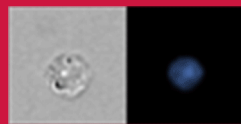


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Mystery Solved: IL-15

Todd A. Fehniger

It was the early 1990s, and there was an unsolved mystery in innate immunity: what cytokine was responsible for supporting the development and survival of NK cells? We understood that the common γ -chain (γ_c) cytokine receptor was critical because patients with SCID due to *IL-2RG* mutations had markedly reduced NK cell function (1, 2). The prevailing thought in the field was that IL-2, which signals via a high-affinity heterotrimeric receptor, IL-2R $\alpha\beta\gamma$, was the most likely candidate. After all, IL-2 resulted in potent NK cell activation, proliferation, and survival in vitro. However, all of the facts did not add up; IL-2 is produced mainly by activated T cells, which are spatially and temporally disconnected from normal NK cell development and homeostasis. Moreover, mice that had disrupted genes for IL-2 (3) or the IL-2R α (4) did not have a defect in NK cells. What clandestine cytokine used the IL-2R to support NK cells?

The mystery was unraveled in landmark reports by two independent groups that employed a simple strategy to search for alternative IL-2R ligands: examining the composition of cell line supernatants capable of supporting IL-2R-dependent growth signals in the presence of anti-IL-2-neutralizing Abs (5, 6). Scientists at Immunex Corporation used chromatography to purify a 14-kDa IL-2-functional mimic from the simian CV-1/EBNA cell line supernatant, sequenced NH₂-terminal residues, and then used degenerate oligonucleotide primers to clone a full-length cDNA. This approach ultimately revealed a novel cytokine designated IL-15 (this article is available at <https://science.sciencemag.org/content/264/5161/965>) (5). IL-15 used the IL-2R β and γ_c (7) but did not require the IL-2R α , and IL-15 promoted NK cell activation and proliferation (8), matching the profile of the mystery cytokine. Concurrently, scientists at the National Institutes of Health, led by Thomas Waldmann, identified functional IL-2-mimicry activity in HTLV-1-transformed HuT-102 cell line supernatant and isolated a cytokine that was initially designated IL-T (this article is available at <https://www.pnas.org/content/pnas/91/11/4935.full.pdf>) (6).

IL-T stimulated T cell proliferation and generated lymphokine-activated killer cells, required the IL-2R β and γ_c but not IL-2R α , and thus also fit the cytokine puzzle. Reconciling IL-T and IL-15, mature IL-T was found to be identical to IL-15. Indeed, the gene encoding IL-T/IL-15 within HuT-102 was fused to the HTLV-1 LTR that had inserted into the 5'UTR of IL-T. This resulted in enhanced transcription, translation, and secretion of IL-T/IL-15 (9). IL-15 mRNA was found in a wide variety of tissues with an expression profile distinct from IL-2, also allowing for greater cytokine access to NK cells and nonclassical T cells. Further analysis revealed disparate IL-2 and IL-15 primary sequences, but protein modeling indicated that IL-15 folded into a 4- α helix bundle cytokine, similar to IL-2. Mystery solved, and a new cytokine supporting NK cell homeostasis was discovered!

Definitive evidence of IL-15's nonredundant role in NK cell development and survival followed from IL-15 gene loss-of-function studies in mice (10). The importance of IL-15 was identified not only for NK cells but also several nonclassical T cell lineages and memory CD8 T cells. Gain-of-function studies using IL-15 transgenic mice complemented these findings, demonstrating an expansion of NK cells and memory-phenotype CD8 T cells but cautioning that chronic, unrestrained IL-15 can contribute to the development of T/NK leukemias (11). Thus, IL-15 is the primary cytokine responsible for IL-2/15R $\beta\gamma_c$ signals that promote and sustain NK cell development and survival.

Although the similarities between IL-2 and IL-15, including shared signaling receptor subunits, were instrumental in the discovery of IL-15, these same parallels influenced our initial understanding of IL-15R biology. After IL-15's discovery, mice with genetic deletions of the γ_c (12) and IL-2/15R β (13) were shown to lack NK cells, solidifying the importance of IL-2/15R $\beta\gamma_c$ signaling in NK cell development. IL-2 interacts with several forms of the IL-2R with differing affinities; it has intermediate (nanomolar) affinity for the heterodimeric IL-2R $\beta\gamma$ and high (picomolar) affinity for the heterotrimeric IL-2 $\alpha\beta\gamma$, which includes a "private" α receptor subunit. However, IL-2R α was not involved in IL-15 signaling, and a new search was on for additional IL-15R components responsible for high-affinity IL-15 binding. Almost immediately, IL-15R α was identified and cloned based on structural homology to the IL-2R α (14, 15). However, unlike IL-2R α , which has low affinity for IL-2 on its own, IL-15R α bound IL-15 with high affinity. This receptor biology leads to a situation in which limiting amounts of IL-2 are preferentially sensed by immune cells expressing IL-2R α in the context of the IL-2R $\alpha\beta\gamma$. Indeed, ultra-low dose IL-2 therapy in humans

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Abbreviation used in this article: γ_c , common γ -chain.

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expanded subsets of NK cells that expressed the IL-2R $\alpha\beta\gamma$ (16) and was later recognized to also expand IL-2R α^+ regulatory T cells (17). In contrast, IL-15R α on its own has high affinity for IL-15, which provides the opportunity to operate in *cis* when interacting with IL-15R $\beta\gamma_c$ on the same cell or in *trans* with IL-15R α presenting IL-15 to IL-15R $\beta\gamma_c$ on a neighboring cell. A new mystery: which was the primary mode of IL-15 interaction with its signaling receptor components? In elegant studies using IL-15 $^{-/-}$ and IL-15R $\alpha^{-/-}$ mice and bone marrow chimera approaches, Averil Ma's group provided clear in vivo evidence that IL-15 *trans*-presentation by IL-15R α to NK cells expressing the IL-2/15R $\beta\gamma_c$ was the major mode supporting NK cell development and homeostasis (18–20). Thus, the initial similarity in biology between IL-2R and IL-15R has been revised, and we now know that these cytokines mediate fundamentally different functions in vivo, in part because of this distinct receptor biology.

Twenty-five years after the discovery of IL-15, the importance of solving these mysteries is more evident than ever with the clinical translation of IL-15 to treat human disease. Initially, the biology of IL-15 was thought to be similar to FDA-approved human rIL-2 and was therefore not pursued as a therapeutic drug. With more appreciation of the distinct biology between IL-2 and IL-15, IL-15R agonists now represent an emerging and exciting immunotherapy strategy to promote NK and CD8 T cell responses in the context of infection and cancer. In 2015, the first clinical trial demonstrated NK and CD8 T cell expansion and activation in response to human rIL-15 (21). Armed with a deeper understanding of IL-15R biology, new therapeutic agents have been developed to improve in vivo pharmacokinetics and mimic physiologic IL-15:IL-15R α *trans*-presentation. These therapeutics have been shown to be safe and promote expansion and activation of NK and CD8 T cells without expanding regulatory T cells (22). Thus, in human patients, IL-15 has now been shown to possess distinct immunomodulatory properties compared with IL-2. These IL-15R agonists hold particular promise as combination agents with other forms of immunotherapy, including cellular therapies, tumor-targeting therapeutic Abs, inhibitory checkpoint blockade, and vaccines. Thus, although IL-15's discovery solved an immunologic mystery in 1994, its recent application as a cancer immunotherapy appears promising, and only time will tell us the story of IL-15's full impact on human health and disease.

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References

- Koren, H. S., D. B. Amos, and R. H. Buckley. 1978. Natural killing in immunodeficient patients. *J. Immunol.* 120: 796–799.
- Noguchi, M., H. Yi, H. M. Rosenblatt, A. H. Filipovich, S. Adelstein, W. S. Modi, O. W. McBride, and W. J. Leonard. 1993. Interleukin-2 receptor γ chain mutation results in X-linked severe combined immunodeficiency in humans. *Cell* 73: 147–157.
- Schorle, H., T. Holtschke, T. Hüning, A. Schimpl, and I. Horak. 1991. Development and function of T cells in mice rendered interleukin-2 deficient by gene targeting. *Nature* 352: 621–624.
- Wallerford, D. M., J. Chen, J. A. Ferry, L. Davidson, A. Ma, and F. W. Alt. 1995. Interleukin-2 receptor α chain regulates the size and content of the peripheral lymphoid compartment. *Immunity* 3: 521–530.
- Grabstein, K. H., J. Eisenman, K. Shanebeck, C. Rauch, S. Srinivasan, V. Fung, C. Beers, J. Richardson, M. A. Schoenborn, M. Ahdieh, et al. 1994. Cloning of a T cell growth factor that interacts with the β chain of the interleukin-2 receptor. *Science* 264: 965–968.
- Burton, J. D., R. N. Bamford, C. Peters, A. J. Grant, G. Kurys, C. K. Goldman, J. Brennan, E. Roessler, and T. A. Waldmann. 1994. A lymphokine, provisionally designated interleukin T and produced by a human adult T-cell leukemia line, stimulates T-cell proliferation and the induction of lymphokine-activated killer cells. *Proc. Natl. Acad. Sci. USA* 91: 4935–4939.
- Giri, J. G., M. Ahdieh, J. Eisenman, K. Shanebeck, K. Grabstein, S. Kumaki, A. Namen, L. S. Park, D. Cosman, and D. Anderson. 1994. Utilization of the beta and gamma chains of the IL-2 receptor by the novel cytokine IL-15. *EMBO J.* 13: 2822–2830.
- Carson, W. E., J. G. Giri, M. J. Lindemann, M. L. Linett, M. Ahdieh, R. Paxton, D. Anderson, J. Eisenmann, K. Grabstein, and M. A. Caligiuri. 1994. Interleukin (IL) 15 is a novel cytokine that activates human natural killer cells via components of the IL-2 receptor. *J. Exp. Med.* 180: 1395–1403.
- Bamford, R. N., A. P. Battista, J. D. Burton, H. Sharma, and T. A. Waldmann. 1996. Interleukin (IL) 15/IL-T production by the adult T-cell leukemia cell line HuT-102 is associated with a human T-cell lymphotropic virus type I region/IL-15 fusion message that lacks many upstream AUGs that normally attenuates IL-15 mRNA translation. *Proc. Natl. Acad. Sci. USA* 93: 2897–2902.
- Kennedy, M. K., M. Glaccum, S. N. Brown, E. A. Butz, J. L. Viney, M. Embers, N. Matsuiki, K. Charrier, L. Sedger, C. R. Willis, et al. 2000. Reversible defects in natural killer and memory CD8 T cell lineages in interleukin 15-deficient mice. *J. Exp. Med.* 191: 771–780.
- Fehniger, T. A., K. Suzuki, A. Ponnappan, J. B. VanDeusen, M. A. Cooper, S. M. Florea, A. G. Freud, M. L. Robinson, J. Durbin, and M. A. Caligiuri. 2001. Fatal leukemia in interleukin 15 transgenic mice follows early expansions in natural killer and memory phenotype CD8+ T cells. *J. Exp. Med.* 193: 219–231.
- DiSanto, J. P., W. Müller, D. Guy-Grand, A. Fischer, and K. Rajewsky. 1995. Lymphoid development in mice with a targeted deletion of the interleukin 2 receptor gamma chain. *Proc. Natl. Acad. Sci. USA* 92: 377–381.
- Suzuki, H., G. S. Duncan, H. Takimoto, and T. W. Mak. 1997. Abnormal development of intestinal intraepithelial lymphocytes and peripheral natural killer cells in mice lacking the IL-2 receptor beta chain. *J. Exp. Med.* 185: 499–505.
- Giri, J. G., S. Kumaki, M. Ahdieh, D. J. Friend, A. Loomis, K. Shanebeck, R. DuBose, D. Cosman, L. S. Park, and D. M. Anderson. 1995. Identification and cloning of a novel IL-15 binding protein that is structurally related to the alpha chain of the IL-2 receptor. *EMBO J.* 14: 3654–3663.
- Anderson, D. M., S. Kumaki, M. Ahdieh, J. Bertles, M. Tometsko, A. Loomis, J. Giri, N. G. Copeland, D. J. Gilbert, N. A. Jenkins, et al. 1995. Functional characterization of the human interleukin-15 receptor α chain and close linkage of IL15RA and IL2RA genes. *J. Biol. Chem.* 270: 29862–29869.
- Khatri, V. P., T. A. Fehniger, R. A. Baiocchi, F. Yu, M. H. Shah, D. S. Schiller, M. Gould, R. T. Gazzinelli, Z. P. Bernstein, and M. A. Caligiuri. 1998. Ultra low dose interleukin-2 therapy promotes a type 1 cytokine profile in vivo in patients with AIDS and AIDS-associated malignancies. *J. Clin. Invest.* 101: 1373–1378.
- Koreth, J., K. Matsuoka, H. T. Kim, S. M. McDonough, B. Bindra, E. P. Alyea, III, P. Armand, C. Cutler, V. T. Ho, N. S. Treister, et al. 2011. Interleukin-2 and regulatory T cells in graft-versus-host disease. *N. Engl. J. Med.* 365: 2055–2066.
- Koka, R., P. R. Burkett, M. Chien, S. Chai, F. Chan, J. P. Lodolce, D. L. Boone, and A. Ma. 2003. Interleukin (IL)-15R[alpha]-deficient natural killer cells survive in normal but not IL-15R[alpha]-deficient mice. *J. Exp. Med.* 197: 977–984.
- Koka, R., P. Burkett, M. Chien, S. Chai, D. L. Boone, and A. Ma. 2004. Cutting edge: murine dendritic cells require IL-15R alpha to prime NK cells. *J. Immunol.* 173: 3594–3598.
- Burkett, P. R., R. Koka, M. Chien, S. Chai, D. L. Boone, and A. Ma. 2004. Coordinate expression and trans presentation of interleukin (IL)-15Ralpha and IL-15 supports natural killer cell and memory CD8+ T cell homeostasis. *J. Exp. Med.* 200: 825–834.
- Conlon, K. C., E. Lugli, H. C. Welles, S. A. Rosenberg, A. T. Fojo, J. C. Morris, T. A. Fleisher, S. P. Dubois, L. P. Perera, D. M. Stewart, et al. 2015. Redistribution, hyperproliferation, activation of natural killer cells and CD8 T cells, and cytokine production during first-in-human clinical trial of recombinant human interleukin-15 in patients with cancer. *J. Clin. Oncol.* 33: 74–82.
- Romee, R., S. Cooley, M. M. Berrien-Elliott, P. Westervelt, M. R. Verneris, J. E. Wagner, D. J. Weisdorf, B. R. Blazar, C. Ustun, T. E. DeFor, et al. 2018. First-in-human phase 1 clinical study of the IL-15 superagonist complex ALT-803 to treat relapse after transplantation. *Blood* 131: 2515–2527.