IL-1R Type 2 Suppresses Collagen-Induced Arthritis by Inhibiting IL-1 Signal on Macrophages

Kenji Shimizu, Akiko Nakajima, Katsuko Sudo, Yang Liu, Atsuhiko Mizoroki, Tetsuro Ikarashi, Reiko Horai, Shigeru Kakuta, Toshiki Watanabe and Yoichiro Iwakura

*J Immunol* published online 27 February 2015
http://www.jimmunol.org/content/early/2015/02/27/jimmunol.1402155

**Supplementary Material**
http://www.jimmunol.org/content/suppl/2015/02/27/jimmunol.1402155.5.DCSupplemental

**Subscription**
Information about subscribing to *The Journal of Immunology* is online at:
http://jimmunol.org/subscription

**Permissions**
Submit copyright permission requests at:
http://www.aai.org/About/Publications/JI/copyright.html

**Email Alerts**
Receive free email-alerts when new articles cite this article. Sign up at:
http://jimmunol.org/alerts
**IL-1R Type 2 Suppresses Collagen-Induced Arthritis by Inhibiting IL-1 Signal on Macrophages**

Kenji Shimizu,*† Akiko Nakajima,‡ Katsuko Sudo,†,1 Yang Liu,†,2 Atsuhiko Mizoroki,† Tetsuro Ikarashi,§ Reiko Horai,‡,3 Shigeru Kakuta,‡,‡ Yoshiki Watanabe,§ and Yoichiro Iwakura*†,‡,‖

IL-1α and IL-1β (in this article referred to as IL-1) play important roles in host defense against infection and inflammatory diseases. IL-1R1 is the receptor for IL-1, and IL-1R2 is suggested to be a decoy receptor, because it lacks the signal-transducing TIR domain in the cytoplasmic part. However, the roles of IL-1R2 in health and disease remain largely unknown. In this study, we generated EGFP-knock-in IIr2−/− mice and showed that they were highly susceptible to collagen-induced arthritis, an animal model for rheumatoid arthritis in which the expression of IL-1R2 is augmented in inflammatory joints. IIr2 was highly expressed in neutrophils but had only low expression in other cells, including monocytes and macrophages. Ab production and T cell responses against type II collagen were normal in IIr2−/− mice. Despite the high expression in neutrophils, no effects of IIr2 deficiency were observed; however, we found that production of inflammatory mediators in response to IL-1 was greatly enhanced in IIr2−/− macrophages. These results suggest that IL-1R2 is an important regulator of arthritis by acting specifically on macrophages as a decoy receptor for IL-1. *The Journal of Immunology, 2015, 194: 000–000.

IL-1α and IL-1β, an inflammatory cytokine that plays important roles in host defense against infection and inflammatory diseases (1, 2). IL-1 is primarily produced by macrophages, dendritic cells (DCs), and monocytes, and it induces cell adhesion molecules, cytokines, and chemokines in various types of cells, including epithelial cells, endothelial cells, synovial cells, and macrophages, to induce inflammation. Especially, IL-1 disrupts vascular stability through induction of vasodilators and internalization of VE-cadherin on epithelial cells, causing exudation of fluid from blood vessels (3). IL-1 is also involved in the development of fever by inducing PGE2 in the hypothalamus (4–6), activation and proliferation of lymphocytes (7), development of autoimmunity by breaking tolerance of T cells (8), bone resorption and erosion by activating osteoclasts (9, 10), regulation of lipid metabolism by regulating insulin secretion (11), and regulation of body weight by regulating sympathetic nerves (12).

Two receptors, IL-1R1 and IL-1R2, and one antagonistic molecule, IL-1R antagonist (IL-1Ra), are known for the IL-1 system, suggesting a complex control of the system. Although IL-1α and IL-1β have low amino acid sequence homology, both bind IL-1R1, followed by recruitment of IL-1RAcP. Then, MyD88 is recruited to the complex via homotypic interaction of the TIR domains of each of the three molecules to activate NF-κB and AP-1 through the IRAK–TRAF6–TAK1 signaling pathway. Although IL-1Ra binds IL-1R1, IL-1Ra does not induce the signal, because IL-1Ra cannot recruit IL-1R AcP. IL-1R2 also binds IL-1; however, IL-1R2 cannot transduce the signal, because it lacks the TIR domain, which is essential for the association with MyD88, in its intracellular region. It was reported that, when IL-1R2 was blocked by anti–IL-1R2 Ab, polymorphonuclear cell survival was prolonged, and the expression of cytokines, chemokines, and adhesion molecules in response to IL-1 was enhanced in monocytes (13). Overexpression of IL-1R2 inhibited the IL-1 signal in the human 8387 sarcoma line (14) and the human keratinocyte cell line HaCaT (15). Knockdown of IL-1R2 by small interfering RNA results in upregulation of chemokine production from HaCaT cells upon stimulation with IL-1 (16). Transgenic mice, which express IL-1R2 in basal keratinocytes under control of the human keratin 14 promoter, showed impaired inflammation during PMA-induced dermatitis (17). Therefore, the expression of IL-1R2 is rather inhibitory to IL-1 signaling, and IL-1R2 is thought to be a decoy receptor to attenuate IL-1 activity. However, the roles of IL-1R2 under physiological conditions and in diseases remain to be elucidated.

Rheumatoid arthritis (RA) is a typical autoimmune disease affecting ∼1% of people worldwide, irrespective of race. RA is...
characterized by chronic inflammation of synovial tissues in multiple joints that causes swelling of joints, pain, joint deformity, and loss of joint function. Studies using animal models demonstrated that IL-1 is involved in the development of RA (18, 19), and blockade of IL-1 signaling is effective in treating RA (20–23), although this cytokine is primarily important in the development of autoinflammatory diseases, such as Familial Mediterranean fever, cryopyrin-associated periodic syndrome, and Still’s disease (19, 24, 25). Because the concentration of IL-1R2 protein is increased in the synovial fluid (26) and plasma (27, 28) of RA patients, and the promoter region of the IL1R2 gene is hypomethylated in PBMCs of RA patients (28), IL-1R2 is suggested to be involved in its pathogenesis.

In this study, we investigated the role of IL-1R2 in the pathogenesis of arthritis using Il1r2−/− mice. Our results demonstrate that IL-1R2 is an important negative regulator of collagen-induced arthritis (CIA) by suppressing IL-1 functions as a decoy receptor.

Materials and Methods

Mice

C57BL/6J mice were purchased from Japan SLC (Hamamatsu, Japan) and kept in mouse rooms of the Research Institute for Biomedical Sciences, Tokyo University of Science for ≥1 wk before use. Age-matched (7–10-wk-old) and sex-matched mice were used for all experiments. Il1r2−/− mice were obtained from the Central Institute for Experimental Animals. Il1r2+/− mice were generated, as previously described (5), and backcrossed to C57BL/6J mice for eight generations. Il1r2−/− and Il1r2+/− mice were obtained by crossing Il1r2−/− mice and Il1r2+/− mice. Il1r2+/− mice were provided by Immunex (Amgen, Thousand Oaks, CA) (29). All mice were kept under specific pathogen-free conditions in environmentally controlled clean rooms in The Institute of Medical Science and The Graduate School of Agricultural and Life Science, The University of Tokyo, and the Research Institute for Biomedical Sciences, Tokyo University of Science. All experiments were approved by the institutional animal use committees and were conducted according to the institutional ethical guidelines for animal experiments and the safety guidelines for gene-manipulation experiments.

Generation of Il1r2−/− mice

Il1r2−/− mice were generated by gene targeting using embryonic stem (ES) cells (Fig. 1). For the cloning of the homologous arms of the targeting vector, Il1r2 genomic clones were isolated from mouse 129 genomic phage libraries from Stratagene (129SvJ Mouse Genomic Library in the FIX II Vector; La Jolla, CA) using the cDNA probe amplified by the following PCR primers: forward, 5′-CCCCATATTACATCGGAAAGGAGCCCA-3′ and reverse, 5′-TCCATGGACGTGTGGATATCA-3′. The 3.9-kb 5′ arm fragment was generated by ligating two DNA fragments; the 5′ side was cloned from a Il1r2 genomic clone digested with EcoRV and EcoRI, whereas the 3′ side was amplified using the following PCR primers: forward, 5′-GAATTCCTAATCTTTCCTGAGTTAACA-3′ and reverse, 5′-GAAGTTGGTGCGAGCACAATTGTC-3′. The 2.6-kb 3′ arm fragment was cloned from the Il1r2 genomic clone by digesting with EcoRV. Then, the chromosome region of E14.1 ES cells containing the second, third, and fourth exons of the Il1r2 gene, which ranged from the initiation codon ATG to an EcoRV site just after the fourth exon, was replaced by the 2.5-kb DNA fragment containing the Il1r2 gene cDNA was amplified by PCR with KOD FX Neo (TOYOBO, Osaka, Japan) and the following primers: for Il1r2, forward (exon 3; P1), 5′-TGGGTGAAGGTGACAATCCTCGGA-3′, forward (exon 5; P2): 5′-GGGCATACATCTTCTGTGAA-3′, and reverse (exon 9; P3): 5′-CGGGCCATGCAGGTGCAT-3′ and for Actb, forward, 5′-GCACATACAGGCGAGGAGG-3′ and reverse, 5′-AACTTGAGCAGGGTAC3′.

Flow cytometry

Cells were treated with an anti-CD16/32 mAb (clone 93; BioLegend, San Jose, CA) for 30 min at 4°C in HBSS containing 2% FBS and 0.01% sodium azide and then stained with Abs described in Table I for 30 min at 4°C. For intracellular staining, cells were stimulated with 50 ng/ml PMA, 500 ng/ml ionomycin, and 2 μM monensin (all from Sigma, St. Louis, MO) for 5 h. After staining surface Abs, cells were fixed and permeabilized with Foxp3/Transcription Factor Staining Buffer (eBioscience, San Diego, CA), according to the manufacturer’s instructions. The intracellular Ags were stained with the Abs described in Table I for 30 min at 4°C, followed by analysis with a FACS Canto II (BD Biosciences, San Jose, CA) and FlowJo software (TreeStar, Ashland, OR). Biotin-conjugated anti-CD40 mAb was stained with Pacific Blue–conjugated streptavidin (Invitrogen, Carlsbad, CA). Dead cells were stained with 7-aminoactinomycin D (Sigma) or an Aqua Dead Cell Stain Kit (Molecular Probes, Eugene, OR).

Collagen-induced arthritis

CIA was achieved as previously described (32). CFA was prepared by mixing 1 mg heat-killed Mycobacterium tuberculosis (H37Ra; Difco Laboratories, Detroit, MI) with 1 ml IFA (Thermo Scientific, Waltham, MA). Chicken type II collagen (ICC; 4 mg; Sigma) was dissolved in 1 ml 10 mM acetic acid overnight at 4°C. An emulsion was formed by combining CFA with an equal volume of ICC solution. Mice were immunized by s.c. injection with 200 μl emulsion at two different sites on each hind flank. Twenty days later, mice received the same immunization plus 10 μg LPS (Sigma) i.p. as a boost. Arthritis severity was scored for each limb with maximum possible score of 12: 0 = normal, 1 = slight swelling and/or erythema, 2 = extensive swelling and/or erythema, 3 = ankylosing change of the joint.

Histological assessment of arthritis

On day 40 of CIA, two hind limbs were fixed with 10% neutral-buffered formalin and decalcified with 10% EDTA. They were embedded in paraffin, and 4-μm thick slices were prepared. Sections were stained with H&E. Four histological parameters—innervation, pannus formation, cartilage damage, and bone damage—were scored with total score 6 for two hind legs for each parameter. Inflammation: 0 = normal, 1 = local infiltration of a few inflammatory cells, 2 = broad local infiltration, and 3 = broad infiltration invading the joint capsule. Pannus formation: 0 = normal, 1 = pannus formation at up to two sites, 2 = pannus formation at up to four sites, and 3 = pannus formation at more than four sites, one broad pannus formation counts as two sites. Cartilage damage: 0 = normal, 1 = small loss of articular chondrocytes, 2 = cartilage degradation in one region, and 3 = cartilage degradation in more than two regions. Bone damage: 0 = normal, 1 = rough surface of tarsus, 2 = shallow loss of tarsus, and 3 = deep loss of tarsus.

Measurement of mRNA levels in inflammatory joints

On day 40 of CIA, two hind limbs were homogenized in Sepasol–RNA I Super, and RNA was purified according to the manufacturer’s instructions. The resulting RNAs were reverse transcribed using a high-capacity cDNA reverse transcription kit (Applied Biosystems, Carlsbad, CA). Quantitative PCR was performed with the SYBR Premix Ex Taq (TaKaRa; Shiga, Japan) using an iCycler system (Bio-Rad, Hercules, CA). The content of mRNA was determined from the appropriate standard curve and normalized to the amount of Gpdp mRNA. The primer sets are shown in Table II.

Measurement of collagen-specific Ab concentrations

Sera were collected from the tail on days 0 and 40 of CIA. A total of 20 μg/ml ICS in PBS was coated on a 96-well plate at 4°C overnight. Then the wells were blocked with 10% FBS in PBS at room temperature for 1 h. Next, diluted serum samples (5000-, 2500-, 500-, 50-, and 10-fold dilution for total IgG, IgG1, IgG2a, IgG2b, and IgG3, respectively) were
applied and incubated at room temperature for 1 h, followed by the addition of 0.8 µg/ml HRP-conjugated goat anti-mouse IgG (Jackson ImmunoResearch, West Grove, PA) or 0.4 µg/ml alkaline phosphatase–conjugated anti-mouse IgG1, IgG2a, IgG2b, and IgG3 (Santa Cruz, Dallas, TX). Then, 3,3′,5,5′-tetramethylbenzidine solutions (Dako, Carpinteria, CA) or p-nitrophenyl phosphate solutions (Sigma) were applied as the substrate, and 1 N HCl or 1 N NaOH was added to stop color development. The absorbancy at 450 or 415 nm was measured using a microplate reader (MTP-300; Corona Electric, Hitachinaka, Japan). The wells were washed with 0.05% Tween 20 in PBS between each steps. The IgG Ab concentration was calculated using an anti-IIC polyclonal IgG standard with a known concentration. The anti-IIC polyclonal IgG was purified from pooled sera obtained from IIC-immunized mice. We used Protein G Sepharose for prepurification and NHS-activated Sepharose (both from GE Healthcare, Waukesha, WI) for affinity-based purification.

IIC-specific proliferation assay and cytokine titration

We immunized mice with IIC and CFA and harvested lymph nodes (LNs) at day 7 after immunization. Then, LN cells (5 × 10^5 cells/well in 96-well plate) were cultured in the absence or presence of 50 µg/ml heat-denatured IIC for 3 d and labeled with [3H]thymidine (0.25 µCi/ml; PerkinElmer, Boston, MA) for 6 h. RPMI 1640 (Wako, Osaka, Japan) containing 10% FBS, 50 µM 2-ME, 100 U/ml penicillin, and 100 µg/ml streptomycin (all from Life Technologies, Big Cabin, OK), so-called “R10 medium,” was used as culture medium. Then, cells were harvested with a Micro 96 cell harvester (Skatron, Tranby, Norway), and radioactivity was measured with Micro Beta (Pharmacia Biotech, Uppsala, Sweden).

To measure cytokine concentrations, we collected the culture supernatants from the culture for proliferation assay after 3 d and measured the concentration of IFN-γ, IL-17, and TNF with the mouse IFN-γ ELISA set, the mouse IL-17 ELISA set (both from R&D Systems, Minneapolis, MN), and the mouse TNF-α ELISA MAX (BioLegend), respectively.

Myeloid cell preparation and culture

Bone marrow macrophages (BMMPs) were prepared from BMCs, as previously described (33). In brief, we seeded BMCs obtained from femurs and tibiae at 1 × 10^6 cells/ml in a 100-mm nontreated dish using R10 medium.
medium supplemented with 20 ng/ml recombinant mouse M-CSF (R&D Systems). On day 3, nonadherent cells were collected and recultured with another 10 ml fresh medium. At day 7, nonadherent cells were discarded, and adherent cells were collected by treating with 0.7 mM EDTA/PBS. Thiglycollate (TG)-elicited peritoneal macrophages (TGCMPs) were prepared by injecting mice i.p. with 1 ml 4% TGC (Nissui, Tokyo, Japan), followed by the collection of peritoneal cells; dish-adherent cells were used as TGCMPs after culture overnight. Neutrophils (Ly6G⁺Ly6C⁻) and monocytes (Ly6G⁺Ly6C⁺) were sorted from BMCS by flow cytometry on a FACSaria (BD Biosciences) or MoFlo XDP IntelliSort II (Beckman Coulter, Brea, CA). Obtained myeloid cells were cultured in R10 medium containing the stimulants described below.

For cell-survival assay, neutrophils were cultured with 10 ng/ml IL-1β, IL-4, or GM-CSF (all from PeproTech, Rocky Hill, NJ) for 48 h. Live cells (7AAD⁻AnnexinV⁻[BioLegend]) were counted by flow cytometry. Survival ratio was calculated by dividing the live cell number for each condition by the cell number without any cytokines. For the reactive oxygen species (ROS)-generation assay, neutrophils were cultured with 1 mM luminol (Santa Cruz) plus either 100 ng/ml IL-1β or 50 ng/ml C5a (R&D Systems). After stimulation, chemiluminescence was measured with an EnVision plate reader (PerkinElmer, Norwalk, CT) at the indicated time points. The monocyte-activation assay, monocytes were stimulated with 100 ng/ml IL-1β or 1 µg/ml Pam3CSK4 (InvivoGen, San Diego, CA). Cells were collected 15 h later, and surface CD40 expression was examined by flow cytometry.

For the cytokine-production assay, neutrophils and monocytes were treated with either IL-1β or LPS (Sigma), whereas BMMPs and TGCMPs were cultured with 10 ng/ml IL-1α (PeproTech), 10 ng/ml IL-1β, or 1 ng/ml LPS. Culture supernatants were collected after 20 h, and the concentrations of TNF and IL-6 were measured with the Mouse TNF-α ELISA MAX and Mouse IL-6 ELISA MAX (both from BioLegend), respectively. For the measurement of mRNAs, cultured BMMPs and TGCMPs were stimulated with 10 ng/ml IL-1α, 10 ng/ml IL-1β, or 1 ng/ml LPS; 3 h later, total RNAs were purified using a GenElute Mammalian Total RNA Miniprep Kit (Sigma). Similarly, mRNAs from monocytes were collected after stimulation with 100 ng/ml IL-1β or 1 µg/ml LPS. Quantitative PCR was performed, as described above, and the content of mRNA was normalized to the amount of Hprt mRNA.

For Western blotting analysis, BMMPs were cultured for 8 h in RPMI 1640 medium and stimulated with 5 ng/ml IL-1α plus 5 ng/ml IL-1β or 1 ng/ml LPS. Then, cells were lysed with sample buffer (62.5 mM Tris-HCl [pH 6.8], 2% SDS, 5% glycerol, 0.003% bromophenol blue, 5% 2-ME, 6.7 mM NaCl, 1.2 mM KCl, 4.5 mM Na2HPO4, and 0.8 mM KH2PO4) and electrophoresed on 12.5% polyacrylamide gels (Wako). After transfer of proteins onto a polyvinyl difluoride membrane (Bio-Rad Laboratories, Shinagawa, Japan), the membrane was incubated with an Ab to JNK1/JNK2, phosphorylated JNK1/JNK2, phosphorylated p38, p38, IκB (Cell Signaling Technology, Danvers, MA), or β-actin (Sigma), followed by incubation with an HRP-conjugated secondary Ab. Chemiluminescence was developed with an ECL detection system (GE Healthcare) and measured by a LAS 4000 (Fujifilm Life Science, Tokyo, Japan).

Peritonitis

For analysis of EGFP-expressing cells, WT and Il1r2⁻/⁻ mice were injected i.p. with 1 ml 4% TGC, and peritoneal cells were collected 15 h later. For the in vivo neutrophil survival assay, BMCS were isolated from WT (CD45.1) and Il1r2⁻/⁻ (CD45.2) mice, mixed at a 1:1 ratio, and resuspended in PBS. A total of 1 × 10⁷ BMCs was transferred i.v. into irradiated (5.5 Gy twice, 3 h apart) Rag2⁻/- mice. Four weeks later, these mice were injected i.p. with 10 ng IL-1α, and peritoneal cells and BMCS were collected 6 h later.

Fibroblast-like synovial cell preparation and culture

We prepared fibroblast-like synovial cells (FLSCs) according to Chou et al. (34). After the ankle was removed, it was digested in 1 mg/ml collagenase (Wako), 2 mg/ml dispase (Life Technologies), and 10% FBS in 1xMEM (Life Technologies) for 1 h at 37˚C with a tube rotator. Then, cells were suspended in DMEM containing 1% FBS, 100 U/ml penicillin, and 100 µg/ml streptomycin and incubated with Clophosome (FormuMax Scientific, Palo Alto, CA) for 24 h to remove contaminating macrophages. FLSCs were stimulated with IL-1α, IL-1β, TNF-α (PeproTech), IL-17A (R&D Systems), and PGE2 (Nacalai Tesque) for 20 h. Cultured supernatants were collected at 20 h, and the concentration of CCL2 and IL-6 was measured with the Mouse CCL2/JE/MCP-1 DuoSet (R&D Systems) and Mouse IL-6 ELISA MAX (BioLegend), respectively.

**FIGURE 1.** Generation of Il1r2⁻/⁻ mice. (A) Structure of the mouse Il1r2 locus (WT locus), the IL-1R2-targeting construct (Targeting vector), and the predicted mutated gene before (Mutant locus) and after (Cre-mediated neo' deletant locus) neomycin resistance gene (neo') deletion. Bold lines indicate arms used for homologous recombination. Gray boxes with roman numerals represent the exons of the Il1r2 gene and its number, respectively. Exons 2, 3, and 4 of the Il1r2 gene were replaced with the EGFP gene and the loxp flanked neo' gene. A DTA gene was bound to the 3' end of the targeting vector for negative selection. (B) Southern blot analysis of the ES cells. The WT (10.5 kb) and targeted band (7.0 kb) were detected in Southern blot analysis using a FACSaria (BD Biosciences) or MoFlo XDP IntelliSort II (Beckman Coulter, Brea, CA). Obtained myeloid cells were cultured in R10 medium containing the stimulants described below.
Statistical analysis

The p values were calculated using the Mann–Whitney U test for clinical and histological scores of CIA, the χ² test for incidence of CIA, and the two-tailed unpaired Student t test for all other experiments. The p values < 0.05 were considered significant.

Results

Generation of Il1r2−/− mice

We generated Il1r2−/− mice by replacing exons 2, 3, and 4 of the Il1r2 gene with the EGFP gene and the neomycin-resistant gene using homologous-recombination techniques (Fig. 1A). Homologous recombination was confirmed by genomic Southern blot hybridization analysis (Fig. 1B). Il1r2 deficiency was verified by RT-PCR using the RNA from bone marrow. Consistent with this, we did not detect the PCR product from exons 3–9. The PCR product from exons 5–9 also was not detected, indicating that there was no truncated mRNA from the targeted gene (Fig. 1C).

When Il1r2+/− mice were intercrossed, Il1r2−/− mice were born at the expected Mendelian ratio. They were fertile and showed no obvious phenotypic abnormalities under specific pathogen–free conditions (data not shown). The content of T cells, B cells, plasmacytoid DCs, conventional DCs, migratory DCs, monocytes, and neutrophils in the LNs and bone marrow of Il1r2−/− mice was normal, as analyzed by flow cytometry (Fig. 1D, 1E).

We analyzed the expression of Il1r2 through the expression of EGFP in Il1r2+/− mice, which express EGFP in place of Il1r2. High EGFP expression was observed in neutrophils (Fig. 2A). Low levels of EGFP expression were detected in macrophages and monocytes from TGC-treated mice, although the expression was very low under physiological conditions. Expression was not detected in T cells or B cells. The EGFP expression level was consistent with the Il1r2 mRNA level determined by quantitative PCR (Fig. 2B). Similarly, Il1rn and Il1rap mRNAs were highly expressed in neutrophils and modestly expressed in other types of cells. In contrast, high levels of Il1r1 mRNA expression were observed in FLSCs, but its expression was low in other cells.

Il1r2−/− mice show increased susceptibility to CIA

Because it was suggested that IL-1R2 is involved in the pathogenesis or progression of RA (26–28), we examined the susceptibility of Il1r2−/− mice to CIA. Il1r2−/− mice showed higher clinical scores and incidence of CIA (Fig. 3A, 3B). Histological severity scores, as evaluated by the infiltration of polymorphonuclear leukocytes, pannus formation, erosion of cartilage, and bone destruction, were higher in arthritic joints of Il1r2−/− mice than in WT mice (Fig. 3C–G). These results suggest that IL-1R2 negatively controls the development of arthritis.

FIGURE 2. Cell type specificity of IL-1R2 expression. (A) EGFP expression in pre-B cells, B cells, T cells, monocytes, macrophages, and neutrophils from bone marrow, blood, and peritoneal cavity of WT and Il1r2+/− mice was analyzed by flow cytometry. Peritoneal B cells, macrophages, monocytes, and neutrophils were collected before and after induction of peritonitis with TGC. (B) The expression of Il1r1, Il1r2, Il1rap, and Il1rn mRNA was analyzed by quantitative PCR in different types of cells from WT mice. The content of mRNA was normalized to the amount of Hprt mRNA. CD4+ T cells, CD8+ T cells, and B220+ B cells were sorted from spleens and Ly6G+ neutrophils and Ly6C+ monocytes were sorted from bone marrow by flow cytometry. Data are mean ± SD of three mice (B) and are representative of two (A and B) independent experiments.
Inflammatory mediator production, but not Ab production or T cell proliferative responses, is enhanced in Il1r2−/− mice after CIA induction

It is well known that both humoral and cell-mediated immunity are involved in the development of CIA (35, 36). To elucidate the mechanism for the exacerbation of CIA in Il1r2−/− mice, we first analyzed anti-IIC IgG concentration in sera. No significant differences were found between WT mice and Il1r2−/− mice (Fig. 4A). The levels of anti-IIC IgGs in each IgG subclass also did not differ between WT and Il1r2−/− mice (Fig. 4B). Next, we examined IIC-specific recall T cell responses; inguinal LN cells were harvested 7 d after immunization, and the cells were incubated or not with IIC for 3 d. The proliferative response against IIC of Il1r2−/− LN cells was similar to WT cells (Fig. 4C). Consistent with the proliferative response, cytokine production, such as IFN-γ, IL-17, and TNF, was normal in LN cells from Il1r2−/− mice upon stimulation with IIC (Fig. 4D–F). Furthermore, the numbers of total LN cells, B cells, CD4+ and CD8+ T cells, IFN-γ+ CD4+ and CD8+ T cells, IL-17+ CD4+ T cells, and Foxp3+ CD4+ T cells were normal in Il1r2−/− mice at day 7 after immunization (Fig. 4G). Interestingly, we found that the expression of Il6, Cxcl2, Nos2, and Il1b mRNAs, which are produced in macrophages and fibroblasts (37–40), was strongly up-regulated in inflammatory joints from Il1r2−/− mice (Fig. 4H).

Il1r2−/− neutrophils show normal phenotype

IIC-specific T cells and Abs are important for the development of arthritis by recruiting neutrophils and monocytes/macrophages, which enhance inflammation by producing cytokines, chemokines, NO, and...
chemical mediators. In addition, joint-resident FLSCs are involved in the development of inflammation (41). Because Ab production and T cell priming were normal in Il1r2−/− mice, we next examined the effect of IL-1R2 deficiency on these cells.

Because neutrophils expressed high amounts of Il1r2, and a previous report suggested involvement of IL-1R2 in neutrophil survival (13), we analyzed the effects of IL-1 on neutrophils. We found that neutrophil survival did not change upon stimulation.
with IL-1β, even in Il1r2−/− neutrophils (Fig. 5A). IL-6 production in WT or Il1r2−/− neutrophils also did not change as a result of treatment with IL-1β (Fig. 5B). In contrast, GM-CSF and LPS prolonged neutrophil survival and induced cytokine production in both WT and Il1r2−/− neutrophils. Although IL-4 is suggested to induce Il1r1 and Il1r2 mRNA in neutrophils (13), IL-4 or IL-1β plus IL-4 did not affect the survival of WT or Il1r2−/− neutrophils (Fig. 5A). IL-1β also failed to induce ROS generation from both WT and Il1r2−/− neutrophils under the conditions in which C5a efficiently induced ROS generation (Fig. 5C).

We also examined the effect of IL-1R2 deficiency on neutrophil survival in vivo. Peritonitis was induced by i.p. injection of IL-1α to

**FIGURE 5.** The response of Il1r2−/− neutrophils to IL-1 is normal. (A) Neutrophils from WT and Il1r2−/− mice were treated with the indicated cytokines (10 ng/ml), and the survival ratio was measured by flow cytometry after a 48-h culture. (B) IL-6 production was measured by ELISA at 20 h after treatment with IL-1β or LPS. (C) ROS generation was measured by chemiluminescence with a luminometer at the indicated time points after IL-1β or C5a stimulation. (D and E) A mixture of CD45.1+ WT and CD45.2+ Il1r2−/− BMCs were transferred into irradiated Rag2−/− mice, and these bone marrow chimeras were injected with IL-1α to induce peritonitis. (D) After 6 h, neutrophil and monocyte infiltration into the peritoneal cavity was examined. (E) The content of neutrophils (Neut), monocytes (Mono), macrophages (Mac), and B cells of each genotype was determined by allelic forms of CD45 Ag in the peritoneal cavity and bone marrow. Data are mean ± SD of triplicate cultures (A–C) or four mice (E) and are representative of two (A, C, and E) or four (B) independent experiments.
CD45.1+ WT and CD45.2+ Il1r2−/− bone marrow chimeras; infiltration of neutrophils and monocytes into the peritoneal cavity was examined 6 h later. As shown in Fig. 5D, neutrophil and monocyte infiltration was clearly observed. However, the WT/Il1r2−/− ratio did not change in the peritoneal cavity or bone marrow, suggesting comparable survival between WT and Il1r2−/− neutrophils in these mice (Fig. 5E).

Excess cytokine production in Il1r2−/− macrophages was observed upon stimulation with IL-1

Next, we examined inflammatory mediator production from FLSCs upon treatment with IL-1, TNF, IL-17A, or PGE2. FLSCs were prepared as described in Materials and Methods. The percentage of contaminated macrophages in the FLSC preparation was <1%, as determined by flow cytometry using anti-CD45 mAb (Supplemental Fig. 1A). mRNAs for proteoglycan 4 (Prg4) or lubricrin, which is a component of synovial fluid, and α1 type 1 collagen (Col1a1), which is the major component of type 1 collagen, were highly expressed in FLSCs (Supplemental Fig. 1B). We found that the production of IL-6 and CCL2 was normal in Il1r2−/− FLSCs (Fig. 6).

In contrast, monocytes from both WT and Il1r2−/− mice did not respond to IL-1 to produce TNF- or IL-6–like neutrophils, although these cells produced those cytokines at similar levels in response to LPS (Fig. 7A, 7B). Although we detected a slight upregulation of Tnf, Il1b, and Cxcl2 mRNA upon IL-1 stimulation, no difference was observed between WT and Il1r2−/− monocytes (Fig. 7C–E). Similarly, cell surface expression of CD40 was marginally upregulated by treatment with IL-1, but its expression levels were comparable between WT and Il1r2−/− monocytes (Fig. 7F, 7G).

In contrast, significantly higher amounts of TNF and IL-6 were produced in Il1r2−/− BMMPs and TGCMPs compared with WT cells in response to IL-1α or IL-1β but not LPS (Fig. 8A, 8B). Quantitative PCR revealed that mRNA expression of Il1a, Il1b, Cxcl2, Nos2 (encoding iNOS), and Ptgs2 (encoding COX-2) also was enhanced in Il1r2−/− BMMPs and TGCMPs (Fig. 8C, 8D).}

Discussion

IL-1R2 is considered a decoy receptor for IL-1α and IL-1β, and many reports support this concept in vitro (13–16). The negative-regulatory function of IL-1R2 against IL-1 signaling is also suggested in Il1r2-transgenic mice, which are designed to express IL-1R2 in keratinocytes (17). However, the physiological and pathological roles of endogenous IL-1R2 have not been elucidated completely. In this study, we have shown that IL-1R2 is functional in macrophages as a negative regulator of IL-1 and suppresses the development of CIA.

Il1r2−/− mice were born healthy and breed normally; however, we found that the development of CIA was enhanced. The severity score and incidence of arthritis were increased. Anti-IIC concentrations in the serum, IIC-specific T cell proliferative responses, and cytokine production in LN cells upon stimulation with IIC were normal in Il1r2−/− mice, suggesting that Il1r2 is not involved in the control of T cell priming and production of Ag-specific Abs. However, we showed that the expression of mRNAs for inflammatory mediators, such as IL-6, CXCL2, Nos2, and IL-1β, which are important for the development of arthritis (42–45), was upregulated in the joints of Il1r2−/− mice. Furthermore, we found that, upon stimulation with IL-1α and IL-1β, cytokine and inflammatory mediator production, including IL-6, CXCL2, Nos2, and IL-1β, was greatly enhanced in macrophages, but not in FLSCs or monocytes, from Il1r2−/− mice compared with WT mice, suggesting that increased inflammatory mediators in Il1r2−/− mouse joints are derived from macrophages. Consistent

![FIGURE 6. Normal inflammatory mediator production in Il1r2−/− FLSCs. FLSCs from WT and Il1r2−/− mice were stimulated with cytokines, as indicated. The concentrations of IL-6 (A) and CCL2 (B) in the culture supernatant were measured by ELISA. No significant difference was observed. Data are mean ± SD of triplicate cultures and are representative of three independent experiments.](http://www.jimmunol.org/Downloaded from)
with this notion, it was reported that macrophage depletion results in the suppression of CIA associated with downregulation of these inflammatory mediators (46). Thus, these observations suggest that IL-1R2 regulates the development of arthritis by suppressing IL-1 activity on macrophages.

We found that IL-1R2 is expressed most prominently in neutrophils among T cells, B cells, monocytes, BMMPs, and FLSCs, consistent with a recent report (47). However, in contrast to macrophages, we did not detect any functional abnormality of Il1r2<sup>−/−</sup> neutrophils. The neutrophil content in the bone marrow was similar between WT and Il1r2<sup>−/−</sup> mice. Survival after treatment with IL-1β, IL-4, or both was not different between WT and Il1r2<sup>−/−</sup> neutrophils. IL-6 and ROS production after treatment with IL-1α or IL-1β also did not change. Furthermore, the content

**FIGURE 7.** The response of Il1r2<sup>−/−</sup> monocytes to IL-1β is normal. (A and B) Monocytes from WT and Il1r2<sup>−/−</sup> mice were stimulated with IL-1β or LPS, and concentrations of TNF and IL-6 were measured by ELISA. (C-E) Monocytes from WT and Il1r2<sup>−/−</sup> mice were stimulated with IL-1β or LPS, and the mRNA levels of Tnf, Il1b, and Cxcl2 were determined by quantitative PCR. (F and G) Monocytes from WT and Il1r2<sup>−/−</sup> mice were stimulated with IL-1β or Pam3CSK4 (Pam3), and cell surface expression of CD40 was analyzed by flow cytometry. Representative graphs (F) and mean fluorescence intensity levels (G) of CD40. Data are mean ± SD of triplicate cultures (A–E and G) and are representative of three (A and B) or two (E and F) independent experiments.
of Il1r2^−/− neutrophils, monocytes, macrophages, and B cells in peritoneal cavity, blood, and bone marrow was similar to that of WT after induction of peritonitis in mixed bone marrow chimera mice. Consistent with our notion, Prince et al. (48) demonstrated that IL-1 is not involved in neutrophil survival, cell adhesion molecule expression, or cytokine production. Although it was reported that IL-1 prolongs neutrophil survival (13), Prince et al. (48) showed that neutrophils purified by the Percoll gradient method, which was used in the previous report (13), were contaminated with other types of cells, and neutrophils were indirectly activated by IL-1 through these contaminated cells. Regarding the reason why Il1r2^−/− neutrophils are refractory to IL-1 stimulation, we first thought that this is because IL-1R antagonist (IL-1Ra) is highly expressed in neutrophils (Fig. 2B).

FIGURE 8. Inflammatory mediator production is increased in Il1r2^−/− macrophages. BMMPs (A and C) and TGCMPs (B and D) from WT and Il1r2^−/− mice were stimulated with 10 ng/ml IL-1α, 10 ng/ml IL-1β, or 1 ng/ml LPS, and the concentrations of TNF and IL-6 in the culture supernatant were measured by ELISA (A and B). (C and D) mRNA levels of Il1a, Il1b, Cxcl2, Nos2, and Ptgs2 were determined by quantitative PCR. (E) Phosphorylation of JNK1/JNK2 and p38, as well as degradation of IκBα, was examined by Western blot analysis of WT and Il1r2^−/− BMMPs at 0, 15, 30, and 60 min after stimulation with IL-1 or LPS. Data are mean ± SD of triplicate cultures (A–D) and are representative of two independent experiments (A and C). *p < 0.05, two-tailed unpaired Student t test.
However, we found that this is not the case, because neutrophils from Il1r2 mice did not respond to IL-1 (Supplemental Fig. 2). Therefore, it is conceivable that IL-1R1 expression on the cell surface is too low to respond to IL-1, some signaling molecules are missing, or some negative regulators of signal transduction are highly expressed in neutrophils. Consistent with the first possibility, IL-1R1 expression was not detected on neutrophils, although the mRNAs for IL-1R1 and IL-1RaP were clearly detected. Unfortunately, we also failed to detect expression of IL-1R1 on macrophages for which we clearly observed the effects of IL-1. IL-1R1 expression on macrophages and neutrophils was not detected, even after amplification using biotin-conjugated anti-PE Ab and after stimulation with GM-CSF, IFNγ, and IL-23 (Supplemental Fig. 3). Because we could easily detect cell surface IL-1R1 expression on peritoneal CD62L−/γδ T cells from WT mice, but not from Il1r1−/− mice, as described previously (36), the expression levels of IL-1R1 on neutrophils and macrophages are much lower than that on peritoneal CD62L−/γδ T cells.

Nonetheless, high expression of IL-1R2 in neutrophils may play some physiological and/or pathological roles. In fact, it was demonstrated that neutrophils scavenge IL-1β through IL-1R2 (49), and both human and mouse neutrophils cleave IL-1R2 to release the extracellular domain of IL-1R2 (47, 50). Because an IL-1R2–expressing T cell line can neutralize IL-1 activity through binding of soluble IL-1R2 to IL-1β (51), we examined the nature of WT or Il1r2−/− neutrophils. Mice deficient for other type IL-1R signal regulators, such as ST2 and SIGIRR, also do not develop arthritis (53, 54). Neutrophils scavenge IL-1α/β (55). Clearly, further investigations are needed to elucidate the role of IL-1RAcP in neutrophils.

We found that IL1rn−/− mice spontaneously develop autoimmune arthritis on the BALB/cA background (52). In contrast, Il1r2−/− mice did not develop autoimmunity on the BALB/cA or C57BL/6 background. This difference between IL1rn−/− and Il1r2−/− mouse phenotypes could be explained by the difference in target cells; IL-1Ra can inhibit IL-1R1 on all cell types, whereas IL-1R2 can only compete with IL-1R1 on macrophages. Mice deficient for other type IL-1R signal regulators, such as ST2 and SIGIRR, do not develop arthritis (53, 54). These observations suggest that IL-1Ra is the most potent and cell type–independent regulator among endogenous IL-1 inhibitors.

We found that Il1r1−/−Il1r2−/− neutrophils, produced higher levels of TNF than did WT neutrophils when cells were treated with LPS (Supplemental Fig. 2). Interestingly, Il1r1−/−Il1r2−/− mice developed more severe skin inflammation and emaciation compared with Il1r1−/− mice (T. Ikarashi and S. Kakuta, manuscript in preparation). Therefore, Il1r1−/−Il1r2−/− neutrophils seem to be constitutively activated as a result of chronic inflammation of the skin, and they produce TNF easily upon stimulation with LPS. Alternatively, IL-1R2 suppresses IL-1 signaling in neutrophils to induce TNF in collaboration with TLR4 signaling (55). Clearly, further investigations are needed to distinguish these possibilities and to elucidate the roles of IL-1R2 during skin inflammation and emaciation.

In summary, we showed that IL-1R2 plays a regulatory role in the progression of CIA through the inhibition of IL-1 action on macrophages. This regulatory function of IL-1R2, unlike IL-1Ra, is strictly cell type specific. These observations may provide clues for the use of IL-1R2 as a treatment for inflammatory diseases.

Acknowledgments
We thank Dr. J. Miyazaki for providing pCAUG-G-Cre plasmid, H. Katoki for supporting generation of the chimera mice, and H. Ishigame, Y. Tanahashi, and S. Azechi for helpful discussions.

Disclosures
The authors have no financial conflicts of interest.

References


