models (57, 58).

The mechanism by which TPA induces disease in K14/F6 mice is not understood. Although IL-1F6 mRNA expression is considerably higher in TPA-treated transgenic skin than it is in TPA-treated wild-type skin (Supplemental Tables 1, 6), it is induced only 3.5-fold by TPA in the transgenic skin, and the F5:F6 antagonist:agonist mRNA ratio is reduced by only 2-fold after TPA induction in the transgenic mouse (Supplemental Table 6). Changes in expression of other IL-1 family genes after TPA treatment are comparably modest. These small changes seem unlikely to account for the marked change in skin phenotype that follows TPA treatment of transgenic mouse skin. We recently found that the sp. act. of IL-1F6 is increased ~10,000-fold by proteolytic processing near the N terminus (J.E. Towne, B.R. Renshaw, J. Douangpanya, B.P. Lipsky, M. Shen, C.A. Gabel, J.E. Sims, unpublished data). TPA is known to induce expression of a large number of genes in mouse skin (Supplemental Table 1) (1), including proteases. It is possible that among these genes are required for activation and secretion of IL-1F6. Alternatively, TPA leads to an influx of macrophages, neutrophils, and T cells into skin (59). One or more of these cell types or their products might be important to complement the action of IL-1F6 in setting up the self-amplifying gene-expression loop. In this scenario, the preconditioning of the K14/F6 transgenic mouse skin to perturbation by TPA would be regarded as analogous to the preconditioning of nonlesional psoriatic skin to respond to trauma in the well-known Koebner reaction.

Transgenic overexpression or knockout of other IL-1 family members in skin has led to various inflammatory conditions (60–62), but none show the many points of similarity to human psoriasis that are seen with overexpression of IL-1F6. Skin conditions resembling psoriasis to varying extents have also been created by manipulation of a number of other genes in mice (3, 4, 63). However, mouse models are perhaps less successful at reconstructing human disease than they are at allowing exploration of genes and pathways plausibly relevant to human conditions. TPA-treated K14/F6 transgenic mice develop a skin inflammation characterized by increased expression of many genes characteristic of human psoriasis, which is ameliorated by agents known to work clinically in psoriasis patients (TNF and IL-12/23p40 inhibitors). Amelioration by an anti–IL-23p19 Ab suggests that IL-23, rather than IL-12, is the dominant p40-containing cytokine involved in the mouse disease. Ligands for IL-1RL2 are also critical for maintaining the psoriatic characteristics of human lesional skin, at least when transplanted onto...
immunodeficient mice. The data presented in this article suggest that agents that block signaling through IL-1R2 could be useful therapeutically in psoriasis.

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H.D., C.D., E.S.T., K.B., D.S., J.E.T., and J.E.S. are employees of Amgen. N.B., N.D.A., and J.V. have no financial conflicts of interest.

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Supplemental Figure and Table Legends

Supplemental Figure 1. Histological analysis of skin from K14/F6 transgenic mice treated with TPA demonstrates features found in human psoriatic skin. (A) A skin section from transgenic mice showing an intracorneal pustule (*), parakeratosis (#), epidermal hyperplasia (@), and a mixed dermal infiltrate (&). Magnification: 10x. (B) Key features of the skin inflammation found in transgenic mice treated with TPA [dilated dermal vessels (lower middle) revealed by CD31 staining]. Magnification: 60x. (C) Low-power sections of wildtype and transgenic mouse skin treated with TPA, vehicle or nothing and stained for cytokeratin 6 (CK6), to show the character and abundance of the CK6+ areas. Magnification: 2x. (D) Quantitation of CD3+ and CD11c+ cells and of the area of CD31+ vessels in response to TPA treatment. Figure A is representative of five experiments; Figures B, C and D are from one experiment with three mice in each group, all of which showed similar features. TG, transgenic; WT, wildtype.

Supplemental Table 1. Analysis of expression of selected genes in the skin of K14/F6 mice treated with TPA. Transgenic mice and non-transgenic controls were treated with nothing (naïve), acetone (vehicle), or TPA as described in Methods. RNA was prepared from full-thickness skin sections harvested either 4 hr or 48 hr after the second TPA treatment. Taqman quantitative PCR was performed and the CT value for each gene normalized to that for HPRT. The normalized value for each gene for each condition is shown +/- standard deviation (SD), as well as relevant ratios of normalized values. Group sizes were: tg + TPA, 4hr: 7 mice; tg + acetone, 4hr: 6 mice; wt + TPA, 4hr: 6 mice; wt + acetone, 4hr: 4 mice; tg + TPA, 48hr: 6 mice; tg + acetone, 48hr: 7 mice;
wt + TPA, 48hr: 6 mice; wt + acetone, 48hr: 4 mice. Ac, acetone; tg, transgenic; wt, wildtype. The data is from one of two similar experiments.

**Supplemental Table 2. Genes induced in wildtype mouse skin by IL-1F6.** Wildtype FVB mice were injected intradermally with IL-1F6 or PBS, either on two successive days or using six injections over 12 days, as described in Methods. RNA was prepared from full-thickness skin sections harvested 4 hr after the last injection. Taqman quantitative PCR was performed and the CT value for each gene normalized to that for HPRT. The normalized value for each gene for each injection series is shown, as well as relevant ratios of normalized values. Five mice were used in each group. SD, standard deviation. The data is from one of four similar experiments.

**Supplemental Table 3. Induction of IL-1 family members in wildtype mouse skin by cytokines.** Wildtype FVB mice were injected intradermally with the cytokines listed or with PBS, using 6 injections over 12 days, as described in Methods. RNA was prepared from full-thickness skin sections harvested 4 hr after the last injection. Taqman quantitative PCR was performed and the CT value for each gene normalized to that for HPRT. The normalized value for each gene for each injection series is shown, as well as relevant ratios of normalized values. Five mice were used in each group. The data is from one of four similar experiments.

**Supplemental Table 4. Effect of inhibitors of TNF, IL-12/23p40, or IL-23p19 on expression of selected genes in K14/F6 mouse skin treated with TPA.** Transgenic
mice were treated with TPA to induce skin inflammation, and also given antagonists of TNF, IL-12/23p40 or IL-23p19, or isotype control antibody, as described in Methods. RNA was prepared from full-thickness skin sections harvested 48hr after the second TPA treatment. Taqman quantitative PCR was performed and the CT value for each gene normalized to that for HPRT. The normalized value for each gene for each condition is shown, as well as relevant ratios of normalized values. Four mice were used in the naïve, TPA only, and TPA plus control antibody groups, while the other groups (TPA plus either TNF antagonist, antibody to IL-12/23p40, or antibody to IL-23p19) contained 5 mice each. The data is from one of two similar experiments for anti-TNF and anti-p40, and six similar experiments for anti-IL23p19.

**Supplemental Table 5. Effect of inhibitors of TNF, IL-12/23p40, or IL-23p19 on expression of selected proteins in K14/F6 mouse skin treated with TPA.**

Transgenic mice were treated with TPA to induce skin inflammation, and also given antagonists of TNF, IL-12/23p40 or IL-23p19, or isotype control antibody, as described in Methods. Lysates were made from full-thickness skin sections harvested 48hr after the second TPA treatment, as described in Methods, and levels of selected proteins determined either by ELISA or by Multi-Analyte Profiling at Rules-Based Medicine. Values shaded gray were below the detection limit of the assay. Numbers entered into those cells, for the purpose of calculation, were either one-half the detection limit of the assay, or the minimum numeric value for the analyte reported by Rules-Based Medicine, whichever was lower. Standard names for chemokines analyzed are: CCL2, MCP-1; CCL3, MIP-1α; CCL4, MIP-1β; CCL5, RANTES; CCL7, MCP-3; CCL9, MIP-1γ;
CCL12, MCP-5; CCL19, MIP-3β; CCL22, MDC; CXCL1, KC/GROα; CXCL2, MIP-2; XCL-1, lymphotactin. The data is from one of two similar experiments for anti-TNF and anti-p40, and six similar experiments for anti-IL23p19. Group sizes were the same as for Supplemental Table 4.

Supplemental Table 6. Changes in IL-1F gene expression in K14/F6 mice treated with TPA. Selected data from the same experiment described in Supplemental Table 1 is presented to emphasize changes in IL-1F mRNA levels as well as changes in the IL-F5 antagonist :: IL-1F agonist mRNA ratios.
A. Intracorneal Pustule
B. Parakeratosis
C. Spongiform Pustule

Mixed Dermal Infiltrate Dilated Blood Vessels Suprapapillary Thinning

C.

Naive Acetone TPA

Tg

Wt

D.

CD3 Quantitation

CD11c Quantitation

CD31 Quantitation