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J Immunol published online 9 December 2013 http://www.jimmunol.org/content/early/2013/12/08/jimmunol.1301513

Supplementary Material http://www.jimmunol.org/content/suppl/2013/12/09/jimmunol.1301513.DC1

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Therapeutic Efficacy of Suppressing the JAK/STAT Pathway in Multiple Models of Experimental Autoimmune Encephalomyelitis

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Pathogenic Th cells and myeloid cells are involved in the pathogenesis of multiple sclerosis (MS) and experimental autoimmune encephalomyelitis (EAE), an animal model of MS. The JAK/STAT pathway is used by numerous cytokines for signaling and is critical for development, regulation, and termination of immune responses. Dysregulation of the JAK/STAT pathway has pathological implications in autoimmune and neuroinflammatory diseases. Many of the cytokines involved in MS/EAE, including IL-6, IL-12, IL-23, IFN-γ, and GM-CSF, use the JAK/STAT pathway to induce biological responses. Thus, targeting JAKs has implications for treating autoimmune inflammation of the brain. We have used AZD1480, a JAK1/2 inhibitor, to investigate the therapeutic potential of inhibiting the JAK/STAT pathway in models of EAE. AZD1480 treatment inhibits disease severity in myelin oligodendrocyte glycoprotein-induced classical and atypical EAE models by preventing entry of immune cells into the brain, suppressing differentiation of Th1 and Th17 cells, deactivating myeloid cells, inhibiting STAT activation in the brain, and reducing expression of proinflammatory cytokines and chemokines. Treatment of SJL/J mice with AZD1480 delays disease onset of PLP-induced relapsing-remitting disease, reduces relapses and diminishes clinical severity. AZD1480 treatment was also effective in reducing ongoing paralysis induced by adoptive transfer of either pathogenic Th1 or Th17 cells. In vivo AZD1480 treatment impairs both the priming and expansion of T cells and attenuates Ag presentation functions of myeloid cells. Inhibition of the JAK/STAT pathway has clinical efficacy in multiple preclinical models of MS, suggesting the feasibility of the JAK/STAT pathway as a target for neuroinflammatory diseases. The Journal of Immunology, 2014, 192: 000–000.

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Multiple sclerosis (MS) is an autoimmune disease of the CNS characterized by demyelination, inflammatory lesions, axonal damage, activation of IFN-γ-producing Th1 cells and IL-17–producing Th17 cells, inappropriate activation of innate immune cells (macrophages, dendritic cells [DCs], neutrophils, and microglia), and aberrant production of cytokines/chemokines (1, 2). Th1 cells, Th17 cells, and innate immune cells are also implicated in experimental autoimmune encephalomyelitis (EAE), an animal model of MS (3, 4). The pathogenesis of MS and EAE is associated with the overexpression of cytokines

Received for publication June 10, 2013. Accepted for publication October 25, 2013.

This work was supported in part by National Institutes of Health Grants NS45290 and NS57563 (to E.N.B.); NS64261 (to P.D.S.); DK84082 (to L.E.H.); AI27767 (to S.S.); and AI76562 (to C.R.); National Multiple Sclerosis Society Grants RG-3891 (to C.R.); CA-1059-A-13 (to E.N.B.); and RG-4885-A-14 (to E.N.B.); and a grant from the American Brain Tumor Foundation (to B.C.M.).

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The online version of this article contains supplemental material.

Abbreviations used in this article: BMDM, bone marrow–derived macrophage; DC, dendritic cell; EAE, experiment autoimmune encephalomyelitis; EdU, 5-ethynyl-2′-deoxyuridine; LFB, luxol fast blue; MOG, myelin oligodendrocyte glycoprotein; MS, multiple sclerosis; qRT-PCR, quantitative RT-PCR; RR, relapsing-remitting; SOCS, suppressor of cytokine signaling; UN, unimmunized.

Copyright © 2013 by The American Association of Immunologists, Inc. 0022-1767/13/S16.00 including IL-12, IFN-γ, IL-6, IL-21, and IL-23, which function in part to promote differentiation of effector Th1 and Th17 cells (1, 3, 5, 6).

The JAK/STAT signaling pathway is used by numerous cytokines and is critical for initiating innate immunity, orchestrating adaptive immunity, and ultimately constraining immune responses (7). Cytokines are of paramount importance in regulating the development, differentiation, and function of myeloid cells and T cells (8, 9); thus, unrestrained activation of the JAK/STAT pathway has pathological implications for autoimmune diseases (7, 10, 11). In MS and EAE, there is evidence for aberrant functionality of the JAK/STAT pathway. T cells and monocytes from MS patients during relapse have elevated levels of activated STAT3 compared with cells from patients in remission (12), and high levels of activated STAT3 in T cells from patients with clinically isolated syndrome predict conversion to clinically defined MS (13). In EAE, IL-6 has a deleterious role by activation of STAT3, which is pivotal for induction of pathogenic Th17 cells (14–16). Loss of STAT3 in T cells renders mice resistant to EAE disease (17, 18). STAT target genes, including IL-23R, IL-6, IL-17A, and IL-17F, are implicated in contributing to MS and EAE. The JAK/STAT pathway has received attention as a therapeutic target in autoimmune diseases and cancers (7, 11). JAK inhibitors have demonstrated clinical efficacy in rheumatoid arthritis and other inflammatory diseases (19–21). Indeed, Bright et al. (22) previously demonstrated that tyrphostin B42, a JAK2 inhibitor, reduced severity of EAE. JAK inhibitors interrupt signaling downstream of multiple cytokines, a useful approach for EAE and MS, which are characterized by a “cytokine storm” in the periphery and CNS. Simultaneous inhibition of cytokine signaling by JAK inhibitors
may break the cycle of inflammation characteristic of neuroinflammatory diseases. AZD1480, an ATP competitive inhibitor of JAK1 and JAK2, has beneficial effects in cancer models by suppressing downstream activation of STATs, particularly STAT3 (23, 24). We demonstrate that AZD1480 is effective in suppressing clinical symptoms in five preclinical models of MS. AZD1480 treatment was associated with diminished STAT activation in the CNS, reduced pathogenic Th1 and Th17 cell responses, alterations in DC and macrophage functionality, decreased infiltration of immune cells into the CNS, reduced demyelination, and suppression of proinflammatory cytokine/chemokine expression in vivo. Using AZD1480 for proof-of-principle, we demonstrate that inhibiting the JAK/STAT pathway has striking clinical efficacy in multiple models of EAE.

Materials and Methods

**Mice**

C57BL/6 and myelin oligodendrocyte glycoprotein (MOG)35–55 TCR-transgenic 2D2 mice (25) were bred in the animal facility at the University of Alabama (Birmingham, AL). SJL/J mice were purchased from the National Cancer Institute–Frederick National Laboratory (Frederick, MD). Suppressor of cytokine signaling (Socs3)-floxed transgenic (Socs3fl/fl) mice (26) were gifts from Dr. W. Alexander (Walter and Eliza Hall Institute of Medical Research, Parkville, VIC, Australia) and were bred at the University of Alabama. Socs3 conditional knockout (LysMcCre-Socs3fl/fl) mice were generated by serial breeding of Socs3fl/fl mice with mice expressing a Cre-recombinase under the control of the Ly-5M promoter (27). All experiments were reviewed and approved by the institutional animal care and use committee of the University of Alabama.

**JAK inhibitor**

AZD1480 (Supplemental Fig. 1), a JAK1/JAK2 inhibitor, was synthesized and provided by AstraZeneca R&D (Waltham, MA) and resuspended in DMSO, as described previously (23, 24).

**Peptides, Abs, and cytokines**

MOG35–55 peptide was synthesized by New England Peptide, and PLP139–151 peptide was from CPC Scientific. LPS was from Sigma-Aldrich. Neutralizing Abs to IL-4 and IFN-γ; Abs against mouse CD3, B220, Gr-1, CD4, IFN-γ, and IL-17A; and human CD3, CD4, CD14, and HLA-DR used for flow cytometry are from eBioscience. Mouse IL-6, IFN-γ, IL-12, IL-23, and GM-CSF and human IFN-γ and TGF-β1 are from R&D Systems. Abs against mouse and human CD3 and CD28 are from BioLegend. Abs against phospho-STAT1 (Tyr701) and phospho-STAT3 (Tyr705) used for immunoblotting are from Cell Signaling Technology. Ab against GAPDH is from Abcam, and Abs against Ly-6C, CD11c, MHC class II, CD40, CD80, CD86, CD11b, and CD45 are from BD Pharmingen.

**EAE induction, assessment and treatment with AZD1480**

Active and adoptive transfer EAE was induced as described previously (27, 28). Eight- to 12-wk-old C57BL/6 or LysMcCre-Socs3fl/fl mice were immunized s.c. with 200 μg MOG35–55 emulsified in CFA (supplemented with 2 mg/ml Mycobacterium tuberculosis) and injected i.p. on days 0 and 2 with 500 ng pertussis toxin. For adoptive transfer EAE, C57BL/6 mice were immunized with MOG35–55 for 10 d, and then, splenic and axillary lymph nodes cells were isolated and restimulated with MOG35–55 (10 μg/ml) in vitro for 7 d. CD4+ T cells from Th1 cell differentiation or with 20 ng/ml IL-12 and 20 ng/ml IL-23, and 3 ng/ml TGF-β1 for Th17 cell differentiation for 3 d. Cells (30 × 105) were transferred into healthy recipients. The frequency of donor CD4+ T cells producing IL-17A or IFN-γ was assessed by FACS prior to transfer (data not shown). Relapsing–remitting (RR) EAE was induced as follows: female SJL/J mice (8–12 wk old) were immunized with a s.c. injection of 150 μg PLP199–211 peptide emulsified in CFA (29). Assessment of EAE severity was as follows: 0, no disease; 1, decreased tail tone; 2, hind-limb weakness or partial paralysis; 3, complete hind-limb paralysis; 4, front- and hind-limb paralysis; and 5, moribund state. Assessment of atypical EAE was done using the scoring system as described previously (27): 0, no disease; 1, hunched appearance, slight head tilt; 2, ataxia, scruffy coat; 3, severe head tilt, slight axial rotation, staggered walking; 4, severe axial rotation, spinning; and 5, moribund. For treatment of mice with active classical EAE, AZD1480 was administered at 25 mg/kg or DMSO (Sigma-Aldrich) as vehicle control i.p. daily from day 9 postimmunization for 10 d, or when disease score reached 1.5 for 12 d, and for atypical active EAE at day 5 postimmunization. For treatment of adoptive transfer EAE, AZD1480 was administered at 25 mg/kg or DMSO as vehicle control i.p. daily from day 7 (Th1) or day 8 (Th17) postimmunization for 10 d. For treatment of RR EAE, mice were administered AZD1480 (25 mg/kg) or vehicle control by oral gavage daily from day 7 postimmunization for 10 d, and again at day 60 for 10 d. Mice were weighed and examined daily for disease symptoms. Mice were killed and perfused, and mononuclear cells were isolated from the CNS (cerebellum and spinal cord) by a 30-min Percoll gradient or from draining cervical lymph nodes and spleen as previously described (27), and cell phenotype was determined by surface and intracellular staining by flow cytometry.

**Cell preparation**

Bone marrow cells were cultured with RPMI 1640 medium containing 10% FBS (v/v) and 20 ng/ml mouse M-CSF for 7 d to obtain bone marrow–derived macrophages (BMDMs) (30). Naive CD4+ T cells were obtained from spleen or lymph nodes of 8- to 12-wk-old mice and isolated by Dynabeads CD4 Positive Isolation Kit (Invitrogen). T cells were cultured in RPMI 1640 medium supplemented with 10% FBS (v/v), 2 mM glutamine, 100 IU/ml penicillin, 0.1 mg/ml streptomycin, 10 mM HEPES, 1 mM sodium pyruvate and nonessential amino acids, and 2 μM 2-ME.

**Human subjects**

Cryopreserved PBMC obtained by standard Histopaque density centrifugation from healthy donors were used in this study. These subjects were recruited from the Alabama Vaccine Research Clinic at the University of Alabama. To obtain monocytes, PBMC (3 × 106 cells/ml) were incubated in serum-free medium (RPMI 1640) supplemented with 2 mM glutamine, 100 IU/ml penicillin, 0.1 mg/ml streptomycin, and 10 mM HEPES for 3 h at 37°C. Adherent cells were obtained by washing four times with prewarmed serum-free medium. For T cells, nonadherent PBMC were collected after the 3-h period of plastic adherence and washed three times with serum-free medium. Written informed consent was obtained from all donors who participated in this study. The Institutional Review Board of the University of Alabama approved the study (IRB number X090708004).

**Stimulation of human PBMC**

For T cell stimulation, nonadherent PBMC were stimulated in vitro with anti-CD3 (5 μg/ml) and anti-CD28 (2 μg/ml) Abs in the absence or presence of AZD1480 (0.25 and 0.5 μM) for 4 d, and tryosine phosphorylation of STAT1, STAT3, and STAT4 was examined in gated CD3+CD4+ T cells. For human monocytes, adherent PBMCs were pretreated in the absence or presence of AZD1480 (0.25 and 0.5 μM) for 2 h and then stimulated with IFN-γ (100 U/ml) for 1 h. The extent of STAT1 and STAT3 tyrosine phosphorylation within the CD5+ CD14+ monocyte population was assessed by intracellular flow cytometry.

**In vitro Th cell differentiation**

Purified murine CD4+CD25+ T cells were stimulated with Abs to CD3 (1 μg/ml) and CD28 (1 μg/ml) under Th1 cell differentiation conditions (10 ng/ml IL-12 and 10 μg/ml anti-IL-4) and Th17 cell differentiation conditions (20 ng/ml IL-6, 10 ng/ml IL-23, 5 ng/ml TGF-β1, 10 μg/ml anti-IL-4, and 10 μg/ml anti-IFN-γ) in the presence of vehicle control (DMSO) or AZD1480. For Ag-specific differentiation, spleen and lymph nodes cells from MOG35–55-immunized mice were restimulated with MOG35–55 (10 μg/ml) for 3 d in Th1 or Th17 cell differentiation conditions in the absence or presence of AZD1480.

**Intracellular staining**

Cells were stimulated for 4 h with PMA/ionomycin (25 ng/ml and 1 μg/ml) plus GolgiStop (BD Pharmingen) and analyzed for intracellular production of cytokines by staining with anti-cytokine Abs and subsequent flow cytometry, as described previously (27). All flow data shown are representative of three independent experiments.

**Immunohistology analysis**

Mice were killed and cerebellum and spinal cord immersion fixed in Bouin’s fixative and paraffin embedded. Sections were stained with H&E and luxol fast blue (LFB). Digital images were captured with a Zeiss.
Axiovision software. Images were collected at the same time using identical settings with respect to image exposure time and image compensation settings as described previously (27).

RNA isolation, RT-PCR, and TaqMan gene expression assays
Total RNA was isolated from the cerebellum and spinal cord of mice, and reverse transcription reactions performed as described previously (27). RNA (500 ng) was used to reverse transcribed into cDNA and subjected to quantitative RT-PCR (qRT-PCR). The data were analyzed using the comparative threshold cycle method to obtain relative quantitation values.

Immunoblotting
Thirty micrograms of cell lysate or brain homogenate was separated by electrophoresis and probed with Abs as described previously (27). All immunoblots are representative of three individual experiments.

Cell proliferation assay in vitro and in vivo
For in vitro proliferation, CFSE-labeled CD4+CD25– T cells were stimulated with anti-CD3 (5 μg/ml) and anti-CD28 (2 μg/ml) Abs, and cell proliferation was assessed by CFSE dilution detected by flow cytometric analysis 72 h poststimulation. For in vivo proliferation, C57BL/6 mice were immunized with MOG35–55 (200 μg) and then administrated vehicle or AZD1480 (25 mg/kg) at day 7 postimmunization. Mice were then injected with 5-ethynyl-2’-deoxyuridine (EdU) on day 13. CNS-infiltrating mononuclear cells were isolated from the cerebellum and spinal cord 12 h later, and EdU incorporation was detected for proliferation.

FIGURE 1. Inhibition of the JAK/STAT pathway suppresses Th1 and Th17 cell differentiation in vitro. Naive CD4+ T cells were isolated from the spleen of C57BL/6 mice. DCs were used as APCs at a 1:5 ratio to CD4+ T cells in coculture. Vehicle control or AZD1480 (0.25 μM) was added simultaneously to the Th1 or Th17 differentiation mixture. (A) Th1 cells were differentiated with anti-CD3 (1 μg/ml), anti-CD28 (1 μg/ml), and IL-12 (10 ng/ml) plus anti-IL-4 Ab (10 μg/ml) for 4 d. STAT1 and STAT4 tyrosine phosphorylation was examined by flow cytometry with isotype control shown in blue. (B) At day 4, cells were stimulated with PMA/ionomycin plus GolgiStop for 4 h and stained for the surface marker CD4 and by intracellular flow to detect IFN-γ expression. For mRNA expression, cells were collected at day 4, and mRNA was analyzed by qRT-PCR for IFN-γ and T-bet. (C) Th17 cells were differentiated with anti-CD3 (1 μg/ml), anti-CD28 (1 μg/ml), TGF-β (5 ng/ml), IL-6 (20 ng/ml), IL-23 (10 ng/ml) plus anti–IFN-γ Ab (10 μg/ml), and anti–IL-4 Ab (10 μg/ml) for 4 d. STAT3 tyrosine phosphorylation (red line) was examined by flow cytometry, with isotype control shown in blue. (D) At day 4, cells were stimulated with PMA/ionomycin plus GolgiStop for 4 h and stained for the surface marker CD4 and by intracellular flow to detect IL-17A expression. For mRNA expression, cells were collected at day 4, and mRNA was analyzed by qRT-PCR for IL-17A, RORγt, IL-22, and IL-23R. **p < 0.001.

Statistical analysis
Levels of significance for comparison between two groups were determined by one-sided two-sample Mann–Whitney rank-sum test and the Student t test distribution. A p value < 0.05 was considered statistically significant.

Results
AZD1480 inhibits Th1 and Th17 cell differentiation in vitro
AZD1480 potently inhibits the JAK/STAT pathway in tumor cells (23, 24) but has not been evaluated in primary immune cells. AZD1480 at 0.25 μM partially inhibits STAT1 and STAT4 tyrosine phosphorylation in T cells cultured in Th1 cell polarizing conditions (Fig. 1A). The differentiation of naive T cells to Th1 cells was strongly inhibited by AZD1480 treatment, as assessed by decreased IFN-γ production, and decreased mRNA levels of IFN-γ and T-bet (Fig. 1B). As some of the key cytokines for Th17 differentiation, including IL-6 and IL-23, signal through JAK1/2, we investigated the influence of inhibiting the JAK/STAT pathway on Th17 cell polarization. Using the Th17 cell differentiation mixture of IL-6, IL-23, and TGF-β, we observed that treatment with AZD1480 partially inhibited STAT3 tyrosine phosphorylation (Fig. 1C), which led to reduced differentiation of naive T cells to Th17 cells (Fig. 1D). AZD1480 treatment also inhibited mRNA levels of the STAT3 target genes IL-17A, RORγt, IL-22, and IL-23R.
23R (Fig. 1D). Th17 cells generated in the absence of TGF-β (with IL-6 + IL-23 or IL-6 + IL-23 + IL-1β), so called Th17 (23) cells, are more pathogenic in vivo (31). The differentiation of naïve T cells to Th17 cells in the presence of IL-6 + IL-23 (Supplemental Fig. 2A) or IL-6 + IL-23 + IL-1β (Supplemental Fig. 2B) also was substantially inhibited by AZD1480 treatment. Furthermore, the expression of STAT3 target genes under those differentiation conditions was inhibited by AZD1480 (Supplemental Fig. 2). These results clearly indicate that inhibition of the JAK/STAT pathway in T cells suppresses differentiation of Th1 and Th17 cells.

To assess whether the inhibitory effect of AZD1480 was a direct or indirect effect, short-term experiments were performed. As shown in Supplemental Fig. 3A and 3B, AZD1480 inhibited the phosphorylation of STAT1, JAK2, STAT3, and STAT5 in naïve T cells, suggesting a direct inhibitory effect on the JAK/STAT pathway.

**AZD1480 blocks JAK/STAT activation and gene expression in macrophages and DCs**

Many cytokines important in activating macrophages and DCs, including IFN-γ and GM-CSF, use the JAK/STAT pathway for signaling (7, 30). BMDMs were stimulated with IFN-γ, a key cytokine in polarizing macrophages to the proinflammatory M1 phenotype (30, 32). IFN-γ stimulation led to strong STAT1 and STAT3 tyrosine phosphorylation, which was inhibited by AZD1480 (treatment Fig. 2A). GM-CSF, another important cytokine in polarization of M1 macrophages and inflammatory DCs (32, 33), induced STAT5 tyrosine phosphorylation, and AZD1480 inhibited this response (Fig. 2B). To examine the effect of AZD1480 on upstream JAK activation, BMDM were stimulated with IL-6/soluble IL-6R to activate JAK2 and STAT3. AZD1480 pretreatment inhibited phosphorylation of both JAK2 and STAT3 (Fig. 2C). We examined the influence of AZD1480 on IL-4 signaling, which through activation of STAT6, induces polarization of macrophages to the M2 anti-inflammatory phenotype (34). IL-4 activation of STAT6 was largely unaffected by AZD1480 (Supplemental Fig. 3C), indicating that there are selective effects of AZD1480 on different STATs, dependent on cytokine stimulation. Interestingly, when BMDM were stimulated with LPS, rapid activation of the NF-κB pathway was not affected, whereas delayed activation of STAT1 and STAT3 at 2–4 h (via IFN induction) was inhibited (Fig. 2D), indicating the specificity of AZD1480 treatment.

NO is typically produced by M1 macrophages upon triggering of IFN-γ and TLR pathways (30). The expression of NO and its reactive derivative peroxynitrite has been implicated in the pathogenesis of MS and EAE (35, 36). Pretreatment of BMDM with AZD1480 significantly inhibited LPS/IFN-γ–induced production of nitrite (Fig. 2E). IFN-γ–inducible MHC class II expression is...
mediated by activation of STAT1 (37). AZD1480, in a dose-dependent manner, inhibited MHC class II expression in BMDM (Fig. 2F) and DCs (Fig. 2G). IFN-γ-inducible CD40 costimulatory molecule expression on DCs was also inhibited by AZD1480 (Fig. 2H). This suggests that AZD1480 may regulate the Ag presentation capability of these cells. Taken together, these findings indicate that AZD1480 inhibits JAK/STAT signaling in macrophages and DCs.

**Inhibition of the JAK/STAT pathway suppresses classical, atypical, and RR EAE**

Given the striking effect of AZD1480 in inhibiting Th1 and Th17 cell differentiation, as well as blocking IFN-γ-inducible gene expression in myeloid cells, we assessed the therapeutic potential of AZD1480 in EAE. First, we evaluated the effect of AZD1480 in naive mice by analyzing proliferation of naive CD4+ T cells and hematological parameters. Dosage was determined based on previous studies (23, 24) and on preliminary dose-response experiments (data not shown). No significant differences were noted in the proliferation of CD4+ T cells stimulated with anti-CD3 and anti-CD28, as well as in any hematological parameters in mice administrated AZD1480 (Fig. 3A, 3B). These results demonstrate that in vivo AZD1480 treatment does not induce toxicity when assessing circulating populations of the major cell types, consistent with studies using other JAK inhibitors, which are well-tolerated in vivo (21, 23, 38).

C57BL/6 mice were immunized with MOG35–55 peptide to induce classical EAE, and AZD1480 (25 mg/kg) or vehicle control was administrated i.p. daily for 10 d starting at the onset of disease on day 9. Animals were scored daily for signs of ascending motor paralysis graded on a scale of 0–5. Most strikingly, AZD1480 treatment resulted in a significant reduction in disease severity compared with vehicle treatment (Fig. 4A). Evaluation of the spinal cord revealed strong inhibition of STAT3 and STAT4 tyrosine phosphorylation, and a modest reduction in expression of STAT1 and STAT3 in the cerebellum and decreased total levels of STAT1 (Fig. 5B). Decreased infiltration of neutrophils, monocytes, microglia, DCs, and B cells in the cerebellum was observed (Fig. 5C), with reduced expression of Th1 markers (IFN-γ and T-bet) and Th17 markers (IL-17A), and reduced expression of cytokines/chemokines in the cerebellum comparable to unimmunized (UN) mice (Fig. 5D). Decreased inflammation and demyelination was observed in the cerebellum of AZD1480-treated mice compared with control mice (Fig. 5E). Oral gavage of AZD1480 produced a comparable protective effect in both classical and atypical EAE as i.p. treatment (data not shown). Collectively, these results indicate that AZD1480 has the desired, selective...
effect on its target (STATs) in the tissue region of interest (the brain) and that suppression of STAT activation by AZD1480 has a protective effect in classical and atypical EAE models.

To determine whether the protective effect of AZD1480 was specific to the MOG-C57BL/6 EAE model, we examined the RR model of EAE induced in SJL/J mice by PLP139–151. We observed that disease onset was significantly delayed with the first AZD1480 treatment at day 7, with no clinical symptoms until ∼day 23 (Fig. 6A). A second AZD1480 treatment at day 60 inhibited further relapse (Fig. 6A). Examination of spinal cord revealed inhibition of STAT1 and STAT3 tyrosine phosphorylation and a reduction in expression levels of STAT3 protein (Fig. 6B). AZD1480 treatment also inhibited IL-6, IL-12p40, and IL-23p19 mRNA expression in the spinal cord (Fig. 6C).

Inhibition of the JAK/STAT pathway influences the priming and expansion phases of EAE

Next, we examined the underlying cellular mechanisms leading to clinical improvement of actively induced EAE by AZD1480 treatment. Because MOG35–55-specific T cells are first primed to differentiate into specialized effector subsets in the periphery (3), the effect of AZD1480 on the priming phase of EAE was investigated. C57BL/6 mice were immunized with MOG35–55, and vehicle or AZD1480 (25 mg/kg) was administered at day 2 postimmunization for 3 d (Fig. 7A). Purified CD4+ T cells were isolated from the draining lymph nodes and spleen of these mice on day 10 postimmunization, restimulated with MOG35–55 peptide under Th1- and Th17-polarizing conditions, and the percentage of MOG35–55-specific Th1 and Th17 cells was measured by flow cytometry. Treatment with AZD1480 led to a significant decrease of MOG35–55-specific Th1 and Th17 cells in the draining lymph nodes and, to a lesser degree, in the spleen (Fig. 7B, 7C). Next, the effect of AZD1480 on expansion of MOG-specific T cells was examined. C57BL/6 mice were immunized with MOG35–55 and administrated vehicle or AZD1480 (25 mg/kg) at day 7 postimmunization for 3 d (Fig. 7D). Purified CD4+ T cells were isolated and stimulated as above. Decreased MOG35–55-specific Th1 cells in the draining lymph nodes were observed (Fig. 7E), and diminished percentages of MOG-specific Th17 cells were detected in draining lymph nodes and spleen (Fig. 7F). These results indicate that inhibition of the JAK/STAT pathway influences both the priming and expansion phases of EAE by suppressing MOG35–55-specific T cell responses in secondary lymphoid tissues.

JAK/STAT pathway inhibition suppresses proliferation of CD4+ T cells and CD11b+ cells

The decreased number of CNS-infiltrating cells in AZD1480 treated mice may be due to reduced cell proliferation; thus, the effect of AZD1480 on T cell and myeloid cell proliferation during EAE development was assessed. C57BL/6 mice were immunized with MOG35–55 and vehicle or AZD1480 (25 mg/kg) was administered at day 2 postimmunization for 3 d. Purified CD4+ T cells were isolated from the draining lymph nodes and spleen of these mice on day 10 postimmunization, restimulated with MOG35–55 peptide under Th1- and Th17-polarizing conditions, and the percentage of MOG35–55-specific T cells was measured by flow cytometry. Treatment with AZD1480 led to a significant decrease of MOG35–55-specific Th1 and Th17 cells in the draining lymph nodes and, to a lesser degree, in the spleen (Fig. 7B, 7C). Next, the effect of AZD1480 on expansion of MOG-specific T cells was examined. C57BL/6 mice were immunized with MOG35–55 and administrated vehicle or AZD1480 (25 mg/kg) at day 7 postimmunization for 3 d (Fig. 7D). Purified CD4+ T cells were isolated and stimulated as above. Decreased MOG35–55-specific Th1 cells in the draining lymph nodes were observed (Fig. 7E), and diminished percentages of MOG-specific Th17 cells were detected in draining lymph nodes and spleen (Fig. 7F). These results indicate that inhibition of the JAK/STAT pathway influences both the priming and expansion phases of EAE by suppressing MOG35–55-specific T cell responses in secondary lymphoid tissues.
with MOG<sub>35-55</sub> and then administrated vehicle or AZD1480 (25 mg/kg) at day 7 postimmunization. Mice were then injected with EdU on day 13 and sacrificed on day 14 (Fig. 7G). CNS-infiltrating mononuclear cells were isolated from the cerebellum of vehicle control or AZD1480-treated mice at day 14 after MOG immunization. Cells were stained with Abs to CD4, CD11b, Gr-1, CD45, CD11c, Ly-6C<sup>e</sup>, and B220, and the percentages of CD11b<sup>+</sup>/Gr-1<sup>+</sup> neutrophils, CD11b<sup>+</sup>/Ly-6C<sup>lo</sup> and CD11b<sup>+</sup>/Ly-6C<sup>hi</sup> macrophages, CD11b<sup>+</sup>/CD45<sup>int</sup> microglia, CD11c<sup>+</sup> DCs, CD4<sup>+</sup> T cells, and B220<sup>+</sup> B cells were gated. The absolute number of cells is shown. (D) Mice were perfused, the cerebellums were removed, and mRNA was analyzed by qRT-PCR. (E) Sections from the cerebellums of vehicle control or AZD1480-treated mice at day 14 were stained with H&E for inflammatory infiltrates and LFB for demyelination. Original magnification ×60. *p < 0.05, **p < 0.001.

The transfer of myelin-specific CD4<sup>+</sup> T cells induces EAE in naive recipients; thus, we tested whether AZD1480 treatment would be effective in treating EAE induced by adoptive transfer of pathogenic CD4<sup>+</sup> Th1 or Th17 cells. C57BL/6 mice were adoptively transferred with MOG<sub>35-55</sub>-specific Th1 or Th17 cells to induce EAE, and AZD1480 was administrated i.p. daily for 10 d starting at day 7 for Th1-induced EAE (Fig. 8A) or on day 8 for Th17-induced EAE (Fig. 8D). Notably, we observed a delay in onset of disease from days 10 to 14 and diminished disease severity in Th1-induced EAE (Fig. 8A). When AZD1480 treatment was discontinued, a protective effect was still noted at days 17 and 18 but was lost at days 19 and 20. A substantial reduction of STAT1 and STAT3 tyrosine phosphorylation in the spinal cord of AZD1480-treated mice was observed (Fig. 8B), and mRNA levels of proinflammatory cytokines were markedly decreased (Fig. 8C). AZD1480 treatment suppressed mRNA levels of IFN-γ and T-bet in Th1-induced EAE at the site of disease in the spinal cords of treated mice (Fig. 8C). It should be noted that IL-17A and ROR<gamma> mRNA expression was not detected in mice with Th1-induced EAE (data not shown). In Th17-induced EAE, disease onset and severity were significantly inhibited by AZD1480 treatment (Fig. 8D), although the mice developed clinical signs of EAE after AZD1480 withdrawal. STAT1 and STAT3 tyrosine phosphorylation was inhibited in Th17-induced EAE by AZD1480 as were total levels of STAT1 and STAT3 protein (Fig. 8E). In addition, markers of Th1 and Th17 cells and proinflammatory cytokine mRNA levels were inhibited by AZD1480 treatment (Fig. 8F). These data indicate that AZD1480, by inhibiting the JAK/STAT pathway, can diminish peripheral T cell–mediated damage against the CNS in vivo.

AZD1480 treatment in vivo affects the function of myeloid cells and Th1 cells

Myeloid cells are pivotal for the development of EAE by modulating T cell responses because of the production of proinflammatory...
cytokines and presentation of Ag. To assess the functional influence of JAK/STAT inhibition in myeloid cells, C57BL/6 mice were infected with vehicle or AZD1480 (25 mg/kg) for 4 d, and BMDM isolated from these mice were examined for JAK/STAT pathway activation. IL-6 induced STAT3 tyrosine phosphorylation was strongly inhibited in BMDM obtained from AZD1480-treated mice (Fig. 9A), as was IFN-γ-induced STAT1 tyrosine phosphorylation (Fig. 9B). In addition, IFN-γ-inducible MHC class II expression was suppressed by in vivo AZD1480 treatment (Fig. 9C). Consistent with the in vitro data, LPS activation of the NF-κB pathway was not affected (Fig. 9D).

To address whether inhibiting the JAK/STAT pathway modulates the APC functions of myeloid cells, purified CD11b+/CD11c− cells from vehicle- or AZD1480-treated C57BL/6 mice were used as APCs for stimulation of naïve MOG35–55-specific 2D2 CD4+ T cells under Th1- or Th17-polarizing conditions. Notably, we found that APCs from AZD1480-treated mice (AZD-APC) were less efficient than those from vehicle-treated mice (V-APC) in promoting MOG35–55-specific Th1 cell differentiation (Fig. 9E, compare panels 1, 2) and Th17 cell differentiation (Fig. 9F, compare panels 1, 2). These results indicate that CD11b+/CD11c− APCs from AZD1480-treated mice are defective in their ability to induce MOG35–55-specific Th1 and Th17 cell differentiation.

Next, the response of purified naïve MOG35–55-specific CD4+ T cells from vehicle- or AZD1480-treated 2D2 mice to CD11b+/CD11c− APCs from vehicle-treated C57BL/6 mice was examined. CD4+ MOG-specific T cells from AZD1480-treated 2D2 mice (AZD-T cells) were unable to differentiate into Th1 cells when provided with vehicle-treated APCs (Fig. 9E, compare panels 1, 3), and there was a further decrease when MOG-specific T cells from AZD1480-treated 2D2 mice were cultured with AZD1480-treated APCs (Fig. 9E, panel 4). However, CD4+ T cells from AZD1480-treated 2D2 mice were capable of differentiating into Th17 cells in the presence of APCs (Fig. 9F, compare panels 1, 3). We next investigated whether the inhibitory effect of AZD1480 on Th1 cell differentiation was Ag specific. Purified naïve CD4+ T cells from vehicle- or AZD1480-treated C57BL/6 mice were stimulated with anti-CD3 and anti-CD28 Abs under Th1 or Th17 differentiation conditions. Consistent with the results from MOG35–55-specific T cells, in vivo treatment with AZD1480 resulted in partial inhibition of Th1 cell polarization (Fig. 9G), whereas Th17 cell polarization was not affected (Fig. 9H). These findings indicate that in vivo inhibition of the JAK/STAT pathway has an inhibitory effect on the ability of CD4+ T cells to become polarized to the Th1 phenotype but does not appear to affect the Th17 polarization program.

AZD1480 inhibits the JAK/STAT pathway in human T cells and monocytes

The findings thus far demonstrate that AZD1480 suppresses the function of murine T cells and macrophages. To extend these findings to humans, PBMCs were stimulated in vitro with anti-CD3 and anti-CD28 Abs in the absence or presence of AZD1480, and phosphorylation of STAT1, STAT3, and STAT4 was examined in gated CD3+CD4+ T cells. Treatment with AZD1480 led to substantially reduced levels of STAT1 and STAT3 tyrosine phosphorylation and partial inhibition of STAT4 tyrosine phosphorylation (Fig. 10A). In addition, mRNA levels of IFN-γ and IL-17A were significantly reduced by AZD1480 (Fig. 10B). We also assessed the effect of AZD1480 on the JAK/STAT signaling pathway in human monocytes. CD3+CD14+ monocytes were stimulated with IFN-γ in the absence or presence of AZD1480, and phosphorylation of STAT1 and STAT3 was examined. IFN-γ-induced strong phosphorylation of STAT1 and moderate phosphorylation of STAT3, both of which were inhibited by AZD1480 (Fig. 10C). Furthermore, consistent with the inhibition of IFN-γ-induced STAT1 phosphorylation, the expression of IFN-γ-inducible HLA-DR expression in AZD1480-treated monocytes was strongly suppressed (Fig. 10D). These in vitro experiments demonstrate that activated human T cells and monocytes are direct targets of AZD1480.

Discussion

Genome-wide association studies have shown that cytokines, their receptors, JAKs, STATs, and SOCS proteins are associated with human autoimmune diseases, especially pathways leading to STAT3...
and STAT4 activation (39, 40). STAT3 has been identified as an MS susceptibility gene (41, 42), and independent replication supports the association between STAT3 and increase in MS risk (43). We demonstrate that inhibition of the JAK/STAT pathway, specifically inhibition of STAT1, STAT3, and STAT4 activation, has broad actions on cells of the innate and adaptive immune systems, leading to amelioration of clinical disease in EAE models. Treatment with AZD1480, an inhibitor of JAK1 and JAK2, effectively prevents infiltration of immune cells into the CNS, inhibits STAT activation in the CNS, suppresses differentiation and proliferation of CD4+ cells, inhibits proliferation and deactivates myeloid cells, and reduces expression of proinflammatory cytokines and chemokines.

In MS and EAE, the pivotal role of pathogenic Th1 and Th17 cells, as well as myeloid cells, has been well documented (44–46). Inhibition of the JAK/STAT pathway by AZD1480 suppressed the in vitro differentiation of naive T cells to Th1 cells and also suppressed in vitro Th17 cell differentiation induced by three different polarization conditions: IL-6 + IL-23 + TGF-β; IL-6 + IL-23; and IL-6 + IL-23 + IL-1β. Because STAT3 signaling is a central component of Th17 cell differentiation and Th17-dependent autoimmune processes (17, 47), using a JAK inhibitor that suppresses STAT3 activation is a promising strategy for therapeutic intervention. Disease supporting functions of macrophages, DCs, and microglia have been described for CNS autoimmune diseases (45, 48, 49). In vitro results indicate that AZD1480 inhibits IFN-γ–induced NO release, which contributes in a significant manner to EAE pathogenesis and axonal damage (35). AZD1480 suppressed IFN-γ induction of MHC

![FIGURE 7](http://www.jimmunol.org/)

**FIGURE 7.** Inhibition of the JAK/STAT pathway regulates the priming and expansion phases of EAE and proliferation of CD4+ T cells and CD11b+ Cells. C57BL/6 mice were immunized with MOG35–55 peptide (200 µg) emulsified in CFA containing *M. tuberculosis*. Vehicle control or AZD1480 (25 mg/kg) was administered i.p. daily for 3 d starting at day 2 postimmunization (A) or at day 7 postimmunization (D), and mice were sacrificed at day 10. Purified CD4 + T cells isolated from the lymph nodes (LN) and spleens of C57BL/6 mice at day 10 were restimulated with MOG35–55 peptide (10 µg/ml) under Th1 and Th17 differentiation conditions for 4 d. At day 4, cells were stimulated with PMA/ionomycin plus GolgiStop for 4 h, stained for the surface marker CD4, and by intracellular flow for IFN-γ (B, E) or IL-17A (C, F). C57BL/6 mice were immunized with MOG35–55 peptide (200 µg) emulsified in CFA containing *M. tuberculosis*. The mice received i.p. injections of 250 ng pertussis toxin at days 0 and 2. Vehicle control or AZD1480 (25 mg/kg) was administered i.p. daily starting at day 7 postimmunization for 7 d. Mice were injected i.p. with 100 µg EdU in PBS at day 13. (G) CNS-infiltrating mononuclear cells were isolated from the cerebellum or spinal cord 12 h later and stained with Abs to CD11b (H) or CD4 (I), and EdU was detected by the Click reaction and flow cytometry. *p < 0.05, **p < 0.001.
class II and CD40, which are critical for Ag presentation functions of macrophages/DCs. IFN-γ induction of MHC class II and CD40 expression is STAT1 dependent (56, 57), indicating that inhibition of the JAK/STAT pathway is an effective means of suppressing these genes. In macrophages, AZD1480 inhibited GM-CSF activation of STAT5, and IL-6 activation of STAT3, while not influencing IL-4 activation of STAT6. IFN-γ, GM-CSF, and IL-6 signaling use JAK1 and/or JAK2, which explains the inhibitory effect of AZD1480. IL-4 signaling involves both JAK1 and JAK3; thus, inhibition of JAK1 by AZD1480 may not suffice to inhibit this pathway because of the kinase activity of JAK3.

Inhibition of the JAK/STAT pathway by AZD1480 has beneficial effects in mitigating clinical symptoms in five EAE models: chronic progressive disease in C57BL/6 mice; atypical disease with involvement of the cerebellum in LysMCre-SOCS3fl/fl mice; RR disease in SJL/J mice; adoptive Th1 cell EAE; and adoptive Th17 cell EAE. Improvement in clinical scores was associated with suppression of inflammatory responses in lymph node and spleen, indicating that inhibition of the JAK/STAT pathway affects the pathogenic potential of T cells outside the CNS. The most reliable parameter of JAK inhibition is that of downstream inhibition of STAT activation (23, 24). AZD1480 treatment inhibited STAT1, STAT3, and/or STAT4 activation within the CNS. These STATs are activated downstream of JAK1 and/or JAK2 and are essential for development and differentiation of Th1 and Th17 cells and maturation of myeloid cells. Thus, inhibition of STAT activation may be responsible for the reduction of inflammatory lesions and diminished expression of proinflammatory mediators in the CNS. Total levels of STAT proteins were also inhibited by AZD1480 treatment, most notably that of STAT1. STAT1 is itself a STAT1-inducible gene (37); therefore, inhibiting STAT activity will function in a negative feedback loop to inhibit STAT expression. Mice deficient in STAT3 in CD4+ T cells are resistant to EAE (17, 18), as are mice deficient in STAT4 (58), whereas mice lacking STAT1 are highly susceptible to EAE (25). It is possible that in the absence of STAT1, STAT3, and/or STAT4 signaling may compensate to drive Th1 cell responses, because these mice were characterized by IFN-γ–producing Th1 cells (25). Nonetheless, our results demonstrate that pharmacologic inhibition of STAT1, STAT3, and STAT4 signaling contributes to significant lessening of EAE disease severity.

The beneficial effects of inhibiting the JAK/STAT pathway on EAE pathogenesis appear to involve a number of mechanisms, including effects on both APCs and T cells. First, both the primary and expansion phases of EAE were inhibited by AZD1480 treatment, as reflected by suppressed Ag-specific Th1 and Th17 responses in secondary lymphoid tissues, indicating that inhibition of the JAK/STAT pathway affects the early stages of the immunologic cascade leading to EAE. Second, this, in turn, leads to diminished infiltration of immune cells into the CNS. Third, of the T cells and myeloid cells that did extravasate into the CNS, AZD1480 treatment diminished their proliferative capacity, which

**FIGURE 8.** Effect of AZD1480 on Th1- and Th17-induced adoptive transfer EAE. (A) C57BL/6 mice were immunized with MOG35–55 peptide (200 μg) for 10 d, splenocytes and lymph node cells were isolated, and then, MOG-specific T cells were cultured with MOG35–55 peptide (10 μg/ml) under Th1 differentiation conditions for 3 d. Cells (30 \( \times 10^6 \)) were injected i.v. into C57BL/6 mice. Vehicle control (n = 10) or AZD1480 (25 mg/kg) (n = 11) was administrated daily by i.p. starting at day 7 for 10 d. Mean ± SD of classical EAE clinical scores. (B) Mice were perfused, the spinal cords were removed, and protein extracts from spinal cord of vehicle control or AZD1480-treated mice at day 17 were immunoblotted with the indicated Abs. (C) Mice were perfused, the spinal cords were removed, and mRNA from vehicle control or AZD1480-treated mice at day 17 was analyzed by qRT-PCR. (D) C57BL/6 mice were immunized with MOG35–55 peptide (200 μg) for 10 d, splenocytes and lymph node cells were isolated, and then, MOG-specific T cells were cultured with MOG35–55 peptide (10 μg/ml) under Th1 differentiation conditions for 3 d. Cells (30 \( \times 10^6 \)) were injected i.v. into C57BL/6 mice. Vehicle control (n = 11) or AZD1480 (25 mg/kg) (n = 10) was administrated daily by i.p. starting at day 8 for 10 d. Mean ± SD of classical EAE clinical scores. (E) Mice were perfused, the spinal cords were removed, and protein extracts from spinal cord of vehicle control or AZD1480-treated mice at day 18 were immunoblotted with the indicated Abs. (F) Mice were perfused, the spinal cords were removed, and mRNA from spinal cord of vehicle control or AZD1480-treated mice at day 18 was analyzed by qRT-PCR. *p < 0.05, **p < 0.001.
we believe contributes to the therapeutic effectiveness of AZD1480 treatment. Fourth, in vivo AZD1480 treatment inhibits both the innate and adaptive arms of the immune system that are associated with EAE. AZD1480 treatment in vivo was effective in inhibiting the ability of T cells to differentiate into Th1 cells, in both Ag-specific and non–Ag-specific manners. This intriguing finding needs to be examined in more detail but may relate to the effectiveness of AZD1480 in inhibiting IFN-γ and IL-12 signaling, which are critical for Th1 cell differentiation. This suggests a direct effect of AZD1480 on Th1 differentiation. In contrast, although our in vitro data documented an inhibitory effect of AZD1480 on Th17 cell differentiation, we did not observe a direct inhibitory effect on Th17 cell polarization in vivo. This may reflect the complexities of promoting Th17 cell differentiation by diverse cytokines including TGF-β1, TGF-β3, IL-1, IL-6, and IL-23 and inhibition by IL-2 and IL-27 (59, 60). We do know that AZD1480 is a strong inhibitor of IL-6 and IL-23 signaling and suppresses expression of the IL-23R on T cells, a critical determinant of Th17 cell polarization. However, the influence of inhibiting the JAK/STAT pathway on TGF-β1, TGF-β3, and IL-27 signaling is unknown, and IL-1β signaling is unaffected (data not shown). Inhibition of the JAK/STAT pathway has potent sup-

FIGURE 9. The JAK1/2 inhibitor AZD1480 has direct effects on myeloid APCs and Th1 cells. C57BL/6 mice were administrated vehicle control or AZD1480 (25 mg/kg) i.p. daily for 4 d. At that time, BMDM were isolated from mice and stimulated with medium (−) or IL-6 (10 ng/ml) plus sIL-6R (25 ng/ml) for 30 min (A), or IFN-γ (10 ng/ml) for 30 min (B), and then, cell lysates were subjected to immunoblotting with the indicated Abs. (C) BMDM isolated from vehicle control or AZD1480-treated mice after 4 d of treatment were stimulated with medium (UN) or IFN-γ (10 ng/ml) for 24 h, and then, MHC class II expression was analyzed by flow cytometry. (D) BMDM obtained as described above were stimulated with medium or LPS (10 ng/ml) for 30 min, and then, cell lysates were subjected to immunoblotting with the indicated Abs. (E and F) C57BL/6 or MOG35–55 TCR-transgenic mice (2D2) were administrated vehicle control or AZD1480 (25 mg/kg) i.p. daily for 5 d. CD11b+ [CD11c]+ cells sorted from the spleen of vehicle control (V) or AZD1480 (AZD)-treated C57BL/6 mice were used as APCs in coculture with naive CD4+ T cells from vehicle- or AZD1480-treated 2D2 mice. The polarization of MOG-specific Th1 cells was determined by intracellular cytokine staining for IFN-γ after 3 d of culture with MOG peptide (10 μg/ml), IL-12, and anti–IL-4 Ab (E). The polarization of MOG-specific Th17 cells was determined by intracellular cytokine staining for IL-17A after 3 d of culture with MOG peptide (10 μg/ml), IL-23, IL-6, TGF-β, anti–IFN-γ, and anti–IL-4 Abs (F). (G and H) C57BL/6 mice were administrated vehicle control or AZD1480 (25 mg/kg) i.p. daily for 3 d. Naive CD4+ T cells were isolated from the spleen. Vehicle control DCs were used as APCs at a 1:5 ratio to CD4+ T cells in coculture under Th1 or Th17 differentiation mixture conditions with anti-CD3 and anti-CD28 Abs for 4 d and then stained for the surface marker CD4 and by intracellular flow for IFN-γ (G) or IL-17A (H).
pressive effects on myeloid cell functions in vivo. AZD1480 treatment renders myeloid cells nonresponsive to IFN-γ and IL-6 signal transduction, resulting in inhibition of STAT1 and STAT3 activation, and downstream inhibition of gene expression such as MHC class II. Furthermore, AZD1480 treatment has a direct effect on APC functions of myeloid cells as demonstrated by the findings that CD11b+/CD11c+ APCs from AZD1480-treated mice inhibited the development of MOG35–55-specific Th1 and Th17 cells. This is in line with observations on the in vivo treatment of mice with glatiramer acetate, an approved therapy for RRMS (61), fumarates (62), which were approved by the U.S. Food and Drug Administration on March 27, 2013, for treatment of RRMS, and laquinimod, which has been successfully evaluated in phase II/III studies of RRMS (63, 64). All of these agents had direct effects on cells of the innate immune system and, importantly, skewed macrophages and DCs toward a regulatory phenotype. Characteristics of the regulatory, anti-inflammatory phenotype included reduced STAT1 phosphorylation, decreased production of IL-6, IL-12, and IL-23, and elevated secretion of IL-10 (61–64). At this point, we do not know whether in vivo treatment with JAK inhibitors skews monocytes/macrophages and DCs toward a regulatory phenotype, but we do demonstrate a suppression of the M1 proinflammatory phenotype by AZD1480. Thus, collectively, AZD1480 functions to promote immune modulation by a direct effect on myeloid APCs, which impacts the differentiation of Th1 and Th17 cells in vivo, as well as a direct effect of AZD1480 in vivo on Th1 cell differentiation.

Studies have implicated the JAK/STAT axis in regulating clinical manifestations of EAE. Peroxisome proliferator activated receptor γ and cyclooxygenase 2 inhibitors suppress EAE severity, in part, by inhibiting IL-12–induced activation of the JAK/STAT pathway and subsequent suppression of Th1 cell differentiation (65, 66). The protective effect of glatiramer acetate in EAE is in part due to inhibition of STAT4 and STAT3 phosphorylation in T cells, altering Th1 and Th17 cell differentiation, respectively (67). Two herbal compounds, plumbagin and berberine, exert protective effects in EAE models by inhibiting STAT activation and Th1 and Th17 cell differentiation (68, 69). In this study, we have used a specific inhibitor of the JAK/STAT pathway, AZD1480, and documented a striking beneficial immunomodulatory effect in five different models of EAE. Importantly, AZD1480 treatment was administered at the onset of disease and in a therapeutic manner after the appearance of clinical symptoms, with potent clinical efficacy. These findings collectively suggest the JAK/STAT axis may serve as a therapeutic target for intervention in MS, as well as other neuroinflammatory conditions such as Parkinson’s disease, spinal cord injury, and Alzheimer’s disease.

**FIGURE 10.** AZD1480 inhibits activation of the JAK/STAT pathway and gene expression in human T cells and monocytes. (A) PBMCs were stimulated with anti-CD3 (5 μg/ml) and anti-CD28 (2 μg/ml) Abs in the absence or presence of AZD1480 (0.25 and 0.5 μM) for 4 d. The extent of STAT phosphorylation within the CD4+ T cell population was assessed by intracellular flow cytometry using anti–phospho-STAT1Tyr701, anti–phospho-STAT3Tyr705, and phospho-STAT4Tyr693 (red), with isotype control shown in blue. (B) For mRNA expression, cells were collected at day 4, and mRNA was analyzed by qRT-PCR for IFN-γ and IL-17A. (C) Adherent PBMCs were pretreated with vehicle control or AZD1480 (0.25 and 0.5 μM) for 2 h and then stimulated with IFN-γ (100 U/ml) for 1 h. The extent of STAT phosphorylation within the CD3+/CD14+ monocyte population was assessed by intracellular flow cytometry using anti–phospho-STAT1Tyr701 and anti–phospho-STAT3Tyr705 (red), with isotype control shown in blue. (D) Adherent PBMCs were pretreated with vehicle control or AZD1480 (0.25 or 0.5 μM) for 2 h and stimulated with medium or IFN-γ (100 U/ml) for 24 h, and then, HLA-DR expression was analyzed by flow cytometry. *p < 0.05, **p < 0.001.

**ACKNOWLEDGMENTS**
We thank the University of Alabama at Birmingham Neuroscience Molecular Detection Core and the Rheumatic Diseases Core Center (P30 Flow Core) for advice and technical assistance, Dr. Dennis Huszar (Cancer Bioscience, AstraZeneca R&D, Waltham, MA) for providing AZD1480 and helpful discussions, Dr. Kui Zhang (Section on Statistical Genetics, Department of Biostatistics, School of Public Health at the University of Alabama at Birmingham) for assistance with statistical analysis of the EAE data, Dr. Warren Alexander for the gift of the SOCS3fl/fl mice, and members of the Benveniste Laboratory for helpful discussions.
Disclosures

The authors have no financial conflicts of interest.

References


