Functional Analysis of a Complement Polymorphism (rs17611) Associated with Rheumatoid Arthritis

Joanna L. Giles, Ernest Choy, Carmen van den Berg, B. Paul Morgan and Claire L. Harris

*J Immunol* published online 27 February 2015
http://www.jimmunol.org/content/early/2015/02/27/jimmunol.1402956

Supplementary Material
http://www.jimmunol.org/content/suppl/2015/02/27/jimmunol.1402956.6.DCSupplemental

Subscription
Information about subscribing to *The Journal of Immunology* is online at:
http://jimmunol.org/subscription

Permissions
Submit copyright permission requests at:
http://www.aai.org/About/Publications/JI/copyright.html

Email Alerts
Receive free email-alerts when new articles cite this article. Sign up at:
http://jimmunol.org/alerts

*The Journal of Immunology* is published twice each month by
The American Association of Immunologists, Inc.,
1451 Rockville Pike, Suite 650, Rockville, MD 20852
Copyright © 2015 The Authors All rights reserved.
Print ISSN: 0022-1767 Online ISSN: 1550-6606.
Functional Analysis of a Complement Polymorphism (rs17611) Associated with Rheumatoid Arthritis

Joanna L. Giles,* Ernest Choy,† Carmen van den Berg,‡ B. Paul Morgan,*1 and Claire L. Harris*1

Complement is implicated in the pathogenesis of rheumatoid arthritis (RA); elevated levels of complement activation products have been measured in plasma, synovial fluid, and synovial tissues of patients. Complement polymorphisms are associated with RA in genome-wide association studies. Coding-region polymorphisms may directly impact protein activity; indeed, we have shown that complement polymorphisms affecting a single amino acid change cause subtle changes in individual component function that in combination have dramatic effects on complement activity and disease risk. In this study, we explore the functional consequences of a single nucleotide polymorphism (SNP) (rs17611) encoding a V802I polymorphism in C5 and propose a mechanism for its link to RA pathology. Plasma levels of C5, C5a, and terminal complement complex were measured in healthy and RA donors and correlated to rs17611 polymorphic status. Impact of the SNP on C5 functionality was assessed. Plasma C5a levels were significantly increased and C5 levels significantly lower with higher copy number of the RA risk allele for rs17611, suggesting increased turnover of C5 V802I. Functional assays using purified C5 variants revealed no significant differences in lytic activity, suggesting that increased C5-V802 turnover was not mediated by complement convertase enzymes. C5 is also cleaved in vivo by proteases; the C5 V802 variant was more sensitive to cleavage with elastase and the “C5a” generated was biologically active. We hypothesize that this SNP in C5 alters the rate at which elastase generates active C5a in rheumatoid joints, hence recruiting neutrophils to the site thus maintaining a state of inflammation in arthritic joints. The Journal of Immunology, 2015, 194: 000–000.

Received for publication November 26, 2014. Accepted for publication January 30, 2015.

This work was supported by Arthritis Research U.K. Reference 19436. The Cardiff Regional Experimental Arthritis Treatment and Evaluation Centre, Section of Rheumatology, School of Medicine, Cardiff University, Cardiff CF14 4XN, United Kingdom; and the National Institute of Social Care and Health Research.

Address correspondence and reprint requests to Dr. B. Paul Morgan, Institute of Infection and Immunity, School of Medicine, Cardiff University, Cardiff CF14 4XN, United Kingdom; E-mail address: MorganBP@cf.ac.uk

The online version of this article contains supplemental material.

Abbreviations used in this article: CFD, complement fixation diluent; CFE, cobra venom factor; HBS, HEPES buffered saline; HNE, human neutrophil elastase; NHS, normal human serum; RA, rheumatoid arthritis; ShEA, sheep erythrocyte; SNP, single nucleotide polymorphism; TCC, terminal complement complex; TRAF, TNFR-associated factor.

This is an open-access article distributed under the terms of the CC-BY 3.0 Unported license.

Copyright © 2015 The Authors 0022-1767/15

www.jimmunol.org/cgi/doi/10.4049/jimmunol.1402956
cleavage by human neutrophil elastase (HNE), generating an active fragment functionally indistinguishable from C5a. We suggest that increased susceptibility to cleavage by HNE, with release of proinflammatory fragments, explains the association of the polymorphism with RA and other inflammatory diseases in which elastase levels are locally elevated.

Materials and Methods

DNA preparation and genotyping

Blood was collected into EDTA from healthy donors and from RA and osteoarthritis patients satisfying American College of Rheumatology/European League Against Rheumatism classification criteria (25). An aliquot was used for gDNA preparation and plasma harvested from the residual stored at −80°C. gDNA was purified using a RELIAPrep gDNA kit (Promega, Southampton, U.K.). Following manufacturer’s instructions, gDNA concentration was determined using a Nanodrop 1000 spectrophotometer (Thermofisher, Loughborough, U.K.). All samples were normalized to 10 ng/ml and genotyped at the rs17611 SNP using KASP technology (Kbiosciences, Teddington, U.K.).

Measurement of complement components

C5, C5a, C3a, and the terminal complement complex (TCC) were measured in EDTA plasma from genotyped donors. C5a and C3a were measured using commercial ELISA kits (Microvieve; Quidel, San Diego, CA), according to the manufacturer’s instructions. TCC levels were measured in an in-house ELISA. Briefly, 96-well Maxisorp plates (Thermofisher) were coated with anti-C9 Ab (aE11, Hycult, The Netherlands) at 2 μg/ml in carbonate bicarbonate buffer (pH 9.6); plates were blocked in 2% BSA and then incubated with plasma samples. Detection was with HRP-conjugated anti-C9 mAb (clone E2) in 2% BSA and color developed using orthophenylenediamine (Sigma-Aldrich, Dorset, U.K.). Plasma C5 levels were measured using a novel in-house C5 ELISA. Briefly, 96-well Maxisorp plates were coated with an in-house polyclonal rabbit anti-human C5 Ab (8 μg/ml IgG in bicarbonate buffer [pH 9.6]), blocked in 2% BSA, and then incubated with plasma samples diluted 1 in 600 in 2% BSA. Bound C5 was detected using a mouse monoclonal anti-human C5 (MBI-C5-3; developed in-house, 5 μg/ml), followed by donkey anti-mouse IgG HRP (Jackson Labs, Stratach, Newmarket, U.K.); 1 in 2500 and color developed using orthophenylendiamine. Plasma levels of complement components were calculated using Graphpad prism 3 (GraphPad, La Jolla, CA), and statistical analysis was performed using one-way ANOVA. For C5, p = 0.0047; for C5a, p < 0.0001; and for TCC, p = 0.6032.

C5 purification

Donors homozygous for the C5V802I polymorphic variants were identified and EDTA plasma collected. C5 was purified by affinity chromatography on monoclonal anti-human C5 (MBI-C5-3; 20 mg) immobilized to a 5 ml 4-12% SDS gradient gel (Invitrogen), stained with Coomassie blue. C5a release was also measured using a C5a ELISA kit as described previously (29). Each C5 variant (30 μg) was incubated with 8 μl of the preformed convertase at 37°C. Aliquots (5 μg protein in a 5-μl volume) were removed from the mix at intervals to separate the C5 convertase from the TCC. After 15 min at 37°C, absorbance at 415 nm was measured as described above.

Cobra venom factor assays

Cobra venom factor (CVF) was purified from cobra venom (Russa naja kaouthia) venom (Sigma-Aldrich). Briefly, 200 μg lyophilized venom was solubilized in 10 ml TBS (pH 8.5). Insoluble material was removed by centrifugation, and filtrated supernatant was applied to a DEAE sepharose column at 4°C on an Akta Purifier. CVF was eluted using a linear gradient to 1 M NaCl in TBS. Fractions were screened on 7.5% SDS-PAGE gels. Fractions corresponding to CVF were dialyzed into PBS (10 mM phosphate and 150 mM NaCl [pH 7.5]) and applied to a SD200 gel filtration column in PBS. C5 convertase (CVFBB) was prepared by incubating 10 μg CVF, 10 μg factor B, 0.12 μg factor D, and 5 mM MgCl2 in 50 μl TBS for 1 h at 37°C (as described previously (29)). Each C5 variant (30 μg) was incubated with 8 μl of the preformed convertase at 37°C. Aliquots (5 μg protein in a 5-μl volume) were collected at intervals (100% control)/(A690 0% control)/(A690 100% control) and rate of lysis plotted as described above.

Calcium Flux assays

U937 cells, stably transfected with the C5aRI or IL-8RB (a gift from E. Prossnitz, Albuquerque, NM) at 105 cells/ml were loaded with 2 μM Fura-2-AM at 30 min at room temperature. Cells were washed and resuspended in Krebs/HEPES/BESA buffer (25 mM HEPES, 120 mM NaCl, 4.8 mM KCl, 1.2 mM KH2PO4, 1.2 mM MgSO4, 1.3 mM CaCl2 [pH 7.4], and 0.1% BSA). Cells (200 μl at 5 × 106/ml) were stimulated with C5a, C5, or C5 preincubated with HNE (2 h at 37°C as described above). Changes in intracellular calcium were measured as described previously (30, 31). To confirm that the observed signal was C5a induced, a C5aRI antagonist peptide (H-Phe-Lys-Pro-D-Che-Arg-OH) (100 nM diluted in Krebs/HEPES/BESA) was added to the cells 10 min before stimulation with C5-derived cleavage products.

Results

Measurement of C5 and C5 activation products in plasma samples

In 110 healthy donors genotyped for the rs17611 SNP, the minor allele frequency was 0.46, in line with published frequency (National Center for Biotechnology Information reported minor allele frequency 0.41). Plasma levels of C5a, C5, and TCC were measured in the cohort; mean levels of these analytes were comparable to published levels: [C5a] = 9.23 ng/ml, [C5] = 86.1 μg/ml, and [TCC] = 0.31 μg/ml. When the cohort was divided on the basis of rs17611 SNP status, risk allele number was positively associated with [C5a] (p < 0.001 by ANOVA) and negatively associated with [C5] (p = 0.005) (Fig. 1). Plasma [TCC] was not significantly
different between the groups. Attempts to detect C5a in patient and control plasma by Western blotting were unsuccessful, likely because of its very low concentration. Synovial fluid was not available from patients in the cohorts studied.

In a second cohort comprising 80 RA patients, [C5a] again showed strong positive correlation with risk allele number ($p < 0.001$), and [C5] showed a trend to negative correlation that did not reach significance; as in healthy subjects (Supplemental Fig. 1), [TCC] was not different between the groups (data not shown).

To assess whether the observed differences in C5 and C5a concentrations were a result of systemic complement activation, C3a levels were measured in 71 RA patients. There was no significant difference between the groups (Supplemental Fig. 1), leading us to conclude that the increased C5 turnover in risk allele carriers was not due to increased complement activation.

Complement lytic activity of the two C5 V802I protein variants

To determine whether the decreased C5 and elevated C5a levels in individuals expressing rs17611 risk alleles was the result of higher turnover by the C5 convertase enzyme, hemolytic assays were performed. C5 purified from healthy individuals homozygous for each variant (C5V802 or C5I802) was titrated back into NHS depleted of C5 (NHSΔC5) and used in hemolysis assays. Neither end point nor rate hemolytic assays showed any significant difference between the C5 variants (Fig. 2).

To increase the sensitivity of this assay and reveal small differences in C5 function, C3/C5 convertase coated sheep erythrocytes were generated and C5 variant proteins were titrated into the cells. Lysis was developed by addition of purified terminal components. No significant difference in the development of lysis was seen between the two C5 variants (Fig. 3).

Finally, we compared the C5 variants in a CVF C3/C5 convertase assay; no significant difference in rate or extent of cleavage of the variants was observed, either by SDS-PAGE or by C5a ELISA (Supplemental Fig. 2).

These data confirm that the C5V802I variants are not differentially processed by the C5 convertase and provoked us to look for alternative explanations for differences in turnover.

HNE cleaves C5V802I variants at different rates to produce an active C5a-like fragment

HNE, an enzyme released from activated neutrophils at inflammatory sites, has been shown to cleave C5 (32), but the resulting fragments have not been fully characterized. To assess the impact of the C5V802I polymorphism on HNE-mediated cleavage, purified C5 variant were digested with HNE. Cleavage of the C5 chains was assessed on SDS-PAGE and appearance of the C5a-like fragment measured by Western blotting with a C5a-specific mAb (Fig. 4A). Both rate of disappearance of intact C5 and appearance of the C5a-like fragment were greater for the C5 risk variant C5V802. To confirm this accelerated cleavage, generation of the C5a-like fragment was measured by ELISA; levels were higher for C5V802 at all time points in the experiment, up to 6-fold at 60 min (Fig. 4B).

The HNE-generated C5a-like fragment migrated at a slightly higher molecular mass compared with recombinant C5a and C5a

---

**FIGURE 1.** C5 and C5 activation products in plasma samples correlate to rs17611 genotype. (A) Genotyping a healthy population for rs17611. The gene frequency of the risk allele (g) in this population is 0.54. (B) and (C), respectively, show C5a and C5 plasma levels in genotyped individuals measured by ELISA. Plasma levels were observed to be significantly different with increasing copy number the risk variant (g) correlating to higher levels of C5a ($**p < 0.001$) and lower levels of C5 ($**p = 0.05$). (D) TCC plasma levels were also measured by ELISA and show no significant trend with genotype in these assays.
generated through cleavage of C5 by the CVFBb convertase, as shown by Western blot with a C5a-specific mAb (Supplemental Fig. 3). To test whether this HNE-generated fragment retained C5a activities, C5 was incubated with or without HNE for 2 h and then added to C5αR1-transfected U937 cells preloaded with Fura2-AM. HNE-digested C5 triggered a prompt Ca²⁺ flux, whereas addition of uncleaved C5 or HNE alone had no effect. As a negative control, U937 cells transfected with an IL-8R did

**FIGURE 2.** Both C5 V802I variants show comparable complement lytic activity in standard hemolysis assays. (A) Two C5 variants were purified by affinity chromatography and subsequent gel filtration. C5 variants were run on a reduced gel to confirm purity. Two bands were present, representing C5α (115 kDa) and C5β (75 kDa). (B) C5 variants were titrated into C5-depleted serum and ShEA. C6–C9 was then added, and RBC lysis was measured by hemoglobin release after 30-min incubation. Representative results from three experiments are shown. (C) The rate of lysis was calculated by measuring the number of intact cells present at 3-min intervals at each concentration of C5 for the C5 add-back assays described. Results were calculated for each concentration in of C5 in (B). The rate curves for two concentrations of C5 are shown. Representative results from three experiments are shown.

**FIGURE 3.** Both C5 V802I variants show comparable complement lytic activity. (A) Hemolysis was measured by first coating sheep erythrocytes with C3bBb and then titrating C5 and inducing lysis by adding remaining complement components. The end-point assay is shown after 30-min incubation. Percentage lysis was calculated by measuring hemoglobin release for each C5 concentration at end point. Representative results from more than six experiments are shown. (B) The rate of lysis was plotted by measuring absorbance of intact cells at 3-min intervals for each concentration of the C5 add-back assays described. Two representative concentrations of C5 are shown. Representative results from six experiments are shown.
not respond to HNE-digested C5 (data not shown). Pre-incubation with a C5aR1 antagonist blocked the HNE treated C5-triggered increase in intracellular Ca\(^{2+}\) (Fig. 5). rC5a caused a Ca\(^{2+}\) flux in C5aR1 expressing U937 comparable to that caused by HNE-digested C5 (Fig. 5).

Discussion

Complement is a key component of the innate immune response, and complement polymorphisms and mutations influence risk in a range of diseases (17). Some complement polymorphisms have been shown to influence complement activity, and the complotype describes the inherited set of complement variants that dictate complement function in an individual (18). Several polymorphisms in C5 have been associated with RA in various populations (4, 20). Multiple SNPs at the C5-TRAF locus have a significant association in C5 have been associated with RA in various populations (4, 20). Some cells were preincubated with C5aR antagonist (ANTA) and then stimulated with HNE-incubated C5 (dotted line). Arrow 1, addition of C5 ± HNE; arrow 2, addition of C5a.

HNE, a neutrophil-derived enzyme, was previously shown to cleave C5 and, in the presence of C6, generate a C5b6-like hemolytic complex (32). In this study, we showed that C5V802, risk for RA, was more readily cleaved by HNE than C5V802. These findings suggest that the observed difference in turnover between the C5 variants lies in their susceptibility to cleavage by HNE and perhaps other noncomplement enzymes. It is compelling that this difference is evident even in normal individuals even though HNE plasma levels are normally low and most are bound in an inactive form to α1-protease inhibitor (36); however, during inflammation, neutrophils activated locally and/or systemically release HNE leading to markedly increased local levels (35). HNE cleaves C5 at a site close to but distinct from the C5a cleavage site [after Arg\(^{751}\) in the α-chain (32)]; the precise cleavage site is not identified. We noted that the HNE-released fragment was \(~1\text{ kDa}\) larger than convertase-generated C5a (Supplemental Fig. 3); knowledge that HNE preferentially cleaves after small side-chain aliphatic amino acids with Val preferred leads us to predict that the HNE cleavage site in C5 follows Val\(^{760}\), nine residues downstream of the C5a terminus at Arg\(^{751}\), generating a fragment with a predicted molecular mass of 9307 Da (921 Da larger than C5a). Efforts to confirm this prediction by MALDI-TOF analysis are ongoing. The HNE-generated C5a-like fragment was capable of binding C5aR on indicator cells and triggering Ca\(^{2+}\) flux similar to that generated by recombinant C5a, confirming that it was biologically active. It is possible that this novel C5a-like molecule will be more stable than C5a in vivo as it will not be inactivated by des-Argination. We have shown that plasma levels of C5a and C5 are affected by C5V802 status, likely a consequence of increased susceptibility of the C5V802 variant to HNE cleavage. In RA, the inflamed joint is full enzyme cleavage during tickover activation and suggest that other mechanisms of C5 cleavage and C5a generation are responsible.

To investigate the cause of these differences, functional assays comparing the C5V802 variants were performed. In multiple hemolytic assays, no differences between the C5 variants was observed, indicating that increased turnover of C5V802 was not caused by increased susceptibility to cleavage by C5 convertase. These findings eliminate the possibility that increased C5 turnover in normal individuals expressing the risk variant is due to increased complement

![FIGURE 4. HNE cleaves C5 V802I variants at different rates. (A) C5 (I802 and V802) variants were incubated with HNE for 24 h. The reaction was stopped at various time points, and samples were run on a gel (top gel) and Western blot (bottom gel) and probed for C5a. A C5a-like molecule appeared over time. (B) C5 (I802 and V802) variants were incubated with HNE for 3 h. The reaction was stopped at various time points, and samples were measured for the presence of C5a by ELISA.](http://www.jimmunol.org/)

![FIGURE 5. Both C5 variants are functionally active when tested in calcium flux assays. C5aR1-transfected cells were loaded with Fura-2-AM and stimulated with C5 variants I801 (A) or V802 (B) after incubation with HNE (solid black line) or buffer alone (not shown). Cells stimulated with C5 not incubated with HNE were subsequently stimulated with C5a (0.8 nM). Some cells were preincubated with C5aR antagonist (ANTA) and then stimulated with HNE-incubated C5 (dotted line). Arrow 1, addition of C5 ± HNE; arrow 2, addition of C5a.](http://www.jimmunol.org/)
of activated neutrophils and other inflammatory cells. HNE and other proteases are abundant in RA synovial fluid (36, 37); this would lead to local cleavage of C5 and generation of functional C5a-like fragments that in turn drive more inflammation, a process that is enhanced for C5aR and explains its association with inflammatory diseases. Our findings build the case for use of HNE inhibitors in the treatment of RA. HNE has long been implicated in chronic inflammatory disease (38, 39); however, although a number of effective agents have been developed for treatment of other conditions, few have been tested in RA. Sivelestat, a small molecule inhibitor of HNE used in therapy of acute respiratory failure in sepsis (40), already has proved effective in the collagen-induced arthritis model (41). The anti-C5 mAb eculizumab was tested in RA but was not progressed; data on the Alexion Web site shows that although the primary end point was missed, a positive response was achieved at 3 mo in patients stratified according to baseline TCC despite the low dose (8 mg/kg) used. Trials of the C5aR antagonist PMX53 also failed, but this agent has failed in all trials likely because of limited bioavailability (42). Our data suggest that effective inhibition of C5a/C5aR would be therapeutic in RA.

Disclosures

C.L.H. has a contract of employment with GlaxoSmithKline, and B.P.M. prohicles are abundant in RA synovial fluid (36, 37); this would lead of activated neutrophils and other inflammatory cells. HNE and other proteases are abundant in RA synovial fluid (36, 37); this would lead to local cleavage of C5 and generation of functional C5a-like fragments that in turn drive more inflammation, a process that is enhanced for C5aR and explains its association with inflammatory diseases. Our findings build the case for use of HNE inhibitors in the treatment of RA. HNE has long been implicated in chronic inflammatory disease (38, 39); however, although a number of effective agents have been developed for treatment of other conditions, few have been tested in RA. Sivelestat, a small molecule inhibitor of HNE used in therapy of acute respiratory failure in sepsis (40), already has proved effective in the collagen-induced arthritis model (41). The anti-C5 mAb eculizumab was tested in RA but was not progressed; data on the Alexion Web site shows that although the primary end point was missed, a positive response was achieved at 3 mo in patients stratified according to baseline TCC despite the low dose (8 mg/kg) used. Trials of the C5aR antagonist PMX53 also failed, but this agent has failed in all trials likely because of limited bioavailability (42). Our data suggest that effective inhibition of C5a/C5aR would be therapeutic in RA.

References

1. Silman, A. J., and M. C. Hochberg. 1993. Epidemiology of the rheumatic dis-
2. Tobón, G. J., P. Youinou, and A. Saraux. 2010. The environment, geo-
epidemiology, and autoimmune disease: Rheumatoid arthritis. J. Autoimmun.
35: 10–14.
dperimental arthritis by targeting synovial endothelium with a neutralizing antibody to C5. Arthritis Rheum. 60: 2559–2567.
Figure S1: C5 and C5 activation products in plasma samples of RA patients also correlate to rs17611 genotype, C3a levels do not.
A) B) and C) respectively show C5a, C5 and C3a plasma levels of genotyped RA individuals. Levels were measured by ELISA and were observed to be significantly different between homozygous individuals for C5a, with C5a levels increasing according to copy number of the risk variant (g). C5 levels show the reverse trend, but with no significance owing to limited sample size. C3a levels show no trend and were not significantly different.
Figure S2. Cobra Venom Factor assays show comparable cleavage of the two variants.

A) Cleavage assays show that both C5 variants can be cleaved to C5α’ using CVFBb enzyme as analysed by SDS PAGE and quantified by densitometry (table), the rate of cleavage is comparable for both C5 variants. Gel shown is representative of three separate experiments.

B) C5a release was also measured in the samples at desired time points by ELISA. There was no significant difference in C5a generation observed.
C5a was generated by incubation of C5 variants with HNE or CVFBB for 1 hour at 37°C. Samples were run on SDS-PAGE, probed with anti-C5a and developed as a western blot. Recombinant C5a was also run as a control.

Figure S3. C5a and C5 cleavage fragments sizes by western blot.