

## Cutting Edge: $V\alpha 14$ - $J\alpha 281$ NKT Cells Naturally Regulate Experimental Autoimmune Encephalomyelitis in Nonobese Diabetic Mice

This information is current as of October 17, 2021.

Lennart T. Mars, Véronique Laloux, Karine Goude, Sabine Desbois, Abdelhadi Saoudi, Luc Van Kaer, Hans Lassmann, André Herbelin, Agnès Lehuen and Roland S. Liblau

*J Immunol* 2002; 168:6007-6011; ;  
doi: 10.4049/jimmunol.168.12.6007  
<http://www.jimmunol.org/content/168/12/6007>

**References** This article **cites 27 articles**, 18 of which you can access for free at:  
<http://www.jimmunol.org/content/168/12/6007.full#ref-list-1>

Why *The JI*? [Submit online.](#)

- **Rapid Reviews! 30 days\*** from submission to initial decision
- **No Triage!** Every submission reviewed by practicing scientists
- **Fast Publication!** 4 weeks from acceptance to publication

*\*average*

**Subscription** Information about subscribing to *The Journal of Immunology* is online at:  
<http://jimmunol.org/subscription>

**Permissions** Submit copyright permission requests at:  
<http://www.aai.org/About/Publications/JI/copyright.html>

**Email Alerts** Receive free email-alerts when new articles cite this article. Sign up at:  
<http://jimmunol.org/alerts>



## Cutting Edge: V $\alpha$ 14-J $\alpha$ 281 NKT Cells Naturally Regulate Experimental Autoimmune Encephalomyelitis in Nonobese Diabetic Mice<sup>1</sup>

Lennart T. Mars,\* Véronique Laloux,<sup>†</sup> Karine Goude,\* Sabine Desbois,\* Abdelhadi Saoudi,<sup>‡</sup> Luc Van Kaer,<sup>§</sup> Hans Lassmann,<sup>||</sup> André Herbelin,<sup>†</sup> Agnès Lehuen,<sup>†</sup> and Roland S. Liblau<sup>2\*‡</sup>

Although deficiencies in the NKT cell population have been observed in multiple sclerosis and mouse strains susceptible to experimental autoimmune encephalomyelitis (EAE), little is known about the function of these cells in CNS autoimmunity. In this work we report that TCR V $\alpha$ 14-J $\alpha$ 281 transgenic nonobese diabetic mice, which are enriched in CD1d-restricted NKT cells, are protected from EAE. The protection is associated with a striking inhibition of Ag-specific IFN- $\gamma$  production in the spleen, implying modulation of the encephalitogenic Th1 response. This modulation is independent of IL-4 because IL-4-deficient V $\alpha$ 14-J $\alpha$ 281 mice are still protected against EAE and independent of NKT cell-driven Th1 to Th2 deviation, because no increased autoantigen-specific Th2 response was observed in immunized V $\alpha$ 14-J $\alpha$ 281 transgenic mice. Our findings indicate that enrichment and/or stimulation of CD1d-dependent NKT cells may be used as a novel strategy to treat CNS autoimmunity. *The Journal of Immunology*, 2002, 168: 6007–6011.

Experimental autoimmune encephalomyelitis (EAE)<sup>3</sup> is a T cell-driven autoimmune demyelinating disease affecting the CNS that serves as a model for multiple sclerosis (MS). EAE arises as a consequence of the breakdown of self-tolerance, induced by the immunization of susceptible animals

with myelin Ags. Self-tolerance of T lymphocytes is instilled by recessive mechanisms as well as by T cell subsets able to dominantly suppress the activation, proliferation, and differentiation of autoreactive T cells (1). These regulatory T cells have recently been suggested to include NKT cells, a T cell subset that has been implicated in the regulation of tumor rejection and infectious immunity (2). This atypical T cell subset is characterized by the expression of classical NK cell markers together with a CD1d-restricted canonical  $\alpha\beta$  TCR, V $\alpha$ 14-J $\alpha$ 281/V $\beta$ 8.2, -7, and -2 in mice and V $\alpha$ 24-J $\alpha$ Q/V $\beta$ 11 in humans (2).

The association of NKT cells with autoimmunity was originally suggested by selectively reduced numbers of NKT cells in mice and patients with systemic lupus erythematosus and insulin-dependent diabetes mellitus (3–6). More detailed studies correlated the spontaneous development of autoimmune diabetes in nonobese diabetic (NOD) mice to qualitative defects in their NKT cell population (4, 7, 8). Moreover, spontaneous autoimmune diabetes could be prevented by the overexpression, *in vivo* activation, or adoptive transfer of NKT cells, suggesting a regulatory function of NKT cells in organ-specific autoimmunity (7, 9, 10).

Indirect evidence suggests that NKT cells may also perform a regulatory role in CNS autoimmunity. In humans, quantitative deficiencies in NKT cells have been observed in MS (11). Similarly, NKT cell dysfunctions have been observed in mouse strains susceptible to EAE (4, 12). Passive EAE can be prevented in TCR knockout mice by the adoptive transfer of DX5<sup>+</sup>TCR $\alpha\beta$ <sup>+</sup> T cells (13). Moreover, active EAE can be controlled either by the prototypical NKT cell agonist  $\alpha$ -galactosylceramide ( $\alpha$ -GalCer) or by administration of an  $\alpha$ -GalCer analog that enhances IL-4 production of NKT cells (14, 15). Although these studies demonstrate the potential of NKT cells to control EAE, it is unclear whether V $\alpha$ 14-J $\alpha$ 281 NKT cells are natural regulators of EAE. Our present data strongly argue that NKT cells naturally inhibit CNS autoimmunity and prevent the accumulation of autoreactive T cells in the spleen and CNS.

### Materials and Methods

#### *Mice and induction of EAE*

V $\alpha$ 14-J $\alpha$ 281 (9) and V $\alpha$ 8 (A. Lehuen, unpublished observation) transgenic (Tg) NOD mice were generated by microinjecting a TCR  $\alpha$ -chain V $\alpha$ 14-J $\alpha$ 281 or a V $\alpha$ 8-J $\alpha$ 37 construct into fertilized NOD eggs. TCR C $\alpha$ <sup>-/-</sup> NOD and V $\alpha$ 14-J $\alpha$ 281 Tg C $\alpha$ <sup>-/-</sup> NOD were described previously (9). V $\alpha$ 14-J $\alpha$ 281 IL-4<sup>-/-</sup> NOD mice and V $\alpha$ 8 Tg C $\alpha$ <sup>-/-</sup> NOD were obtained by two backcrosses onto IL-4<sup>-/-</sup> NOD and C $\alpha$ <sup>-/-</sup> NOD, respectively (16). CD1d-deficient mice were backcrossed at least seven times onto the NOD strain (17). Mice were housed under specific pathogen-free conditions.

\*Institut National de la Santé et de la Recherche Médicale, Unité 546, Hôpital Pitié-Salpêtrière, and <sup>†</sup>Institut National de la Santé et de la Recherche Médicale, Unité 25, Hôpital Necker, and Unité 561, Hôpital Saint Vincent de Paul, Paris, France; <sup>‡</sup>Institut National de la Santé et de la Recherche Médicale, Unité 563, Hôpital Purpan, Toulouse, France; <sup>§</sup>Department of Microbiology and Immunology, Howard Hughes Medical Institute, Vanderbilt University School of Medicine, Nashville, TN 37232; and <sup>||</sup>Brain Research Institute, University of Vienna, Vienna, Austria

Received for publication October 23, 2001. Accepted for publication April 15, 2002.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked *advertisement* in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

<sup>1</sup> This work was supported by Institut National de la Santé et de la Recherche Médicale and by the Association de Recherche sur la Sclérose en Plaques. L.T.M. was supported by a postdoctoral fellowship from the Fondation pour la Recherche Médicale. V.L. was supported by a studentship from the Fondation pour la Recherche Médicale.

<sup>2</sup> Address correspondence and reprint requests to Dr. Roland S. Liblau, Institut National de la Santé et de la Recherche Médicale, Unité 546, 105 Boulevard de l'Hôpital, Paris 75013, France. E-mail address: 106063.1005@compuserve.com

<sup>3</sup> Abbreviations used in this paper: EAE, experimental autoimmune encephalomyelitis;  $\alpha$ -GalCer,  $\alpha$ -galactosylceramide; LN, lymph node; MOG, myelin oligodendrocyte glycoprotein; MS, multiple sclerosis; NOD, nonobese diabetic; Tg, transgenic.

Eight- to 12-wk-old male mice were immunized s.c. with 100  $\mu$ g of myelin oligodendrocyte glycoprotein (MOG)<sub>35–55</sub> peptide (MEVGWYRSPFSRVVHLYRNGK; Mimotopes, Clayton Victoria, Australia) emulsified in CFA (Difco, Detroit, MI) and supplemented with 5 mg/ml *Mycobacterium tuberculosis* (strain H37RA; Difco). Pertussis toxin (List Biological Laboratories, Campbell, CA) was injected i.v. at day 0 (200 ng) and day 2 (400 ng) postimmunization. Clinical scores were recorded daily.

### Histological analysis

Animals were anesthetized and perfused with 2% paraformaldehyde in PBS. Tissues were embedded in paraffin, and tissue sections were stained with H&E or Luxol fast blue.

### T cell purification, cell transfer, proliferation assays, and ELISA

For negative selection of T cells (80–85% pure) by MACS, single-cell suspensions were incubated with F4/80 mAb (Caltag Laboratories, Burlingame, CA) and B220 mAb (BD Pharmingen, San Diego, CA), followed by microbeads (Miltenyi Biotech, Bergisch Gladbach, Germany) coated with anti-rat IgG mAb. For cell transfer  $50 \times 10^6$  purified T cells were injected i.v. 48 h preimmunization. Splenic single-cell suspensions ( $10^7$  cells/ml) or cocultures of draining lymph node (LN)-derived purified T cells ( $7.5 \times 10^5$ /ml) and irradiated syngeneic splenocytes ( $37.5 \times 10^5$ /ml) were stimulated with anti-CD3 mAb (BD Pharmingen),  $\alpha$ -GalCer (a generous gift from Kirin Brewery, Gunma, Japan), or MOG<sub>35–55</sub>. Proliferation was assessed after 48 h by incorporation of [<sup>3</sup>H]thymidine (1  $\mu$ Ci/well). Cytokines were detected by ELISA after 24 and 48 h (9).

### Statistical analysis

Proliferation and cytokine production were compared using the unpaired Student *t* test, whereas EAE day of onset, cumulative severity, and maximal severity were compared using the log rank test, Mann-Whitney *U* test, and relative to an identified distribution (RIDIT) analysis (18), respectively.

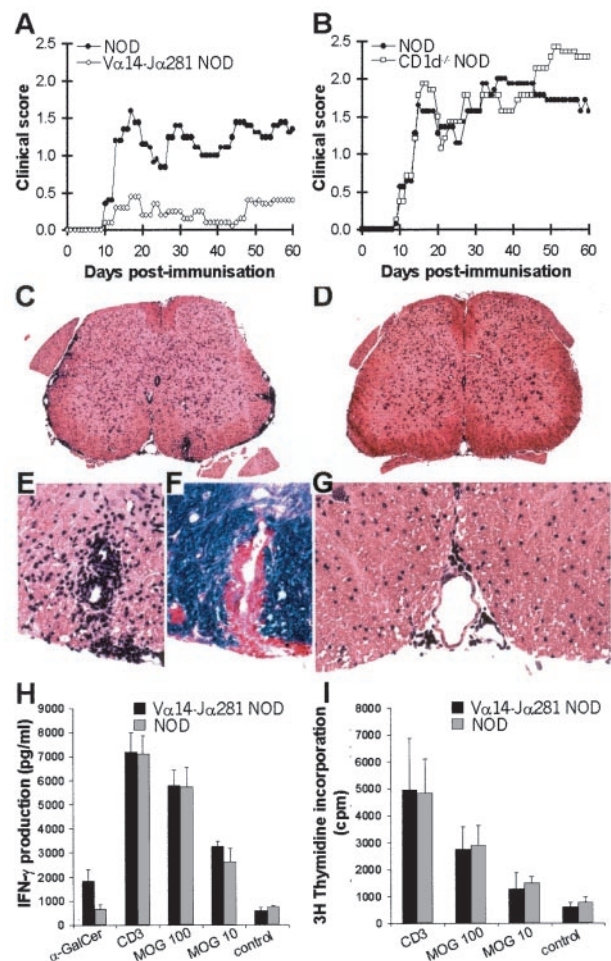
## Results and Discussion

### V $\alpha$ 14-J $\alpha$ 281 NKT cells regulate EAE

To address whether a regulatory function of CD1d-dependent NKT cells is naturally exploited to curb the encephalitogenic process in NOD mice, we undertook two simultaneous approaches. First, using V $\alpha$ 14-J $\alpha$ 281 Tg mice, we evaluated whether an increase in the number of NKT cells could reduce the severity of EAE. Second, we tested whether CD1d-deficient mice, which lack V $\alpha$ 14-J $\alpha$ 281 NKT cells (17), develop an exacerbated EAE.

NOD mice develop, 11 days (on average) after immunization with MOG<sub>35–55</sub>, a relapsing-remitting EAE resulting in limb paralysis (Fig. 1A and Table I). In stark contrast, the EAE observed in V $\alpha$ 14-J $\alpha$ 281 Tg NOD mice is significantly delayed and milder (Fig. 1A and Table I). This difference in clinical severity is corroborated by the histological analysis, as a clear inflammatory infiltrate is observed in the CNS of NOD mice (Fig. 1, C and E), while only a minimal, but definite, infiltrate is noted in the V $\alpha$ 14-J $\alpha$ 281 Tg NOD mice (Fig. 1, D and G). Moreover, perivascular demyelination is observed only in the CNS of non-Tg mice, but not V $\alpha$ 14-J $\alpha$ 281 Tg mice (Fig. 1F). These observations were reaffirmed by experiments in which  $50 \times 10^6$  purified T cells from normal NOD were transferred into T cell-deficient  $\text{Ca}^{-/-}$  NOD, V $\alpha$ 8 Tg  $\text{Ca}^{-/-}$  NOD lacking NKT cells, and V $\alpha$ 14-J $\alpha$ 281 Tg  $\text{Ca}^{-/-}$  NOD enriched in NKT cells (Fig. 2A). Following MOG immunization 48 h posttransfer, the severity of EAE was indistinguishable in the two control groups ( $\text{Ca}^{-/-}$  and V $\alpha$ 8 Tg  $\text{Ca}^{-/-}$  recipients) but was significantly reduced in V $\alpha$ 14-J $\alpha$ 281 Tg  $\text{Ca}^{-/-}$  recipients. Taken together, these data indicate that a quantitative enhancement of NKT cells conveys protection against EAE.

We did not observe any difference in the severity of EAE in CD1d<sup>-/-</sup> mice as compared with NOD littermates (Fig. 1B and Table I). This was not unexpected, as NOD mice are known to harbor a quantitatively and qualitatively defective NKT cell population. Moreover, the regulatory function of the already compromised NKT cell population might be overwhelmed by the ability



**FIGURE 1.** V $\alpha$ 14-J $\alpha$ 281 NKT cells control EAE. A and B, Wild-type NOD (●; A, *n* = 10; B, *n* = 7), V $\alpha$ 14-J $\alpha$ 281 Tg NOD (A, ◇; *n* = 10) and CD1d<sup>-/-</sup> NOD (B, □; *n* = 7) male mice were immunized with 100  $\mu$ g of MOG<sub>35–55</sub> emulsified in CFA. The clinical severity of EAE was monitored daily. C–G, Histological severity of EAE. The spinal cord of NOD mice (C, E, and F) and V $\alpha$ 14-J $\alpha$ 281 Tg NOD mice (D and G) was analyzed 20 days after immunization. Pronounced inflammation is observed in the spinal cord of NOD mice (C and E), associated with some perivascular demyelination (F). V $\alpha$ 14-J $\alpha$ 281 Tg NOD mice develop a low-level inflammation in the meninges (D and G). Samples were stained with 30 $\times$  H&E (C and D), 150 $\times$  H&E (E and G), and 150 $\times$  Luxol fast blue (F). H and I, NKT cells do not prevent T cell priming in the draining LN. T cells were purified from the draining LN of V $\alpha$ 14-J $\alpha$ 281 Tg NOD mice (filled bars) and NOD littermates (shaded bars) 10 days after immunization with 100  $\mu$ g of MOG<sub>35–55</sub> emulsified in CFA. IFN- $\gamma$  production (H) and the proliferation (I) of T cells cocultured with irradiated APCs and  $\alpha$ -GalCer (0.05  $\mu$ g/ml), anti-CD3 mAb (0.5  $\mu$ g/ml), or MOG<sub>35–55</sub> (100 and 10  $\mu$ g/ml) were measured for each individual mouse. The data represent mean  $\pm$  SEM of eight individual mice per group. Similar results were obtained in three independent experiments.

of the bacterial adjuvants, used to induce EAE, to strongly activate the innate and adaptive immune systems. The spontaneous occurrence of autoimmune diabetes (no requirement for adjuvants) in NOD mice might explain why NOD CD1d<sup>-/-</sup> mice do develop an exacerbated diabetes (19).

### Overexpression of NKT cells does not prevent efficient priming of MOG-specific T cells in V $\alpha$ 14-J $\alpha$ 281 Tg animals

The introduction of a rearranged TCR transgene in V $\alpha$ 14-J $\alpha$ 281 Tg mice may have resulted in limited T cell repertoire diversity

Table I. Clinical severity of EAE<sup>a</sup>

| Mice                    | n  | Day of Onset (SEM)       | Mean Maximum Severity (SEM) | Mean Cumulative Severity (SEM) | Relapses (SEM) | Mortality (%) |
|-------------------------|----|--------------------------|-----------------------------|--------------------------------|----------------|---------------|
| NOD                     | 23 | 10.9 (±0.6)              | 2.7 (±0.2)                  | 83.1 (±9.2)                    | 0.7 (±0.2)     | 1/23 (4.3)    |
| NOD Vα14-Jα281          | 22 | 21.9 (±3.3) <sup>b</sup> | 1.4 (±0.3) <sup>c</sup>     | 26.1 (±6.7) <sup>d</sup>       | 0.2 (±0.2)     | 1/22 (4.5)    |
| NOD                     | 7  | 12.9 (±1.0)              | 2.6 (±0.5)                  | 81.1 (±26.0)                   | 0.6 (±0.2)     | 1/7 (14.3)    |
| NOD CD1d <sup>-/-</sup> | 7  | 11.2 (±0.9)              | 2.9 (±0.7)                  | 89.1 (±23.3)                   | 0.4 (±0.3)     | 2/7 (28.5)    |

<sup>a</sup> Shown are the pooled data from three independent experiments comparing Vα14-Jα281 Tg vs non-Tg NOD and one experiment involving the CD1d<sup>-/-</sup> and CD1d<sup>+/+</sup> NOD mice.

<sup>b</sup> Value of  $p < 0.0003$  (log rank test).

<sup>c</sup> Value of  $p < 0.001$  (RIDIT analysis).

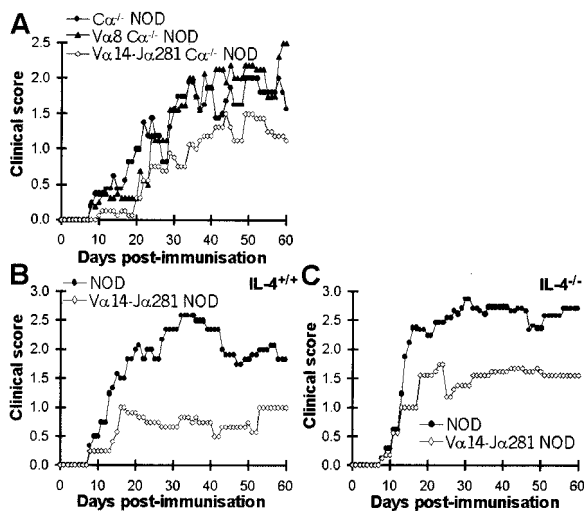
<sup>d</sup> Value of  $p < 0.0001$  (Mann-Whitney  $U$  test).

due to the preferential expression of the rearranged Vα14-Jα281 transgene, resulting in reduced Ag priming. Although this has carefully been excluded by previous studies (9, 20), we confirmed that both the intensity and the quality (MOG-specific T cell proliferation and IFN-γ production, IL-4 being undetectable) of the autoreactive T cell response induced in the draining LN upon MOG immunization was comparable in Vα14-Jα281 Tg and non-Tg NOD mice (Fig. 1, *H* and *I*). These data in combination with the adoptive transfer experiments (Fig. 2A) indicate that the striking inhibition of actively induced EAE observed in the Vα14-Jα281 Tg mice is not due to an infringement of the MOG-specific T cell repertoire but rather due to the immune regulation mediated by the enriched NKT cell population.

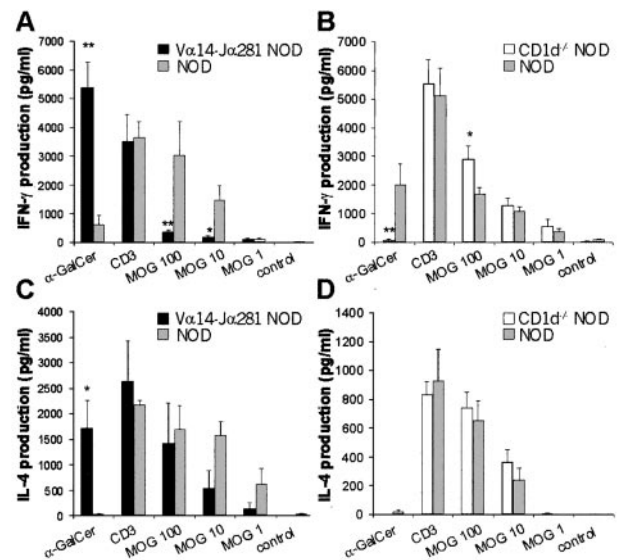
#### IL-4 is not required for the regulatory function of Vα14-Jα281 NKT cells in EAE

To address the mechanism by which NKT cells control EAE we focused on the ability of NKT cells to explosively release IL-4 and IFN-γ upon TCR ligation. By analogy to conventional T cells, the cytokine profile of NKT cells can be polarized, with those NKT cells polarized to produce IL-4 being suggested to control autoimmune diabetes and EAE (7, 10, 14, 15, 20, 21).

To address the role of IL-4 in our model we crossed Vα14-Jα281 Tg and non-Tg littermates with IL-4-deficient NOD mice. The disruption of IL-4 expression by itself did not significantly affect EAE severity (Fig. 2, *B* and *C*). Importantly, as shown in Fig. 2C, the protection mediated by the overexpression of NKT cells is preserved in mice lacking functional IL-4 expression. These data indicate that the efficacy of NKT cells to regulate EAE is not affected by the lack of IL-4. Moreover, no evidence of a MOG-specific Th2 bias was obtained when assessing the MOG-specific IL-4 release in splenocyte cultures (Fig. 3C) and the



**FIGURE 2.** Vα14-Jα281 NKT cells control EAE induced by an identical donor-derived T cell repertoire. *A*, The severity of MOG-induced EAE was assessed in male Cα<sup>-/-</sup> (●), Vα8 Tg Cα<sup>-/-</sup> (▲), and Vα14-Jα281 Tg Cα<sup>-/-</sup> NOD (◇) mice following i.v. transfer of 50 × 10<sup>6</sup> purified T cells from normal NOD males. Data represent the mean of eight mice per group. The day of onset ( $p = 0.01$ , log rank test) and the mean cumulative score ( $p = 0.02$ , Mann-Whitney  $U$  test) were significantly delayed or reduced in Vα14-Jα281 Tg Cα<sup>-/-</sup> NOD recipients as compared with the pooled control groups. *B* and *C*, The regulatory function of Vα14-Jα281 NKT cells does not require IL-4. The severity of MOG<sub>35-55</sub>-induced EAE was compared in IL-4-sufficient (*B*) and IL-4-deficient (*C*) Vα14-Jα281 Tg (*B* and *C*, ◇) and non-Tg NOD littermates (*B* and *C*, ●). Pooled data of two independent experiments including 8–12 mice per group are presented. The day of onset (log rank test), cumulative severity (Mann-Whitney  $U$  test), and maximum severity of EAE (RIDIT analysis) were all significantly delayed or decreased in IL-4<sup>-/-</sup> Vα14-Jα281 Tg as compared with IL-4<sup>-/-</sup> NOD mice ( $p < 0.05$ , one-tailed analysis).



**FIGURE 3.** NKT cells inhibit the MOG-induced IFN-γ response in the spleen. The MOG<sub>35-55</sub>-specific IFN-γ (*A* and *B*) and IL-4 (*C* and *D*) response of individual splenocyte cultures obtained from NOD Vα14-Jα281 Tg (*A* and *C*, filled bars) and NOD CD1d<sup>-/-</sup> mice (*B* and *D*, open bars) was compared with wild-type NOD mice (shaded bars). Individual spleen cultures were generated 10 days after immunization and stimulated in vitro for 24 (IL-4) and 48 (IFN-γ) h with α-GalCer (0.05 μg/ml), anti-CD3 mAb (0.5 μg/ml), or MOG<sub>35-55</sub> (100 and 10 μg/ml). The results represent the mean ± SEM of eight mice per group. The experiment was performed three times with similar results. \*,  $p < 0.05$ ; \*\*,  $p < 0.001$  (Student  $t$  test).

MOG-specific serum IgG isotypes of immunized V $\alpha$ 14-J $\alpha$ 281 Tg mice (data not shown). Consequently, our data reveal that the natural regulatory function of NKT cells in EAE is independent of IL-4 and an autoantigen-specific Th2 switch; this in contrast to the control of EAE induced by exogenous NKT cell activation (14, 15).

#### *V $\alpha$ 14-J $\alpha$ 281 NKT cells inhibit the encephalitogenic autoantigen-specific IFN- $\gamma$ response*

Because activated encephalitogenic T cells migrate from the LN to the spleen before their infiltration into the CNS, we contrasted the MOG-specific T cell response in the LN with that of the spleen to address whether the MOG-specific T cell population is exposed to NKT cell-mediated regulation. As compared with wild-type NOD, splenic cultures of CD1d-deficient NOD mice exhibited an unaltered IL-4 and IFN- $\gamma$  response following MOG<sub>35-55</sub> stimulation, confirming that the impaired NKT cell population in NOD mice does not alter the generation of encephalitogenic T cells (Figs. 1B and 3, B and D). Strikingly, stimulation of T cells with MOG<sub>35-55</sub> resulted in a significantly reduced production of IFN- $\gamma$  in spleen cultures from V $\alpha$ 14-J $\alpha$ 281 Tg mice, compared with those of non-Tg littermates (Fig. 3A). As no such reduction in the autoantigen-specific IFN- $\gamma$  response was observed in the draining LN (Fig. 1H), these data indicate that the autoreactive T cell response is exposed to NKT cell-mediated regulation once they have migrated away from the draining LN, where the relative number of NKT cells is low (20). Indeed, splenocytes from MOG-immunized V $\alpha$ 14-J $\alpha$ 281 Tg mice, but not from non-Tg littermates, produced large amounts of IFN- $\gamma$  and IL-4 in response to  $\alpha$ -GalCer (Fig. 3, A and C), confirming the overrepresentation of functional CD1d-dependent NKT cells in the spleen.

Having excluded the induction of an autoantigen-specific Th2 bias, alternative mechanisms able to account for a reduced MOG-induced IFN- $\gamma$  response in the spleen could include an altered migratory behavior of autoreactive T cells, or exposure to NKT cell-produced immunoregulatory cytokines other than IL-4. Indeed, NKT cell-produced IL-10, IL-13, and TGF- $\beta$  have been implicated in the down-regulation of tumor surveillance, the prevention of autoimmunity, and the preservation of immune privilege in the eye (2, 22). In addition, as activated T cells up-regulate CD1d (23), a direct cytotoxic NKT cell response might be induced against activated autoreactive T cells, as has been reported for thymocytes (24).

It is interesting to note that the regulatory function of NKT cells can be enhanced by their ability to recruit cell populations with the capacity to control autoimmune/inflammatory reactions. Indeed, NKT cells rapidly activate components of the innate immune response, including NK cells, which in turn are able to regulate EAE (2, 25). Alternatively, NKT cells might stimulate the generation of Ag-specific regulatory T cell populations able to suppress autoimmune processes (22).

Whether the regulatory function of NKT cells is used locally in the CNS during episodes of inflammation remains to be defined. However, NKT cells are not only detectable in the CNS of MS patients (11); they might be triggered in situ as the expression of CD1d is up-regulated on murine glial cells during CNS inflammation (26).

In conclusion, this study shows that V $\alpha$ 14-J $\alpha$ 281 NKT cells naturally regulate EAE in NOD mice. Preliminary results suggest that this property also applies to other mouse strains, as EAE severity is moderately reduced in V $\alpha$ 14-J $\alpha$ 281 Tg C57BL/6 mice (our unpublished observation). Although the precise regulatory mechanisms of V $\alpha$ 14-J $\alpha$ 281 NKT cells remain to be defined, our data indicate that they include preventing the accumulation of au-

toactive T cells in the spleen. This study further strengthens the concept of developing pharmacological stimuli for CD1d-dependent NKT cells as a novel strategy to treat CNS autoimmunity (15, 27, 28).

## Acknowledgments

We thank Dr. David Serreze for the IL-4<sup>-/-</sup> NOD mice, the Kirin Brewery for kindly providing  $\alpha$ -GalCer, Dr. Jean-Paul Charlet for help with statistical analyses, and Philippe Delis and Isabelle Cissé for animal care.

## References

- Roncarolo, M. G., and M. K. LeVings. 2000. The role of different subsets of T regulatory cells in controlling autoimmunity. *Curr. Opin. Immunol.* 12:676.
- Bendelac, A., M. Bonneville, and J. F. Kearney. 2001. Autoreactivity by design: innate B and T lymphocytes. *Nat. Rev. Immunol.* 1:177.
- Mieza, M. A., T. Itoh, J. Q. Cui, Y. Makino, T. Kawano, K. Tsuchida, T. Koike, T. Shirai, H. Yagita, A. Matsuzawa, et al. 1996. Selective reduction of V $\alpha$ 14<sup>+</sup> NK T cells associated with disease development in autoimmune-prone mice. *J. Immunol.* 156:4035.
- Gombert, J. M., A. Herbelin, E. Tancrede-Bohin, M. Dy, C. Carnaud, and J. F. Bach. 1996. Early quantitative and functional deficiency of NK1<sup>+</sup>-like thymocytes in the NOD mouse. *Eur. J. Immunol.* 26:2989.
- Wilson, S. B., S. C. Kent, K. T. Patton, T. Orban, R. A. Jackson, M. Exley, S. Porcelli, D. A. Schatz, M. A. Atkinson, S. P. Balk, et al. 1998. Extreme Th1 bias of invariant V $\alpha$ 24J $\alpha$ Q T cells in type 1 diabetes. *Nature* 391:177.
- Oishi, Y., T. Sumida, A. Sakamoto, Y. Kita, K. Kurasawa, Y. Nawata, K. Takabayashi, H. Takahashi, S. Yoshida, M. Taniguchi, et al. 2001. Selective reduction and recovery of invariant V $\alpha$ 24J $\alpha$ Q T cell receptor T cells in correlation with disease activity in patients with systemic lupus erythematosus. *J. Rheumatol.* 28:275.
- Hammond, K. J., L. D. Poulton, L. J. Palmisano, P. A. Silveira, D. I. Godfrey, and A. G. Baxter. 1998.  $\alpha$ / $\beta$ -T cell receptor (TCR)<sup>+</sup>CD4<sup>+</sup>CD8<sup>-</sup> (NKT) thymocytes prevent insulin-dependent diabetes mellitus in nonobese diabetic (NOD)/Lt mouse by the influence of interleukin (IL)-4 and/or IL-10. *J. Exp. Med.* 187:1047.
- Falcone, M., B. Yeung, L. Tucker, E. Rodriguez, and N. Sarvetnick. 1999. A defect in interleukin 12-induced activation and interferon  $\gamma$  secretion of peripheral natural killer T cells in nonobese diabetic mice suggests new pathogenic mechanisms for insulin-dependent diabetes mellitus. *J. Exp. Med.* 190:963.
- Lehuen, A., O. Lantz, L. Beaudoin, V. Laloux, C. Carnaud, A. Bendelac, J. F. Bach, and R. C. Monteiro. 1998. Overexpression of natural killer T cells protects V $\alpha$ 14-J $\alpha$ 281 transgenic nonobese diabetic mice against diabetes. *J. Exp. Med.* 188:1831.
- Sharif, S., G. A. Arreaza, P. Zucker, Q. S. Mi, J. Sondhi, O. V. Naidenko, M. Kronenberg, Y. Kozuka, T. L. Delovitch, J. M. Gombert, et al. 2001. Activation of natural killer T cells by  $\alpha$ -galactosylceramide treatment prevents the onset and recurrence of autoimmune type 1 diabetes. *Nat. Med.* 7:1057.
- Illes, Z., T. Kondo, J. Newcombe, N. Oka, T. Tabira, and T. Yamamura. 2000. Differential expression of NK T cell V $\alpha$ 24J $\alpha$ Q invariant TCR chain in the lesions of multiple sclerosis and chronic inflammatory demyelinating polyneuropathy. *J. Immunol.* 164:4375.
- Yoshimoto, T., A. Bendelac, J. Hu-Li, and W. E. Paul. 1995. Defective IgE production by SJL mice is linked to the absence of CD4<sup>+</sup>, NK1.1<sup>+</sup> T cells that promptly produce interleukin 4. *Proc. Natl. Acad. Sci. USA* 92:11931.
- Fritz, R. B., and M. L. Zhao. 2001. Regulation of experimental autoimmune encephalomyelitis in the C57BL/6J mouse by NK1.1<sup>+</sup>DX5<sup>+</sup> $\alpha$  $\beta$ <sup>+</sup> T cells. *J. Immunol.* 166:4209.
- Pal, E., T. Tabira, T. Kawano, M. Taniguchi, S. Miyake, and T. Yamamura. 2001. Costimulation-dependent modulation of experimental autoimmune encephalomyelitis by ligand stimulation of V $\alpha$ 14 NK T cells. *J. Immunol.* 166:662.
- Miyamoto, K., S. Miyake, and T. Yamamura. 2001. A synthetic glycolipid prevents autoimmune encephalomyelitis by inducing Th2 bias of natural killer T cells. *Nature* 413:531.
- Serreze, D. V., H. D. Chapman, C. M. Post, E. A. Johnson, W. L. Suarez-Pinzon, and A. Rabinovitch. 2001. Th1 to Th2 cytokine shifts in nonobese diabetic mice: sometimes an outcome, rather than the cause, of diabetes resistance elicited by immunostimulation. *J. Immunol.* 166:1352.
- Mendiratta, S. K., W. D. Martin, S. Hong, A. Boesteanu, S. Joyce, and L. Van Kaer. 1997. CD1d1 mutant mice are deficient in natural T cells that promptly produce IL-4. *Immunity* 6:469.
- Fleiss, J. L. 1981. Comparing proportions from independent samples. In *Statistical Methods for Rates and Proportions*. J. L. Fleiss, ed. J. L. Wiley, New York, p. 150.
- Shi, F. D., M. Flodstrom, B. Balasa, S. H. Kim, K. Van Gunst, J. L. Strominger, S. B. Wilson, and N. Sarvetnick. 2001. Germ line deletion of the CD1 locus exacerbates diabetes in the NOD mouse. *Proc. Natl. Acad. Sci. USA* 98:6777.
- Laloux, V., L. Beaudoin, D. Jeske, C. Carnaud, and A. Lehuen. 2001. NK T

- cell-induced protection against diabetes in V $\alpha$ 14-J $\alpha$ 281 transgenic nonobese diabetic mice is associated with a Th2 shift circumscribed regionally to the islets and functionally to islet autoantigen. *J. Immunol.* 166:3749.
21. Singh, N., S. Hong, D. C. Scherer, I. Serizawa, N. Burdin, M. Kronenberg, Y. Koezuka, and L. Van Kaer. 1999. Cutting edge: activation of NK T cells by CD1d and  $\alpha$ -galactosylceramide directs conventional T cells to the acquisition of a Th2 phenotype. *J. Immunol.* 163:2373.
  22. Sonoda, K. H., D. E. Faunce, M. Taniguchi, M. Exley, S. Balk, and J. Stein-Streilein. 2001. NK T cell-derived IL-10 is essential for the differentiation of antigen-specific T regulatory cells in systemic tolerance. *J. Immunol.* 166:42.
  23. Salamone, M. C., G. A. Rabinovich, A. K. Mendiguren, G. V. Salamone, and L. Fainboim. 2001. Activation-induced expression of CD1d antigen on mature T cells. *J. Leukocyte Biol.* 69:207.
  24. Arase, H., N. Arase, Y. Kobayashi, Y. Nishimura, S. Yonehara, and K. Onoe. 1994. Cytotoxicity of fresh NK1.1<sup>+</sup> T cell receptor  $\alpha/\beta$ <sup>+</sup> thymocytes against a CD4<sup>+</sup>8<sup>+</sup> thymocyte population associated with intact Fas antigen expression on the target. *J. Exp. Med.* 180:423.
  25. Zhang, B., T. Yamamura, T. Kondo, M. Fujiwara, and T. Tabira. 1997. Regulation of experimental autoimmune encephalomyelitis by natural killer (NK) cells. *J. Exp. Med.* 186:1677.
  26. Busshoff, U., A. Hein, A. Iglesias, R. Dorries, and A. Regnier-Vigouroux. 2001. CD1 expression is differentially regulated by microglia, macrophages and T cells in the central nervous system upon inflammation and demyelination. *J. Neuroimmunol.* 113:220.
  27. Jahng, A. W., I. Maricic, B. Pedersen, N. Burdin, O. Naidenko, M. Kronenberg, Y. Koezuka, and V. Kumar. 2001. Activation of natural killer T cells potentiates or prevents experimental autoimmune encephalomyelitis. *J. Exp. Med.* 194:1789.
  28. Singh, A. K., M. T. Wilson, S. Hong, D. Olivares-Villagomez, C. Du, A. K. Stanic, S. Joyce, S. Sriram, Y. Koezuka, and L. Van Kaer. 2001. Natural killer T cell activation protects mice against experimental autoimmune encephalomyelitis. *J. Exp. Med.* 194:1801.