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*J Immunol* published online 11 March 2015
http://www.jimmunol.org/content/early/2015/03/10/jimmunol.1402254

Supplementary Material
http://www.jimmunol.org/content/suppl/2015/03/10/jimmunol.1402254.4.DCSupplemental

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Deficiency of MALT1 Paracaspase Activity Results in Unbalanced Regulatory and Effector T and B Cell Responses Leading to Multiorgan Inflammation


The paracaspase MALT1 plays an important role in immune receptor-driven signaling pathways leading to NF-κB activation. MALT1 promotes signaling by acting as a scaffold, recruiting downstream signaling proteins, as well as by proteolytic cleavage of multiple substrates. However, the relative contributions of these two different activities to T and B cell function are not well understood. To investigate how MALT1 proteolytic activity contributes to overall immune cell regulation, we generated MALT1 protease-deficient mice (Malt1PD/PD) and compared their phenotype with that of MALT1 knockout animals (Malt1−/−). Malt1PD/PD mice displayed defects in multiple cell types including marginal zone B cells, B1 B cells, IL-10–producing B cells, regulatory T cells, and mature T and B cells. In general, immune defects were more pronounced in Malt1−/− animals. Both mouse lines showed aberrant B cell responses upon immunization with T-dependent and T-independent Ags. In vitro, inactivation of MALT1 protease activity caused reduced stimulation-induced T cell proliferation, impaired IL-2 and TNF-α production, as well as defective Th17 differentiation. Consequently, Malt1PD/PD mice were protected in a Th17-dependent experimental autoimmune encephalomyelitis model. Surprisingly, Malt1PD/PD animals developed a multiorgan inflammatory pathology, characterized by Th1 and Th2/0 responses and enhanced IgG1 and IgE levels, which was delayed by wild-type regulatory T cell reconstitution. We therefore propose that the pathology characteristic of Malt1PD/PD animals arises from an immune imbalance featuring pathogenic Th1- and Th2/0-skewed effector responses and reduced immunosuppressive compartments. These data uncover a previously unappreciated key function of MALT1 protease activity in immune homeostasis and underline its relevance in human health and disease.  The Journal of Immunology, 2015, 194: 000–000.

Uncontrolled NF-κB activation can lead to lymphoproliferative disorders and autoimmune diseases associated with chronic inflammation, underscoring the need for appropriate regulation of signaling via this commonly used pathway (1–4). The paracaspase MALT1 plays an important role in the regulation of NF-κB (5). Upon activation by Ag receptors, Dectin receptors, or G protein–coupled receptors, MALT1 and the caspase recruitment domain (CARD)–containing protein BCL10 assemble with another CARD-containing protein: CARD11 (also known as CARMA-1), CARD9, or CARD10, respectively. The resulting CARD-BCL10-MALT1 (CBM) complex functions as a hub for triggering a signaling cascade that culminates in activation of the IkB kinase (IKK) complex, IkB degradation, and NF-κB activation (6, 7).

MALT1 regulates NF-κB activity in at least two ways. First, its scaffolding function is essential for NF-κB activation (8, 9).
Second, MALT1 has proteolytic activity targeting several substrates important in NF-κB signaling (10). These include BCL10, whose cleavage by MALT1 appears to regulate integrin-dependent T cell adhesion (11). The other known substrates of MALT1 are all negative regulators of canonical NF-κB signaling. MALT1-dependent cleavage of the two ubiquitin editing enzymes, A20 and cylindromatosis (CYLD), downregulates their activity (12–15). MALT1 also cleaves RelB, priming it for proteasome-dependent degradation (16) and resulting in modulation of canonical NF-κB activation via the release of RelA and c-Rel. In addition, Regnase-1, an RNase that destabilizes a subset of NF-κB–dependent mRNAs by cleaving their 3′ termini (e.g., c-Rel and IL-2), is also proteolyzed and inactivated by MALT1 (17). Thus, MALT1 regulates the NF-κB cascade by acting as a scaffolding partner within the CBM complex enabling signaling, and as a protease by cleaving key regulatory proteins, thereby sustaining NF-κB activation.

Previous reports using MALT1, CARD11-, and BCL10-deficient cells have established that the CBM complex is essential for NF-κB signaling in T and B cells (8, 9, 18–20). However, the relevance of MALT1 protease versus scaffolding activities in relation to immune function and the long-term consequences associated with the abrogation of MALT1 enzymatic activity in vivo are still unclear. To investigate this point, we generated and characterized a novel MALT1 protease dead (PD) mouse line (Malt1PD/PD). In this study, we describe an unexpected phenotype of Malt1PD/PD mice, as well as the impact of constitutive disruption of MALT1 enzymatic activity on lymphocyte compartments and general immune homeostasis. A direct comparison with MALT1 knockout (Malt1−/−) mice enabled us to delineate the respective contributions of scaffolding and proteolytic functions of MALT1 in the regulation of T and B cell–mediated immunity.

Materials and Methods

**Generation of Malt1PD/PD and Malt1−/− mice and genotyping**

To generate a targeting vector for homologous recombination, we amplified MALT1 genomic sequences from C57BL/6 mouse genomic DNA. First, a 1.6-kb DNA fragment containing Malt1 exons 10 and 11 was amplified by PCR, and a cysteine-to-alanine mutation was introduced at aa position 472 by overlapping PCR. The mutated fragment was cloned into vector pRAY 2loxP 2FRT in between the two loxp sites. Next, a 2.5-kb genomic fragment corresponding to Malt1 intron 9 was amplified and cloned in front of the 5′ loxp site. Finally, 1.7-kb Malt1 genomic DNA downstream of exon 11 was amplified and subcloned behind the FRT-flanked neomycin cassette of pRAY 2loxP 2FRT, resulting in the final Malt1 targeting construct for homologous recombination. Subcloned sequences were compared with sequences available from the Ensembl database (Ensembl Gene ID ENSMUSG00000032688).

After introduction into C57BL/6 embryonic stem cells, neomycin-resistant clones were screened by PCR for homologous recombination. Correct targeting was confirmed by Southern blot using a neomycin-specific probe that allowed the exclusion of random integration events of the targeting vector. Selected targeted embryonic stem cells were injected into BALB/c blastocysts allowed the exclusion of random integration events of the targeting vector. Selected targeted embryonic stem cells were injected into BALB/c blastocysts. Malt1PD/PD animals were generated by the interbreeding of Malt1PD/WT animals. Malt1−/− animals were bred Malt1PD/PD mice with a Cre5 mouse line to excise exons 10 and 11 of the Malt1 gene (Supplemental Fig. 1A). Successive breeding was used to generate homozygous Malt1−/− animals devoid of the Cre recombination gene. Genotyping was performed using the following primer sets: Forward 5′-CCGAGCCATGCTAGCCT-GAAGC-3′, and Reverse 5′-GGCGTCTTGGTGATTGCGTC-3′, 5′-ACACGCGGATATGTCAGGCA-3′.

**Generation of bone marrow chimeras**

To generate bone marrow (BM) chimeras, we injected 106 BM cells derived from female CD45.2 wild-type (WT), Malt1PD/PD, or Malt1−/− mice i.v. into 10-wk-old sublethally irradiated (2 times 450 rad 4 h apart) CD45.1 female recipients (B6.SJL-PtgRc5Pepc5Boyd; Taconic). BM cells were isolated by flushing femur and tibiae with RPMI 1640 media and subsequently depleted of RBC by hypotonic shock using ACK buffer; 0.82% NH4Cl, 0.1% KHC2O3, and 0.0372% EDTA.

**DNP-keyhole limpet hemocyanin and 2,4-dinitrophenyl-β-αnyl-glycyl-glycyl-N-[2-aminoethyl]carbamyl-methylated–Ficol immunization and tier measurement**

Keyhole limpet hemocyanin (KLH; Calbiochem) conjugated to DNP (Sigma-Aldrich) was prepared, at a ratio of 1:20, as DNP-KLH stock solution (5 mg/ml) and stored at −20°C. DAGG (2,4-DNP-β-αnyl-glycyl-glycyl-N-[2-aminoethyl]carbamyl-methylated)–Ficol conjugate was prepared as described previously (21). For induction of T-dependent (TD) responses, DNP-KLH was adsorbed onto colloidal aluminum hydroxide (Alu-Gel-S; SERVA; 1 ml DNP-KLH/5 ml Alu-Gel-S/ml PBS) and injected i.p. into mice at 100 μg/0.2 ml/mouse. For induction of T-independent (TI) type 2 (TI-2) responses, DAGG-Ficol was injected in PBS i.v. into mice at 20 μg/0.2 ml/mouse. DNP-specific IgG and IgM levels were measured in serum on days 8 (DNP-KLH) or 6 (DAGG-Ficol) postimmunization. Ab titers were measured by ELISA and are expressed as mean ± SEM log10 dilution values using 50% of the maximal extinction at 405 nm as the end point.

**T cell activation, proliferation, and differentiation**

For T cell proliferation assays, total CD3+ T cells purified from spleens using the CD3 isolation kit (Miltenyi Biotec) were cultured in IMDM (Life Technologies), 10% FCS (Amimed), and 50 U/ml Pen-strep. For differentiation of CD4+ T cells into Th17 cells, naive Th cells were stimulated for 5 days with 1 μg/ml plate-bound anti-CD3 Ab (clone 37-51; BD Biosciences). Three days after stimulation, plates were pulsed with 1 μCi/ml [3H]thymidine (Amersham) for 5 h and incorporation of [3H]thymidine was measured using a β-plate counter (Wallac).

For ex vivo T cell stimulation for cytokine release or intracellular cytokine staining was performed on total LN or spleen cells. In brief, 2.5 × 106 cells/ml were stimulated for 4 h at 37°C in 24-well plates in the presence of 100 ng/ml PMA (Sigma-Aldrich) and 100 ng/ml ionomycin (Sigma-Aldrich). For cytokine secretion assays, supernatants were collected 4 h after stimulation and cytokine levels were analyzed using Meso Scale Discovery following the manufacturer’s instructions and an MSD Sector Imager 6000. Cell stimulation for intracellular cytokine staining was performed in the presence 1 μl/ml GolgiStop (BD Biosciences).

**In vitro Th cell differentiation assays, naive CD4+CD25− T cells were isolated from spleens and LN, and sorted using a magnetic bead cell purification kit according to the manufacturer’s instructions (Miltenyi Biotec). For priming, enriched Th cells were stimulated for 96 h with 1 μg/ml plate-bound anti-CD3 Ab plus 1–2 μg/ml anti-CD28 Ab (BD Biosciences). naive cells were induced to differentiate into regulatory T cells (Tregs) by addition of 3 ng/ml recombinant human TGF-β (rhTGF-β; R&D), 30 U/ml rh-IL-2 (Peprotech), and 5 μg/ml anti–IFN-γ Ab (BD Biosciences), and into Th1 cells by addition of 4 ng/ml IL-12 (Peprotech) plus 50 U/ml rhIL-2. For differentiation of Th17 cells, naive Th cells received 5 μg/ml anti–IFN-γ Ab, 30 ng/ml rhIL-6 (Peprotech), 2 ng/ml rhTGF-β, and 50 U/ml rhIL-2.

**FACS analysis and reagents**

FACS stainings of cell suspensions from spleen, LN, and Peyer’s patches (PP) were performed by passing tissues through a 70-μm sieve followed by RBC lysis using ACK buffer. Cells were washed once in FACS buffer (PBS containing 2% FCS, 0.05% NaN3) blocked with mouse Fc Block (BD Biosciences), and stained for 30 min at 4°C with the indicated combination of fluorochrome-conjugated Abs. After staining, the cells were washed twice and resuspended in 200 μl buffer before acquisition on a FACSCanTol or FACSFortessa flow cytometer. Data were analyzed using the FlowJo software.

The Abs were used obtained from different sources: B220, CD25, CD3, CD4, CD11b, CD45.1, CD45.2, Fasp3, GL7, IgG1, IgM, PD1, TNF-α, and TNF-β (BD Biosciences); CD4; CD11c, CD19, CD44, CXCR5, IFN-γ, IL-2, IL-17A, and TNF-α from BD Biosciences; CD3, CD5, CD8, CD21/CD35, CD23, CD62L, and IgG1 from Biologend; peanut agglutinin–FITC from Vector Laboratory.

**Histology, immunofluorescence, and serum Ig measurements**

Histological analyses were performed on formalin-fixed organs using standard protocols and Ig levels assessed by the Luminox technology.
Immunofluorescence stainings were performed on OCT-embedded snap-frozen tissues with antibodies against JNK, anti-phospho–stress-activated protein kinase/JNK (Thr183/Tyr185) and CYLD, a reported MALT1 substrate important for the regulation of NF-κB signaling.

For immunofluorescence stainings, spleens and mesenteric LN (mesLN) were snap frozen, OCT-embedded, and 5-μm sections were prepared. Sections were fixed in acetone for 10 min and then air-dried for 60 min. Spleen sections were stained with anti-IgM-FITC, anti-IgD-Alexa 647, and Moma-PE. mesLN sections were stained with anti–IgM-FITC, anti-IgD-Alexa 647, and peanut agglutinin–PE.

Serum and intestinal lavage Ig concentrations were determined by the Luminex xMAP technology using the IgE Single Plex Magnetic Bead Kit and the Ig Isotyping Magnetic Bead Panel (Merck Millipore) with a Luminex 200 instrument according to manufacturer’s instructions.

**Treg adoptive transfer**

For Treg reconstitution experiments, transferred cells were FACS sorted from LN and spleen cells collected from EGFP-Foxp3 knock-in mice (Jackson Laboratory). In brief, pooled LN and spleens were stained with fluorescently labeled CD4 Ab and EGFP CD4 (i.e., Foxp3+), and EGFP+ CD4+ (Foxp3+) cell populations were sorted onto a FACSAria. Freshly sorted cells were then injected i.v. (5 × 10^5 cells/mouse) into 8- to 9-wk-old female Malt1PD/PD mice. Mice were subsequently monitored for weight loss and signs of pathology development.

**B cell activation**

B cell proliferation assays were performed on total splenic B cells purified by negative selection using an EasySep Mouse B Cell Enrichment Kit (Stemcell Technologies). Cells were plated at 10^6 cells/well on flat-bottom 96-well plates in RPMI 1640, 1 mM sodium pyruvate, nonessential amino acids, 50 μM 2-ME, 100 U/ml penicillin and 100 μg/ml streptomycin (Life Technologies), and 10% FCS (Amimed). Cells were stimulated with 10 μg/ml CpG1826 (Invivogen), 25% of the CD40L-containing supernatant from cell line S49-40-19 with or without 125 U/ml IL-4 (derived from CHO-mouse IL-4 KI-5; Novartis), 50 ng/ml LPS from Salmonella enterica (Sigma-Aldrich) or 25 μg/ml anti-IgM Ab (Jackson Immunoresearch) supplemented with 5 μg/ml IL-4 (Peprotech). On day 3 of culture, cells were pulsed with 1 μCi/well [3H]thymidine (Amersham), and 5 h later incorporation of [3H]thymidine was measured using a β-plate counter (Wallac).

For experiments assessing TLR-ligand–induced cytokine release, cells were stimulated overnight with ssRNA40 (200 μg/ml) and CpG1826 (20 μM) (Invivogen). Levels of TNF-α, IL-6, and IL-10 in culture supernatants were measured using a plate counter.

**Inmunoblotting**

CD3+ T cells were isolated from pooled LN using the mouse T cell enrichment kit (Stemcell Technologies). CD19+ B cells were isolated from pooled spleens using the mouse B cell enrichment kit (Stemcell Technologies). A total of 1.4 × 10^7 purified CD3+ T cells or CD19+ B cells were stimulated for the indicated times using 40 ng/ml PMA and 1 μM ionomycin. Cells were collected by centrifugation and resuspended in lysis buffer (36 μM β-glycerophosphate pH 7.5, 1% Nonidet P-40, 0.5% Na cholate, 0.1% SDS, 2 mM DTT, 1% phosphatase inhibitor mixture 2 and 3 (Sigma-Aldrich) supplemented with complete protease inhibitor mixtures (Roche)). After 5-min incubation on ice, cell lysates were cleared by centrifugation at 13,000 × g for 10 min. Proteins were separated on 4–12% Bis-Tris SDS-PAGE gradient gels (Life Technologies) and transferred to polyvinylidene difluoride membranes (Life Technologies), and immunobLOTS were incubated with the indicated Abs. Finally, proteins were detected by fluorescence using an Odyssey imager following the instructions provided by the manufacturer (Li-COR Biosciences).

Abs used for Western blot detection were anti-MALT1 (H-300) and anti-CYLD (E-10) from Santa Cruz Biotechnology; anti-RelB (C1E4), anti–phospho–Rac1 (Ser21/22) (G9), anti–phospho–p44/42 MAPK (Erk1/2) (Thr202/Tyr204) (D13.14.4E) from Cell Signaling Technology; anti–Regnase (MCPIP1) (604421) from R&D, and anti–phospho-p44/42 MAPK (Erk1/2) (Thr202/Tyr204) from Sigma-Aldrich. Western blot analysis was performed using the QuantTec Reverse Transcription Kit (Qiagen). For quantitative PCR, cDNA fragments were amplified through the use of TaqMan Fast Universal PCR Master Mix and TaqMan probes for mouse c-Rel and IL-2 (Applied Biosystems). Fluorescence was detected with a 7500 fast real-time PCR system (Applied Biosystems). The mRNA expression level was normalized to the expression level of β2-microglobulin, and changes were calculated using the ΔΔct cycle threshold method.

**Experimental autoimmune encephalomyelitis induction**

Nine-week-old male WT or Malt1PD/PD mice (n = 10/group) were s.c. immunized at the lower back with myelin oligodendrocyte glycoprotein (MOG35-125, in-house produced; 200 μg/100 μl) emulsified in 4 mg/ml CFA (Sigma-Aldrich). Pertussis toxin (Fluka; 200 ng/mouse) was administered i.p. on days 0 and 2. Experimental autoimmune encephalomyelitis (EAE) development was monitored daily using the well-established scoring system (0, normal appearance; 1, complete tail paralysis; 2, unilateral partial hind-limb paralysis; 3, complete bilateral hind-limb paralysis; 4, quadriplegia; 5, death).

**Statistics**

Bar graphs in the figures represent average values ± SEM unless indicated otherwise. Statistical significance between groups was calculated using a two-tailed unpaired Student t test or a Kruskal–Wallis one way ANOVA using GraphPad Prism (GraphPad Software) and is indicated in the graphs as follows: *p < 0.05, **p < 0.01, ***p < 0.001. Nonsignificant differences were not indicated.

**Study approval**

Procedures involving animals were carried out on Experimental Animal Licenses approved by the regional governmental authorities. All efforts were made to minimize animal suffering, and Malt1PD/PD mice were humanely killed before or as soon as the first signs of pathology were noticed.

**Results**

Malt1PD/PD mice express catalytically inactive MALT1 and display normal activation of the IKK, JNK, and ERK pathways in T and B lymphocytes

To study the physiological relevance of MALT1 enzymatic function, we generated and characterized a novel PD MALT1 knock-in mouse line (Malt1PD/PD), where the protease function of MALT1 was specifically ablated by mutating the active site cysteine (C472A). Because of the specific design of the targeting vector, it was possible to generate a control Malt1+/- mouse line that originated from the same targeted offspring as the Malt1PD/PD mice (Supplemental Fig. 1A). Malt1PD/PD and Malt1-/- mice were both born at rates close to the expected Mendelian ratio (Supplemental Fig. 1B) and displayed a normal growth and behavior until at least 8–9 wk of age.

To confirm the impact of the alterations in the Malt1 gene introduced by the targeting strategy on the expression and function of MALT1 protein, we assessed purified splenic T and B cells from the different mouse strains for MALT1 protein levels and function, as well as for PMA/ionomycin-induced activation of NF-κB, ERK, and JNK pathways. As expected, MALT1 protein was undetectable in Malt1-/- lymphocytes, whereas the C472A MALT1 mutant was expressed in Malt1PD/PD lymphocytes to a similar level as in WT cells (Fig. 1A and 1B). Cleavage of CYLD, a reported MALT1 substrate important for the regulation of JNK, was abrogated in Malt1PD/PD T cells, confirming that the C472A mutation rendered MALT1 catalytically inactive. Consistently, cleavage of two further MALT1 substrates, RelB and Regnase-1, was also fully blocked in Malt1PD/PD T cells (Fig. 1A).

Phosphorylation of the IKK substrate IκBα was clearly detectable in Malt1PD/PD T cells but blocked in the absence of...
MALT1 protein (Fig. 1A), indicating that IKK activity is largely independent of MALT1 proteolytic function. Consistent data were obtained with other IKK substrates such as p65 and p105 (data not shown). Similarly, normal phosphorylation of JNK and ERK kinases in Malt1PD/PD T cells indicated independency of these pathways from MALT1 proteolytic function. These results show that the proteolytic function of MALT1 is not essential for acute IKK, JNK, and ERK signaling. Furthermore, they suggest that catalytically inactive MALT1 is able to scaffold properly. Supporting this, PMA/ionomycin stimulation induced high m.w. MALT1 species (Fig. 1A) consistent with monoubiquitinated MALT1, a recently identified modification of MALT1 that results from scaffold-induced MALT1 processing by a mechanism that remains to be elucidated (22).

Similarly to the observations in T lymphocytes, PMA/ionomycin-stimulated purified B cells displayed normal acute phosphorylation of IkBα and JNK in the absence of MALT1 proteolytic function, whereas the same process was suppressed in the absence of MALT1 protein (Fig. 1B). Phosphorylation of ERK was also confirmed to occur independently of MALT1 as both Malt1PD/PD and Malt1−/− B cells displayed normal ERK phosphorylation in response to PMA/ionomycin (Fig. 1B). As observed in T cells, increased levels of putative monoubiquitinated MALT1 species were observed upon stimulation of Malt1PD/PD B cells, supporting an intact scaffolding ability of catalytically inactive MALT1. Cleavage of RelB, Regnase-1, and CYLD was also abolished in Malt1PD/PD B cells (Fig. 1B).

Collectively, these biochemical data provide evidence for normal expression of the catalytically inactive MALT1 C472A mutant protein in Malt1PD/PD mice and show that activation-induced IKK, ERK, and JNK activation can occur independently of the proteolytic function of MALT1.

Malt1PD/PD mice develop a spontaneous multiorgan inflammatory pathology

Similarly to other Malt1−/− mouse lines (8, 9), the Malt1−/− mice described in this article were viable and displayed normal behavior and life span. Unexpectedly, at the age of 9–10 wk, Malt1PD/PD animals started developing pathological symptoms, including weakness, hunched posture, eye inflammation, and hind-limb paralysis, ultimately leading to weight loss and death. The incidence of the pathology was more apparent in female Malt1PD/PD mice (Supplemental Fig. 1C). Extensive macroscopic and histopathology analyses, performed on 8- to 12-wk-old Malt1PD/PD mice, revealed minimal-to-moderate lymphoid cell infiltration in a variety of organs, mainly glandular stomach, lacrimal, salivary, and Harderian glands, lungs, and peripheral nerves (Fig. 2A–L, Supplemental Fig. 2, and data not shown). Affected nerves included the sciatic and spinal nerves, as well as small peripheral nerve fibers such as the pharyngeal nerve (Supplemental Fig. 2 and data not shown). No alterations were observed in brain, spinal cord, or optic nerve (data not shown). Minimal-to-moderate lymphoid hyperplasia and plasma cell hyperplasia were evident in lymph nodes (LN), particularly mandibular LN and mesLN (Fig. 2F). Lymphoid hyperplasia was also present in the GALT of the small intestine (data not shown). A predominance of B cells over T cells was observed in the glandular stomach, salivary and lacrimal glands, and the GALT. Lymphoid infiltrates in the lungs, the Harderian glands, and the small nerve fibers consisted mainly of T cells. Ex vivo analysis of thymus, spleen, pooled cervical and mandibular LN (cerLN), mesLN, inguinal LN, PP, and peritoneal exudate cells (PEL) confirmed the increased cellularity of mesLN, cerLN, and PP in Malt1PD/PD mice and revealed normal splenic, thymic, and inguinal LN size (Fig. 2M). Malt1−/− mice also exhibited a significant increase in mesLN and cerLN cellularity, but the extent was less pronounced when compared with Malt1PD/PD mice.

Analysis of serum Ig levels in Malt1−/− mice confirmed the previously reported reduction in serum IgM and IgG, and revealed reduced IgE levels (Fig. 2N) (8, 9). Malt1PD/PD mice had reduced IgM, IgG2a, IgG2b, and IgG3 levels, but in contrast displayed a significant elevation in both serum IgG1 and IgE. Serum and intestinal IgA levels were not altered in Malt1PD/PD mice.

Thus, the selective abrogation of the proteolytic function of MALT1 resulted in a progressive inflammatory pathology associated with partial lymphadenopathy, multiorgan infiltration by T and B cells, and increased IgG1 and IgE levels.

Malt1PD/PD mice display partial impairment of the B cell compartment and follicular T cells and abrogated B cell responses to active Ag immunization

Given the high prevalence of B cells in affected tissues of Malt1PD/PD mice and the increased IgG1 and IgE levels, we characterized in more detail the composition and function of the B cell compartment in these mice.

In line with previous reports, Malt1−/− mice displayed a normal development of splenic follicular (FO) B cells and a severe reduction of marginal zone (MZ) B cells and peritoneal B1 B cells (8, 9) (Fig. 3A and 3B). Similarly, peritoneal B1 B cells were absent in Malt1PD/PD mice, indicating that proteolytic function of
MALT1 was essential for the development and/or survival of this innate B cell population. Although splenic FO B cells were not affected, a reduced, but detectable MZ B cell population was present in Malt1<sup>PD/PD</sup> mice (Fig. 3B, Supplemental Fig. 3A and 3B). Further characterization of germinal center (GC) B cells revealed an almost complete absence of GC B cells in all lymphoid organs of Malt1<sup>−/−</sup> mice, including PP and mesLN (Fig. 3C, Supplemental Fig. 3C and 3D, and data not shown). The absence of GC B cells correlated with a severe reduction in CD4<sup>+</sup>CXCR5<sup>+</sup>PD1<sup>+</sup> follicular Th (T<sub>FH</sub>) cells (Fig. 3D). By contrast, Malt1<sup>PD/PD</sup> mice retained normal or partially reduced proportions of GC B cells in PP, mesLN, and cerLN (Fig. 3C, Supplemental Fig. 3C and 3D). In line with the increased serum IgG1 levels, GCs in Malt1<sup>PD/PD</sup>
mice contained increased proportions of IgG1+ B cells (Fig. 3C, Supplemental Fig. 3D). Similarly, to the GC B cell population, the TFH compartment was only partially reduced in Malt1PD/PD mice with a selective abrogation of CXCR5/PD1high TFH cells, but detectable levels of CXCR5/PD1dim TFH cells (Fig. 3D). Thus, although the combined absence of the scaffolding and enzymatic functions of MALT1 severely abrogated the generation of innate B1, MZ B cells, TFH, and GC B cells, the selective lack of MALT1 enzymatic activity disrupted the B1 B cell compartment, but only partially affected the MZ, GC B cells, and TFH cells.

To better understand the relevance of MALT1 proteolytic activity on B cell function, we assessed in vitro proliferative responses of purified splenic B cells in response to major triggers such as BCR, CD40, and TLR agonists. After IgM, CD40L, or CD40 triggering, proliferation of Malt1+/− B cells was significantly reduced (Fig. 3E). For anti-IgM and anti-CD40 stimulation, the defects were partially rectified by addition of exogenous IL-4. Malt1PD/PD B cells also showed defective proliferation in response to activation through TLR4 and CD40, but not by anti-IgM (Fig. 3E). Similarly to Malt1+/− B cells, addition of IL-4 restored the proliferation of Malt1PD/PD B cells in response to CD40 stimulation. TLR9-induced proliferation of B cells from Malt1+/− and Malt1PD/PD was normal.

Although Malt1PD/PD mice had a detectable MZ B cell population, ex vivo restimulation of MZ B cells with a TLR9 agonist revealed that the Malt1PD/PD MZ B cells did not produce IL-10 and secreted lower levels of IL-6 (Fig. 3F). Similar defects in IL-10 production, but normal or enhanced production of TNF-α was observed after ex vivo stimulation of total purified Malt1PD/PD splenic B cells (Fig. 3G), suggesting that certain immunoregulatory functions of MZ-like B cells are also abrogated in the absence of the proteolytic activity of MALT1.

The multiple defects characterizing the TFH and B cell compartments of Malt1+/− mice have been associated with defective B cell responses to TD and TI-2 Ags (8, 9), a finding we could confirm (Supplemental Fig. 3E). Malt1PD/PD mice were also unable to mount IgM and IgG responses when immunized with the TI-2 Ag DNP-KLH (Fig. 3H). Interestingly, Malt1PD/PD mice also displayed abrogated Ab responses to the TD Ag DNP-KLH conjugate. The reduced signaling through CD40 combined with reduced TFH cell proportions likely contributed to the defective TD B cell responses observed in Malt1PD/PD mice.
Catalytic inactivation of MALT1 increases the frequency of effector T cells and alters the cytokine expression profile

In view of the partial impact of the MALT1 C472A mutation on Tfh differentiation, we next analyzed the composition and function of other T cell subsets present in lymphoid organs. Malt1PD/PD mice displayed a partial reduction in the proportion of CD4+ and CD8+ T cells (Supplemental Fig. 3F), mainly in LN displaying increased cellularity (i.e., mesLN and cerLN). Of note, proportions of effector memory (TEM) and central memory (TCM) CD4+ and CD8+ T cells in LN and spleen of Malt1PD/PD mice were increased (Fig. 4A and data not shown). A similar increase in CD44 levels and the TCM subtype was also apparent in Malt1−/− mice.

The functional ex vivo characterization of splenic T cells indicated that Malt1−/− T cells displayed severely impaired proliferative responses after activation by CD3 and CD28 Abs (Fig. 4B), confirming previous findings (8, 9). By contrast, [3H]thymidine incorporation by Malt1PD/PD T cells was reduced compared with WT cells, but not to the extent observed for the NF-κB detected in PMA/ionomycin-activated T cells. Previous work using T cell lines reconstituted with an enzymatically inactive form of MALT1 suggested that the proteolytic function of MALT1 was important for TCR-triggered IL-2 production, a major survival factor required for in vitro T cell proliferation (11). Consistent with this, we observed that Malt1PD/PD CD4+ and CD8+ T cells produced lower amounts of IL-2 and TNF-α after ex vivo restimulation with PMA/ionomycin (Fig. 4C and 4D). Once again, defects of Malt1−/− T cells were more profound than those of Malt1PD/PD T cells.

Optimal production of IL-2 and TNF-α relies on the activity of the NFκB member c-Rel (23, 24). MALT1 substrates such as RelB and Regnase-1 can impact the activity and expression of c-Rel and IL-2 either by modulating c-Rel localization or by promoting c-Rel and IL-2 mRNA degradation, respectively (16, 17). Consistent with the accumulation of uncleaved RelB and Regnase-1 detected in PMA/ionomycin-activated Malt1PD/PD and Malt1−/− T cells (Fig. 1A), Malt1PD/PD and Malt1−/− T cells failed to up-regulate the mRNAs encoding for c-Rel and IL-2 upon activation (Fig. 4E).

Collectively, the accumulation of uncleaved MALT1 substrates in Malt1PD/PD T cells was sufficient to indirectly impact expression of certain activation-induced genes resulting in defective production of cytokines such as IL-2 and TNF-α and leading to partially impaired proliferative responses in vitro. The normal activation of the MALT1 scaffolding function-dependent IKK axis combined with the defective modulation of c-Rel activity and Regnase-1-regulated genes might lead to an impaired immune homeostasis.

Functional defects in Malt1PD/PD T cells lead to EAE resistance and skewed Th1 and Th2 responses in vivo

The enhanced cerLN and mesLN size and accumulation of memory T cells in lymphoid organs of Malt1PD/PD mice indicated that T cell activation and expansion in vivo occurred irrespective of the defective IL-2 and TNF-α production identified in vitro. Interestingly, the cytokine expression pattern of spleen cells stimulated with PMA/ionomycin ex vivo revealed higher amounts of the Th1 cytokine IFN-γ and strongly elevated Th2 cytokines such as IL-4 and IL-13 in Malt1PD/PD splenocytes (Fig. 4D). These data suggested that the immune defects associated with selective deficiency of MALT1 proteolytic activity were associated with the expansion of both Th1- and Th2-skewed T cells in vivo.

In view of the Th1- and Th2-skewed T cell responses observed in Malt1PD/PD mice and the previously reported defective differentiation of Malt1−/− T cells into Th17 cells (25), we investigated the Th1 and Th17 differentiation potential of Malt1PD/PD T cells.

In vitro skewing of Malt1PD/PD and Malt1−/− CD4+ T cells toward Th1 cells was normal (Fig. 5A). By contrast, Malt1PD/PD and Malt1−/− CD4+ T cells displayed a severe reduction in IL-17 production after differentiation into Th17 cells (Fig. 5A). With the aim to further elucidate the relevance of defective Th17 differentiation in vivo, we analyzed Malt1PD/PD mice in a Th17-dependent EAE model. Consistent with our in vitro findings, Malt1PD/PD mice were completely protected from EAE development induced by immunization with MOG1-125 in adjuvant (Fig. 5B).

Overall, the lack of MALT1 protease activity in T cells altered the activation process and cytokine expression profile of T cells induced upon stimulation. Although IL-2, TNF-α, and IL-17 production were reduced or impaired, secretion of IFN-γ and Th2 cytokines was not affected. The IL-4– and IL-13–rich environment identified in lymphoid organs of Malt1PD/PD mice also suggested that Th2-skewed responses promoted the IgG1 and IgE B cell responses observed in these mice.

Reconstitution of Malt1PD/PD mice with WT Tregs overcomes a partial Treg deficiency and delays the multiorgan inflammatory pathology

The spontaneous and progressive pathology characterizing Malt1PD/PD mice shared many similarities with the disease occurring in certain mouse strains characterized by an impaired Treg compartment. The CBM complex is critical for the development and survival of natural Treg cells, and Malt1−/− mice were previously reported to display a severe impairment in peripheral Tregs (26–28). Whereas Malt1−/− mice displayed the expected severe reduction in Foxp3+ Tregs (Fig. 6A), Malt1PD/PD mice were characterized by an ~60% decrease in Foxp3+ cells in the CD4+ T cell compartment compared with WT littermate controls in all lymphoid organs tested (Fig. 6A). In vitro Treg differentiation experiments performed in the presence of exogenous IL-2 indicated that Malt1PD/PD CD4 T cells have the potential to differentiate into Tregs (Fig. 6B).

Considering the reduced Treg proportions and the functional alterations of Malt1PD/PD T cells and B cells, we hypothesized that the spontaneous multiorgan pathology occurring in Malt1PD/PD mice developed as a result of improper Treg-mediated suppression. To test this, we adoptively transferred WT Tregs isolated from EGFP-Foxp3 animals into Malt1PD/PD mice. The transfer of 0.5 × 10^6 WT Tregs into female Malt1PD/PD mice delayed the weight loss and the lethal phenotype (Fig. 6C). By contrast, Malt1PD/PD mice receiving PBS or WT EGFP−/−CD4+Foxp3 cells displayed the typical symptoms and started to succumb to the immune pathology.

A major role of the defective Treg compartment in the development of the spontaneous pathology was also supported by additional observations made in BM chimeric mice in which sublethally irradiated CD45.1 WT recipients were reconstituted with CD45.2 BM cells from WT or Malt1PD/PD mice. Interestingly, the recipient mice receiving Malt1PD/PD BM cells were characterized by similar defects in immune compartments to those observed in Malt1PD/PD mice (Supplemental Fig. 4A–D). However, none of the Malt1PD/PD BM chimeras developed any of the symptoms of pathology occurring in constitutive Malt1PD/PD mice. All animals did not develop any sign of hypercellularity, multiorgan T and B cell infiltration, TEm and TCM cell expansion, or increased IgG1 and IgE levels for up to 38 wk after reconstitution (Supplemental Fig. 4E–H and data not shown). As expected, the majority of the CD4+ and CD8+ T cell, B cell, and myeloid compartments were derived from the BM donor cells (Supplemental Fig. 4E–G). Remarkably, WT Tregs derived from
Dysfunctional responses of Malt1PD/PD and Malt1−/− T cells including reduced IL-2 and TNF-α. (A) Representative staining and proportions of naive, CD44highCD62Llow TEM, and CD44highCD62Lhigh TCM subset distribution among CD8+ and CD4+ T cell populations present in cerLN. Bars represent mean ± SEM (n = 5 mice/group). Data are representative of three independent experiments. (B) Proliferation of purified Malt1PD/PD and Malt1−/− CD3+ T cells stimulated with different concentrations of plate-bound CD3 and CD28 Abs. Data are representative of two independent experiments. (C) Intracellular cytokine staining of LN CD4+ and CD8+ T cells stimulated ex vivo for 4 h with PMA/ionomycin. (D) Cytokine expression pattern of spleen cells stimulated ex vivo for 4 h with PMA/ionomycin. Bars represent mean ± SEM of data obtained on spleen cells from three independent mice per group and a representative of two independent experiments. (E) Modulation of c-Rel and IL-2 mRNA levels in WT, Malt1−/−, and Malt1PD/PD purified splenic CD4+ T cells. mRNA levels were evaluated by Q-PCR at different times after PMA/ionomycin stimulation. The mRNA expression level was normalized to the expression level of β2-microglobulin, and fold changes were calculated using the ΔΔ cycle threshold method. Bars indicate average ± SD. Data are representative of at least two independent experiments. Statistical significance was calculated using a two-tailed unpaired t test (*p < 0.05, **p < 0.01, ***p < 0.001).
the CD45.1+ recipient accounted for ~80% of the Treg repertoire of the chimeras reconstituted with the Malt1PD/PD BM cells compared with ~20% of the Treg compartment in chimeras receiving WT BM cells (Supplemental Fig. 4I). These data highlighted that a fraction of the recipient’s Treg compartment survived the sublethal irradiation process and probably had a survival and/or homeostatic proliferation advantage over the Tregs derived from the Malt1PD/PD hematopoietic precursors. In line with the Treg adoptive transfer data, the presence of WT Treg likely prevented disease development despite the immune defects characterizing the Malt1PD/PD hematopoietic compartment.

Therefore, these data strongly suggest that the immune defects and partial Treg deficiency characterizing Malt1PD/PD mice, combined with the capacity to mount certain types of T cell and B cell immune responses, contribute to the development of the multiorgan inflammatory pathology observed.

Discussion

In this study, we investigated the relative contributions of the protease versus scaffolding functions of the paracaspase MALT1 in the regulation of T and B cell immune function in vivo by characterizing novel MALT1 PD mice (Malt1PD/PD) and comparing them with Malt1−/− mice. Our findings indicated that the selective lack of MALT1 protease activity prevented the formation of a functional peritoneal B1 B cell compartment and led to defective CD40 and TLR4 signaling in B cells, defective TLR9-induced IL10 production by MZ B cells, and defective Th17 differentiation. In addition, Malt1PD/PD mice were unable to mount B cell responses to TD and TI-2 Ags and were protected in a Th17-dependent EAE model similarly to Malt1−/− mice. By contrast, MALT1 proteolytic activity only partially affected other immune cell subsets and functions. This was highlighted first by the development of MZ B cells, which was relatively undisturbed in...
Malt1PD/PD but defective in Malt1−/− mice. Second, TFH and GC B cell compartments were only moderately diminished in Malt1PD/PD animals, but completely abrogated in Malt1−/− mice. Third, Malt1PD/PD T cells displayed a milder defect in proliferative responses and in IL-2 and TNF-α production compared with Malt1−/− T cells. These defects were associated with subtle alterations in NF-κB activation observed in Malt1PD/PD T cells in contrast with the severe abrogation detected in Malt1−/− T cells. Finally, natural Treg proportions in the CD4+ T cell compartment were reduced by ~60% in Malt1PD/PD mice, but were almost completely absent in Malt1−/− mice. Unexpectedly, we found that the immune alterations associated with the selective absence of MALT1 protease function resulted in a lethal inflammatory pathology associated with accumulation of TEM and TCM CD4+ and CD8+ T cells in lymphoid organs, Th1- and Th2-skewed responses, enhanced IgG1 and IgE levels, and lymphocytic infiltrations in multiple organs.

The pathology observed in the Malt1PD/PD mouse is reminiscent of the phenotype associated with partial or complete reduction in Treg numbers due to defective IL-2 signaling or Foxp3 deficiency (29–34). However, the pathology of Malt1PD/PD mice was associated with a slower onset, mandibular- and mesLN-restricted lymphadenopathy, and lymphocytic infiltration in a more limited set of organs. Because TCR- and CD28-mediated signals, as well as IL-2 levels, are important for natural Treg development and peripheral survival, it is likely that the alteration of CBM-driven signaling events and the incomplete abrogation of IL-2 production as IL-2 levels, are important for natural Treg development and associated with a slower onset, mandibular- and mesLN-restricted lymphocyte compartments were only moderately diminished in Malt1PD/PD mice. In line with a Treg development defect, we observed a reduction in the Foxp3+ Treg proportions also among Malt1PD/PD CD4 single-positive thymocytes (data not shown).

The potential association between an imbalanced Treg compartment and the inflammatory pathology in Malt1PD/PD mice was supported by the delay in symptoms of disease after reconstitution of Malt1PD/PD mice with WT Tregs. Although the Treg transfer was sufficient to delay weight loss and the appearance of symptoms over the observation time, histopathology analyses performed on the various animals at the time of sacrifice revealed no or only limited reduction of the inflammatory infiltrates and cerLN cellularity (data not shown). We believe that the reason for limited histopathological amelioration is related to the pre-existing asymptomatic pathology already present in the mice at time of the Treg transfer. The different ages at which the animals were analyzed (i.e., most of the Treg transferred mice were analyzed at later time points than the other groups) may account as well. Altogether, these conditions may limit the overall effect of the Treg transfer. However, additional experiments involving BM chimeras bearing a Malt1PD/PD hematopoietic system suggested that WT Tregs surviving the sublethal irradiation of recipient animals likely prevented disease development despite the immune defects in the Malt1PD/PD hematopoietic compartment. In line with this, radiation-resistant Tregs were reported previously to prevent T cell–mediated autoimmunity after syngeneic BM transplantation (35). Detailed histopathology analyses revealed only limited effects on nonimmune cells, but we cannot fully exclude that additional alterations outside the hematopoietic compartment also contribute to disease development in Malt1PD/PD mice. More specific analyses in affected peripheral tissues are required to investigate this question.

In contrast with Malt1PD/PD mice, Malt1−/− mice displayed an almost complete absence of peripheral Tregs, yet did not develop any overt sign of immune pathology. Of note, we did observe mild lymphadenopathy of mesLN and cerLN and an increased proportion of TCM-like cells in the lymphoid organs of Malt1−/− mice, as well as moderate lymphoid cell infiltration in lacrimal and salivary glands (data not shown). In line with the critical role of CBM-driven signals and IL-2 in promoting Treg development and survival, Malt1−/− T cells displayed a more severe abrogation of CBM-mediated NF-κB activation resulting in strongly impaired IL-2 and TNF-α production. Therefore, although abrogation of the Treg compartment in Malt1−/− animals potentially sensitized them to develop a similar pathology as Malt1PD/PD mice, the more severe functional defects in the T and B cell compartments likely limited the expansion and activation of pathogenic effector cells and restrained the extent and severity of the disease. In contrast, we believe that the pathology occurring in Malt1PD/PD mice was facilitated by the combination of impaired immunosuppressive compartments (e.g., Treg and IL-10–producing B cells) and partially dysfunctional T and B cells, which retained the capacity to generate Th1- and Th2-skewed effectors in response to yet unidentified Ags (Fig. 6D). Interestingly, analyses involving FACS-based intracellular cytokine staining aiming at assessing the identity of IL-4− and IFN-γ− producing cells suggest that the main sources of these cytokines among Malt1PD/PD T cells are CD4high TEM and TCM CD8+ cells of a Th1-like phenotype (IFN-γ−IL4−) and CD4high TEM and TCM CD4+ cells including a mix of Th0 (IFN-γIL4+), Th1 (IFN-γIL4−), and Th2 (IFN-γIL4+) phenotypes (data not shown). Given the accumulation of these multiple T cell subsets, additional work will be required to assess how these different subsets contribute to the complex multilogan pathology occurring in Malt1PD/PD mice and which Ags drive the expansion of these different lymphocyte populations. In addition, given the central defect in thymic Treg development observed in Malt1PD/PD mice, it remains to be assessed whether alterations in the thymic selection process associated with defective MALT1 proteolytic activity may also promote pathology development by releasing autoreactive T cell clones (normally deleted in WT animals) in the peripheral T cell effector pool.

The accumulation of IL-4− and IL-13−producing Th0/Th2 cell subsets in Malt1PD/PD mice was also highlighted by increased CD23 (FceRII) levels on FO and MZ B cells (Fig. 3B, Supplementary Fig. 3B). Indeed, IL-4−, IL-13, and IgE were previously reported to upregulate CD23 expression (discussed by Dierks et al. [36]). The Th2-skewing observed in Malt1PD/PD mice may also explain the enhanced IgG1 and IgE levels observed in these animals. However, it was intriguing to observe that Malt1PD/PD mice could not mount Ab responses to TD and TI-2 Ags upon immunization. Further investigation is required to assess whether endogenous GC B cell responses and IgG1 and IgE secretion rely on help provided by the CXCXR5(PD)dim T FH cells remaining in Malt1PD/PD mice, or whether alternative T cell–independent pathways also contribute to this process (37). Innate B1 and MZ B cells are key mediators of TI-2 B cell responses (38). Because DAGG-Ficoll was administered in the absence of additional adjuvants, we believe that the defective anti-DNP IgM and IgG responses observed in Malt1PD/PD mice were a consequence of the absent B1 B cells and reduced/dysfunctional MZ B cell compartment observed in these mice. It is therefore likely that the GC B cells retained in mesLN and PP of Malt1PD/PD mice are directed to other types of Ags present at these sites and triggering alternative B cell costimulatory pathways independent of MALT1 protease activity such as TLR7 or TLR9 agonists associated with, for example, Ti-1 Ags present in the intestinal microflora.

The selective effect on cytokines such as IL-2, TNF-α, and IL-17, and the expansion of Th1-like IFN-γ+ T cells, as well as Th0/Th2 cytokine-producing cells, observed in Malt1PD/PD mice sug-
gests that MALT1 protease function modulates selected transcriptional programs downstream of the TCR. Our analysis of the signaling events downstream of the CBM complex revealed minor changes in IkBa, p65, or p105 phosphorylation and IkBa degradation in both Malt1<sup>PD/PD</sup> T cells and B cells. This suggests that MALT1 protease activity modulates signaling events downstream of the CBM complex without completely abrogating canonical NF-κB signaling (via the IKK-IκB axis). It is interesting to observe that the alterations in the T and B cell compartments identified in Malt1<sup>PD/PD</sup> mice closely resemble those reported in c-Rel–deficient animals (23, 39–42). Moreover, c-Rel was also shown to regulate IL-21 production and T<sub>FH</sub> cell differentiation (43). These similarities are consistent with recent reports showing that cleavage of ReIB and Regnase-1 by MALT1 can influence c-Rel levels and activity (16, 17). Our biochemical analyses could demonstrate the accumulation of uncleaved ReIB and Regnase-1 in Malt1<sup>PD/PD</sup> T cells and B cells. Consistent with this, analysis of the levels of several Regnase-1–regulated mRNA in T cells clearly indicated that c-Rel as well as IL-2 mRNA induction was abrogated in the absence of MALT1 proteolytic activity. These data strongly indicate that reduced c-Rel–mediated transcription contributed to the functional defects identified in Malt1<sup>PD/PD</sup> T cells. However, the lack of overt inflammatory pathology in c-Rel–deficient mice indicates that additional defects contribute to the pathology characterizing Malt1<sup>PD/PD</sup> animals.

In conclusion, this work revealed a previously unappreciated role for the MALT1 protease activity in finely regulating immune function and homeostasis. The defective Th17 and B cell responses observed in Malt1<sup>PD/PD</sup> mice highlight the therapeutic potential of drugs targeting MALT1 protease function. In line with this, treatment with meprazine, which inhibits MALT1 protease activity in a micromolar range, was recently reported to attenuate EAE symptoms (44). However, the reduced Treg compartment and multiorgan pathology occurring in Malt1<sup>PD/PD</sup> mice provide a warning for the potential risks associated with sustained abrogation of MALT1’s enzymatic activity. Development of inducible PD mice or selective MALT1 inhibitors will help clarify whether chronic inhibition of MALT1 activity in autoimmune pathologies is of therapeutic value. The spontaneous pathology and the complexity of the immune alterations identified in this study open new exciting questions that deserve further investigation. In particular, it is still unclear which Ags and pathways drive the accumulation of T<sub>EM</sub> and T<sub>CM</sub> cells and the IgG1/IgE-skewed B cell responses in Malt1<sup>PD/PD</sup> mice, and what is their contribution to the overall pathology. More importantly, what is the relevance of MALT1 protease activity in humans? Polymorphisms in the Malt1 gene have been associated with combined immunodeficiency (45–47). However, in all cases the mutations resulted in a complete deficiency of the protein or very low MALT1 levels because of protein instability. Further work is required to assess whether specific mutations in the catalytic function of MALT1 contribute to multiorgan inflammatory disorders.

Note added in proof. During the revision of this manuscript, two studies reporting the generation and characterization of novel Malt1<sup>PD/PD</sup> mouse lines were published (48, 49). Both articles described Malt1<sup>PD/PD</sup> mice characterized by cerLN and mesLN lymphadenopathy and very similar immune defects to the ones we describe in our manuscript. Jaworski et al. (48) claim that Malt1<sup>PD/PD</sup> mice succumb to autoimmune gastritis and provide evidence that the disease can be prevented by the adoptive transfer of WT Tregs into 3– to 8-d-old Malt1<sup>PD/PD</sup> mice. Giewes et al. (49) describe an inflammatory pathology characterized by lymphocytic infiltrates in the glandular stomach but also a neurological disorder associated with dystonia and progressive ataxia that was prevented by a deficiency in Bcl110, T and B cell compartments (Rag1KO mice), or IFN-γ production.

Acknowledgments

We thank Patricia Brande Weber for the initial histopathological evaluations as well as Danielle Anastasi, Maureen Bardet, Karine Bigot, Vanessa Cornacchione, Elena Degl’Innocenti, Nicolas Dubois, Esther Erard, Yvonne Hager, Catherine Huck, Bernhard Jest, David Ledieu, Daniel Kaiser, Sonja Kerstin Frenzel, Bernadette Kleylein-Sohn, Claire Malinverni, Barbara Metzler, Gautier Robert, Regina Stanton, Philippe Scheubel, Patrick Schmutz, Pamela Ramsay, Roland Sollberger, Marie-Catherine Stutz, Adeline Unterreiner, and Catherine Wieland for excellent technical assistance and additional contributions. We also thank Stephan Gadola, Chauncey Spooner, and Michael W. Tuschl for comments and discussions.

Disclosures


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