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Small Molecule Inhibitor of Antigen Binding and Presentation by HLA-DR2b as a Therapeutic Strategy for the Treatment of Multiple Sclerosis

Niannian Ji,* Animesh Somanaboeina,* Aakanksha Dixit,* Kazuyuki Kawamura,* Neil J. Hayward,† Christopher Self,‡ Gary L. Olson,‡ and Thomas G. Forsthuber*†

The strong association of HLA-DR2b (DRB1*1501) with multiple sclerosis (MS) suggests this molecule as prime target for specific immunotherapy. Inhibition of HLA-DR2b–restricted myelin-specific T cells has the potential to selectively prevent CNS pathology mediated by these MHC molecules without undesired global immunosuppression. In this study, we report development of a highly selective small molecule inhibitor of peptide binding and presentation by HLA-DR2b. PV-267, the candidate molecule used in these studies, inhibited cytokine production and proliferation of myelin-specific HLA-DR2b–restricted T cells. PV-267 had no significant effect on T cell responses mediated by other MHC class II molecules, including HLA-DR1, -DR4, or -DR9. Importantly, PV-267 did not induce nonspecific immune activation of human PBMC. Lastly, PV-267 showed treatment efficacy both in preventing experimental autoimmune encephalomyelitis and in treating established disease. The results suggest that blocking the MS-associated HLA-DR2b allele with small molecule inhibitors may be a promising therapeutic strategy for the treatment of MS. The Journal of Immunology, 2013, 191: 000–000.

A widely accepted concept of autoimmune disease is that self-derived cellular proteins trigger activation of pathogenic T cells via the presentation of autoantigens by MHC class II molecules. The resulting recognition of MHC:Ag complexes triggers proliferation and cytokine production of autoreactive T cells, resulting in inflammatory processes and subsequent destruction of normal tissues such as the CNS in multiple sclerosis (MS). In the case of MS, peptides derived from myelin Ags such as myelin basic protein (MBP) or myelin oligodendrocyte glycoprotein (MOG) bind to the strongly disease-associated MHC class II allele HLA-DR2b (DRB1*1501) (1–3), resulting in an immune response and attack on the myelin nerve sheath, leading to disease symptoms and eventual disability. Despite the understanding of this fundamental mechanism and its inherent potential, no drugs based on the direct inhibition of Ag binding have been successfully developed to date.

First-generation MS therapeutics (e.g., IFN-βs and glatiramer acetate) are widely used (4–6), but do not act on the critical mechanism of Ag presentation, but rather modify downstream inflammatory responses. IFN-βs have multiple targets, including modifying multiple inflammatory cytokines, whereas glatiramer acetate shifts the population of T cells from proinflammatory Th1 types to regulatory Th2 types and may also mimic myelin. However, these compounds have only modest effects on the disease in some patients, but, on the positive side, are relatively safe.

Over the past several decades, much attention has been directed toward the concept of specific immunotherapy, that is, the removal or silencing of specific disease-inducing pathogenic T cell clones in a manner that would not compromise the ability of the rest of the immune system to respond to foreign pathogens. Among the strategies that have been previously pursued are induction of oral tolerance via ingestion of myelin proteins, altered peptide ligands, T cell vaccination, and rTCR ligands (7–14). Clinical trials of these approaches in MS have met with limited success, due in part to unexpected immune reactions to the agents used (15).

In contrast to these approaches at specific immunotherapy, development of disease-modifying drugs that affect T cell migration or agents that deplete immune cells in general or that alter cytokine profiles has been more efficacious and has led to approval of several second-generation therapeutics for MS, although posing significant safety risks. Natalizumab (a VLA-4 inhibitor) blocks T cell migration and is effective in MS, but the drug has led to serious safety concerns over progressive multifocal leukoencephalopathy, an infectious brain disorder due to activation of John Cunningham virus in patients treated with the drug (16). The first oral MS drug, fingolimod, acts on S1P receptors and prevents certain activated T cells from leaving lymph nodes, thus suppressing their entry into the brain (17, 18). Whereas initial results from clinical trials support therapeutic efficacy, fingolimod has concerns with cardiac side effects, increased incidence of certain malignancies, and a somewhat increased risk of infection. Teriflunomide and alemtuzumab have migrated to MS from cancer chemotherapy and suppress MS, but their overall utility is also compromised by safety concerns (19–21). A number of other drugs are in development or recently approved, including dimethyl fumarate (Tecfidera, BG-00012), an oral agent originally used in the treatment of psoriasis (22).

An attractive alternative strategy that focuses on the key step of Ag binding would be to block the activation of pathogenic T cells at...
the critical step of Ag binding to the disease-associated MHC class II molecule HLA-DR2b (DRB1*1501). In those MS patients who express the DR2b gene [~60% of all MS patients (23)], selectively blocking this target is expected to disrupt the autoimmune process but not generally compromise the immune system. Early attempts at developing peptide inhibitors of MHC binding for HLA-DR1 and DR4 had been made but failed for several reasons. First, their natural peptide nature left them susceptible to rapid degradation by proteolytic enzymes in plasma and particularly toward cathepsins in the loading compartment of APCs. Initially, it was hypothesized that a large molecular size (similar in peptide length to the Ags) would be required to satisfy all the molecular recognition sites in the binding groove of the MHC (24, 25). However, such long peptides turned out to be unstable, as noted, but in addition were also potentially immunogenic. Finally, these investigations and compounds also did not achieve sufficiently high binding affinity to compete efficiently with Ags in cellular assays (26). In this study, we describe an approach to design MHC inhibitors guided by structure-based methods to generate compounds with high affinity and specificity for DRB1*1501. The compounds are designed as small molecular structures and are stabilized against proteolytic cleavage. We describe the design, binding studies, structure–activity relationships, activity in animal models, immunological evaluations, and efficacy in human cells that have led to the selection of a development candidate, PV-267.

Materials and Methods

Human PBMCs and cell line

Characterized, HLA-typed cryopreserved human PBMCs were purchased from Cellular Technology and Astarte Biologics. Donor 4 PBMCs (DRB1*0101/DRB1*1501) and donor 9 PBMCs (DRB1*0401/DRB1*0901) were from Cellular Technology. Donor 003 (DRB1*1501/DRB1*14) PBMCs were from Astarte Biologics. A cryopreserved MBP-specific T cell line (BC3) was purchased from Astarte Biologics (originally derived from donor 003 PBMCs). All cryopreserved human cells were thawed using Cellular Technology thaw solution and were resuspended in Cellular Technology test medium before being used in experiments.

PV-267 and HLA-DR–restricted peptides

Compound PV-267 and all modified low-affinity compounds were designed and synthesized by Provid Pharmaceuticals. HLA-DR2b (DRB1*1501)–restricted MBP peptide 87–99 (VHFKNIVTPRT) for recall of BC3 T cells was purchased from United Biochemical Research. HLA-DR1 (DRB1*0101)–restricted influenza A matrix protein peptide 17–31 (SGLPKAEIAQRLEDV), HLA-DR9 (DRB1*0901)–restricted tetanus toxin peptide 830–843 (QYIKANSKFPITGE), and HLA-DR4 (DRB1*0401)–restricted influenza hemagglutinin peptide 307–319 (PKYKVQNTKLAT) were purchased from GenScript. HLA-DR2b–restricted tetanus toxin peptide 830–844 (QYIKANSKFPITGE) was purchased from AnaSpec.

Cathepsin stability assay

Test compounds (1 mg) were dissolved in 20 μl DMSO and diluted to 1 ml with acetate (pH 5.5) or citrate (pH 3.5) buffer. Cathepsin enzymes (B, Sigma-Aldrich; L, S, CalBio) were diluted in water to generate desired optimal working concentrations. Enzyme was added to the compound solution and incubated, as indicated in Table II, and aliquots were quenched with 50% trifluoroacetic acid and frozen at −20°C until HPLC analysis.

Human cytokine ELISPOT assays

Primary anti-human IFN-γ mAb (Thermo Scientific; B213.5) was diluted in PBS and coated on ELISPOT plate (Millipore) at 4°C overnight. After washing and blocking, autologous APCs (irradiated PBMCs) pretreated with different concentrations of PV-267 for 45 min were added on the plate (0.3–0.5 × 10⁶ cells/well), followed by the addition of Ag or positive control using PHA (Sigma-Aldrich) or anti-human CD3 mAb (Bio X Cell; OKT3) along with BC3 T cells or enriched PBMC T cells (4,000–20,000 cells/well). Plates were incubated at 37°C for 24 h, and then washed and biotinylated anti-human IFN-γ mAb (Thermo Scientific; B213.5) was added at 4°C overnight. After that, plates were incubated with streptavidin alkaline phosphatase (Invitrogen) and developed with 5-bromo-4-chloro-3-indolyl phosphate/NBT phosphatase substrate (Kirkegaard & Perry Laboratories) after washing. After developing of the plates, image analysis of ELISPOT assays was performed on a Series 2 ImmunoSpot analyzer and software (Cellular Technology), as described previously (27).

Generation of HLA-DR–restricted Ag-specific T cell lines from human PBMCs

Thawed human PBMCs were cultured in fresh Cellular Technology test medium at 3 × 10⁶ cells/ml and stimulated with peptide Ag at 2 μg/ml for 4 d. Viable cells were counted and plated at 3 × 10⁶ cells/ml in Cellular Technology test medium with human rIL-2 at 2 U/ml. After 5 d, cells were again counted and plated at 3 × 10⁶ cells/ml with fresh IL-2 at 2 U/ml, or alternatively, used for the experiments, as detailed.

Flow cytometry and mAbs

Thawed or cultured human PBMCs were washed in cold flow buffer containing 2% FCS (Atlanta Biologicals) and PBS (Beckman Coulter), followed by blocking with human FeR-binding inhibitor (eBioscience) for 20 min and staining in 100 μl vol with 0.3–1 × 10⁶ cells per sample at 4°C for 45 min in dark. The cells were then washed twice and fixed before examination by flow cytometry (BD FACs Aria II). Most of the mAbs for staining were obtained from eBioscience, as follows: allophycocyanin anti-human CD40 (5C5), PE anti-human CD69 (FN50), allophycocyanin anti-human/mouse CD44 (IM7), PerCP-Cy5.5 anti-human CD28 (CD28.2), PE anti-human CD25 (BC96), PerCP-Cy5.5 anti-human CD45 (2D1), PerCP–EFluor710 anti-human CD86 (IT2.2), allophycocyanin anti-human HLA-DR (LN3), and allophycocyanin anti-human CD4 (OKT4). Anti-human CD3 mAb (Bio X Cell; OKT3) was conjugated with either Alexa Fluor 488 or Alexa Fluor 594 dye using a mAb labeling kit, according to the manufacturer’s instructions (Invitrogen/Life Technologies).

Cytokine multiplex assay (Bio-Plex)

Supernatants from human PBMC or cultured BC3 T cells were collected and analyzed for human cytokine levels by cytokine multiplex assay following the Bio-Plex Pro Human Cytokine Standard Group I 27-Plex manufacturer’s protocol (Bio-Rad).

Cell proliferation assay by CFSE

BC3 T cells (Astarte Biologics) were resuspended in prewarmed 0.1% BSA/PBS (Sigma-Aldrich) at 1 × 10⁶ cells/ml and incubated with CFSE (CellTrace CFSE Cell Proliferation kit; Invitrogen) solution for a final concentration of 10 μg/ml at 37°C for 10 min. Stock CFSE was prepared fresh in DMSO following the manufacturer’s instructions. The staining was quenched by adding 5× vol of ice-cold culture media to the cells, followed by incubation on ice for 5 min. CFSE-labeled BC3 T cells were centrifuged and washed twice with culture media, then cultured with autologous APCs (irradiated human PBMCs; Astarte Biologics), 10 μg/ml MBP87–99 peptide, or mitogens such as PHA, anti-human CD3 mAb (OKT3) for 5 d at 37°C. Cells were collected and stained with allophycocyanin anti-human CD4 mAb (OKT4), followed by flow cytometry analysis.

Mice

HLA-DR2b (DRB1*1501) transgenic mice (27) backcrossed to MHC class II–deficient mice (28) were maintained under specific pathogen-free conditions, and all animal procedures were conducted according to the guidelines of the Institutional Animal Care and Use Committee of the University of Texas at San Antonio.

Experimental autoimmune encephalomyelitis induction and treatment with PV-267

Active experimental autoimmune encephalomyelitis (EAE) was induced in HLA-DR2b transgenic mice by s.c. injection of 200 μg MOG35–55 peptide (United Biological Research) in 50 μl CFA. Mice also received i.p. injections of 200 ng Pertussis toxin (PTX) on days 0 and 2. A total of 30 mg/kg PV-267 or vehicle (30% Captisol or Tween 80) was injected i.p. into mice once daily starting either 1 d before disease induction for 20 d or with 50 mg/Kg in PBS once daily upon disease onset for 10 d. Adoptive transfer EAE model was induced in HLA-DR2b transgenic mice by injecting recipient mice i.p. with 20 × 10⁶ encephalitogenic donor cells. Briefly, HLA-DR2b donor mice were immunized s.c. with 50 μg MOG35–55
in CFA. Splenocytes and draining lymph node cells were harvested from donors on day 10 and cultured for 3 d in complete DMEM (10% FBS, 1-glutamine) in the presence of MOG35–55 peptide (20 μg/ml), mouse rIL-12 (30 ng/ml; R&D Systems), and anti-mouse IFN-γ mAb (10 μg/ml; R4-6A2; Bio X Cell). After the culture, donor cells were harvested and resuspended in warm serum-free DMEM and injected i.p. into HLA-DR2b recipients. A total of 50 mg/kg PV-267 or control (0.02% PBS Tween 80) was injected i.p. into the mice daily starting 2 d before disease induction for 15 d.

**EAE evaluation**

Mice were monitored and graded daily for clinical signs of EAE using the following scoring system (27, 29): 0, no abnormality; 1, limp tail; 2, moderate and hind limb weakness; 3, complete hind limb paralysis; 4, quadriplegia or premoribund state; 5, death.

**Mouse cytokine ELISPOT**

ELISPOT plates (Millipore) were coated with functional-grade purified anti-mouse IFN-γ mAb (eBioscience; AN-18) and anti-mouse IL-17 mAb (BioLegend; TC11-8H4) were then added to the plate overnight. Mouse splenocytes (1 × 10^6 cells/well) and brain-infiltrating mononuclear cells (isolated by Lympholyte-M software (Cellular Technology), as described previously (27). ELISPOT assays was performed on a Series 2 ImmunoSpot analyzer and Perry Laboratories). After developing of the plates, image analysis of IL-17 mAb (Bio X Cell; 17F3) at 4˚C overnight. Mouse splenocytes (1 × 10^6 cells well) and brain-infiltrating mononuclear cells (isolated by Lympholyte-M media; 2–5 × 10^6 cells/well; Cedarlane) were collected and restimatated with MOG35–55 peptide or PV-267 as control on ELISPOT plates in HL-1 medium (Lonza BioWhittaker, Fisher Scientific) at 37°C for 24 h. Biotinylated anti-mouse IFN-γ mAb (eBioscience; R4-6A2) and anti-mouse IL-17 mAb (BioLegend; TC11-8H4) were then added to the plate overnight at 4°C, followed by incubation with streptavidin alkaline phosphatase (Invitrogen) for 2 h at room temperature and developing with 5-bromo-4-chloro-3-indolyl phosphate/NBT phosphate substrate (Kirkegaard & Perry Laboratories). After developing of the plates, image analysis of ELISPOT assays was performed on a Series 2 ImmunoSpot analyzer and software (Cellular Technology), as described previously (27).

**Serum Ag-specific IgG ELISA**

ELISA plates (Corning Costar 9018) were coated with MOG35–55 peptide or PV-267 compound at 20 μg/ml and incubated at 4°C overnight. After washing with 1% BSA in PBS/Tween 20 buffer, serum titrations were added to the plates and incubated at 4°C overnight. The plates were then washed with PBS/Tween 20 and incubated with detection Ab (HRP-conjugated goat anti-mouse IgG; Thermo Scientific) for 1 h at room temperature. Plates were then washed and incubated with 1× tetramethylbenzidine substrate solution (eBioscience) for 15 min and stopped by 2 N H_2SO_4. Plates were read at OD 450 nm for absorbance on a Synergy HT microplate reader (BioTek).

**Statistical analysis**

Sigma Stat (Systat Software) was used to perform all tests of significance. Statistical analysis was performed on individual animals, considering Mice with at least one clinical score of 2 or higher by day 14 as having a clinical EAE disease endpoint. Differences in disease onset were compared with a Fishers’ exact test. The cumulative EAE score was calculated by adding up daily EAE scores from the onset of disease until the end of study or before recovery for individual mice and divided by the number of mice within each group (31). Statistical significance was calculated by t test.

**Results**

**Generation of small molecule inhibitors for HLA-DR2 molecules**

The interactions between peptides and MHC molecules have been previously well characterized in studies of high-resolution crystal structures (25, 32, 33). The Wiley group’s determination of the HLA-DR2 (DRA*0101/DRB1*1501) structure with the MBP85–99 peptide bound provided a basis for the design of small molecule inhibitors used in this study. The 15-aa MBP peptide lies in the binding groove with parts extending beyond the binding pocket (4–5 residues at each terminus) (25). Key interactions are with the hydrophobic pockets of DRB1*1501 designated as P1 and P4. Because much of the interaction energy of a DR2 inhibitor was expected to be associated with hydrophobic interactions (dominant in P1 to P4), we explored truncation of the sequences, optimizing interactions by filling of the P1 and P4 pockets, and constraining the P3 group into a cyclic structure (for potency, reducing its exposure on the surface of the complex, and for cation stabilization) through a series of 185 analogs. In analyzing the DR2-binding pocket, we noted that the region extending from the P5 pocket to the peptide Ag C terminus was primarily polar. We reasoned that because those regions also mainly were interacting via a network of hydrogen bonds to the backbone of the MBP peptide, it might be possible to truncate the peptide ligand further from P4 to the C terminus, essentially permitting solvent water to replace this region of the peptide. The N terminus also seemed to extend out of the binding pocket and was associated with T cell activation, so was also a candidate for truncation. As shown in Table I, removal of four residues from each of the N and C termini in the context of the MBP85–99 peptide (5 nM binding affinity) gave a minimally active sequence of only seven residues, with a binding affinity of 1213 nM (compound 1). The 7-mer formed the basis for further modifications. Replacing the His at P2 by Arg (compound 2) enhanced binding (from 1213 to 707 nM) as anticipated from previous studies on DR4 ligands (32). We then examined the P1 pocket, which in MBP is valine (Val). Because the isopropyl side chain of Val does not fully occupy the P1 hydrophobic pocket, introducing cyclopentylglycine (compound 3) to reduce its exposure on the surface of the complex, and for cation stabilization) through a series of 185 analogs. In analyzing the DR2-binding pocket, we noted that the region extending from the P5 pocket to the peptide Ag C terminus was primarily polar. We reasoned that because those regions also mainly were interacting via a network of hydrogen bonds to the backbone of the MBP peptide, it might be possible to truncate the peptide ligand further from P4 to the C terminus, essentially permitting solvent water to replace this region of the peptide. The N terminus also seemed to extend out of the binding pocket and was associated with T cell activation, so was also a candidate for truncation. As shown in Table I, removal of four residues from each of the N and C termini in the context of the MBP85–99 peptide (5 nM binding affinity) gave a minimally active sequence of only seven residues, with a binding affinity of 1213 nM (compound 1). The 7-mer formed the basis for further modifications. Replacing the His at P2 by Arg (compound 2) enhanced binding (from 1213 to 707 nM) as anticipated from previous studies on DR4 ligands (32). We then examined the P1 pocket, which in MBP is valine (Val). Because the isopropyl side chain of Val does not fully occupy the P1 hydrophobic pocket, introducing cyclopentylglycine (compound 3)
or cyclohexylglycine (Chg, compound 4) in this 7-mer context to fill this space increased affinity to 16 and 4 nM, respectively (Table I). Constraining the backbone and P3 side chain by replacing the Phe of compound 2 by Tic (a constrained tetrahydroisoquinoline amino acid) increased binding affinity for compound 5 to 17 nM compared with the original reference sequence (707 nM, compound 2). Extending the chain cap at the N terminus to include Val at P(-1) enabled further truncation at the C terminus (compound 7). Combining all of the features [P(-1) Val, P1 Chg, and P3 Tic] in a five-residue sequence, we obtained PV-267 with an affinity of 2 nM versus DRB1*1501 and with high selectivity for DRB1*1501 versus DRB1*0101 and other alleles tested (data not shown). These modifications (particularly the Tic group that introduces a cyclic amino acid) impart high resistance to cathepsins, as outlined in Table II. Based on the potency of PV-267, this compound was selected as the lead for further studies.

Inhibition of HLA-DR2b–restricted T cell responses by PV-267

PV-267 was chosen as a candidate MHC II inhibitor due to its high in vitro binding affinity for HLA-DR2b (2 nM) and selectivity against other alleles (>400-fold) combined with stability to degradation by cathepsins (Tables I, II) and with good t_{1/2} and pharmacokinetics following i.p. or s.c. administration (data not shown). To test the inhibitory effect of PV-267 on HLA-DR2b–restricted T cell responses, we activated a human MBP83–99–specific T cell line (BC3) derived from a HLA-DR2b+ MS patient with its cognate MBP peptide in the presence or absence of the inhibitor and determined T cell responses by cytokine ELISPOT and CFSE-based proliferation assays.

As shown in Fig. 1A, BC3 T cells cultured with the minimal-epitope MBP87–99 peptide in the presence of autologous APCs showed strong production of IFN-γ as measured by cytokine ELISPOT assay. Importantly, incubation of BC3 T cells with MBP87–99 and autologous APCs pretreated with increasing concentrations of PV-267 (0–200 μg/ml) significantly inhibited cytokine production (Fig. 1A, 1B). Optimal inhibition was achieved at 50 μg/ml PV-267 (Fig. 1A, 1B). Importantly, no significant inhibitory effect by PV-267 was observed when BC3 T cells were stimulated with anti-CD3 mAb, indicating that the compound did not affect viability at the concentrations used and that inhibition was specific for MHC:peptide-mediated signaling (Fig. 1C). Additionally, PV-267 did not affect the viability of resting and Ag-activated BC3 T cells or autologous PBMCs at concentrations of up to 400 μg/ml for 24 h (data not shown).

To test whether PV-267 inhibited production of other proinflammatory cytokines by BC3 T cells, supernatants were collected after stimulation of the cells with MBP87–99 peptide in the presence of autologous APCs and incubation with PV-267 at increasing concentrations. Supernatants were assayed by cytokine multiplex assay (human 27-multiplex; Fig. 1D–I, Table III). The results show that PV-267 also inhibited the production of TNF-α and IL-1ra (Fig. 1D, 1E). Production of IL-6, IL-1β, MIP-1α, and IL-17 showed a trend to be decreased as PV-267 concentration increased (Fig. 1F–I), but the results did not reach statistical significance. Similarly, several other cytokines and chemokines, including IFN-γ, showed a trend to be decreased by PV-267, but the results did not achieve statistical significance (Table III). Of note, BC3 T cells produced very low levels of IL-17, which we confirmed by cytokine ELISPOT assay (data not shown). No significant inhibition was observed for other cytokines tested (data not shown).

Next, we tested whether PV-267 inhibited Ag-specific proliferation of BC3 T cells. As shown in Fig. 1I, PV-267 inhibited MBP87–99–induced proliferation of BC3 T cells in a dose-dependent manner, as measured by CFSE-based proliferation assay. In contrast, proliferation induced by anti-CD3 mAb or PHA was not inhibited by PV-267.

Taken together, the data showed that PV-267 inhibited Ag-induced cytokine production and proliferation of human HLA-DR2–restricted T cells.

Inhibition by PV-267 is specific for HLA-DR2b molecules

PV-267 inhibited human T cell responses to myelin peptides presented by HLA-DR2b molecules. To determine the specificity of the inhibition for HLA-DR2b as compared with other human MHC class II alleles, T cells were enriched from PBMC of healthy donors that expressed other MHC molecules, including HLA-DR1 (DRB1*0101), HLA-DR4 (DRB1*0401), HLA-DR9 (DRB1*0901), with or without HLA-DR2b (DRB1*1501). The PBMCs were activated with influenza or tetanus toxin peptides restricted by HLA-DR1, –DR4, –DR9, or –DR2b, respectively, and tested by IFN-γ ELISPOT assay in the presence of increasing concentrations of PV-267.

As shown in Fig. 2A, PV-267 did not inhibit M17-31 peptide-induced IFN-γ cytokine production by HLA-DR1–restricted T cells. Similarly, T cell responses to the HLA-DR4–restricted HA307–319 and the HLA-DR9–restricted TT830–843 peptide, respectively, were also not inhibited by PV-267 in PBMCs coexpressing these MHC molecules (Fig. 2B). In strong contrast, T cell responses to the HLA-DR2b–restricted TT830–844 peptide were significantly inhibited by PV-267 (p < 0.05), confirming the selectivity of the MHC blocker for HLA-DR2b. In addition, in vivo studies using HLA-DR4 (DRB1*0401) transgenic mice (34) showed that PV-267 treatment did not inhibit Ag-specific cytokine response restricted by non-DR2b HLA molecules, consistent with its HLA-DR2b–specific inhibitory effect (Supplemental Fig. 1).

Taken together, the results showed that PV-276 specifically inhibited T cell responses mediated by HLA-DR2b, but not by PV-267.
HLA-DR1, -DR4, or -DR9 molecules. The data support the selectivity of the inhibitor for HLA-DR2b.

PV-267 does not induce nonspecific immune activation of human PBMC

An important concern with immunomodulatory treatments is the induction of undesired detrimental side effects (16, 35–37). Therefore, we asked whether PV-267 had nonspecific immune-activating effects on T cells or other PBMCs. To address this issue, HLA-DR2b–expressing PBMCs were incubated with PV-267–pretreated autologous APCs and MBS87–99 peptide for human IFN-γ ELISpot assay, as described in Materials and Methods. Shown are representative results of (A) quantified spots at each PV-267 titration, (B) percentage of inhibited response from at least three independent experiments, and (C) effect of PV-267 on BC3 T cells in response to anti-CD3 mAb stimulation as control (mean ± SD). After MBP-specific BC3 T cells were cocultured with PV-267–pretreated autologous APCs and medium, Ag, or positive control (PTX/PHA) for 24 h, supernatants were collected and analyzed using Bio-Plex assay for human cytokines. Shown are pooled results of proinflammatory cytokine production of (D) TNF-α, (E) IL-1ra, (F) IL-1β, (G) IL-6, (H) IL-17, and (I) MIP-1α from three independent experiments (mean ± SD). (J) MBP-specific BC3 T cells were labeled with CFSE and then cocultured with PV-267–pretreated autologous APCs and MBS87–99 or mitogen for 5 d, followed by flow cytometry analysis, as described in Materials and Methods. Shown are pooled results (top) of percentage of proliferated BC3 T cells in response to MBP87–99, anti-CD3 mAb, and PHA (mean ± SD), as well as representative flow cytometry histograms (bottom) from three to four independent experiments. *Indicates significant difference between 0 μg/ml PV-267– and 200 μg/ml PV-267-treated groups (p < 0.05, t test).

Collectively, the results show that PV-267 did not have undesired nonspecific effects on the activation of human T cells or other PBMCs (e.g., monocytes or NK cells) and the production of proinflammatory cytokines. In addition, mice treated daily with up to 400 mg/Kg PV-267 for 2 wk did not show a dose-limiting toxicity.
PV-267 ameliorates EAE in HLA-DR2b transgenic mice

Based on the property of PV-267 to inhibit proliferation and cytokine production by HLA-DR2b–restricted human myelin Ag-specific T cells in vitro, we asked whether the compound showed efficacy in the prevention and/or treatment of EAE in HLA-DR2b transgenic mice as a preclinical model for human MS.

First, we tested whether PV-267 inhibited cytokine production by HLA-DR2b–restricted T cells generated in HLA-DR2b (DRB1*1501) transgenic mice deficient for endogenous murine MHC class II molecules on the C57BL/6 background (27). To address this question, PV-267 was titrated against increasing concentrations of the cognate MBP peptide using a T cell clone specific for MBP84–102 generated from the HLA-DR2b transgenic mice. The results showed that PV-267 inhibited production of IFN-γ by MBP84–102 peptide-specific murine T cells in a dose-

### Table III. Summary of cytokine production of BC3T cells cultured with Ag and PV-267

<table>
<thead>
<tr>
<th>Ag-Stimulated Cytokine Production (pg/ml)</th>
<th>0 µg/ml PV-267</th>
<th>200 µg/ml PV-267</th>
<th>Statistical Difference (*p Value or NS)</th>
</tr>
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<tbody>
<tr>
<td>IL-8</td>
<td>17,427.5 ± 10,983.3</td>
<td>14,791.0 ± 7,191.8</td>
<td>NS</td>
</tr>
<tr>
<td>MIP-1β</td>
<td>3,137.5 ± 2,642.7</td>
<td>2,409.0 ± 652.9</td>
<td>NS</td>
</tr>
<tr>
<td>MCP-1 (MCAF)</td>
<td>1,701.7 ± 1,272.3</td>
<td>2,001.4 ± 1,079.3</td>
<td>NS</td>
</tr>
<tr>
<td>IL-6</td>
<td>1,228.6 ± 729.9</td>
<td>738.8 ± 261.0</td>
<td>NS</td>
</tr>
<tr>
<td>RANTES</td>
<td>905.5 ± 350.3</td>
<td>1,124.9 ± 268.4</td>
<td>NS</td>
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<tr>
<td>IP-10</td>
<td>703.8 ± 734.8</td>
<td>283.3 ± 302.2</td>
<td>NS</td>
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<td>MIP-1α</td>
<td>399.5 ± 258.8</td>
<td>233.2 ± 60.2</td>
<td>NS</td>
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<tr>
<td>PDGF-bb</td>
<td>312.8 ± 50.5</td>
<td>415.9 ± 291.7</td>
<td>NS</td>
</tr>
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<td>TNF-α</td>
<td>225.8 ± 79.3</td>
<td>45.7 ± 34.9*</td>
<td>p = 0.023</td>
</tr>
<tr>
<td>IL-1α</td>
<td>215.6 ± 27.2</td>
<td>128.2 ± 31.5*</td>
<td>p = 0.022</td>
</tr>
<tr>
<td>IL-13</td>
<td>126.7 ± 55.5</td>
<td>60.1 ± 78.5</td>
<td>NS</td>
</tr>
<tr>
<td>VEGF</td>
<td>96.2 ± 20.4</td>
<td>90.7 ± 33.9</td>
<td>NS</td>
</tr>
<tr>
<td>IFN-γ</td>
<td>77.5 ± 22.9</td>
<td>46.2 ± 17.8</td>
<td>NS</td>
</tr>
<tr>
<td>IL-9</td>
<td>35.3 ± 9.9</td>
<td>27.3 ± 9.1</td>
<td>NS</td>
</tr>
<tr>
<td>IL-1b</td>
<td>30.6 ± 13.2</td>
<td>19.7 ± 5.7</td>
<td>NS</td>
</tr>
<tr>
<td>IL-10</td>
<td>17.9 ± 3.1</td>
<td>6.5 ± 2.4</td>
<td>NS</td>
</tr>
<tr>
<td>G-CSF</td>
<td>16.5 ± 9.5</td>
<td>11.8 ± 4.3</td>
<td>NS</td>
</tr>
<tr>
<td>IL-17</td>
<td>15.5 ± 3.6</td>
<td>11.0 ± 1.2</td>
<td>NS</td>
</tr>
<tr>
<td>Eotaxin</td>
<td>&lt;16.4</td>
<td>&lt;23.5</td>
<td>NS</td>
</tr>
<tr>
<td>IL-12 (P70)</td>
<td>9.7 ± 5.4</td>
<td>7.2 ± 5.2</td>
<td>NS</td>
</tr>
<tr>
<td>FGF</td>
<td>6.0 ± 1.1</td>
<td>2.1 ± 0.6*</td>
<td>p = 0.005</td>
</tr>
<tr>
<td>IL-2</td>
<td>4.4 ± 0.4</td>
<td>&lt;2.7*</td>
<td>p = 0.018</td>
</tr>
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<td>IL-4</td>
<td>3.7 ± 0.5</td>
<td>2.5 ± 0.9</td>
<td>NS</td>
</tr>
<tr>
<td>IL-7</td>
<td>2.0 ± 0.4</td>
<td>&lt;3.7</td>
<td>NS</td>
</tr>
<tr>
<td>IL-5</td>
<td>&lt;3.3</td>
<td>&lt;0.3</td>
<td>NS</td>
</tr>
<tr>
<td>GM-CSF</td>
<td>&lt;0.5</td>
<td>&lt;2.8</td>
<td>NS</td>
</tr>
<tr>
<td>IL-15</td>
<td>&lt;0.3</td>
<td>&lt;0.2</td>
<td>NS</td>
</tr>
</tbody>
</table>

MBP-specific BC3 T cells were cultured with Ag MBP87–99 or PHA as positive control and autologous APCs pretreated with PV-267 at different concentrations for 24 h. Supernatants were collected and analyzed using Bio-Plex assay for human cytokines. Shown are the pooled result of the production of 27 human cytokines when BC3 T cells were cultured with Ag and PV-267 (0–200 µg/ml) from at least three independent experiments (mean ± SD).

*Statistically significant.

IP-10, IFN-γ–induced protein 10; MCP-1 (MCAF), monocyte chemotactic protein-1 (macrophage/monocyte chemotactic and activating factor); PDGF-bb, platelet-derived growth factor bb; VEGF, vascular endothelial growth factor.

(data not shown), suggesting a promising general as well as immunological safety profile.

**PV-267 ameliorates EAE in HLA-DR2b transgenic mice**

Based on the property of PV-267 to inhibit proliferation and cytokine production by HLA-DR2b–restricted human myelin Ag-specific T cells in vitro, we asked whether the compound showed efficacy in the prevention and/or treatment of EAE in HLA-DR2b transgenic mice as a preclinical model for human MS.

First, we tested whether PV-267 inhibited cytokine production by HLA-DR2b–restricted T cells generated in HLA-DR2b (DRB1*1501) transgenic mice deficient for endogenous murine MHC class II molecules on the C57BL/6 background (27). To address this question, PV-267 was titrated against increasing concentrations of the cognate MBP peptide using a T cell clone specific for MBP84–102 generated from the HLA-DR2b transgenic mice. The results showed that PV-267 inhibited production of IFN-γ by MBP84–102 peptide-specific murine T cells in a dose-

**FIGURE 2.** PV-267 specifically inhibits DR2- but not DR1-, DR4-, or DR9-mediated T cell responses. T cell–enriched human PBMCs were cocultured with PV-267–pretreated autologous APCs and the indicated Ag and tested by human IFN-γ ELISPOT assay, as described in Materials and Methods. Shown are representative results of the percentage of inhibited response of (A) Cellular Technology donor 4 PBMCs (DRB1*0101/DRB1*1501) to DR1-restricted Ag influenza A matrix protein (M17-31) or DR2-restricted Ag tetanus toxin (TT830–844), and (B) Cellular Technology donor 9 PBMCs (DRB1*0401/DRB1*0901) to DR4-restricted Ag influenza A hemagglutinin (HA307–319) or DR9-restricted Ag tetanus toxin (TT830–843) from at least three independent experiments (mean ± SD). *Indicates significant difference between the inhibition of response to DR1- and DR2-restricted peptide from donor 4 PBMCs (p < 0.05, t test).
dependent fashion (Fig. 4A). In contrast, a non-DR2b–binding control compound (PV-015) did not inhibit T cell responses (data not shown).

Next, we investigated whether PV-267 was efficacious in treating EAE induced by active immunization with the MOG35–55 peptide in the HLA-DR2b transgenic mice. To test whether the compound was efficacious in a prevention mode, the HLA-DR2b transgenic mice were injected once daily with PV-267 (30 mg/kg) or vehicle control (30% Captisol) starting on day 21 before induction of EAE and for 20 d after induction. Disease was induced with MOG35–55 peptide, and the animals were observed for clinical signs of disease for 30 d, as described (29). As shown in Fig. 4B and Supplemental Table I (analysis of cumulative EAE scores), PV-267 efficiently prevented EAE and significantly delayed disease onset and lowered disease scores as compared with the control group. To test the efficacy of PV-267 for treatment of established EAE, disease was induced as outlined above and the mice randomized at the onset of EAE (day 11) into two groups. Subsequently, mice were injected once daily over 10 d with PV-267 (50 mg/kg) or carrier control and observed for clinical EAE. Importantly, treatment with PV-267 significantly ameliorated disease after onset of EAE and the animals recovered faster (Fig. 4C). Of note, the PV-267–treated mice did not show rebound disease after termination of the treatment (Fig. 4C). Analysis of cumulative EAE scores of PV-267–treated and control mice confirmed treatment efficacy of the treatment (Supplemental Table I). Lastly, PV-267 was efficacious when EAE was induced by adoptive transfer of MOG35–55–reactive T cells (Fig. 4D).

Lastly, we examined serum from the EAE mice for possible Ab production against PV-267 due to the injections. However, we did not detect substantial serum IgG production against PV-267 as compared with production of IgG against MOG35–55 in PV-267–treated DR2b transgenic mice (Supplemental Fig. 2).

Taken together, the results showed that PV-267 was safe and highly effective in preventing and treating EAE in HLA-DR2b transgenic mice.

Discussion

In this study, we report development of a small molecule inhibitor of human HLA-DR2b molecules that can curtail myelin-reactive
Table IV. Summary of cytokine production of human PBMCs cultured with PV-267

<table>
<thead>
<tr>
<th>Cytokine Production (pg/ml)</th>
<th>0 μg/ml PV-267</th>
<th>250 μg/ml PV-267</th>
<th>500 μg/ml PV-267</th>
<th>Statistical Difference (p Value or NS)</th>
</tr>
</thead>
<tbody>
<tr>
<td>IL-8</td>
<td>21,481.3 ± 10,022.3</td>
<td>15,684.8 ± 14,390.2</td>
<td>13,472.2 ± 14,245.7</td>
<td>NS</td>
</tr>
<tr>
<td>IL-6</td>
<td>5,303.1 ± 7,899.7</td>
<td>964.0 ± 1,373.6</td>
<td>488.7 ± 677.1</td>
<td>NS</td>
</tr>
<tr>
<td>MIP-1α (MCAF)</td>
<td>4,283.9 ± 3,034.5</td>
<td>4,848.5 ± 5,346.3</td>
<td>4,789.8 ± 4,894.8</td>
<td>NS</td>
</tr>
<tr>
<td>MIP-1β</td>
<td>2,064.8 ± 283.6</td>
<td>917.5 ± 133.1*</td>
<td>618.1 ± 168.3*</td>
<td>* p ≤ 0.003</td>
</tr>
<tr>
<td>RANTES</td>
<td>586.4 ± 860.6</td>
<td>100.5 ± 129.7</td>
<td>46.4 ± 51.8</td>
<td>NS</td>
</tr>
<tr>
<td>IP-10</td>
<td>388.0 ± 328.9</td>
<td>549.3 ± 458.9</td>
<td>6,991.3 ± 8,147.6</td>
<td>NS</td>
</tr>
<tr>
<td>IL-1α</td>
<td>255.9 ± 166.4</td>
<td>1,219.2 ± 961.2</td>
<td>8,307.3 ± 4,559.8</td>
<td>NS</td>
</tr>
<tr>
<td>IL-2</td>
<td>178.8 ± 90.4</td>
<td>201.1 ± 109.9</td>
<td>151.2 ± 69.4</td>
<td>NS</td>
</tr>
<tr>
<td>PDGF-bβ</td>
<td>153.4 ± 114.4</td>
<td>250.0 ± 255.2</td>
<td>1,251.4 ± 709.6</td>
<td>NS</td>
</tr>
<tr>
<td>TNF-α</td>
<td>133.2 ± 144.8</td>
<td>12.5 ± 15.3</td>
<td>7.0 ± 3.1</td>
<td>NS</td>
</tr>
<tr>
<td>VEGF</td>
<td>116.6 ± 38.1</td>
<td>111.3 ± 61.7</td>
<td>125.3 ± 58.3</td>
<td>NS</td>
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<tr>
<td>G-CSF</td>
<td>61.9 ± 86.6</td>
<td>9.6 ± 12.8</td>
<td>6.0 ± 7.6</td>
<td>NS</td>
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<tr>
<td>IL-1b</td>
<td>57.8 ± 66.7</td>
<td>20.5 ± 25.7</td>
<td>12.7 ± 15.9</td>
<td>NS</td>
</tr>
<tr>
<td>IFN-γ</td>
<td>49.7 ± 42.9</td>
<td>28.0 ± 18.4</td>
<td>26.9 ± 6.2</td>
<td>NS</td>
</tr>
<tr>
<td>Eotaxin</td>
<td>&lt;21.3</td>
<td>&lt;15.7</td>
<td>&lt;1.6</td>
<td>NS</td>
</tr>
<tr>
<td>IL-3</td>
<td>19.7 ± 22.2</td>
<td>4.3 ± 2.9</td>
<td>2.0 ± 1.7</td>
<td>NS</td>
</tr>
<tr>
<td>IL-9</td>
<td>16.7 ± 7.0</td>
<td>16.0 ± 8.1</td>
<td>12.3 ± 4.8</td>
<td>NS</td>
</tr>
<tr>
<td>IL-17</td>
<td>11.5 ± 4.9</td>
<td>5.5 ± 4.7</td>
<td>4.3 ± 3.1</td>
<td>NS</td>
</tr>
<tr>
<td>FGF</td>
<td>9.9 ± 3.3</td>
<td>&lt;4.5*</td>
<td>2.7 ± 12.8*</td>
<td>* p ≤ 0.03</td>
</tr>
<tr>
<td>GM-CSF</td>
<td>&lt;9.8</td>
<td>&lt;6.9</td>
<td>&lt;1.3</td>
<td>NS</td>
</tr>
<tr>
<td>IL-12 (p70)</td>
<td>9.9 ± 0.9</td>
<td>4.5 ± 1.2*</td>
<td>2.5 ± 2.0*</td>
<td>* p ≤ 0.004</td>
</tr>
<tr>
<td>IL-2</td>
<td>3.0 ± 1.8</td>
<td>6.9 ± 3.3</td>
<td>6.4 ± 2.3</td>
<td>NS</td>
</tr>
<tr>
<td>IL-7</td>
<td>2.6 ± 1.7</td>
<td>0.3 ± 0.5</td>
<td>0.6 ± 0.5</td>
<td>NS</td>
</tr>
<tr>
<td>IL-4</td>
<td>2.0 ± 1.3</td>
<td>1.2 ± 0.6</td>
<td>1.4 ± 0.2</td>
<td>NS</td>
</tr>
<tr>
<td>IL-13</td>
<td>1.3 ± 0.6</td>
<td>&lt;0.3*</td>
<td>&lt;0.3*</td>
<td>* p ≤ 0.043</td>
</tr>
<tr>
<td>IL-15</td>
<td>&lt;0.5</td>
<td>&lt;0.5</td>
<td>&lt;0.19</td>
<td>NS</td>
</tr>
<tr>
<td>IL-5</td>
<td>&lt;0.2</td>
<td>&lt;0.2</td>
<td>&lt;0.2</td>
<td>NS</td>
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</tbody>
</table>

**Human PBMCs were cultured for 24 h with PV-267 at different concentrations, or with positive control (PTX or PHA). Supernatants were collected and analyzed using Bio-Plex assay for human cytokines. Shown are the summary of the level and profile of 27 human cytokines when PBMCs were cultured with PV-267 (0-250 μg/ml or maximum 500 μg/ml) from at least three independent experiments of three human PBMC donors.**

*Statistically significant.

**IP-10, IFN-γ-inducd protein 10; MCP-1 (MCAF), monocyte chemotactic protein-1 (macrophage/monocyte chemotactic and activating factor); PDGF-bβ, platelet-derived growth factor bb; VEGF, vascular endothelial growth factor.**

T cell responses. The lead compound used in these studies, PV-267, inhibited cytokine production and proliferation by MBP87–99-specific HLA-DR2b–restricted CD4+ T cells derived from a MS patient. Furthermore, PV-267 similarly inhibited murine MBP-reactive T cells in a HLA-DR2b transgenic mouse model and ameliorated EAE both in prevention and treatment mode. Importantly, PV-267 did not have nonspecific immune-activating effects on human PBMCs in vitro and did not result in upregulation of activating/costimulatory receptors and production of cytokines.

**FIGURE 4.** PV-267 inhibits IFN-γ production by HLA-DR2b–restricted, MBP84–102–specific T cells and ameliorates EAE in HLA-DR2b mice. MBP84–102–specific T cell clone derived from HLA-DR2b transgenic mice was cultured with APCs, cognate MBP peptide at increasing concentrations, and PV-267 was titrated as indicated, and cytokine production was assessed by IFN-γ ELISPOT assay (A). HLA-DR2b transgenic mice were immunized with MOG35–55 peptide, then monitored and scored daily for clinical EAE, as described in Materials and Methods. Mice were injected i.p. with (B) 30 mg/kg PV-267 or control daily starting on day −1 for 20 d, or (C) 50 mg/kg PV-267 or control daily starting upon disease onset (day 11) for 10 d (n = 10–12 mice per group, mean ± SD). (D) HLA-DR2b transgenic recipient mice were adoptively transferred with encephalitogenic MOG35–55 peptide–specific donor cells, as described in Materials and Methods. Recipients were also injected i.p. with 50 mg/kg PV-267 or control daily starting on day −2 for 15 d (n = 4 or 5, mean ± SEM). Shown are the representative results of three independent experiments. *Indicates significant difference between PV-267– and vehicle-treated groups, *p < 0.05, **p < 0.01, t test.
The mode of action of PV-267 is proposed to be via specific binding of the molecule to the peptide groove of the HLA-DR2b molecule and thus blocking of Ag recognition by T cells. This interpretation is based on the observation that inhibition of T cell responses was only observed with HLA-DR2b–restricted T cell responses, but not with T cell responses mediated by other human MHC II molecules such as HLA-DR1, DR4, or DR9. In line with specific inhibition of HLA-DR2b:peptide presentation, PV-267 did not show an effect when T cells or PBMC were activated with anti-CD3 mAb, which circumvents peptide recognition by T cells. In addition, incubation of human PBMC or T cell lines with PV-267 did not result in increased T cell death, arguing against cytotoxic effects of the compound.

Silencing pathogenic T cells via inhibition of HLA-DR2b–mediated presentation of neuroantigens, as shown in this study, advances the concept of specific immunotherapy without invoking undesired immunomodulatory effects observed in clinical trials of oral tolerance, altered peptide ligands, T cell vaccination, rTCR peptide therapy, etc. (8, 12, 13). Most of these approaches have shown activity in preclinical animal models but were of limited efficacy in clinical trials (8, 15, 38). Concerns were failure to show sufficient clinical efficacy as compared with placebo as well as development of serious side effects. Thus, most of these approaches have been abandoned and strategies have shifted to broad-based immunomodulatory therapies targeting widely expressed molecules, such as homing receptors and inflammatory mediators. However, targeting of molecules expressed by many cell types, such as VLA-4, brings with it the inherent risk of global immunosuppression, as well as the potential to have significant undesired immunomodulatory and/or immune-activating effects. The potentially dire consequences were clearly evidenced by development of progressive multifocal leukoencephalopathy in MS patients treated with natalizumab due to reactivation of John Cunningham virus (16).

HLA-DR2b has repeatedly been confirmed as the most important gene complex linked to MS (39). Although the exact reasons for this genetic linkage have not yet been elucidated, it is conceivable that the critical contribution of these molecules is via presentation of peptide Ags to myelin-reactive T cells, resulting in their activation, proliferation, and production of pathogenic cytokines. Thus, inhibition of Ag presentation specifically by the MS-associated HLA-DR2b molecules is a conceptually attractive means to prevent the activation of pathogenic T cells in MS patients. This inhibition could take effect both in peripheral lymphatic tissues as well as directly in the CNS and would be independent of the nature of the autoantigenic peptides.

To block HLA-DR2b, several key factors needed to be addressed. First, an inhibitor would need to bind to HLA-DR2b with high affinity and selectivity to be able to compete with Ag binding in the endosomal loading compartment of APCs and in the presence of HLA-DM, a catalyst for peptide–MHC exchange (40, 41). Second, because the endosomal compartment of the APCs is rich in cathepsin enzymes that process MHC molecules and degrade protein Ags into peptide fragments that bind the MHC, an inhibitor would need to be stabilized against cathepsins as well as show stability in plasma. Of note, once a peptide (Ag or inhibitor) is loaded, the MHC complex translocates to the cell surface, where it has a very low off-rate and remains bound and is subject to a turnover rate ranging in days. Next, an effective inhibitor, to be differentiated from the other strategies described, needs to be highly selective in binding to the disease-associated DR2b allele (in MS) to avoid interfering with immune responses to pathogens via other MHC molecules. In effect, this would make this a personalized medicine approach targeted toward MS patients carrying the HLA-DR2b allele. Finally, the inhibitor needed to be immunologically inert as to not stimulate an immune response on its own.

In keeping with the above outlined key requirements, development of a HLA-DR2b–selective inhibitor appeared to be feasible based on previously established differences in key positions in the Ag-binding pocket between different MHC class II molecules. The unique characteristics of the binding pocket of DRB1*1501 may be attributed to the small Ala residue at position 71 (p71) of the MHC β-chain. This opens up the space for a larger P4 residue in the inhibitor sequence as a way of achieving selectivity versus DRB1*0101 and DRB1*0401 (where p71 is positively charged [Arg or Lys] in these DR1/DR4 alleles, respectively, and the chain length blocks off space so that the Phe residue is a good fit to DR2b, but too large to bind DR1/DR4). Also, the hydrophobic nature of the P1 pocket is a characteristic of DR alleles, whereas this site is polar in MHC DQ alleles. Thus, a Chg group at P1 selects for DR versus DQ, and a Phe at P4 selects for DR2b versus DR1/DR4. The small p71 Ala group is unique to DR2b, so selectivity can be achieved with ligands that have larger P4 groups (e.g., Phe) that would not fit the corresponding pocket in DR1, where p71 is Arg, or DR4, where it is Lys.

An important consequence of selectively blocking HLA-DR2b in MS patients is that it should inhibit the autoimmune T cell responses associated with MS, but not affect T cell responses mediated via other MHC alleles. Thus, a HLA-DR2b inhibitor should not impair protective immunity toward microbial pathogens mediated by other MHC class II alleles. Our results suggest the feasibility of this presumption by showing that PV-267 did not impair T cell responses to peptides derived from microbial pathogens restricted by other MHC class II molecules such as HLA-DR1, -DR4, and -DR9.

Another key conceptual benefit of inhibiting Ag presentation by the MS-associated DR2b allele is its utility in globally inhibiting autoreactive T cells irrespective of their specificity for particular autoantigenic peptides. In other words, by blocking HLA-DR2b itself, it becomes irrelevant whether autoimmune T cells respond to MBP, MOG, or proteolipid protein peptides, etc., as long as they are restricted by HLA-DR2b. Furthermore, this approach eliminates potential concerns over epitope spreading (42, 43).

Because MS and other autoimmune diseases are chronic, lifelong diseases, they may require long-term treatment, and thus, accumulation of long-term side effects is a major concern. For example, treatments that deplete immune cells, even if specific for certain subsets of cells such as B cells, will incur the potential to cause significant harm if applied over a longer period of time. Often, it is difficult to predict serious side effects, as, for example, evidenced by development of lymphomas in rheumatoid arthritis or Crohn’s disease patients treated with TNF immunomodulators (44–46). Thus, treatments with as few side effects as possible are highly desirable, and, conceptually, MHC II inhibitors selective for one MHC class II allele could fulfill that tenet. First, as outlined earlier, protection from infectious microorganisms would still be mediated via the other MHC class II alleles expressed in an individual. Furthermore, because the only function of MHC blockers is to occupy the peptide-binding groove of one specific MHC II allele without having any other effect on the immune system, including being nontoxic, detrimental side effects are not expected even over a prolonged course of treatment. Our results showing a lack of effect of PV-267 on the activation or cytokine production by T cells or other PBMCs support this view and suggest that long-term treatment with this class of molecules may be feasible without resulting in detrimental side effects found with immunomodulatory drugs. Importantly, lack of side effects such as flu-like symptoms in patients treated with IFN-β will facilitate treatment adherence and conceivably improve long-term outcomes.
Furthermore, the lack of side effects may facilitate development of prevention strategies for people at risk but not yet affected by MS, such as monozygotic twins of MS patients who have a 25% risk to develop the disease (47, 48) or in patients with diagnoses of clinically isolated syndrome.

Acknowledgments

We acknowledge the following who have contributed to the efforts described in this work: 1) peptide synthesis, Nallaganchu Rao, Hong Chen, Bernard Maillere and Lora Hamuro; 3) modeling, Charles Cook; 4) DR2b described in this work: 1) peptide synthesis, Nallaganchu Rao, Hong Chen, Bernard Maillere and Lora Hamuro; 3) modeling, Charles Cook; 4) DR2b.

Disclosures

G.L.O. is the president and chief executive officer of Provio, and C.S. is an employee of Provio.

References

SUPPLEMENTARY FIGURE 1. PV-267 does not inhibit INF-γ production by
DRB1*04:01-restricted, MOG97-108 specific T cell responses. HLA-DR4 transgenic
mice were immunized with MOG97-108 peptide in CFA, and injected i.p. with 30 mg/kg
of PV-267 or control daily starting one day before immunization and continuing for 12 d.
On d 10 splenocytes and DLN cells were harvested and tested for Ag-specific IFN-
responses by mouse cytokine ELISPOT assay as described in Materials and Methods.
Shown are representative results of 2 independent experiments (n = 4 mice per group;
mean ± SEM). NS, no significant difference between PV-267 and control groups (t-
test).

SUPPLEMENTARY FIGURE 2. Treatment with PV-267 does not induce substantial
production of specific Ab in serum of mice. Sera were collected from adoptive transfer
recipient HLA-DR2b transgenic mice around d 15 and measured for Ag-specific mouse
IgG against MOG35-55 or PV-267 by ELISA assay as described in Materials and
Methods. Shown are representative result of 2 independent experiments (n = 4 mice
per group; mean ± SEM).
Cumulative EAE score was calculated by adding total EAE scores from onset until end of study or before recovery for individual mice and dividing by the number of mice.

### Supplementary Table 1: cumulative EAE scores and statistical analysis of active immunized EAE mice treated with PV-267

<table>
<thead>
<tr>
<th>Disease study / corresponding Figure</th>
<th>Time period for cumulative scores</th>
<th>Control (mean ± SD)</th>
<th>PV-267 (mean ± SD)</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Preventive / Fig. 4B</td>
<td>d 12 – 30</td>
<td>28.2 ± 19.8</td>
<td>7.8 ± 11.6 *</td>
<td>P = 0.009</td>
</tr>
<tr>
<td>Treatment / Fig. 4C</td>
<td>d 9 – 40</td>
<td>35.1 ± 16.3</td>
<td>19.2 ± 10.2 *</td>
<td>P = 0.023</td>
</tr>
</tbody>
</table>
Supplementary figure 2

Graph showing the mean OD (450 nm) against different serum titrations. The graph compares various control and antibody treatments. The legend indicates:
- Control: MOG35-55 Ab
- Control: PV-267 Ab
- PV-267: MOG35-55 Ab
- PV-267: PV-267 Ab

The x-axis represents the serum titration (1/200 to 1/3200), while the y-axis shows the mean OD values from 0.0 to 1.0.