Steady-State Antigen Scavenging, Cross-Presentation, and CD8+ T Cell Priming: A New Role for Lymphatic Endothelial Cells


*J Immunol* published online 2 May 2014
http://www.jimmunol.org/content/early/2014/05/02/jimmunol.1302492
Steady-State Antigen Scavenging, Cross-Presentation, and CD8\(^+\) T Cell Priming: A New Role for Lymphatic Endothelial Cells

Sachiko Hirose,*† Ethlynia Vokali,*† Vidya R. Raghavan,* Marcela Rincon-Restrepo,* Amanda W. Lund,* Patricia Corthésy-Henrioud,* Francesca Capotosti,* Cornelia Halin Winter,† Stéphanie Hugues,* and Melody A. Swartz*†‡

Until recently, the known roles of lymphatic endothelial cells (LECs) in immune modulation were limited to directing immune cell trafficking and passively transporting peripheral Ags to lymph nodes. Recent studies demonstrated that LECs can directly suppress dendritic cell maturation and present peripheral tissue and tumor Ags for autoreactive T cell deletion. We asked whether LECs play a constitutive role in T cell deletion under homeostatic conditions. In this study, we demonstrate that murine LECs under noninflamed conditions actively scavenge and cross-present foreign exogenous Ags to cognate CD8\(^+\) T cells. This cross-presentation was sensitive to inhibitors of lysosomal acidification and endoplasmic reticulum–golgi transport and was TAP1 dependent. Furthermore, LECs upregulated MHC class I and the PD-1 ligand PD-L1, but not the costimulatory molecules CD40, CD80, or CD86, upon Ag-specific interactions with CD8\(^+\) T cells. Finally, Ag-specific CD8\(^+\) T cells that were activated by LECs underwent proliferation, with early-generation apoptosis and dysfunctionally activated phenotypes that could not be reversed by exogenous IL-2. These findings help to establish LECs as APCs that are capable of scavenging and cross-presenting exogenous Ags, in turn causing dysfunctional activation of CD8\(^+\) T cells under homeostatic conditions. Thus, we suggest that steady-state lymphatic drainage may contribute to peripheral tolerance by delivering self-Ags to lymph node–resident leukocytes, as well as by providing constant exposure of draining peripheral Ags to LECs, which maintain tolerogenic cross-presentation of such Ags. The Journal of Immunology, 2014, 192: 000–000.

The lymphatic system transports interstitial fluid, Ags, solutes, and immune cells from the periphery and returns them to the blood circulation after surveillance through lymph nodes (LNs), thereby initiating adaptive immune responses (1–3). In addition to effector immune responses, LNs are important sites for the maintenance of peripheral tolerance. LN stromal cells, which include lymphatic endothelial cells (LECs) and blood endothelial cells (BECs), as well as fibroblastic reticular cells (FRCs) in the T cell zone, are thought to contribute to tolerance induction of autoreactive T cells that escape central memory (4), as well as to regulate the contraction of inflammatory responses (5). Indeed, the lymphatic endothelium is emerging as an important player in shaping immunity and tolerance (1–3, 6–10). For example, LECs were shown to suppress maturation of dendritic cells (DCs) (1, 4, 11) and their subsequent priming of CD8\(^+\) T cells in a contact-dependent manner (4, 5, 9). In addition, LECs, as well as FRCs, can directly prime CD8\(^+\) T cells (5); they express components of the Ag-presentation machinery, including MHC class I and II molecules (6–9, 12), and were shown to directly contribute to peripheral tolerance by expression and presentation of endogenous peripheral tissue Ags (PTAs), leading to compromised CD8\(^+\) T cell activation (6–9). They are also sensitive to pathogen-associated molecular patterns via the expression of various members of the TLR family (8, 11). Together, these studies established LECs as contributors to the maintenance of peripheral tolerance to endogenously expressed self-Ags.

However, little is known about whether LECs as APCs have the ability to capture and process exogenous Ags for CD8\(^+\) T cell deletion. Although so-called “professional” APCs, such as CD8\(^+\) DCs, can process exogenous Ags for cross-presentation to CD8\(^+\) T cells, some nonhematopoietic cell types also were shown to be capable of cross-presentation (13). For example, liver sinusoidal endothelial cells (LSECs) are thought to capture and cross-present circulating Ag to CD8\(^+\) T cells, leading to CD8\(^+\) T cell deletion and the establishment of a tolerogenic environment (14). This is especially important in the liver, where LSECs are among the first cells to encounter the large diversity of foreign Ags from food, as well as TLR agonists from commensal sources (15).
Similarly, LECs are the first cells to contact extracutaneous Ags that arise in the periphery and drain into lymphatic vessels after, for example, tissue damage, inflammation, or infection. We recently showed that a foreign Ag (OVA) expressed by an orthotypically implanted tumor could be cross-presented by tumor-associated LECs that, when isolated, could drive dysfunctional activation of cognate CD8 T cells and promote tumor progression (16). Because tumors use physiological mechanisms to promote tolerance for their survival (17), we hypothesized that a similar mechanism of Ag cross-presentation by LECs may exist under steady-state conditions to promote tolerance against self-Ags.

In this article, we demonstrate that, under homeostatic conditions, LECs constitutively uptake and cross-present exogenous Ags to CD8 T cells. We further show that LEC-activated T cells are more rapidly apoptotic, upregulate so-called “exhaustion markers” (PD-1, CTLA-4, and CD80), secrete less IFN-γ and IL-2, and express lower levels of the activation markers CD25, CD44, and CD69 compared with T cells activated by mature DCs. Together, these data suggest that LECs help to maintain CD8 T cell tolerance to exogenous Ags that are encountered in lymph under steady-state conditions, which may be important for preventing autoimmune reactions against self-Ags after infection or injury.

Materials and Methods

Reagents

All chemicals were from Sigma-Aldrich (Buchs, Switzerland), unless otherwise noted. The mature MHC class I epitope, OVA256–264 (SINIFELK), peptide, was from GenScript (Piscataway, NJ). Endotoxin-free OVA was from Hyglos (Bernried am Starnberger See, Germany). Abs used in flow cytometry were from eBioscience (Vienna, Austria) or BioLegend (Lucerne, Switzerland) unless otherwise noted.

Mice

The following mice strains were used in this study at age 6–12 wk, unless noted otherwise. Female C57BL/6 wild-type mice and OT-I-transgenic mice, C57BL/6-Tg(TcraTcrb)1100Mjb/J, were purchased from Harlan (Bicester, UK). TAP1−/− mice, C57BL/6-Tg(TcraTcrb)1100Mjb/J, were purchased from The Jackson Laboratory (Farmington, CT). All mice were on a C57BL/6 background and were 8–10 wk old at the time of experimentation. Male C57BL/6 mice (B6.129S2-Ifngtm1Sor/J) were purchased from The Jackson Laboratory (Gannat, France). TAPI-1−/− mice (B6.129S2-Tap1tm1ArgJ) were purchased from The Jackson Laboratory (Farmington, CT). Animals were housed in pathogen-free facilities, and all procedures were approved by the Cantonal Veterinary Committee of Vaud, Switzerland (Protocol number 2518).

Cell lines

Conditionally immortalized dermal LECs (iLECs; Immortomice) were isolated and cultured as previously described (18). Cell culture surfaces used in all assays were coated with collagen (10 μg/ml PureCol; Advanced Biomatrix, San Diego, CA) and 10 μg/ml human fibronectin (Millipore, Billerica, MA) prior to seeding. Cells were grown in 40% DMEM low glucose and 20 mM HEPES (pH 7.2) at 33 °C. Prior to all experiments, cells were transcardially perfused with a heparinized saline solution containing 1 g/l glucose and 20 mM HEPES (pH 7.2). For immunostaining, brachial LNs were removed and fixed overnight in 2% paraformaldehyde in PBS (pH 7.4). After three washes in PBS, LNs were embedded in a block of 2% agarose and sectioned (150 μm) using a using a vibratome (Leica, Wetzlar, Germany). Sections were blocked in 0.5% casein and further labeled using Abs against CD3ε (BD Pharmingen; clone 500A2) and LYE-1 (Reliatech, San Pablo, CA). Images were acquired on a Leica SP5 confocal microscope using a 20× or 60× objective and processed with Imaris software (Bitplane, Zürich, Switzerland). For flow cytometric analysis, brachial LNs from individual mice were pooled and digested with 1 mg/ml Collagenase D and 200 Kunitz/ml DNase I (Sigma-Aldrich). After LNs were fully digested, as described (19), the single-cell preparations were enriched for nonhematopoietic stromal cells by CD45 cell depletion using CD45 MicroBeads (Miltenyi Biotec, Bergisch Gladbach, Germany). Enriched stroma and the CD45+ fraction were counted, stained with gp38, CD31, CD45, and CD11c, and CD11b, and MHC class II, respectively, and analyzed by flow cytometry (CyAn ADP Flow Cytometer; Dako). Data analysis was performed using FlowJo software v9.2 (TreeStar, Ashland, OR).

Intracellular localization studies

To determine the intracellular pathways of Ag trafficking, we incubated iLECs with fluorescently labeled OVA protein and stained them with various markers in the LN after 90 min. Endotoxin-free OVA was labeled with Alexa Fluor 647 NHS (OVA-AF647; Dyomics) and purified by size-exclusion chromatography using a Sephadex G-25 column with PBS as eluent. C57BL/6 mice were injected intradermally (i.d.) with 15 μg OVA-AF647 in the limbs. After 90 min, mice were transcardially perfused with a heparinized saline solution containing 1 g/l glucose and 20 mM HEPES (pH 7.2). For immunostaining, brachial LNs were removed and fixed overnight in 2% paraformaldehyde in PBS (pH 7.4). After three washes in PBS, LNs were embedded in a block of 2% agarose and sectioned (150 μm) using a using a vibratome (Leica, Wetzlar, Germany). Sections were blocked in 0.5% casein and further labeled using Abs against CD3ε (BD Pharmingen; clone 500A2) and LYE-1 (Reliatech, San Pablo, CA). Images were acquired on a Leica SP5 confocal microscope using a 20× or 60× objective and processed with Imaris software (Bitplane, Zürich, Switzerland). For flow cytometric analysis, brachial LNs from individual mice were pooled and digested with 1 mg/ml Collagenase D and 200 Kunitz/ml DNase I (Sigma-Aldrich). After LNs were fully digested, as described (19), the single-cell preparations were enriched for nonhematopoietic stromal cells by CD45 cell depletion using CD45 MicroBeads (Miltenyi Biotec, Bergisch Gladbach, Germany). Enriched stroma and the CD45+ fraction were counted, stained with gp38, CD31, CD45, and CD11c, and CD11b, and MHC class II, respectively, and analyzed by flow cytometry (CyAn ADP Flow Cytometer; Dako). Data analysis was performed using FlowJo software v9.2 (TreeStar, Ashland, OR).

Synthesis of peptide-conjugated nanoparticles

To explore the mechanisms of cross-presentation, poly (propylene sulfide) nanoparticles (NPs) with ∼30 nm diameter were synthesized and characterized as described (20). The long peptide containing the mature MHC class I epitope SINIFELK–Cys-OVA256–264 (OVA256–264) was synthesized in-house and activated with a 2-pyridyldithiol, as previously described (22). Core sulfhydryls on NPs were reacted with the activated peptide to achieve conjugation of the peptide to the NPs (NP-ss-OVA256–264) via a reducible disulfide bond (–SS–) and purified on a Sepharose CL6B column (Sigma-Aldrich). To fluorescently label the NPs, they were exposed to Dy-649 maleimide (Dyomics, Jena, Germany) after dialysis in a 1:60 molar ratio of dye/NP sulfhydryl groups in PBS at room temperature for 24 h (22). Free dye was removed by gel filtration, as above, but in endotoxin-free water (B. Braun Medical, Sempach, Switzerland) as eluent. Endotoxin levels of NPs were routinely assessed by a colorimetric assay based on the HEK-Blue TLR4 cell line (InvivoGen, San Diego, CA), according to the manufacturer’s protocol using a standard curve generated from the E-Toxate endotoxin standard (Sigma-Aldrich).

In vivo Ag drainage

To determine whether LN LECs can actively capture Ags in vivo, we injected fluorescently labeled OVA protein into the limbs of mice and determined its distribution within various cells in the LN after 90 min. Endotoxin-free OVA was labeled with Alexa Fluor 647 NHS (OVA-AF647; Dyomics) and purified by size-exclusion chromatography using a Sephadex G-25 column with PBS as eluent. C57BL/6 mice were injected intradermally (i.d.) with 15 μg OVA-AF647 in the limbs. After 90 min, mice were transcardially perfused with a heparinized saline solution containing 1 g/l glucose and 20 mM HEPES (pH 7.2). For immunostaining, brachial LNs were removed and fixed overnight in 2% paraformaldehyde in PBS (pH 7.4). After three washes in PBS, LNs were embedded in a block of 2% agarose and sectioned (150 μm) using a using a vibratome (Leica, Wetzlar, Germany). Sections were blocked in 0.5% casein and further labeled using Abs against CD3ε (BD Pharmingen; clone 500A2) and LYE-1 (Reliatech, San Pablo, CA). Images were acquired on a Leica SP5 confocal microscope using a 20× or 60× objective and processed with Imaris software (Bitplane, Zürich, Switzerland). For flow cytometric analysis, brachial LNs from individual mice were pooled and digested with 1 mg/ml Collagenase D and 200 Kunitz/ml DNase I (Sigma-Aldrich). After LNs were fully digested, as described (19), the single-cell preparations were enriched for nonhematopoietic stromal cells by CD45 cell depletion using CD45 MicroBeads (Miltenyi Biotec, Bergisch Gladbach, Germany). Enriched stroma and the CD45+ fraction were counted, stained with gp38, CD31, CD45, and CD11c, and CD11b, and MHC class II, respectively, and analyzed by flow cytometry (CyAn ADP Flow Cytometer; Dako). Data analysis was performed using FlowJo software v9.2 (TreeStar, Ashland, OR).
In vitro Ag cross-presentation

To determine whether LECs can cross-present Ag in vitro, cells were plated at 5 × 10^5 well in 24-well plates and stimulated with 2.5 μM OVA257-264 (SIINFEKL), 2.5 μM NP-ss-COV A250-264, or equivalent concentrations of unconjugated NP for 18 h in medium buffered with 25 mM HEPES (pH 7.4) at 4 or 37°C. Cell surface H2K-H2OVA257-264 complexes were detected with the Ab 25d1.16 using flow cytometry. To characterize the kinetics of OVA accumulation in LECs, cells were stimulated with 1 μM OVA-AF647 for up to 90 min, washed, and analyzed for OVA uptake by flow cytometry. To demonstrate CD8+ T cell priming, APCs were seeded at 10^4 cells/well for up to 90 min, washed, and analyzed for OVA uptake by flow cytometry.

In vitro T cell coculture assays

To determine the outcome of CD8+ T cell interaction with cross-presenting LECs, we performed coculture assays. CD8α+ T cells were purified from the spleen of an OT-I mouse by negative selection (CD8α Kit II; Miltenyi Biotec). For LEC–T cell or DC–T cell coculture studies, 10^4 LECs or DCs were cocultured with naïve CD8+ T cells from OT-I mice (1:10 ratio) in 96-well round-bottom (BMDC) or flat-bottom (LEC) plates.

Statistical analysis

Statistical analysis was performed using one-way ANOVA, followed by the Bonferroni posttest, with Prism software (GraphPad, San Diego, CA) unless otherwise stated. Results are shown as mean ± SD, with significance indicated as *p ≤ 0.05, **p ≤ 0.01, and ***p ≤ 0.001.

Results

LECs scavenge exogenous Ag in vivo and in vitro

Although LECs transport Ags from the periphery to the lymph, we asked whether they also could scavenge and process Ags. To this end, we injected fluorescently labeled OVA protein (OVA-AF647) i.d. in the forearm and observed its distribution in the brachial-draining LNs after 90 min. The use of a foreign protein allowed us to determine specifically the immune response against an exogenous versus self-expressed TTA. Using confocal microscopy of thick sections of the brachial LN, we observed OVA in the lymphatic-rich, LYVE-1+ sinuses of the LN (Fig. 1A). Upon magnification of the LYVE-1+ regions, we observed that much of the OVA was contained within LYVE-1+ cells, suggesting intracellular accumulation in LECs (Fig. 1B, 1C). Flow cytometric analysis validated the observed scavenger activity and demonstrated that LN LECs (CD45+gp38+CD31+LYVE-1+ cells), as well as professional APCs, contained soluble OVA (Fig. 1D, 1E, Supplemental Fig. 1). Among the CD45+ stromal cells, LECs took up the most OVA (50 ± 8%). When considered as a percentage of each cell population that took up OVA, LECs were on par with DCs for their scavenging ability (30 ± 20% versus 30 ± 5%, respectively).

In vitro, we could follow the accumulation of fluorescent OVA by iLECs (18). The degree of OVA-AF647 accumulation by iLECs was similar to that of BMDCs over 90 min at 37°C, reaching a plateau within 40 min, as observed by flow cytometry (Fig. 1F, 1G, Supplemental Fig. 1C). This exogenous Ag uptake was an active or energy-dependent process, because OVA uptake at 4°C by iLECs was minimal compared with that at 37°C (Fig. 1G, Supplemental Fig. 1D). These results confirm that exogenous proteins are actively scavenged by LECs, both in vivo and in vitro.

LECs process and route Ag for cross-presentation on MHC class I in a TAP1-dependent manner

Accumulation of exogenous proteins inside LECs allows for the possibility of Ag processing and cross-presentation on MHC class I by these cells. We asked whether hallmarks of cross-presentation could be observed in LECs under controlled in vitro conditions.

First, we determined whether uptake of exogenous Ags could lead to peptide loading onto MHC class I molecules and presentation on the cell surface by immunostaining cells with the mAb 25d1.16, which specifically binds the MHC class I–bound CD8+ dominant epitope of OVA, SIINFEKL (OVA257-264). To avoid SIINFEKL peptide binding directly to surface MHC class I, and thus bypass the need for intracellular processing and cross-presentation, we used a N-terminally elongated SIINFEKL peptide conjugated onto synthetic poly(propylene sulfide) 30-nm NPs, a tool that we recently developed in our laboratory for more efficient SIINFEKL/MHC class I cross-presentation by 25d1.16 compared with OVA (22). Although SIINFEKL peptide can bind to MHC class I without cell internalization and processing, this 16-aa peptide on the NPs (NP-ss-COV A250-264) minimally binds to surface MHC class I, and thus requires uptake and intracellular processing for MHC class I loading in BMDCs (22). Exogenously applied NP-ss-COV A250-264 resulted in the detection of MHC class I peptide complexes in an energy-dependent manner in both iLECs and ex vivo–cultured primary LN LECs (Fig. 2A, 2B).

By conducting this study at both 4 and 37°C, we confirmed that the cross-presentation of NP-ss-COV A250-264 by LECs requires active processing, because cells that received SIINFEKL, but not NP-bound peptide Ag, showed elevated 25d1.16 staining at 4°C (Fig. 2A, 2B).

We then applied inhibitors of Ag uptake and intracellular trafficking to elucidate the relevant steps in LEC cross-presentation. We cocultured inhibitor-treated, Ag-loaded LECs with OT-I CD8+ T cells and assessed T cell proliferation by CFSE dilution as a measure of MHC class I/SIINFEKL presentation on the LEC surface. To confirm that LEC-induced T cell stimulation was dependent on intracellular uptake of NP-ss-COV A250-264, we pretreated iLECs with dynasore, an inhibitor of dynamin that affects both clathrin- and caveolin-mediated uptake (24), or LY294002, a PI3K inhibitor that affects macropinocytosis (25). Both inhibitors led to reduced OT-I T cell proliferation in a concentration-dependent manner when LECs were treated with NP-conjugated Ag, but not free SIINFEKL peptide (Fig. 2C), confirming active uptake mechanisms contributing to MHC class I presentation.

We next asked whether intracellular transport processes were important in LEC cross-presentation. To this end, we treated iLECs with BFA, which inhibits Ag transport from the ER to golgi (26), or chloroquine, which inhibits acidification and vesicle fusion to late endosomes/lysosomes (27). When LECs were pretreated with either of these agents, it also resulted in concentration-dependent inhibition of T cell proliferation (Fig. 2C).
Because ER–golgi transport, as well as endosome acidification, was observed to be important in cross-presentation by LECs, we next asked whether exogenous Ag processing in LECs depends on the canonical TAP1 pathway, in which cytoplasmic peptide fragments are loaded onto MHC class I in the ER after translocation by TAP1 (28). Using LN LECs and DCs isolated from TAP1-null mice exposed to whole OVA protein, we found a substantial reduction in OT-I T cell proliferation after coculture compared with those exposed to LECs or DCs isolated from wild-type (WT) mice (Fig. 2D). As expected, T cell proliferation was not significantly altered between WT and TAP1-null mice with SIINFEKL stimulation, which binds externally to MHC class I, suggesting that the density of SIINFEKL/MHC class I complexes on LECs derived from both strains were comparable (Fig. 2D, Supplemental Fig. 2). Together, these data suggest that cross-presentation pathways are active in LECs.
tracellular compartments. Subsequently, LECs efficiently process Ags for cross-presentation through TAP1-dependent cytoplasmic–ER import of peptides. Both ER–golgi transport of peptide-loaded MHC class I and endosome acidification–dependent MHC class I trafficking are important for LEC cross-presentation. This suggests that some internalized Ags traffic and are loaded onto MHC class I through acidified vesicles, whereas others reach the cytosol to be imported by TAP1 for loading onto MHC class I (29).

**Direct Ag-specific CD8⁺ T cell interactions drive upregulation of MHC class I and PD-L1 on LECs**

Having demonstrated the scavenger activity of LECs and efficient processing and cross-presentation of exogenous Ags, we next explored the costimulatory functions of steady-state LECs in the presence of naive CD8⁺ T cells. We compared LEC expression of Ag-presentation molecules and costimulatory molecules with those of professional APCs (DC). As expected, DCs clearly demonstrated constitutive expression of CD40, CD86, CD80, and MHC class I that were further upregulated upon addition of the mature epitope peptide SIINFEKL (Fig. 3A, lower panels, OT-I and OT-I + SIINFEKL peptide, respectively). The increase in the expression levels of costimulatory molecules and receptors on the DC surface upon Ag-specific interactions with OT-I CD8⁺ T cells might appear surprising in the absence of TLR stimulation. However, TLR-independent pathways exist that can drive maturation, and it was reported that cognate interactions between DCs and CD8⁺ T cells alone can induce upregulation of CD80 and CD86 expression on DCs (30).
Furthermore, the transport of peptide-loaded MHC class I to the cell surface was suggested to be accompanied by an increased expression of costimulatory molecules (31). Thus, peptide loading of MHC class I and subsequent engagement of TCR and T cell activation can indirectly upregulate costimulatory molecules on DCs. Because our peptide was not contaminated with endotoxin, and the observed changes in maturation markers were not induced when DCs were incubated with the cognate peptide in the absence of CD8+ T cells (Fig. 3A, lower panels, SIINFEKL), our data suggest that Ag presentation by the APC and subsequent recognition by the T cell leads to the altered expression. In contrast to expression by DCs, ex vivo–cultured primary LN LECs expressed low levels of CD40 and CD80 and undetectable levels of CD86 in either the presence or absence of Ag-specific interactions with CD8+ T cells (Fig. 3A, upper panels). Similarly, constitutive expression of costimulatory molecules in human LECs was lower than that of human blood-derived DCs (data not shown). However, LECs significantly upregulated MHC class I (*p* < 0.01) in a manner that was dependent on Ag-specific CD8+ T cell interactions (Fig. 3). Consistent with previous reports (32, 33), both LECs and DCs constitutively expressed the inhibitory ligand PD-L1 (or B7.H-1, Fig. 3A), which was reported to attenuate T cell proliferation and abrogate effector T cell differentiation (34). Furthermore, both LECs and DCs expressed higher levels of costimulatory molecules in human LECs was lower than that of human blood-derived DCs (data not shown). However, LECs significantly upregulated MHC class I (*p* < 0.01) in a manner that was dependent on Ag-specific CD8+ T cell interactions (Fig. 3).

More importantly, this change was not accompanied by increased costimulatory molecule expression in LECs, as it clearly was in DCs. The same trends also were observed in cultured iLECs (data not shown). Collectively, these data demonstrate an evidently different balance between costimulatory and coinhibitory ligand expression in LECs versus DCs and further suggest that Ag-specific interactions between LECs and CD8+ T cells result in dynamic regulation of the LEC phenotype to favor coinhibitory signaling.

**Cross-presentation of exogenous Ag by LECs leads to impaired activation of naive CD8+ T cells in an Ag-specific manner**

Having shown that Ag-presenting LECs can upregulate PD-L1 in the presence of Ag-specific CD8+ T cells in vitro, we next asked whether cross-presentation by LECs and engagement of Ag-specific TCRs could lead to a tolerized phenotype of CD8+ T cells under steady-state conditions. To this end, we investigated the functional capacity of CD8+ T cells after cross-priming by LECs compared with cross-priming by DCs in vitro. Upon incubation with iLECs in the presence of 1 nM NP-ss-COV A250–264, OT-I CD8+ T cells proliferated strongly; however, these iLEC-primed CD8+ T cells displayed a dysfunctionally activated phenotype characterized by high levels of the apoptotic marker annexin V in early generations of proliferating T cells compared with DC-stimulated T cells (Fig. 4A). The upregulation of PD-L1 on LECs upon Ag-specific interactions with CD8+ T cells (Fig. 3) led us to analyze the expression of PD-L1–binding partners PD-1 and CD80 on cocultured CD8+ T cells. PD-1 is a member of the B7/CD28 superfamily and plays...
a central role in the regulation of T cell immunity; its activation results in decreased proliferation, reduced IFN-γ and IL-2 production, and increased apoptosis (34). We observed that PD-1 expression was consistently high on iLEC-stimulated CD8⁺ T cells from the early proliferative generations, whereas only later generations of DC-stimulated CD8⁺ T cells expressed elevated levels of PD-1 (Fig. 4A). In addition to PD-1, recent studies showed that PD-L1 binds CD80 at a distinct site (35) to deliver inhibitory signals to T cells (36); in those studies, CD80 expression was observed on anergic T cells and was further upregulated after re-exposure to the Ag. In contrast to DC-stimulated CD8⁺ T cells, we detected a high percentage of CD80 expression on proliferating iLEC-primed CD8⁺ T cells (Fig. 4A). In addition to PD-1, CTLA-4 was substantially upregulated in early generations of CD8⁺ T cells stimulated by LECs versus DCs (Fig. 4A). CTLA-4 is another member of the CD28/B7 superfamily, which is implicated in tolerogenic responses with a distinct, nonredundant regulatory role (34, 37), and competes with CD28 for binding to CD80 and CD86 on APCs to impede costimulatory signaling and increase CD86 degradation, resulting in impaired T cell activation (38). CTLA-4 also disrupts positive signaling through recruitment of phosphatases to the immunological synapse and subsequent dephosphorylation of key signaling molecules without direct engagement to CD80 and CD86 (38). In addition to CTLA-4 upregulation, we found reduced expression of the surface activation markers CD25, CD44, and CD69 in iLEC-primed T cells compared with DC-primed T cells (Fig. 4B). Finally, OT-I CD8⁺ T cells primed with iLECs in the presence of NP-ss-COVA250-264 produced significantly less IFN-γ and IL-2 compared with T cells primed with DCs (Fig. 4C).

Taken together, these data indicate that iLECs can efficiently cross-present Ag and directly interact with CD8⁺ T cells to induce Ag-specific proliferating T cells with a tolerized phenotype in vitro. The functional outcome of T cell priming differs significantly from that of T cells primed by conventional APCs, suggesting a tolerizing role for LECs under steady-state conditions.

IL-2 does not rescue the dysfunctional phenotype of CD8⁺ T cells activated by LECs

Because we observed diminished levels of IL-2 production by LEC-stimulated versus DC-stimulated T cells (Fig. 4C), and because IL-2 is essential for CD8⁺ T cell expansion, we asked whether the T cell phenotype could be rescued by exogenous IL-2, as was shown for exhausted T cells in chronic viral infection (39). Interestingly, supplementation of the iLEC–T cell cocultures with IL-2 (50 U/ml) resulted in increased expression of the activation markers CD25, CD44, and CD69 on CD8⁺ T cells (Fig. 5A). However, no effect was seen on T cell proliferation or IFN-γ production (Fig. 5B), nor did IL-2 significantly decrease the percentages of annexin V⁺ or PD-1⁺ cells per generation (Fig. 5C). These trends were similar when LN LECs were treated with NP-ss-COVA250-264 and when iLECs were stimulated with 1 μM OVA protein instead of NP-ss-COVA250-264 (data not shown). Together, these data suggest that CD8⁺ T cells cross-primed by LECs were not merely exhausted, because they could not be rescued by IL-2.

**Discussion**

In addition to carrying Ags to the LN for uptake by immature DCs for immune surveillance, this study highlights an important role for lymphatic drainage in the maintenance of peripheral tolerance: the constant exposure of LECs to lymph-borne peripheral Ags, which they scavenge and cross-present for tolerance induction under steady-state conditions. Ag cross-presentation was dependent on ER–golgi trafficking, endosome acidification, and TAP1. We observed that Ag-specific interactions with CD8⁺ T cells resulted in upregulation of MHC class I and PD-1 on LECs and dysfunctional activation of CD8⁺ T cells, which displayed early apoptosis and diminished cytokine production. Thus, in addition to the previously described role of LECs in presenting endogenous PTAs for autoreactive CD8⁺ T cell deletion (6–8), the current study demonstrates that LECs can efficiently scavenge and cross-present foreign Ags draining from the periphery; thus, they play an immunoprotective role against a broader range of peripheral Ags.

Until recently, the cross-presentation mechanism in the induction of peripheral tolerance to exogenous Ags by nonhematopoietic stromal cells has been almost exclusively attributed to LSECs,
which line the hepatic sinusoidal wall and come into close contact with foreign Ags and leukocytes passing through the liver (40). Similarly, LECs are positioned in a strategic anatomical site where crucial interactions determining the fate of an immune response take place. In this study, we demonstrated that Ag scavenging by LECs occurs under noninflammatory steady-state conditions, adding to our previous observation that tumor-associated LECs cross-present OVA expressed by B16-F10 tumors (16). Murine LECs in the skin-draining LNs actively took up peripherally administered OVA protein under steady-state conditions (Fig. 1A–C), consistent with an earlier study (41), and could be mediated through LEC expression of the mannose receptor (42, 43). This is consistent with continual scavenging of inflammatory CC chemokines (44) by D6 on afferent and subcapsular LECs, which line the peripheral lymph entering the LNs. Although LSECs displayed several similarities to LSECs with regard to Ag processing and cross-presenting capacity, LECs displayed some phenotypic differences to LSECs. For example, under steady-state conditions, LSECs were shown to express the costimulatory molecules CD40, CD80, and CD86 (30), LECs lack an immunostimulatory phenotype with remarkably low expression of costimulatory molecules (Fig. 3). Because the endotoxin levels found in portal blood under physiological conditions are presumably higher than in peripheral lymph, it is not surprising that the steady-state set point of costimulatory molecules in LSECs is higher than in LECs. The inability of LECs to upregulate the costimulation machinery, together with PD-L1 upregulation upon Ag-specific T cell interactions (Fig. 3, Supplemental Fig. 3), suggest that LEC cross-presentation may be nonactivating, because lack of costimulation is one mechanism of a dysfunctional CD8 T cell response that is reminiscent of the classical mechanism of peripheral tolerance induction by immature DCs under noninflammatory conditions by T cell anergy and deletion (54).

The functional discrepancy and fate of LEC-primed versus DC-primed CD8 T cells (Fig. 4C) was coupled with the reciprocal upregulation of inhibitory molecular partners on CD8 T cells: the PD-L1 partners PD-1 and CD80, as well as CTLA-4 (Fig. 4A). In accordance with our observations, Tewalt et al. (6) recently demonstrated a key role for the PD-L1/PD-1–signaling pathway in the absence of costimulation in LEC-induced peripheral tolerance of endogenously expressed PTAs, where blocking PD-1 in LEC-educated, tyrosinase-specific CD8 T cells resulted in autoimmune vitiligo. Although exogenous IL-2 compensated for PD-L1–mediated coinhibitory signaling in the absence of costimulatory molecules in LSEC–T cell cocultures (55), supplementation of LEC–T cell cocultures with IL-2 did not alter the phenotype of LEC-primed CD8 T cells (Fig. 5). Our data suggest that other regulatory pathways might be involved, such as the engagement of PD-L1 by CD80 or signaling through CTLA-4, or additional ligands that are reported to be expressed on LEC surface, including the B and T lymphocyte attenuator molecule or the lymphocyte activation gene 3 (6). Interestingly, there may be more than one differentiation state of CD8 T cells; apparently tolerated LSEC cross-primed CD8 T cells (30), upon inflammatory recall, were capable of becoming effector cells, reminiscent of central memory T cell activity (56). This may also apply in steady-state LEC cross-primed CD8 T cells. Further detailed mechanistic studies must be conducted on the coordination (57) of Ag processing, presentation, and costimulatory/coinhibitory molecule pathways to shape the fate of these cells.

**FIGURE 5.** The LEC-educated T cell phenotype is only partially reversed by IL-2. Naive CFSE-labeled OVA-specific OT-I CD8 T cells were cocultured with iLECs for 3 d in the presence of Ag (1 nM NP-ss-COVA250–264) and supplemented with 50 U/ml IL-2. (A) Representative flow cytometry graphs showing OT-I surface expression of activation markers after 3 d of priming by iLECs in the absence or presence of IL-2. Data are representative of three independent experiments (n = 4 each). (B) Division index of proliferating OT-I CD8 T cells (left panel) and IFN-γ release (right panel) were affected only slightly by IL-2. Data are mean ± SD from one representative of four independent experiments (n = 4 each). (C) Percentages of annexin V+ and PD-1+ OT-I CD8 T cells/generation after 3 d of coculture with iLECs are unaffected by IL-2. Data are mean ± SD from two independent experiments (n = 7 each).
Because LECs are situated in one of the prime anatomical sites for immunological sampling, our findings support the idea that organ-draining LECs are the first to sample and present the exogenous peptides (3, 58), proteins, and particulates present in lymph. Thus, LECs may play an important role in the context of immunomodulation. Several findings from the literature support this concept that flow from the periphery and the presence of lymphatics are important in shaping the adaptive immune responses in the LN. For example, Friedlaender and Baer (59) showed in 1972 that skin missing lymphatic connections was more readily sensitized to dyes than was intact skin, suggesting that the presence of lymphatics in intact skin contributes to a dampened delayed-type hypersensitivity response. Using the K14–VEGFR-3–Ig mouse model, we previously showed how impaired lymph drainage and the absence of dermal LECs resulted in impaired acquired tolerance to contact hypersensitivity, although these mice could mount a systemic T cell response (60). In the experimental autoimmune encephalomyelitis model, impaired lymphatic contraction and fluid drainage were reported to result in an autoimmune response (61). Together, this suggests that impaired lymphatic drainage translates into an inappropriately activated immune response. Reciprocally, steady-state drainage of Ag to LN seems to favor tolerogenic responses. For example, the clinical success of allergen-specific immune therapy is based on a regimen of long-term s.c. low-dose allergen injection (62). We showed that VEGF-C–expanded tumor-draining LN LECs impede a robust Ag-specific CD8 T cell response, which promotes tumor growth (16). In keeping with the idea that peripheral tolerance requires persistent Ag (63), continuous access to draining peripheral Ags newly establishes LECs as active players in the maintenance of a tolerogenic LN environment. It implies that, after injury or infection, when self-Ags drain to the LN, together with TLRs and other danger signals, the steady-state tolerization by LECs can act as a dampening mechanism to prevent later potential autoimmune reactions. In other words, LEC cross-presentation of draining Ags helps to amplify the signal-to-noise ratio between dangerous Ags and those that have been encountered under steady-state conditions. This work demonstrates that priming by LECs, via direct cross-presentation of scavenged exogenous Ags, has a tolerizing effect on CD8 T cells. In addition to T cell tolerization against LEC-presentation of scavenged exogenous Ags, proteins, and particulates present in the endoplasmic reticulum.

References
15. von Oppen, N., A. Schurich, S. Hegenbarth, D. Stabenow, R. Tolba, W. Olszewski, N. E. Miller, W. A. Muller, and Z. A. Cohn. 1983. Endo-
16. Steinman, R. M., I. S. Mellman, W. A. Muller, and Z. A. Cohn. 1983. Endo-
17. Steinman, R. M., I. S. Mellman, W. A. Muller, and Z. A. Cohn. 1983. Endo-
Supplementary Figure 1. Gating strategy and representative flow cytometry plots for assessing OVA uptake by cells in the lymph node and for showing intracellular accumulation of fluorescent OVA in cultured cells. 90 min after i.d. injection of OVA AF-647, brachial lymph nodes were digested and the single cell preparation was first magnetically separated using anti-CD45 microbeads before analyzing each fraction by flow cytometry for data shown in Fig. 1D-E. (A) Using CD45 staining, any remaining CD45+ cells in the magnetically sorted CD45- fraction were excluded from the analysis before the OVA+ subsets were determined for the different populations, with the use of a naive non-injected control. The CD45+ OVA+ populations were subsequently further subdivided in the 4 stromal subsets using the gp38, CD31 antibodies. The CD45+OVA+ population was further subdivided in different APC populations using the CD11b, CD11c and MHC II surface markers. In order to determine the percentage of each cell population out of all OVA positive cells, the counts of OVA+ gp38+CD31+ gated LECs, gp38+CD31- gated fibroblastic reticular cells (FRCs), gp38-CD31+ gated blood endothelial cells (BECs), gp38-CD31- gated double negative cells (DNs) in the CD45+ fraction and OVA+ APCs in the CD45- fraction were separately calculated and subsequently, divided to the overall (added) counts of OVA+ cells in both fractions. Here, the gating for the resident CD11b-CD11c+MHC IImed DCs is shown. (B) The CD45+ stroma was divided in the four LN stromal cell subpopulations using the gp38, CD31 antibodies and the fluorescence of OVA was determined in the LEC (gp38+CD31+) population. LECs from a naive non-injected mouse (no OVA) were used as a reference for the OVA gating. The CD45+ fraction was divided in CD11b- and CD11b+ cells and the CD11b+ subpopulation was further separated with the use of MHC II and CD11c surface markers in the MHC IImedCD11c+ DC subpopulations. The fluorescence of OVA was determined for the resident CD11b-CD11c+ MHC IImed DC subpopulation. The same DC subset from a naive non-injected mouse (no OVA) was used as a reference for the OVA gating. Representative flow cytometry plots showing intracellular accumulation of fluorescent ovalbumin (OVA) in cultured cells. (C) Representative dot plots of immortalized lymphatic endothelial cells (iLEC) or bone marrow-derived dendritic cells (DC) after incubation with 1μM OVA AF-647 for the data shown in Fig. 1F-G. Shown on the far right is the no-antigen control (No Ag) used to determine the positive OVA gating. (D) Representative histograms for the OVA fluorescence are shown for iLECs (top) and DCS (bottom) at 4oC versus 37oC showing different time points as indicated. Data shown are representative of 2 independent experiments, n=3 each.
Supplementary Figure 2. TAP1⁻/⁻ LECs can equally trigger OT-I proliferation at 1 and 5µM SIINFEKL compared to WT LECs despite lower MHCI levels. (A) MHC I levels by flow cytometry analysis, with isotype control (filled histogram). Lymph node lymphatic endothelial cells (LN LEC) and bone marrow derived dendritic cells (DC) from wild type C57Bl/6 (WT) or TAP1⁻/⁻ mice. Representative histograms of n=3 mice. (B) CFSE labeled OT-I CD8 T cells were co-cultured with LN LECs or DCs derived from WT or TAP1⁻/⁻ mice, pulsed with no peptide to 0.001, 0.01, 0.1, 1, and 5µM SIINFEKL peptide. Histogram of CFSE dilution after 3 days shows LEC and OT-I T cell co-cultures, with gray filled histogram representing no peptide, and the respective concentrations from light to dark gray. (C) Percent OT-I proliferation. n=3 in 2 independent experiments.
Supplementary Figure 3. MHC I and PD-L1 expression levels in the presence versus absence of antigen are higher in LN LEC compared to DC. Naïve OVA-specific OT-I CD8⁺ T cells were co-cultured with ex-vivo expanded lymph node lymphatic endothelial cells (LN LEC) or bone marrow derived DCs (DC) from C57/Bl6 mice in the presence (+ SIINFEKL) or absence (no SIINFEKL) of 1nM immunodominant MHC I peptide of OVA (SIINFEKL). After 24h or 72h of T cell/LEC, or after 24h of T cell/DC co-culture, the expression levels of costimulatory molecules CD40, CD86 (B7-2), CD80 (B7-1), major histocompatibility complex class I (MHC I) and Programmed Death Ligand 1 (PD-L1 or B7-H1) were determined by flow cytometric analysis. The Mean Fluorescent Intensity (MFI) was determined for CD40, CD86, CD80, MHC I and PD-L1 in the respective positive populations in LN LECs and DCs. Normalized MFI was calculated as follows: (MFI⁺[SIINFEKL]) / (MFI⁻[SIINFEKL]). Values depict mean ± SD from two representative experiment with n=3-4. ** P<0.01 and * P<0.05, two-way ANOVA followed by Bonferroni post-test.
Supplementary Figure 4. Ovalbumin colocalization analysis shows clathrin colocalization is not by chance, and that OVA can be found within the LAMP1 vesicles. (A, C) Cells were exposed to AF647-labeled OVA for 15 min or 90 min and stained for clathrin heavy chain or Lysosomal-Associated Membrane Protein 1 (LAMP-1), respectively. Images were acquired by confocal microscopy. Arrowheads indicate co-localizing pixels. The regions of interest shown to the right are indicated as white squares on the cell. The scale bar corresponds to 4 and 2μm, respectively. (B) Object-based colocalization method of Fletcher et al. (2010) was used to determine that those OVA+ vesicles, which are also clathrin+ occur more than by chance by comparison to a randomly generated spots. % relative frequency is plotted against %OVA+ vesicles. 14 images were analyzed. (D) A typical Imaris analysis for quantifying LAMP-1 enclosed OVA. Spots (red) were assigned to OVA fluorescence maxima, and the white surfaces were drawn around LAMP-1 fluorescence structures. The ImarisXT plugin “split spots into surface objects” was used to determine the % OVA inside LAMP-1+ surfaces. Scale bar = 3μm. (E) The result of such analysis for 5 images shows that approximately 30% of OVA can be found within LAMP-1+ vesicles.