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Syk Tyrosine Kinase Is Critical for B Cell Antibody Responses and Memory B Cell Survival

Jochen A. Ackermann,* Josquin Nys,* Edina Schweighoffer,* Scott McCleary,† Nicholas Smithers,‡ and Victor L. J. Tybulewicz*

Signals from the BCR are required for Ag-specific B cell recruitment into the immune response. Binding of Ag to the BCR induces phosphorylation of immune receptor tyrosine-based activation motifs in the cytoplasmic domains of the CD79a and CD79b signaling subunits, which subsequently bind and activate the Syk protein tyrosine kinase. Earlier work with the DT40 chicken B cell leukemia cell line showed that Syk was required to transduce BCR signals to proximal activation events, suggesting that Syk also plays an important role in the activation and differentiation of primary B cells during an immune response. In this study, we show that Syk-deficient primary mouse B cells have a severe defect in BCR-induced activation, proliferation, and survival. Furthermore, we demonstrate that Syk is required for both T-dependent and T-independent Ab responses, and that this requirement is B cell intrinsic. In the absence of Syk, Ag fails to induce differentiation of naïve B cells into germinal center B cells and plasma cells. Finally, we show that the survival of existing memory B cells is dependent on Syk. These experiments demonstrate that Syk plays a critical role in multiple aspects of B cell Ab responses. The Journal of Immunology, 2015, 194: 000–000.
more, by deleting Syk after a primary immunization, we show that the kinase is also required for the secondary memory Ab response. Finally, we show that Syk is required for the survival of memory B cells.

**Materials and Methods**

**Mice**

Mice carrying a conditional allele of Syk (Syk<sup>fl/flRMCM</sup>) are a gift from R. Brink and consist of a knock-in targeting a CRE recombinase (MerCreMer, RMCM) in which exon 11 is flanked by loxP sites (Syk<sup>K<sub>12</sub></sup>) and a tamoxifen-inducible Cre recombinase expressed from the ROSA26 locus (Rosa26<sup>Rosa26CreER<sub>TM1Mom</sub></sup>, RMCM) (13). These strains and mice bearing a deleted allele of Syk (Syk<sup>−/−</sup>) (10) were obtained from the breeding facility at the National Institute for Medical Research. To induce Cre activity, mice were injected i.p. for 5 d with 2 mg/day of tamoxifen (Sigma). To generate mixed radiation chimeras, bone marrow was harvested from Syk<sup>−/−</sup>-RMMC or Syk<sup>−/−</sup>-RMMC mice, treated with ACK lysis buffer and mixed in a 1:2 ratio with bone marrow cells from μMT (Igh<sup>μm1Cgcm1Cgcm</sup>) mice, and injected intravenously at 1 × 10<sup>6</sup> cells per recipient into Rag1-deficient animals that were irradiated with 5 Gy using a 137Cs source. Mice expressing an anti–hen egg lysozyme (HEL) BCR were a gift from R. Brink (17) and were obtained from the breeding facility at the National Institute for Medical Research. To induce Cre activity, mice were injected i.p. for 5 d with 2 mg/day of tamoxifen (Sigma). To generate mixed radiation chimeras, bone marrow was harvested from Syk<sup>−/−</sup>-RMMC or Syk<sup>−/−</sup>-RMMC mice, treated with ACK lysis buffer and mixed in a 1:2 ratio with bone marrow cells from μMT (Igh<sup>μm1Cgcm1Cgcm</sup>) mice, and injected intravenously at 1 × 10<sup>6</sup> cells per recipient into Rag1-deficient animals that were irradiated with 5 Gy using a 137Cs source. Mice received Baytril in their drinking water (0.02%, Bayer Healthcare) for at least 4 wk after transplantation, and were used for further studies no less than 6 wk after reconstitution.

**Cell culture**

Splenitic B cells were purified from mice treated 21 d earlier with tamoxifen, by magnetic negative depletion using biotinylated Abs against CD43 (BioR22/60), CD11c (N418), CD11b (M1/70), and CD3c (14-5-2C11; eBioscience) and streptavidin Dynabeads (Life Technologies). B cells (10<sup>6</sup> cells/ml) were incubated in PBS containing 0.5 μM CFSE (Molecular Probes) at 37°C for 10 min. Cells were washed twice in DMEM plus medium (DMEM with 10% FCS, 100 U/ml penicillin, 100 μg/ml streptomycin, 100 μM nonessential amino acids, 20 mM HEPES buffer, 2 mM l-glutamine, and 50 mM 2-ME) and then cultured at 1–1.5 × 10<sup>5</sup> cells/ml in DMEM-plus alone or supplemented with 10 μg/ml anti-IgM Fab<sub>2</sub> (Jackson Immunoresearch) or 100 ng/ml CD40L (R&D Systems) and 20–100 ng/ml IL-4 (PeproTech). Flow cytometry was used to assess surface expression of proteins, proliferation by CFSE dilution, and number of live cells by a LIVE/DEAD fixable near-infrared dead cell stain.

**Immunizations**

For T-independent responses, mice were immunized i.p. with 10 μg TNP-Ficoll. For T-dependent immune responses, mice were immunized i.p. with 50–100 μg Alum-precipitated (Thermo Scientific) 4-hydroxy-3-nitrophenylacetetyl conjugated to chicken-γ-globulin (NP-CGG), with a ratio of NP to CGG ranging from 1 to 27 (BioSearch Technologies). For the recall responses, mice were injected i.p. with 50 μg NP-CGG in PBS, TNP or NP-specific Abs in the serum were measured by ELISA using Maxisorp plates (Nunc) coated with NP<sub>16</sub>-BSA as previously described (16).

**Flow cytometry**

Single-cell suspensions of splenocytes were treated with ACK lysis buffer to remove RBCs before staining in PBS, containing LIVE/DEAD fixable near-IR dead cell stain (Life Technologies) and appropriate, pretetred Abs. Abs used, indicating Ag and fluorophore (and clone): CD69-FITC (H1.2F3), CD86-PE (GL-1), streptavidin-PerCP (Becton Dickinson), MHC class II-bio (I-Ab; M5/114.15.2), IgM-MECJ7 (II/41), B220-eFluor450 (RA3-62B; eBioscience), CD19-FITC, B220-PerC, IgM-PE, IgG1-APC, PNA-FTC, CD38-PE/Cy7, CD45.2-eFluor450, CD45.1-FITC, CD138-PE, IgD-eFluor450, CD4-FITC, CD8-FITC, CD45.2-PerCP-cy5.5, CD45.1-FITC, B220-BV605, CD4-BV650, CD8a-BV650, CD38-bio, Streptavidin-PacO, FAS-PE, Anti-SyK Ab (SF5; BioLegend) and NP-BSA were labeled using LYNX Rapid APC, RPE, and PerCP Ab Conjugation Kits (AbD Serotec). Ab staining of intracellular Syk was performed as described previously (13).

**ELISPOT**

Nitrocellulose 96-well filtration plates (Millipore) were coated overnight at 4°C with 50 μg/ml NP<sub>16</sub>-BSA (Biosearch Technologies) and then blocked with DMEM. Splenocytes or bone marrow cells were serially diluted (maximum concentration of 4 × 10<sup>4</sup> cells/well) on the coated plates and cultivated overnight in DMEM-plus with 5% FCS at 37°C. 5% CO<sub>2</sub>. Plates were washed twice with PBS, 0.05% Tween-20 and twice with PBS, stained with biotinylated anti-IgG1 or anti-IgM Abs for 2 h at room temperature, washed four times with PBS-Tween, and stained with Avidin τ-alanine phosphatase for 1 h at room temperature. Alkaline phosphatase activity was visualized using BCIP/NPT substrate (BioFX). Spots, each representing a single Ab-secreting cell, were counted using an ImmunoSpot reader.

**Cell transfers and immunization with HEL**

To induce primary responses, 2.5 × 10<sup>7</sup> HEL-binding B cells from tamoxifen-treated SWHEL/Syk<sup>fl</sup> or SWHEL/Syk<sup>−/−</sup>-RMMC mice were transferred into B6.SJL recipient mice together with 5 × 10<sup>7</sup> HEL-conjugated sheep RBCs (SRBC). To generate a memory pool, splenocytes containing an equivalent of 5 × 10<sup>7</sup> HEL-binding B cells from SWHEL/Syk<sup>fl</sup>-RMMC or SWHEL/Syk<sup>−/−</sup>-RMMC mice were transferred into (B6 x B6.SJL)F1 recipient mice together with 1 × 10<sup>7</sup> HEL<sup>K<sub>12</sub></sup>-conjugated SRBC. HEL (Sigma-Aldrich) or HEL<sup>K<sub>12</sub></sup> (a gift from R. Brink) (17) was conjugated to SRBC (Patricell) as described previously (18). The germinal center reaction was blocked by three injections of 300 μg anti-CD40L (MR1) or control Hamster IgG (BioXcell) every second day.

**Statistical analysis**

All statistical comparisons used the nonparametric two-tailed Mann–Whitney U test or Student t test. Statistically significant differences are indicated in the figures.

**Results**

Syk is required for in vitro BCR-induced activation

Initially, we investigated whether Syk was required for Ag receptor-induced activation of B cells in vitro, using mice containing a conditional allele of Syk in which exon 11 is flanked by loxP sites (Syk<sup>K<sub>12</sub></sup>) and a tamoxifen-inducible Cre recombinase expressed from the ROSA26 locus (Rosa26<sup>Rosa26CreER<sub>TM1Mom</sub></sup>, RMCM) (13). These strains and mice bearing a deleted allele of Syk (Syk<sup>−/−</sup>) (10) were intercrossed to generate control (Syk<sup>+/+</sup>-RMMC) and conditional mutant (Syk<sup>fl/fl</sup>-RMMC) mice. Treatment with tamoxifen resulted in deletion of the Syk<sup>fl</sup> allele in both strains, leaving control and mutant mice with either one or no functional alleles of Syk. As established previously, by 10 d after tamoxifen treatment, Syk protein is no longer detectable in splenic B cells from mutant mice, and by 21 d ~80% of the B cells are lost because of impaired survival (13). Despite this loss of B cells, ~20% of follicular B cells persist in the mutant but have no detectable Syk. Thus, we were able to use these Syk-deficient B cells to investigate whether Syk was required for BCR-induced activation in vitro. We found that Syk was required for BCR-induced increase in cell surface expression of CD69, CD86, and MHC class II proliferation and survival (Fig. 1A, 1B). In contrast, Syk-deficient B cells were able to respond at least as well as control cells to stimulation with CD40L plus IL-4, as measured by the same parameters, demonstrating that Syk-deficient cells retained the capacity to become activated and to proliferate. However, Ig isotype class switching to IgG1 and secretion of IgM and IgG1 in response to CD40L plus IL4 were greatly reduced in the absence of Syk (Fig. 1C, 1D). Thus, Syk was required in all BCR-induced activation in vitro responses we measured, but only in a select subset of responses induced by CD40L plus IL-4.

**Defective T-independent responses in the absence of Syk**

Next, we investigated the requirement for Syk during in vivo B cell responses to Ag. The Rosa26<sup>Rosa26CreER<sub>TM1Mom</sub></sup> allele is expressed ubiquitously; therefore, to rule out secondary effects on Ab responses because of deletion of Syk in non-B cells, we limited the loss of Syk to B cells by generating mixed chimeras in which the hematopoietic system of irradiated Rag1-deficient mice was reconstituted with...
a mixture of bone marrow from μMT (B cell–deficient) mice and either Syk-expressing (Syk<sup>fl/+RMCM</sup>) or Syk-deficient (Syk<sup>fl/2RMCM</sup>) mice. In the resulting mutant mice, all B cells lost Syk expression after tamoxifen treatment, whereas Syk was maintained in other cell types. Initially, we determined whether Syk was required for T-independent type-II (TI-II) responses to TNP-Ficoll, a hapten coupled to highly repetitive polysaccharide, by immunizing chimeric mice 21 d after treatment with tamoxifen. We found that in the absence of Syk, there was a significant reduction in Ag-specific IgM and IgG3 (Fig. 2), demonstrating a B cell-intrinsic requirement for Syk in TI-II Ab responses.

### Defective T-dependent Ab response in the absence of Syk in B cells

To determine whether Syk was also required for T-dependent B cell responses, Syk<sup>fl/+RMCM</sup> and Syk<sup>fl/2RMCM</sup> mice were treated with tamoxifen, immunized 21 d later with NP-CGG precipitated in alum, and then analyzed after an additional 10 d (Fig. 3A). We found that loss of Syk resulted in a significant decrease in Ag-specific IgM, IgG1, IgG2b, and IgG2c in the serum, and in the induction of far fewer germinal center B cells (Fig. 3B). Notably, although most B cells in the mutant mice had no detectable Syk expression, the few remaining germinal center B cells in the same mice still expressed Syk (Fig. 3B). This strong selection against cells that have deleted Syk indicates that B cells require Syk to differentiate into germinal center cells.

To evaluate whether the requirement for Syk in T-dependent B cell responses was B cell-intrinsic, we immunized chimeric mice with NP-CGG. We found that restricted loss of Syk in B cells resulted in a large decrease in Ag-specific IgM and IgG1 in the serum (Fig. 3C), and a large reduction in the numbers of Ag-specific IgG1-expressing B cells and germinal center B cells, as well as a reduction in Ab-secreting cells producing Ag-specific IgM or IgG1 (Fig. 3D).

### Figure 1.
Syk is required for activation of B cells in response to anti-IgM and class switching in response to CD40L + IL-4. (A) Mean (±SEM) expression of CD69, CD86, and MHC class II on B cells of the indicated genotypes stimulated with anti-IgM or CD40L plus 100 ng/ml IL-4 or left unstimulated (–) for 36 h. (B) Mean (±SEM) percentage of divided cells and percentage of live cells in cultures of B cells stimulated as in (A) for 60 h. (C) Dot plots of surface staining of IgM and IgG1 on B cells of the indicated genotypes stimulated for 60 h with CD40L plus 20 ng/ml IL-4 or left unstimulated (–). Numbers indicate the percentage of cells falling into the gate. Graph of mean (±SEM) percentage of IgG1<sup>+</sup> B cells in cultures, determined as in dot plots. (D) Mean (±SEM) levels of IgM and IgG1 secreted into the medium of B cells cultured as in (C) for 6 d. *p < 0.05, **p < 0.01, ***p < 0.001. n.s., not significant.

### Figure 2.
Defective T-independent Ab response in the absence of Syk in B cells. Irradiated Rag1-deficient mice reconstituted with a mixture of bone marrow from μMT (B cell–deficient) mice and either Syk-expressing (Syk<sup>fl/+RMCM</sup>) or Syk-deficient (Syk<sup>fl/2RMCM</sup>) mice. In the resulting mutant mice, all B cells lost Syk expression after tamoxifen treatment, whereas Syk was maintained in other cell types. Initially, we determined whether Syk was required for T-independent type-II (TI-II) responses to TNP-Ficoll, a hapten coupled to highly repetitive polysaccharide, by immunizing chimeric mice 21 d after treatment with tamoxifen. We found that in the absence of Syk, there was a significant reduction in Ag-specific IgM and IgG3 (Fig. 2), demonstrating a B cell-intrinsic requirement for Syk in TI-II Ab responses.

To determine whether Syk was also required for T-dependent B cell responses, Syk<sup>fl/+RMCM</sup> and Syk<sup>fl/2RMCM</sup> mice were treated with tamoxifen, immunized 21 d later with NP-CGG precipitated in alum, and then analyzed after an additional 10 d (Fig. 3A). We found that loss of Syk resulted in a significant decrease in Ag-specific IgM, IgG1, IgG2b, and IgG2c in the serum, and in the induction of far fewer germinal center B cells (Fig. 3B). Notably, although most B cells in the mutant mice had no detectable Syk expression, the few remaining germinal center B cells in the same mice still expressed Syk (Fig. 3B). This strong selection against cells that have deleted Syk indicates that B cells require Syk to differentiate into germinal center cells.

To evaluate whether the requirement for Syk in T-dependent B cell responses was B cell-intrinsic, we immunized chimeric mice with NP-CGG. We found that restricted loss of Syk in B cells resulted in a large decrease in Ag-specific IgM and IgG1 in the serum (Fig. 3C), and a large reduction in the numbers of Ag-specific IgG1-expressing B cells and germinal center B cells, as well as a reduction in Ab-secreting cells producing Ag-specific IgM or IgG1 (Fig. 3D).
FIGURE 3. Defective T-dependent Ab response in the absence of Syk in B cells. (A and B) Syk-expressing (Syk^{fl/+}RMCM) or Syk-deficient (Syk^{fl/-}RMCM) mice were treated with tamoxifen, immunized 21 d later with NP-CGG in Alum, and analyzed 10 d later. (A) Graph shows anti-NP Ig of the indicated isotype in serum of mice of the indicated genotypes either just before immunization (day 0) or 10 d later (dots, individual mice; line, mean). (B) Dot plots of splenocytes from mice 10 d after immunization gated on B220+ cells showing staining with Abs to Fas and GL7. Numbers indicate the percentage of cells falling into each gate. Column graph shows levels of Syk in T cells (B220^-), B cells (B220^+), and germinal center B cells (GCs). T cells show no detectable Syk by this assay. (C and D) Irradiated Rag1-deficient mice reconstituted with a mixture of μMT bone marrow and Syk-expressing (Syk^{fl/+}RMCM) or Syk-deficient (Syk^{fl/-}RMCM) bone marrow were treated with tamoxifen, immunized 21 d later with NP-CGG in alum or alum alone, and analyzed 10 d after that. (C) Graph of anti-NP IgM or IgG1 in mice of the indicated genotypes (dots, individual mice; line, mean). (D) Mean (±SEM) number of Ag-specific switched B cells (B220^+NP^+IgG1^+) or germinal center B cells (B220^+Fas^+GL7^+NP^+IgG1^+; Supplemental Fig. 1A) or Ab-secreting cells (ASCs) making NP-specific IgM or IgG1. (E) SWHEL, Syk^{fl/+}, or Syk^{fl/-}RMCM mice were treated with tamoxifen and 21 d later B cells were transferred into B6.SJL mice. Recipient mice were immunized with SRBC or HEL-SRBC and analyzed 5 d later. Graphs show mean (±SEM) number of total splenic donor-derived B cells, non-GC B cells, GC B cells, and plasma cells (Supplemental Fig. 1B). *p < 0.05, **p < 0.01, ***p < 0.001.
The reduced T-dependent B cell response could have been caused by a requirement for Syk in B cells to differentiate into germinal center and plasma cells. Alternatively, it could have resulted from Syk deficiency reducing the number of naive B cells, leading to a lower number of precursor cells able to respond to the immunizing Ag. To address this issue, we bred the conditional Syk mutation to SWHEL mice expressing a BCR-specific for HEL (15). In this strain, a recombined VDJ region from a HEL-specific Ab has been inserted into the IgH locus, and the mice also contain a transgene expressing an Igκ L chain from the same Ab. We transferred equal numbers of control and Syk-deficient SWHEL B cells into wild-type recipients, and immunized them with HEL-coupled SRBC or SRBC alone (Fig. 3E). Analysis 5 d later showed that control B cells expanded strongly in mice that were immunized with HEL-SRBC compared with SRBC alone, but that this did not occur in the absence of Syk (Fig. 3E). Subdividing these B cells into nongerminal center and germinal center B cells showed the same result—a large expansion of control Syk-expressing B cells, but not of mutant Syk-deficient cells. Ag-induced expansion of plasma cells was also strictly Syk-dependent. Therefore, B cell-expressed Syk is required for the Ag-induced expansion of B cells and their differentiation into germinal center and plasma cells in response to a T-dependent Ag.

Defective Ab recall response in the absence of Syk in B cells

A characteristic feature of the adaptive immune response to T-dependent Ags is a secondary, or recall, response that is typically faster and stronger than the primary response. This is due to the formation of memory B and T cells during the primary response, which respond more rapidly and more strongly to Ag than naive lymphocytes (19). To evaluate whether Syk is also required...
by memory B cells for their response to Ag, we immunized chi-
meric mice in which deletion of Syk could be limited to the B cell
lineage, but that had not yet been treated with tamoxifen. The
mice were immunized twice with NP-CGG precipitated in Alum,
treated with tamoxifen 53 or 99 d later to delete Syk in B cells,
and given a secondary challenge of NP-CGG in PBS or just PBS
alone a further 21d later (Fig. 4A). Analysis of splenocytes 5 d
after this secondary boost showed that although the number of
control Ag-specific switched B cells or plasma cells increased
greatly in response to the secondary challenge with NP-CGG, in
the absence of Syk this expansion was largely absent (Fig. 4A).
Notably, the few switched B cells or plasma cells seen in these
mice still expressed Syk, showing again a strong selection against
the loss of Syk in responding B cells. Further analysis showed
similar large reductions in Ab-secreting cells making NP-specific
IgM and IgG1 and in Ag-specific switched IgG1+ B cells (Fig. 4B,
4C). These results indicate that the absence of Syk in B cells
greatly diminishes the secondary recall response.

Syk is required for the survival of memory B cells

This requirement for Syk in the secondary T-dependent response
could have been due to a role for Syk within memory B cells for
their proliferation and differentiation into plasma cells. Alterna-
tively, Syk could have been required for the survival of memory
B cells. We have previously shown that the survival of naïve B cells
is dependent on Syk, in part because it transduces survival signals
from BAFFR and the BCR (13). However the signals controlling
the survival of memory B cells are largely unknown. It has been
established that BAFF is not required for their survival (20), but it
is not known whether the BCR is required. Thus, we next inves-
tigated the potential role for Syk in memory B cell survival. We
again made use of SWHEL mice bred to either control or con-
ditional Syk mutant mice and transferred equal numbers of
SWHEL B cells from each strain into wild-type recipients, and
immunized with HEL3×SRBC or SRBC alone (Fig. 5). HEL3×
is a variant of HEL with a lower affinity for the anti-HEL BCR in
SWHEL mice that gives longer-lasting immune responses (17).
The donor mice were not treated with tamoxifen; therefore, the
SWHEL B cells from both control and conditional mutant mice
still expressed Syk and thus could mount a strong immune re-
sponse, including formation of similar numbers of memory B cells
(not shown). Fifty-four days after immunization, the mice were
immunized with tamoxifen to delete Syk in the conditional mutant
memory B cells, and the numbers of these cells was evaluated 21 d

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**FIGURE 5.** Syk is required for the survival of memory B cells. B cells from SWHEL Sykfl/+RMCM or Sykfl/flRMCM mice were transferred into (B6 x B6.
SJL) F1 mice, and recipient mice were immunized with SRBC or HEL3×SRBC. After 48 d, mice were given three injections of anti-CD40L (MR1) or an
isotype control Ab and then tamoxifen 54 d after immunization, and they were analyzed 21 d later. Dot plots in the top row show expression of CD38 and
Fas on donor-derived splenic B cells (CD45.1–B220+), and the gate indicates naïve and memory B cells (CD38+Fas+); these in turn were analyzed for
expression of IgD and IgG1 in the lower dot plots, where the gate shows donor-derived IgG1+ memory B cells. Numbers indicate the percentage of cells
falling into the gate. The graph shows the number of splenic donor-derived IgG1+ memory B cells, with each dot representing a single mouse and the line
showing the mean. **p < 0.01.

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showed that deletion of Syk resulted in a large reduction in the number of IgG1+ memory B cells (Fig. 5). Thus, Syk is required for the survival of memory B cells.

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Disclosures

S.M. and N.S. are employees of GlaxoSmithKline. The other authors have no financial conflicts of interest.

References

Syk tyrosine kinase is critical for B cell antibody responses and memory B cell survival

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Supplemental Figures
Supplemental Figure 1. Defective T-dependent antibody response in the absence of Syk in B cells. (A) Irradiated Rag1-deficient mice reconstituted with a mixture of μMT bone marrow and Syk-expressing (Syk^fl/fl^RMCM) or Syk-deficient (Syk^fl/^RMCM) bone
marrow were treated with tamoxifen, immunized 21d later with NP-CGG in Alum or Alum alone and analyzed 10d after that, as shown in the time-line. Dot plots show binding of antigen (NIP) and expression of IgG1 on the surface of splenic B cells or germinal center (GC) B cells. Gates indicate antigen-specific switched cells (NIP⁺IgG1⁺). Numbers indicate % of cells falling into gate and were used to determine data shown in Fig 3D. (B) SWHEL Syk⁺/+ or SWHEL Syk⁻/⁻ RMCM mice were treated with tamoxifen and 21d later B cells were transferred into B6.SJL mice, recipient mice were immunized with SRBC or HEL-SRBC and analyzed 5d later, as shown in the time-line. Dot plots in top row show expression of B220 and CD138 on donor (CD45.2⁺) splenocytes and gates indicate B cells (B220⁺CD138⁻) and plasma cells (B220⁻CD138⁺). Dot plots in lower row indicate surface binding of PNA and expression of CD38 on donor-derived splenic B cells (CD45.2⁺B220⁺CD138⁻). Gates indicate non-germinal center (PNA⁺CD38⁺) and germinal center (PNA⁺CD38⁻) cells. Numbers indicate % of cells falling into gate and were used to determine data shown in Fig 3E.
Supplemental Figure 2. Defective antibody recall response in the absence of Syk in B cells. Irradiated Rag1-deficient mice reconstituted with a mixture of μMT bone marrow and Syk-expressing ($\text{Syk}^{fl/+}$RMCM) or Syk-deficient ($\text{Syk}^{fl/-}$RMCM) bone marrow were immunized twice with NP-CGG in Alum. 85d after the second immunization the mice were treated with tamoxifen, and a further 21d later immunized with NP-CGG in PBS or PBS alone and analyzed 5d after that, as shown in the time-line. Dot plots show surface binding of antigen (NIP) and expression of IgD on splenic non-T cells (CD4'CD8'), which are mainly B cells, and on plasma cells (CD138'). Gates show switched antigen-specific cells (NIP'\text{IgD}^{-}) and were used to determine data shown in Fig 4A.