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Cutting Edge: T Cells Respond to Lipopolysaccharide Innately via TLR4 Signaling

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LPS, a molecule produced by Gram-negative bacteria, is known to activate both innate immune cells such as macrophages and adaptive immune B cells via TLR4 signaling. Although TLR4 is also expressed on T cells, LPS was observed not to affect T cell proliferation or cytokine secretion. We now report, however, that LPS can induce human T cells to adhere to fibronectin via TLR4 signaling. This response to LPS was confirmed in mouse T cells; functional TLR4 and MyD88 were required, but T cells from TLR2 knockout mice could respond to LPS. The human T cell response to LPS depended on protein kinase C signaling and involved the phosphorylation of the proline-rich tyrosine kinase (Pyk-2) and p38. LPS also up-regulated the T cell expression of suppressor of cytokine signaling 3, which led to inhibition of T cell chemotaxis toward the chemokine stromal cell-derived factor 1α (CXCL12). Thus, LPS, through TLR4 signaling, can affect T cell behavior in inflammation. The Journal of Immunology, 2007, 179: 41–44.

Mindful of these aspects of T cell function, we tested whether LPS might directly affect T cell adherence or migration and can now report this to be the case. These previously unsuspected effects of LPS on T cells are likely to be important in the host-parasite interactions in Gram-negative infections and in AIDS.

Materials and Methods

Reagents

The reagents and chemicals were purchased from the sources previously described (6–8). Salmonella minnesota LPS purified by phenol extraction was purchased from Sigma-Aldrich. The neutralizing mAbs anti-human TLR2 and anti-human TLR4 were obtained from eBioscience.

Mice

Female C57BL/6J were obtained from Harlan Olac. C3HeB/FeJ and C3H/HeJ mice were obtained from The Jackson Laboratory. Dr. S. Akira (Osaka University, Osaka, Japan) provided TLR2−/− (B6.129P2-Tlr2tm1Aki) and MyD88−/− (B6.129P2-MyD88tm1Aki) mice (10, 11).

Human and mouse T cells

CD3+ T cells were purified from the peripheral blood of healthy human donors (Blood Bank, Tel-Hashomer, Israel), or from mouse spleens as previously described (6, 8).

T cell adhesion and migration assays

Analysis of T cell adhesion was determined as previously described (7). T cell chemotaxis was assayed as described using the Transwell system (8).

Western blot analysis of T cell lysates

T cells, 5 × 10^6 per sample, were activated with different concentrations of LPS (37°C in a 7% CO2 humidified atmosphere). Total cell lysates were prepared and analyzed for protein content as previously described (6, 7).

RNA interference

We synthesized a silent RNA (siRNA)3 sequence targeting suppressor of cytokine signaling (SOCS) 3 positions 80–101 relative to the start codon, 5’-AA GAGCGAGTACCAGCTGGTG-3’. a dsRNA targeting luciferase (GL-2) was used as a control (Dharmacon Research). Transfections of freshly purified T cells were performed using the Human T cell Nucleofector kit (Ammax Biosystems) as previously described (8). Cell migration was evaluated as described 48 h after transfection. Transfection efficiency was controlled by evaluating SOCS3 levels by Western blotting.

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5 Abbreviations used in this paper: siRNA, silent RNA; FN, fibronectin; Pyk-2, proline-rich tyrosine kinase 2; SDF, stromal cell-derived factor; SOCS, suppressor of cytokine signaling.

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FIGURE 1. LPS induces T cell adhesion to FN via β1 integrin and activation of PKC and p38 signaling pathways in a TLR4/MyD88-dependent manner. A. Purified human CD3⁺ T cells were labeled with chromium 51. The cells were then pretreated (30 min) with different concentrations of LPS and seeded onto FN-coated microtiter wells for an additional 30 min. The nonadherent cells were washed away and the remaining FN-bound cells were lysed. The radioactivity of lysates, representing the amount of FN-adherent cells, was determined. The mean ± SD of six independent experiments using different donor bloods is shown. B. Time-kinetic analysis of LPS-induced T cell adhesion. T cells were exposed to 100 ng/ml LPS for different periods of time. The results are expressed as the percentage of adherent T cells. The mean ± SD of five independent experiments is shown. C and D. T cells were pretreated with the indicated mAb (20 μg/ml) or FN peptides (10–15 μg/ml) or the intracellular signal transduction inhibitors GF109203X (GF; 20 nM), SB 203580 (SB; 50 nM), or pertussis toxin (PTX; 2 μg/ml). The cells were then activated with LPS (100 ng/ml), and tested for adhesion to FN. The mean ± SD of four different experiments is shown. E and F, T cells were exposed to LPS (100 ng/ml, 15 min) and lysed. The lysates were run on SDS-polyacrylamide gels for electrophoresis, transferred to nitrocellulose membranes, and immunoblotted with mAb anti-phospho-Pyk2 (pPyk2), anti-total Pyk2 (tPyk2), anti-phospho-p38 (pp38), and anti-total p38 (tp38). The blot of one representative experiment of five is presented. Phosphorylation levels of the experiments were estimated by densitometry, and an average percentage of phosphorylation ± SD was calculated as (pPyk2/tPyk2) or (pp38/tp38) × 100. G. T cells were pretreated with mAb anti-TLR2 or anti-TLR4 (20 μg/ml). The cells were then treated with LPS (100 ng/ml), and percentage of adhesion to FN was determined. The mean ± SD of four independent experiments is shown. H, CD3⁺ T cells purified from the spleens of wild-type C3HeB/FeJ, TLR4-mutated C3H/HeJ, MyD88⁻/⁻, or MyD88⁻/⁺ mice were radiolabeled, pretreated with LPS (100 ng/ml) for 60 min, and seeded onto FN-coated microtiter wells. The mean ± SD of three different experiments is shown. *, p < 0.05.
Results and Discussion

LPS induces T cell adhesion to fibronectin via TLR4/MyD88 signaling

To assess whether LPS directly affects T cell adhesion and migration, we purified CD3⁺ T cells (> 99% purity) from the peripheral blood of healthy human donors. The purified T cells were incubated with various concentrations of LPS for various times, and we assayed the adhesion of the T cells to immobilized extracellular matrix components, namely fibronectin (FN), laminin, or type 1 collagen. We found that LPS enhanced T cell adhesion specifically to FN but not to laminin or collagen type 1 (not shown). Fig. 1A shows that LPS induced T cell adhesion to FN in dose-dependent manner; 100 ng/ml produced the maximal effect, and we used this standard concentration. Time-kinetic experiments revealed that treatment for 90–150 min with LPS induced peak levels of T cell adhesion to FN (Fig. 1B). Interestingly, longer periods (270 min) reduced T cell adherence.

APCs and B cells are very sensitive to LPS activation (1, 2). It was therefore critical to learn whether the contamination of T cells with other cell types might have been responsible for LPS-induced adhesion in our experiments. FACS analysis confirmed that our T cell preparations did not contain other cell types such as B cells, dendritic cells, or macrophages (data not shown). Moreover, incubation of purified T cells with a wide range of LPS doses (0.001–100 μg/ml) did not result in secretion of the cytokines typically secreted by APCs and B cells such as TNF-α, IL-12, IL-6, or IL-10 (data not shown). Thus, LPS can directly induce T cell adhesion to FN without involving APCs or B cells.

T cell adhesion to FN is mediated primarily by the α5β1 and αvβ3 integrins (CD29) that recognize Leu-Asp-Val (LDV) and Arg-Gly-Asp (RGD), respectively. Indeed, we found that adhesion was inhibited by the RGD and LDV peptides and by anti-CD29 mAb but not by the control peptides Arg-Gly-Glu (RGE) and Leu-Glu-Val (LEV) or anti-VLA3 mAb (Fig. 1C).

The FN adhesion of T cells requires intracellular signaling to activate β1 integrins (12, 13). Fig. 1D shows that preincubation of T cells with a protein kinase C (PKC) inhibitor, GF109203X (RGE) and Leu-Glu-Val (LEV) or anti-VLA3 mAb (Fig. 1C).

FIGURE 2. LPS inhibits T cell chemotaxis toward SDF-1α through the up-regulation of SOCS3. A and B, Human T cells were incubated with LPS (100 ng/ml) for 1 h without (A) or with pretreatment with mAb anti-TLR2 or TLR4 (B), labeled, and added to the upper chambers of Transwell apparatus. The lower chambers contained the same media with or without SDF-1α (100 ng/ml). After 3 h, the cells that had transmigrated into the lower wells were collected, centrifuged, and lysed, and the radioactivity in the resulting supernatants was determined. The mean ± SD of five different experiments is shown. C and D, T cells without treatment (C) or after transfection with siRNA targeting SOCS3 or control siRNA (D) were incubated with LPS at 100 ng/ml for 1 h and tested for chemotaxis or lysed. Cell lysates were immunoblotted with anti-SOCS3 and anti-total ERK (tERK; evaluation of total ERK served as a control) or with anti-pSTAT3 and anti-STAT3. The levels of SOCS3, total ERK, phospho-STAT3 (pSTAT3), and total STAT3 (tSTAT3) were estimated by densitometry, and the average percentage (± SD) of the five experiments was calculated by the OD of SOCS3/tERK (pSTAT3/tSTAT3) × 100. *p < 0.05.

LPS activates various types of leukocytes via TLR2 and TLR4 signaling (18, 19). However, the removal of contaminants in commercial LPS preparations eliminates signaling through TLR2 (20). In this study we used LPS from S. minnesota purified by phenol extraction. It was recently reported that T cells express low but consistent levels of both TLR2 and TLR4 on their surfaces (5, 9, 21). Hence, we tested whether TLR2 or TLR4 are involved in the proadhesive effects of LPS on T cells. Fig. 1G shows that T cell adhesion to FN induced by LPS was inhibited by a neutralizing Ab to TLR4, but not by an Ab to TLR2.
the C3HeB/FeJ mice to adhere to FN; T cells from the C3H/HeJ mice did not respond. Moreover, T cells purified from spleens of TLR2 knockout mice responded to LPS, as did T cells from wild-type C57BL/6 mice (not shown).

LPS can signal both through MyD88-dependent and MyD88-independent pathways (10, 22). Fig. 1A demonstrates that MyD88-deficient T cells did not respond to LPS. Thus, T cell adhesion to FN induced by LPS requires TLR4 signaling that depends on MyD88.

LPS inhibits human T cell chemotaxis to stromal cell-derived factor (SDF)-1α via TLR4

We incubated human T cells with various concentrations of LPS for 1 h and assayed the effect on T cell chemotaxis directed to SDF-1α. Fig. 2A shows a dose-dependent inhibition of chemotaxis. We incubated purified CD4+ and CD8+ subsets of T cells with LPS and found no differences in the ability of LPS to inhibit their migration (data not shown). Pretreatment with a neutralizing anti-TLR4 mAb, but not an anti-TLR2 mAb (20 μg/ml), abrogated the inhibitory effect of LPS on chemotaxis (Fig. 2B). Thus, TLR4 is also required for LPS to inhibit T cell migration.

SOCS3 mediates the inhibitory effect of LPS on human T cell chemotaxis to SDF-1α

SOCS family proteins are negative feedback regulators of cytokine-induced JAK/STAT activation through their binding to SOCS family proteins are negative feedback regulators of cytokine-induced JAK/STAT activation through their binding to SOCS family proteins. Moreover, the up-regulation of SOCS3 expression by LPS; transfection with control siRNA requires SOCS3, we specifically silenced SOCS3 expression using RNA interference. This treatment abrogated the induction of SOCS3 expression by LPS; transfection with control siRNA

To confirm that the inhibitory effect of LPS on chemotaxis requires SOCS3, we specifically silenced SOCS3 expression using RNA interference. This treatment abrogated the induction of SOCS3 expression by LPS; transfection with control siRNA

In summary, SOCS3 regulates T cell chemotaxis to SDF-1α via TLR4 signaling that depends on MyD88.