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Fast Dendritic Cells Stimulated with Alternative Maturation Mixtures Induce Polyfunctional and Long-Lasting Activation of Innate and Adaptive Effector Cells with Tumor-Killing Capabilities

Chiara Massa and Barbara Seliger

The clinical usage of dendritic cells (DC) for tumor immunotherapy still requires improvements. In this study, three alternative maturation mixtures were compared with the cytokine-based gold standard, and the overall interaction of the resulting DC with effector cells from the innate as well as the adaptive immunity was evaluated in healthy donors. Stimulation with the TLR-4 ligand monophosphoryl lipid A together with IFN-γ (alt-2 DC) resulted in DC with the highest levels of costimulatory molecule expression and IL-12p70/IL-10 ratio. Whereas all alternative DC were able to induce NK and γδ T cells to acquire cytotoxic properties and secrete type 1 and proinflammatory cytokines, after both short (20-h)- and long (5–8 d)-time coculture, secretion of IFN-γ by the innate populations was induced in response to alt-2 and alt-1 DC (TNF-α, IFN-γ, IFN-γ, IL-1β, poly IC), but not to alt-3 DC (TNF-α, IFN-γ, IL-1β, CL097). Regarding CD8⁺ T cell–mediated Ag-specific immune responses, a heterogeneous pattern of responses was obtained among the healthy donors, suggesting rather a competition than a synergy among the different effector cells. Our data promote further evaluation of alt-2 fast DC for translatability into clinical immunotherapy trials, while also fostering the need to identify biomarkers for immune cell responsiveness and tumor susceptibility to be able to select for each patient the best possible DC-based therapy. The Journal of Immunology, 2013, 190: 000–000.

Dendritic cells (DC) are considered the most potent APCs of the immune system and have been investigated to employ their adjuvant activity in tumor immunotherapy. Clinical trials in different tumor entities have revealed safety and tolerability of such treatment. However, despite a frequent vaccine-induced expansion of precursor cells in the blood and/or the induction of delayed-type hypersensitivity, the rate of objective clinical responses obtained in tumor patients is very limited (1, 2). For this reason, an optimization of the protocol for vaccine DC production as well as their deep functional characterization is urgently needed to improve their clinical implementation.

The major protocol currently used for the production of vaccine DC is a two-step process, in which monocytes are first differentiated toward immature DC by a 5-d culture in the presence of GM-CSF and IL-4 (3), followed by their stimulation with the so-called gold standard cytokine mixture composed of TNF-α, IL-1β, IL-6, and PGE₂ (4). The major disadvantages of this protocol are the labor- and time-consuming procedures together with an impaired production as well as their deep functional characterization is urgently needed to improve their clinical implementation. The protocol currently used for the production of vaccine DC is a two-step process, in which monocytes are first differentiated toward immature DC by a 5-d culture in the presence of GM-CSF and IL-4 (3), followed by their stimulation with the so-called gold standard cytokine mixture composed of TNF-α, IL-1β, IL-6, and PGE₂ (4). The major disadvantages of this protocol are the labor- and time-consuming procedures together with an impaired secretion of the bioactive IL-12p70 that is critical for the induction of a type 1–oriented, cell-mediated immune response (5, 6). Therefore, alternative protocols for a faster differentiation of monocytes (7, 8) and a better maturation toward vaccine DC (9–12) have been developed during the last decade.

Despite the fact that the major aim of vaccination is the induction of a CD8⁺ T cell–mediated, Ag-specific immune response, innate effector cells have recently rekindled interest. NK, NK-T, and γδ T cells have been frequently defined as a first line of response influencing the outcome of the adaptive immunity (13). For example, NK cells have been functionally and phenotypically dissected into two different subsets, the cytotoxic NK cells characterized by a dim expression of the CD56 marker and positivity for CD16 (CD56dimCD16⁺) and a CD16-negative CD56bright (CD56brCD16⁻) cytokine producer subset (14). Moreover, murine models of viral infection as well as immunomonitoring of infected humans suggest that NK cells might develop memory-like properties and can thus provide stronger and quicker responses at a second encounter (15–17).

Based on these data, the potency of newly proposed protocols for vaccine DC should be evaluated regarding the adaptive and innate immunity. Therefore, a comparative analysis of the effects of different maturation mixtures on the interactions of fast DC with the different immune cell populations of the human blood was performed.

Materials and Methods

Maturation stimuli, Abs, and ELISA kits

Monophosphoryl lipid A (MLA) and the imidazoquinoline compound CL097 were purchased from InvivoGen (San Diego, CA), IL-1β from BioSource International (Camarillo, CA), IFN-α2b from ProSpec (Rehovot, Israel), and IFN-γ from R&D Systems (Minneapolis, MN). IFN-γ Fc chimera (IFN-γ-Fc) was purchased from BD Biosciences (San Diego, CA). Purified monoclonal Abs against human CD14 (clone M60, BD Biosciences), CD16 (clone 3G8, BD Biosciences), CD56 (clone NKK-L1/3C11, BD Biosciences), CD8 (clone S14, BD Biosciences), PD-1 (clone G6.2C5, BD Biosciences), PD-L1 (clone L200, BD Biosciences), and MCRC-1 (clone 2B4, BD Biosciences) were purified from hybridoma supernatants by protein A affinity chromatography. The following Abs were purchased from BD Biosciences: anti-CD3 (clone OKT3), anti-CD4 (clone 1F4), and anti-CD8 (clone Q5/1.2). Anti-CD19 (clone HIB19) and anti-CD20 (clone L27) were purchased from BD Biosciences and Beckman Coulter (Brea, CA), respectively. Anti-CD69 (clone H1.2F3), anti-CD80 (clone 25–17–1), anti-CD86 (clone GL1X3), and anti-CD83 (clone LA27) were purchased from BD Biosciences. Anti-CD11c (clone B11.41E6), anti-CD58 (clone CD58/13), and anti-CD141 (clone 141/3A5) were purchased from BD Biosciences and Beckman Coulter, respectively. Anti-CD123 (clone 8G12) Abs were purchased from BD Biosciences and InvivoGen. Anti-CD141 Abs were purchased from BD Biosciences and Beckman Coulter. Anti-CD27 (clone M-TAC/27A), anti-CD31 (clone 2B11), and anti-CD49d (clone IA4) Abs were purchased from BD Biosciences. Anti-CD161 (clone 3B5) Abs were purchased from BD Biosciences and eBioscience (San Diego, CA). Anti-CD25 (clone PC61), anti-gamma (clone 3G5), and anti-CD161 (clone 3B5) Abs were purchased from BD Biosciences. Anti-aHLA (clone 173), anti-aHLA (clone 151), anti-aHLA (clone aHLA110), and anti-aHLA (clone aHLA116) Abs were purchased from BD Biosciences. Anti-CD161 (clone 3B5) Abs were purchased from BD Biosciences and eBioscience. Anti-CD27 (clone M-TAC/27A), anti-CD31 (clone 2B11), and anti-CD49d (clone IA4) Abs were purchased from BD Biosciences. Anti-CD161 (clone 3B5) Abs were purchased from BD Biosciences and eBioscience. Anti-CD27 (clone M-TAC/27A), anti-CD31 (clone 2B11), and anti-CD49d (clone IA4) Abs were purchased from BD Biosciences. Anti-CD161 (clone 3B5) Abs were purchased from BD Biosciences and eBioscience.

The online version of this article contains supplemental material.

Abbreviations used in this article: BD, Becton Dickinson; DC, dendritic cell; HSD, honestly significant difference; KIR, killer cell Ig-like receptor; MPLA, monophosphoryl lipid A; polyIFC, polyinosinic-polyricydilyic acid.

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voll, Israel), IFN-γ from Immunotools (Friesoythe, Germany), polyclinics: polycystic acid (polyC) and PGE2 from Sigma-Aldrich (St. Louis, MO), TNF-α from PeproTech (Hamburg, Germany), whereas IL-6 was a gift of S. Rose-John (Institute of Biochemistry, University of Kiel, Kiel, Germany).

The following Abs were used: Alexa Fluor700 anti-CD3, eFluor450 anti-CD8a, PE-Cy7 anti-CD14, FITC anti-CD80, FITC anti-CD83, PE anti-CD86 (from eBioscience/NatuTec, Frankfurt, Germany), PE anti-CD-40, FITC anti-CD54, PE anti-CD60, PE-Cy7 anti-CD56, allophycocyanin, Alexa Fluor750 anti-CD16 (from Beckman Coulter, Brea, CA), FITC anti-granzyme B, FITC and PerCP anti-V62, allophycocyanin anti-CD107a (from BioLegend, San Diego, CA), PE anti-CD69, and allophycocyanin anti-CD25 (from Becton Dickinson [BD] Biosciences, Franklin Lakes, NJ).

A minimum of 10,000 events gated on live, propidium iodide–negative cells was acquired either on a FACScan (BD Biosciences) or a Navios (Beckman Coulter) flow cytometer and evaluated using CellQuest (BD Biosciences) or Kaluza (Beckman Coulter) software. The general gating strategy for the flow cytometric acquisition is shown in Fig. 1.

Concentration of IL-12p70 and IL-10 in DC supernatants was evaluated using commercially available ELISA kits (BioLegend and Immunotools, Friesoythe, Germany, respectively). For lymphocyte characterization, IFN-γ and TNF-α secretion were evaluated using the secretion assay kit from Miltenyi Biotec (Bergisch Gladbach, Germany), whereas the cytokine content of the culture supernatants was evaluated by means of the flow-cytomix Th1/Th2/Th9/Th17/Th22 kit from ebioscience/NatuTec following manufacturer’s instructions.

**DC differentiation and maturation**

 Buffy coats from HLA-A–typed healthy donors were obtained from the blood bank of the University Hospital in Halle upon written informed consent. PBMCs were purified by a density gradient centrifugation on a Percoll layer (Pau-Biotec, Aidenbach, Germany), and then monocytes were isolated using anti-CD14–conjugated microbeads (Miltenyi Biotec). Differentiation was performed by seeding 10⁶ cells/ml in serum-free CellGiro medium (CellGenix, Freiburg, Germany) with 200 ng/ml GM-CSF (Leukine; Bayer HealthCare, Seattle, WA) and 1000 U/ml IL-4 (Immunotools) for 4–24 h to obtain undifferentiated fast DC. Maturation was induced by 18-h incubation in the presence of the gold standard (TNF-α + IL-1β + IL-6 + PGE2) and the three alternative mixtures, alt-1 (also known as α-type one polarizing; TNF-α + IL-1β + IFN-α + IFN-γ + polyIC) (10), alt-2 (MPLA + IFN-γ) (11), and alt-3 (TNF-α + IL-1β + IFN-γ + CL097; adapted from Ref. 12). The concentrations used in the maturation mixtures were as follows: CL097, 3 μg/ml; IFN-α, 3000 U/ml; IFN-γ, 500 U/ml; IL-1β, 5 ng/ml; IL-6, 5 U/ml; MIP-α, 4 μg/ml; PGE2, 1 μg/ml; polyIC, 1 μg/ml; TNF-α, 100 ng/ml.

At the end of the maturation period, supernatants were frozen for cytokine evaluation by ELISA, whereas DC were thoroughly washed and used for functional (flow cytometry) and functional characterization (coulter assays).

**DC lymphocyte coculture**

 All cocultures were performed in serum-free X-vivo15 medium (Lonza, Basel, Switzerland), using as effector cells the autologous PBL corresponding to the fraction of PBMCs remaining after CD14 monocyte purification. When specified, CD8+ T cells were purified and CD56+ cells were depleted using microbeads following manufacturer’s instructions (Miltenyi Biotec).

For short-term cultures (4–21 h), 4 × 10⁶ DC/well were seeded in U-bottom 96-well plates with 10⁶ PBL in 200 μl, whereas for long-time culture (5–10 d), 10⁵ DC were incubated with 10⁶ PBL in 1 ml in 24-well plate.

For mechanisms evaluation, DC were preincubated with blocking Abs directed against IL-12p, IL-10 (Gen-probe, San Diego, CA) for 30 min at 37°C before lymphocytes were added; the mAb concentration in the final volume was 10 μg/ml.

For evaluation of Ag-specific immune responses, mature DC were pulsed with synthetic peptides for 2 h at 37°C before incubation with effector cells for 10 d. To evaluate the expansion of naïve CTL, the MelanA/Mart-1 peptide (AAGIGILTV; Peptides & Elephants, Potsdam, Germany) was used due to the high frequency of MelanA/Mart-1 precursors in healthy donors (18). In such case, purified CD8+ T cells were purified and total, or NK-depleted PBL were incubated with peptide-pulsed DC, and 50 μU/ml IL-2 (Prolengo, Biotest, Friesoythe, Germany) for 24 h (24-h cytotoxicity assay) or 10 ng/ml IL-7 (ImmunoTools) were added on days 3, 4, 5, and 7. On day 10, cultures were stained with PE MelanA/Mart-1–specific tetramers provided by Beckman Coulter following manufacturer’s instructions. Cultures containing expanded MelanA/
of these molecules, whereas alt-3 induced the lowest levels, which were for CD83 and CD86 even lower than that of the gold standard.

In line with published results, all alternative mixtures employed were able to induce the secretion of the bioactive IL-12p70 during DC maturation, whereas the gold standard induced no or very low levels of this cytokine (Fig. 2B, left). Furthermore, alt-1 and alt-3 induced high levels of IL-10 that were absent in the supernatants of gold standard and alt-2 DC (Fig. 2B, right). Upon reseeding in the absence of exogenous stimuli, the alt-1 and alt-2 DC were still able to secrete significant, but much lower levels of IL-12, whereas alt-1 DC also maintained a significant IL-10 secretion (Fig. 2C). Interaction with autologous PBL was able to induce IL-
12 secretion by alt-1 and alt-2 DC both when occurring immediately after maturation and after 24 h, during which the DC were left untreated (Fig. 2D), a setting mimicking the time required by vaccine DC for migration to the lymph node upon in vivo injection.

Alternative DC strongly stimulate innate cells in short-time culture

To analyze the functional properties of the various preparations, DC were incubated overnight (16–21 h) with autologous lymphocytes, and the culture supernatants were evaluated for the presence of cytokines corresponding to the different types of immune polarization, namely proinflammatory (IL-6 and TNF-α), type 1 (IL-1β, IL-2, and IFN-γ), type 2 (IL-4, IL-5, IL-10, and IL-13), type 9 (IL-9), type 17 (IL-17A), and type 22 (IL-22) cytokines. A potent secretion of IL-1β, IL-6, IFN-γ, and TNF-α was found in response to the coculture with the alternative DC, with alt-1 and alt-2 inducing higher levels than alt-3 DC, whereas almost no cytokines were detected in response to stimulation with gold standard DC. Regarding the other tested cytokines, only IL-10 was consistently detected among different donors and significantly produced uniquely in response to alt-1 DC (Fig. 3A). To dissect the responsible immune cell population(s), a cytokine secretion assay was performed and immune cells were then counterstained with CD3, CD56, and V82 Abs to identify NK (CD3-CD56+) and γδ T cells (CD3- Vδ2+) following the gating strategy illustrated in Fig. 1. Despite the highly variable frequencies in the different donors, NK and γδ T cells responded to the overnight incubation with the alternative DC by secreting IFN-γ and TNF-α (Fig. 3B). A consistent secretion of IFN-γ by the innate populations was observed already after 4-h incubation with the alt-1 and alt-2 DC (Fig. 3C). Further staining with the CD16 Ab to dissect NK cell subsets revealed that both the CD56dimCD16+ and CD56dimCD16- subpopulations secreted IFN-γ after 4 (Fig. 3D) as well as after 16 h (data not shown).

Overnight stimulation with the alternative DC also resulted in the upregulation of activation markers like CD25 and CD69 (Supplemental Fig. 1) as well as in the induction of cytolytic functions. Indeed, innate cells stimulated with the alternative DC displayed an enhanced degranulation in response to Daudi cells in comparison with gold standard–stimulated effectors (Fig. 4A). Concerning the NK cell subpopulations, not only the CD56dimCD16+ population, but also the CD56+NK CD16- cells were induced by alt-1 (data not shown) and alt-2 DC to degranulate upon incubation with Daudi cells (Fig. 4B, top and center). Intracellular staining revealed a dramatic induction of granzyme B in the CD56dimCD16- NK population by alt-1 (data not shown) and alt-2 DC (Fig. 4C, top), suggesting that their degranulation could also be associated with target killing. In contrast, in CD56dimCD16+ NK and γδ T cells, the already high expression of granzyme B was not significantly augmented by stimulation with the alternative DC (Fig. 4C, center and bottom).

Regarding the γδ T cell population, a highly donor-dependent expression of the CD56 marker, ranging from 0–50% of positive cells, was found (data not shown). Its expression did not strictly correlate with the effector function, because the frequency of degranulation was comparable in both CD56-positive and -negative cells, or was even slightly more pronounced in the negative subset (Fig. 4C, bottom).

Alternative DC have long-time effects on innate cells

Interaction between alternative DC and innate cells also had long-term consequences, as follows: an enhanced proliferation of innate cells was demonstrated by staining of lymphocytes with a prolifer-

![FIGURE 3.](http://www.jimmunol.org/) Alternative DC induce cytokine secretion from innate lymphocytes. Mature DC were thoroughly washed and seeded with autologous total PBL in X-vivo 15 medium for 16–21 (A, B) or 4 h (C, D). At the end of the incubation, supernatants were collected and evaluated for the concentration of cytokines corresponding to the different immune polarization using multiplex technology (A), whereas lymphocytes were washed and evaluated for production of IFN-γ and TNF-α by means of a cytokine secretion assay (B–D) using the gating strategy described in the legend of Fig. 1. (A–C) Reported are the mean ± SE of at least three different experiments. (D) Histogram overlay of one representative donor evaluated after 4-h coculture. *p < 0.05, **p < 0.01 in the Tukey HSD test.

Expanded innate effector cells also retained the enhanced responsiveness to target stimulation. A higher percentage of lymphocytes secreting IFN-γ was recorded after 5–8 d of coculture with the alternative DC upon challenge with Daudi cells (Fig. 6A).
This was accompanied by an enhanced degranulation and also effective killing of the target cells as measured in a classical [51Cr] release assay (Fig. 6B). Effector cells also acquired the ability to kill tumor cells, as representatively shown for the melanoma cell line Mel1379 (Fig. 6C).

**FIGURE 4.** Alternative DC enhance cytotoxic potential of innate lymphocytes. (A and B) PBL were incubated for 16–21 h with mature DC in X-vivo 15, thoroughly washed, and then tested in a CD107a degranulation assay upon 4-h incubation at a 1:1 ratio with Daudi cells. The different effector populations were identified upon counterstaining with mAb directed against CD3, CD56, CD16, and V62, as described in legend to Fig. 1. (A) The mean ± SE of at least three different experiments of target-induced degranulation (i.e., upon subtraction of the background CD107a exposition obtained with PBL incubated alone) are shown. *p < 0.05, **p < 0.01 in the Tukey HSD test. (B) Representative plot from alt-2 DC-stimulated PBL is shown for CD56dim and CD16+CD56dim NK as well as γδ T cells. Gates were set based on the background CD107a staining of PBL incubated alone. (C) PBL stimulated 16 h with mature DC were fixed, permeabilized, and stained for granzyme B. Representative histogram overlays of one of three donors are shown for CD56dim and CD16+CD56dim NK as well as γδ T cells upon stimulation with gold standard and alt-2 DC.

**FIGURE 5.** Sustained expansion of innate cells by alternative DC. PBL were labeled with cell proliferation dyes (CFSE or eFluor670) and incubated with autologous DC (10:1 ratio) in X-vivo 15 in the absence of exogenous Ags or cytokines. After 5 d, proliferation was evaluated by flow cytometry as dye fluorescence dilution in the differently gated lymphocyte populations (A–C), whereas absolute cell counts were calculated using counting beads and reported upon normalization to PBL in the absence of any DC stimulation (D). The results are represented as the mean ± SE of 10 (A) and four different experiments (D), respectively. Representative dot plots of gold standard and alt-2 DC-stimulated NK (B) and γδ T cells (C) are also shown. *p < 0.05, **p < 0.01 in the Tukey HSD test.

Different requirements for cytokine secretion and development of cytotoxicity

To dissect the mechanisms of the enhanced innate activation, DC were incubated with PBL in the presence of the isotype control or blocking Abs directed against IL-12p70 prior to their functional
Upon 16-h incubation with alt-2 DC in the presence of the anti–IL-12p70 Ab, NK and γδ T cells almost completely lost their ability to secrete IFN-γ (Fig. 7A), whereas no statistically significant alteration in their degranulation in response to Daudi cells was found (Fig. 7B). In addition to the loss of IFN-γ secretion, a slight and statistically not significant increase of IL-10 (p = 0.068) and TNF-α (p = 0.12) was found in the coculture supernatants, whereas IL-1β and IL-6 secretion remained unaffected (Fig. 7C) and cytokines corresponding to other immune polarizations were still not detected (data not shown). Blockade of IL-12p70 in the long-term coculture confirmed the independence from IL-12p70 for the degranulation in response to Daudi cells (Fig. 7D, left), but highlighted a partial requirement for the responsiveness of NK and even more γδ T cells to the melanoma Mel1379 (Fig. 7D, right).

Excessive innate activity is counterproductive for Ag-specific CTL expansion

To determine the consequences of the enhanced activation of innate effector cells on the adaptive immunity, DC were pulsed with synthetic peptides and used to stimulate CD8+ T cells either purified or within total PBL. To investigate a cancer-related setting, HLA-A2+ DC were pulsed with the MelanA/Mart-1 peptide and cocultured with effector cells for 10 d in the presence of IL-2 and IL-7. When the DC were used to stimulate purified CD8+ T cells, alt-1 and alt-2 DC were able to strongly expand MelanA/Mart-1–specific T cells, as evaluated by staining with specific tetramers (Fig. 8A). In contrast, in the presence of the other immune cell populations of total PBL, a reduced expansion of MelanA/Mart-1–specific CD8+ T cells by alternative DC was obtained, whereas there was an improvement in gold standard DC performance (Fig. 8A).

FIGURE 6. Alternative DC-promoting effects on innate cells are long lasting. Total PBL were stimulated with autologous mature DC (10:1 ratio) for 8 d in X-vivo 15 medium before functional evaluation against target cells. The stimulated PBL were incubated for 4 h with Daudi cells (A, B) or the melanoma cell line Mel1379 (C). Secretion of IFN-γ (A) and CD107a degranulation assays (B, C, top) were performed at a 1:1 ratio between PBL and target cells and effector populations identified upon counter staining, as described in the legend of Fig. 1. Represented are the mean ± SE among five different experiments. Daudi and Mel1379 were also labeled with [51Cr] and incubated for 4 h at different E:T ratios with PBL in triplicates for a standard chromie release assay (B, C; bottom). Represented are the mean ± SD of one experiment of four with similar results. *p < 0.05, **p < 0.01 in the Tukey HSD test.

FIGURE 7. IL-12p70 is required for induction of IFN-γ secretion. (A–C) PBL were incubated for 16–21 h with alt-2 DC in the presence of 10 μg/ml isotype control or anti–IL-12p70 mAb. Secretion of IFN-γ (A), degranulation to Daudi cells (B), and cytokine content in the supernatants (C) were evaluated, as described in legend to Figs. 3 and 5. Data represent mean ± SE of four different experiments. *p < 0.05, **p < 0.01 in paired t test. (D) PBL were incubated for 8 d with alt-1 or alt-2 DC in the presence of 10 μg/ml isotype control or anti–IL-12p70 mAb before the degranulation in response to Daudi (left) or Mel1379 cells (right) was evaluated. Data represent mean ± SE of four to six different donors. *p < 0.05, **p < 0.01 in the Tukey HSD test.
Alternative DC ability to expand naive CD8+ T cells is hindered by the presence of other immune populations. (A) Mature fast DC were pulsed with MelanA/Mart-1 peptide and incubated with purified CD8+ T cells or total PBL for 10 d in the presence of IL-2 and IL-7. Expansion of MelanA/Mart-1–specific CTL was evaluated by tetramer staining via flow cytometry. Represented are the mean ± SE of six different experiments. *p < 0.05 in the Tukey HSD test (within purified CD8+ T cells) or Student t test (for CD8+ T cells versus total PBL). (B) Stimulation of purified CD8+ T cells by MelanA/Mart-1 peptide-pulsed alt-1 and alt-2 DC was evaluated in the absence (CD8) or presence of other immune cell populations, namely the whole negative fraction of the PBL remaining after CD8+ T cell purification [CD8+ PBL(−)], or the fraction left after further depletion of CD56+ cells [CD8+ PBL(−)CD56−]. Reported are the mean ± SE of four different experiments. *p < 0.05 in the Tukey HSD test (for CD8+ T cells versus total PBL). (C) CD8+ T lymphocytes stimulated for 10 d with MelanA/Mart-1–pulsed DC were restimulated with irradiated MelanA/Mart-1–pulsed PBL before being evaluated in a CD107a assay against T2 cells pulsed with MelanA/Mart-1 or irrelevant peptide (left) as well as against the HLA-A2+ melanoma cell lines Mel1379 (top, right) and Na8 (bottom, right) that are MelanA/Mart-1–positive and –negative, respectively. Representative dot plots from alt-2 DC-expanded CD8+ T cells of one of three experiments are shown.

To dissect a possible negative role of NK cells on the expansion of Ag-specific T cells, which might be either due to the adaptation of CD56+ NK cell to lyse MHC class I–positive T cells and mature DC (19, 20) or due to a NKG2D ligand-dependent killing of recently activated CD8+ T (21, 22), the same coculture experiment was performed with PBL depleted of NK cells. Because in some donors the MelanA/Mart-1 tetramer-positive CD8+ T lymphocytes were CD56+ (data not shown), the following strategy was used: CD8α+ cells were first purified from the total PBL and the remaining negative fraction either was left untreated or underwent depletion of CD56+ cells before being added to the DC–CD8+ T cell coculture. The absence of CD56+ NK cells was not enough to restore the expansion of MelanA/Mart-1–specific T lymphocytes to the levels of the purified setting, but there was a trend, even if not reaching statistical significance toward an increased expansion (Fig. 8B).

The CD8+ T cell cultures containing expanded amounts of MelanA/Mart-1–specific T cells were restimulated with peptide-pulsed, irradiated HLA-A2+ PBL for 10 d and tested in a degranulation assay. Ag-specific degranulation was detected not only in response to peptide-pulsed T2 cells (Fig. 8C, top left), but also to endogenous Ag presentation from the Mel1379 cell line (Fig. 8C, top right). The low levels of degranulation in response to T2 cells pulsed with an unrelated peptide as well as to the MelanA/Mart-1–negative melanoma cell line Na8 (Fig. 8C, bottom) respectively confirm and support the Ag specificity of the induced degranulation.

In the next step, pools of viral peptides were used to pulse mature DC to stimulate memory CD8+ T cells within total PBL. [35Cr] release (data not shown) and CD107a degranulation assays revealed a high level of variability in the Ag-specific immune response among different donors, not only in its intensity, but particularly in the behavior of the different DC, as representatively shown for two donors with opposite situations (Fig. 9). Higher Ag-specific CD8+ T cell responses were obtained in donors with a low reactivity of innate effector cells, as determined by the killing of Mel1379 as well as of Daudi cells. In such donors, alternative DC induced the highest levels of Ag-specific CD8+ T cell–mediated responses (Fig. 9, top panels). In contrast, donors with a higher responsiveness of innate effectors to Daudi cells exerted lower CD8+ T cell–mediated responses with alt-2 DC inducing the lowest levels of Ag-specific responses, whereas the gold standard induced the highest (Fig. 9, bottom panels).

Discussion

The limited clinical objective responses obtained in tumor immunotherapy in general and also in the DC-based vaccination indicate the urgent need to optimize the currently used protocols and to increase the knowledge of the induced immune responses to select for each patient the most effective vaccine schedule.

In this study, different alternative maturation mixtures were compared with the clinical gold standard for their effect on the ability of monocytes differentiated with a fast protocol toward DC to interact with effector cells of the innate and adaptive immune system. The experiments employed fast DC because in preliminary experiments comparison with standard day 5 monocyte-derived DC highlighted not only an increased cell yield, but also a more potent interaction of effector cells with the innate effector cells, as determined by the killing of Mel1379 as well as of Daudi cells. In such donors, alternative DC induced the highest levels of Ag-specific CD8+ T cell–mediated responses (Fig. 9, top panels). In contrast, donors with a higher responsiveness of innate effectors to Daudi cells exerted lower CD8+ T cell–mediated responses with alt-2 DC inducing the lowest levels of Ag-specific responses, whereas the gold standard induced the highest (Fig. 9, bottom panels).

Concerning the phenotype of the fast DC, as determined by the expression of costimulatory molecules and by the IL-12p70/IL-10 ratio, and their ability to activate the immune system, the alt-3 DC (TNF-α + IL-1β + IFN-γ + CL097) were almost comparable to the gold standard DC (TNF-α + IL-1β + IL-6 + PGE2). In contrast, alt-2 DC (MPLA + IFN-γ) exerted a similar and, particularly...
The functional activity of effector cells within the PBL was evaluated by
with autologous PBL for 8–10 d in the absence of additional cytokines.
DC were pulsed with corresponding viral peptide pools and incubated
unpulsed (top) and Daudi cells (right). The results are represented as the
percentage of CD107a+ cells upon removal of background staining (i.e.,
unpulsed cells for CD8+ T cells and absence of target for innate effector
cells) from two donors representative of the extreme situations (top
versus bottom panels).

FIGURE 9. Different levels of innate activity affect the intensity of the
memory response of CD8+ T cells. HLA-A2* or HLA-A3+ mature fast DC
were pulsed with corresponding viral peptide pools and incubated with
autologous PBL for 8–10 d in the absence of additional cytokines.

The functional activity of effector cells within the PBL was evaluated by
means of a CD107a degranulation assay. Target cells were the HLA-A2+
and A3+ Mel1379 melanoma cells pulsed with the viral peptides of left
unpulsed (left) and Daudi cells (right). The results are represented as the
percentage of CD107a+ cells upon removal of background staining (i.e.,
unpulsed cells for CD8+ T cells and absence of target for innate effector
cells) from two donors representative of the extreme situations (top
versus bottom panels).

in respect to γδ T cells, even better activity than alt-1 DC, the
α-type 1–polarizing DC initially proposed by Mailliard et al. (10)
(TNF-α + IL-1β + IFN-α + IFN-γ + polyIC). A possible explanation is that, despite secreting similar levels of IL-12 with a
donor-dependent prevalence of one DC type over the other, alt-1 DC
secrete throughout all donors significantly higher amounts of IL-
10 that might dampen the type 1–oriented, IL-12–induced immune
response.

Regarding the functional activation of the immune system, two
different processes could be identified. The first one, induced to a
different extent by all alternative DC, consists in the acquisition of the
toxic activity and secretion of proinflammatory and type 1 cytokines via a mainly IL-12–dependent signaling. Indeed, only γδ T cell cytotoxicity revealed an IL-12 dependency and only in
response to difficult targets like melanoma, but not Daudi cells. In
the second process, IFN-γ secretion is induced by a highly IL-12–
dependent mechanism that is performed only by alt-1 and alt-2 DC.

Despite the highly variable frequencies in the different donors, it
is noteworthy that both NK and γδ T cells were activated by the
alternative DC, thus leading to the activation of multiple effector
mechanisms that can be useful in the coordinated action against
the tumor.

The important role of NK cells for the treatment of cancer has
been initially highlighted by their involvement in the graft-versus-
leukemia effects obtained in acute myeloid leukemia patients
undergoing haploididentical stem cell transplantation (23). More
recently, solid tumors like melanoma and renal cell carcinoma
also have been evaluated in vitro for their susceptibility to matched
and mismatched NK cells, confirming an increased cytotoxicity
upon usage of NK cells with at least one killer cell Ig-like receptor
(KIR) mismatch (24). Because in our setting a KIR typing of the
PBL was not performed, the intensity of the obtained tumor killing
might be biased by a mismatched situation that would not exist in
vaccinated patients, in which DC-activated autologous NK would
interact with the autologous tumor. In our study, alt-1 and alt-2 DC
induced a potent activation of CD56dim and CD56bright NK subsets
regarding their cytokine production and cytotoxic activity, a finding
that is in line with other reports describing IFN-γ secretion of the
CD56bright subset (25, 26), as well as cytotoxic activity of the
CD56bright population (20). In contrast to Jiang et al. (20) demon-
strating a granzyme K–mediated killing in the absence of gran-
zyme B in the CD56bright population, a significant induction of
granzyme B upon alt-1 and alt-2 DC stimulation was detected.
Due to the limited amount of CD56bright cells in the peripheral blood,
this subset could not be purified to validate target killing by standard [51Cr] release assay and to assess the involved effector
molecule(s). In line with the increasing reports of memory-like
NK cells (27), the enhanced activity of NK was preserved 5–
10 d after the stimulation with DC. Whether this correlates with
the induction of memory-like NK cells, or represents a result of an
expanded memory population in some of our healthy donors is
currently under investigation.

In contrast to the more intraepithelial V61+ cells, the V82+ cells are normally regarded as the blood-borne subset of γδ T cells. De-
spite that, they have been found within the infiltrating lymphocytes of different tumor histotype (28–30), and their ability to infiltrate
human skin and perform endothelial transmigration has been
documented (31, 32). For these reasons, their role in the antitumor
therapy is currently being investigated in phase I/II clinical trials of
adoptive transfer therapy (33, 34). In our setting, alt-2 DC were
more efficiently activating the V82+ γδ T cells, particularly re-
garding proliferation and degranulation to the melanoma cell line.
Although the presentation of the lipid A moiety of LPS in associ-
ation with CD1b or CD1c to γδ T cells has been proposed (35), a
direct stimulation of γδ T cells by MPLA could be excluded be-
cause classical monocyte-derived DC or Langerhans-like DC
stimulated with the alt-2 mixture was not able to induce IFN-γ
secretion in γδ T cells despite their higher expression of CD1b and/or
CD1c in comparison with fast DC (data not shown). The results
from the long-term blockade of IL-12 highlighted a statistically
significant inhibition of the degranulation in response to Mel1379,
but only upon stimulation with alt-2 DC. Whereas the response to
alt-1 DC by γδ T cells was in general low, and thus the effect of
inhibition less evident, the reduction of alt-2 DC-induced killing to
levels similar to the basal alt-1 DC suggested a synergy between IL-
12 and another to date undefined factor/molecule for the enhanced
stimulation of γδ T cells by alt-2 DC. Among the molecules already
involved in the interaction of γδ T cells with phasootiigen-pulsed
tumor cells (36–38), CD86 is the only marker significantly stronger
expressed in alt-2 DC compared with the other DC populations
investigated, and might thus be a possible candidate for the synergic
partner. Whereas the constitutive and short-time incubation with
DC-induced CD56 expression was highly donor variable and not
 correlating with function, a consistent CD56 upregulation was
found in all donors upon long-term incubation with alternative DC
and correlated with enhanced proliferation/degranulation, as previ-
ously reported for in vitro expanded γδ T cells from neoplastic and
infectious setting (39, 40). Thus, CD56 might represent a marker for
immunomonitoring γδ T cell vaccine-induced immune responses.

Evaluation of the consequences of the potent innate activation on
the induction of Ag-specific immune response highlights a complex
immune context and underscores the importance of evaluating the
potency of new vaccine formulations within total blood lymphocytes and not with purified cell populations. Whereas alt-2 DC significantly improved the expansion of MelanA/Mart-1–specific T cells within purified CD8+ lymphocytes over gold standard DC, this advantage was almost completely lost within total PBL. Usage of viral Ags to evaluate the functionality of the expanded CD8+ T cells within total PBL highlighted a broad variability among different donors regarding the potency of the memory responses, ranging from <1% to >30% of responding cells, and the relative efficiency of the various DC formulations. Donors with a reduced activation of innate cells exerted higher CD8+ T cell–mediated immune responses, which were stronger upon alternative DC stimulation. In contrast, in donors with higher innate activity, a much lower frequency of degranulating CD8+ T cells existed, with the best Ag-specific responses detected with gold standard or alt-3 DC. However, the killing induced by alt-2 DC either via CD8+ T cells or innate cells was consistently higher when compared with the other DC preparations, as highlighted by the [51Cr] release assay.

The mechanisms responsible for this dichotomy between an Ag-specific and an innate mediated tumor killing might be due to the following: 1) Competition of effector cells for target during the assay. Despite the 1:1 ratio used in the degranulation assay, it is much lower frequency of degranulating CD8+ T cells existed, with the best Ag-specific responses detected with gold standard or alt-3 DC. However, the killing induced by alt-2 DC either via CD8+ T cells or innate cells was consistently higher when compared with the other DC preparations, as highlighted by the [51Cr] release assay.

Disclosures

The authors have no financial conflicts of interest.

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Online Material

Figure S1. Alternative DC induce activation markers on innate lymphocytes

PBL were incubated for 16 h with mature DC in X-vivo15 and thoroughly washed before the expression levels of the activation markers CD25 and CD69 were evaluated by flow cytometry. The different effector populations were identified upon counter staining with mAb directed against CD3, CD56, CD16 and Vδ2 as described in legend to Figure 1. The mean ± SE of four different donors are shown for CD25 (A) and CD69 (B). C) Representative dot plots of “gold-standard” and “alt-2” DC stimulated CD56\textsuperscript{br} (top) and CD16\textsuperscript{+}56\textsuperscript{dim} NK (center) as well as γδ T cell (bottom) from one representative donor are shown.
Figure S1