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Leukotriene B$_4$ Enhances Innate Immune Defense against the Puerperal Sepsis Agent *Streptococcus pyogenes*

Elyara M. Soares,* Katie L. Mason,* Lisa M. Rogers,* Carlos H. Serezani,§ Lucia H. Faccioli,† and David M. Aronoff*†,∥

Puerperal sepsis is a leading cause of maternal mortality worldwide. *Streptococcus pyogenes* (group A *Streptococcus; GAS) is a major etiologic agent of severe postpartum sepsis, yet little is known regarding the pathogenesis of these infections. Tissue macrophages provide innate defense against GAS, and their actions are highly regulated. The intracellular second messenger cAMP can negatively regulate macrophage actions against GAS. Because leukotriene (LT) B$_4$ has been shown to suppress intracellular CAMP in macrophages, we hypothesized that it could enhance innate defenses against GAS. We assessed the capacity of LTB$_4$ to modulate antirstreptococcal actions of human macrophages, including placental and decidua macrophages and used a novel intrauterine infection model of GAS in mice lacking the 5-lipoxygenase enzyme to determine the role of endogenous LTs in host defense against this pathogen. Animals lacking 5-lipoxygenase were significantly more vulnerable to intrauterine GAS infection than were wild-type mice and showed enhanced dissemination of bacteria out of the uterus and a more robust inflammatory response than did wild-type mice. In addition, LTB$_4$ reduced intracellular cAMP levels via the BLT1 receptor and was a phosphatase, NADPH oxidase-dependent intracellular killing of GAS. Importantly, interference was observed between the macrophage immunomodulatory actions of LTB$_4$ and the CAMP-inducing lipid PGE$_2$, suggesting that interplay between pro- and anti-inflammatory compounds may be important in vivo. This work underscores the potential for pharmacological targeting of lipid mediator signaling cascades in the treatment of invasive GAS infections. *The Journal of Immunology*, 2013, 190: 000–000.

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Abbreviations used in this article: DM, decidual macrophage; 8E, 10E-trienoic acid; GAS, group A *Streptococcus*; 12-HHT, 12(S)-hydroxyheptadeca-5Z; HK, heat killed; SLO, 5-lipoxygenase; LT, leukotriene; NADPHox, NADPH oxidase; PM, placental macrophage; PTX, pertussis toxin; RFU, relative fluorescence unit; ROI, reactive oxygen intermediate; RPMI$^{10^5}$, RPMI 1640 supplemented with 1% antibiotic–anti-mycotic and 10% charcoal/dextran-treated FBS; RPMI$^{10^6}$, RPMI 1640 supplemented with 1% antibiotic-anti- mycotic in the absence of serum; THY, Todd Hewitt Broth plus 5% of yeast; TSA–blood, tryptic soy agar plus 5% sheep blood.

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capacity of macrophages to respond to GAS have not been well characterized. Defining the mechanisms governing antimicrobial functions of tissue macrophages provides an opportunity to develop innovative measures for preventing or treating invasive bacterial infections in susceptible hosts. Recently, host-derived lipid mediators have been found to regulate the capacity for macrophages to respond to GAS (12), but this has not been explored in the context of reproductive immunology.

Eicosanoids are immunoregulatory lipids that are oxygenated metabolites of the cell membrane constituent molecule arachidonic acid. They are potent modulators of innate immunity, including macrophage behaviors (13–17). For example, recent studies suggest that the cyclooxygenase-derived PG compound, PGE$_2$, suppresses macrophage phagocytosis and intracellular killing of GAS through acutely activating the synthesis of the intracellular second messenger molecule cAMP (18). In general, receptor-dependent increases in cAMP suppress macrophage immune responses, possibly an evolved mechanism for limiting the amplitude and duration of inflammatory responses (19). In contrast, the 5-lipoxygenase (5LO)–derived eicosanoid leukotriene (LT) B$_4$ has been reported to stimulate macrophage phagocytosis and intracellular killing of several pathogens in association with suppression of intracellular cAMP production (17, 20). The ability of LTB$_4$ to enhance host elimination of streptococci has been most clearly defined in a model of lung infection caused by S. pneumoniae (21–24). In addition, the administration of aerosolized LTB$_4$ to mice with preexisting pneumococcal pneumonia increased mononuclear phagocyte/macrophage accumulation in the lungs, NADPH oxidase (NADPHox) subunit expression in pulmonary macrophages, and pulmonary bacterial clearance (25). Leukotriene B$_4$ is synthesized from arachidonic acid when an acute Ca$^{2+}$ flux is induced in cells following activation by inflammatory mediators or pathogens (26, 27). Two G protein–coupled receptors for LTB$_4$ inhibit adenylyl cyclase. The BLT1 receptor (high affinity) is expressed primarily in leukocytes, including granulocytes, eosinophils, macrophages, and differentiat$^2$ed T cells, whereas the BLT2 receptor (low affinity) is expressed in many cell types (28–32).

It is notable that both PGE$_2$ and LTB$_4$ are found in the gravid uterus (33–35), and both have immunoregulatory actions that might play a role in defining a postpartum woman’s vulnerability to invasive bacterial infection. Because PGE$_2$ stimulates intracellular cAMP to suppress macrophage immune defenses against GAS (18), we hypothesized that LTB$_4$ would stimulate macrophage actions against this organism because its actions on cAMP are opposite to those of PGE$_2$. We further speculated that mice unable to generate LTB$_4$ because they lack the gene encoding 5LO could be susceptible to intrauterine GAS infection. Both in vivo and in vitro models of infection were used to assess the role of LTs as immunoregulators of innate defense against GAS.

Materials and Methods

Reagents

RPMI 1640 culture medium and antibiotic solution (penicillin and streptomycin) including an antmycotic (aminophoterin) were from Invitrogen (Carlsbad, CA). Charcoal-stripped and dextran-treated PBS was from HyClone Laboratories (Waltham, MA). Pertussis toxin (PTX), cytochalasin D from Zygosporium mansonii, and apocynin were obtained from EMD Chemicals–Calbiochem (Gibbstown, NJ). FITC, trypsin blue, DNase, hyaluronidase, collagenase, Percoll, citrate buffer, ficucin, saponin, PMA, nonenzymatic cell dissociation solution, NaHCO$_3$ buffer, and sodium bicarbonate were from Sigma–Alrich (St. Louis, MO). LTB$_4$, PGE$_2$, 12(S)–hydroxyeicosatetrae–5Z, 8E, 10E–trienoic acid (12–HHT), U75302, and hydroxyheptadeca–5Z, 8E, 10E–trienoic acid (12–HHT) were from Cayman Chemicals (Ann Arbor, MI). Todd Hewitt broth, trypsin soy agar, and yeast extract were from BD Biosciences (San Jose, CA). Sheep blood was from Remel (Lenexa, CA). Ficoll–Hypaque was obtained from GE Healthcare (Piscataway, NJ). Lysis buffer for erythrocytes was purchased from eBioscience (San Diego, CA). FLUO–4 NW used for intracellular calcium assays and 6-carboxy–2‘,7‘–dichloro- dihydrofluorescine diacetate (carboxy H2DFDA) for reactive oxygen species measure were provided from Molecular Probes (Grand Island, NY). ELISA kits to measure intracellular cAMP were from Enzo Life Sciences (Farmingdale, NY). Magnetic MACS CD14 microbeads (human) were from Miltenyi Biotec (Auburn, CA). Required dilutions of all compounds were prepared immediately before use, and equivalent quantities of vehicle were added to the appropriate controls. Compounds requiring reconstitution were dissolved in DMSO (Sigma–Aldrich).

Animals

Age-matched, 6– to 8-wk–old, female, nonpregnant, C57BL/6 wild-type or 5LO-deficient mice (5LO$^{−/−}$) mice on a C57BL/6 background were purchased from The Jackson Laboratory (Bar Harbor, ME). Animals were treated according to National Institutes of Health guidelines for the use of experimental animals, with approval of the University of Michigan Committee for the Use and Care of Animals.

Bacteria

The GAS M1T1 strain 5448 was generously provided by Dr. Malak Kotb of the University of Cincinnati (Cincinnati, OH), and aliquots were grown in Todd Hewitt Broth plus 5% of yeast (THY). Estimates of bacterial concentrations in inocula were made by comparing the absorbance of light at 600 nm (OD$_{600}$) and using a standard curve of CFU per milliliter versus OD$_{600}$. The actual CFU per milliliter concentrations were confirmed for all bacterial suspensions used in these studies by culturing serial 10-fold dilutions on tryptic soy agar plus 5% sheep blood (TSA–blood) plates. For phagocytosis experiments, heat-killed (HK) FITC–labeled GAS were used. These were generated as previously described for S. pneumoniae (36).

Intrauterine infection

GAS cultures were grown 18 h, shaking at 200 rpm, at 37°C in 10 ml THY; cultures were then centrifuged 3000 rpm for 10 min, and the pellet was washed twice in sterile 1× PBS and resuspended in 1 ml total volume. Dilutions were prepared in 1× PBS to obtain the correct approximate inoculum of CFUs per mouse. The actual inoculum was determined by plating serial 10-fold dilutions of the PBS suspension onto TSA–blood plates. The infection was performed in anesthetized animals. A low 2.0-cm midline abdominal incision exposed the right uterine horn, the uterine horn was ligated at the cervical junction to prevent inoculum loss, and 20 μl of prepared culture was directly injected into the horn (37, 38). For survival experiments, mice were monitored every 2 h on weekdays and 2× per day on weekends for 1 wk after inoculation.

Quantitation of bacterial loads

After 24 h of infection, mice were euthanized, and bacterial load was determined in the blood, spleen, and uterus. The blood was serially diluted in sterile 1× PBS and cultured on TSA–blood at 37°C for 18 h. The spleen and uterus were homogenized with a handheld tissue teaser in 1 ml sterile 1× PBS, serially diluted in PBS, and then cultured on TSA–blood at 37°C for 18 h. Colonies were counted as previously described (37).

Human subjects

Following appropriate informed consent, human decidual tissue was obtained from healthy adult women aged 18–44 y who were undergoing elective surgical termination of pregnancy during the first trimester. Human placental tissue was obtained from healthy women undergoing cesarean section at the term of pregnancy. Human peripheral blood was obtained from healthy adults who were not smokers, not taking nonsteroidal anti-inflammatory drugs, not taking oral or inhaled steroids, and not pregnant. Blood was collected by venous puncture into heparinized collection tubes. These studies were reviewed and approved by the University of Michigan Institutional Review Board.

Cell culture

THP–1 monocytes were obtained from the American Type Culture Collection (ATCC; TIB–202; Manassas, VA) and cultured in RPMI 1640 supplemented with 1% antibiotic–antimycotic and 10% charcoal/dextran–treated fetal bovine serum (RPMI$^{+/+}$). These were prepared by passage only through the 10th passage, at which time a new culture was started. THP–1 cells were differentiated into macrophage–like cells by culturing with 100 nM of PMA in RPMI$^{+/+}$ for 18 h at 37°C with 5% CO$_2$. Cells
were detached from the flask with nonenzymatic cell dissociation solution, scrapped after 5 min of incubation at 37 °C with 5% CO2. The protocol was adapted as described before (39, 40). PBMCs, placental macrophages (PMs), decidual macrophages (DMs), and peritoneal macrophages were cultured in RPMI 1640 for 18 h at 37 °C with 5% CO2, at a density of 2 × 10⁵ cells per well in 6-, 96-, or 384-well plates (Costar, Corning, NY), depending on the experiment.

**Isolation and culture of PMs and DMs**

The procedure for DM and PM culture was similar to that used for THP-1 cells above. Isolation was adapted from a previously described protocol (38, 41). Briefly, the tissue was collected in 50-ml conical tubes containing 15 ml sterile PBS and washed three times with 50 ml PBS at 1500 rpm for 10 min. The tissue was weighed and minced into small pieces with autoclaved scissors and of the experimental well to determine well final grams collected. After this, the fragments were placed into 50-ml conical tubes with digestion solution containing DNase, collagenase, and hyaluronidase at 10 ml per gram of tissue. Cells were filtered through a 280-μm autoclaved metal sievel, followed by 180- and 80-μm autoclaved nylon screens. Cells were centrifuged again and re-suspended in 25% Percoll diluted in cold RPMI+/- and overlaid onto 50% Percoll plus 2 ml PBS on top of the density gradient. CD14⁺ cells were isolated using magnetic MACS small-cell separation column system (Miltenyi Biotec) according to the manufacturer’s instructions and adapted as previously reported (38).

**Human PBMC isolation**

Human PBMCs were isolated from heparinized blood by Ficoll-Hypaque gradient density centrifugation by 20 min at 1700 rpm, 22°C, without active breaking. The PBMCs were collected, transferred to new tubes, and washed with PBS at 1400 rpm × 10 min at room temperature. The pellets were broken and pooled together in one tube and washed again. The erythrocytes were lysed after the addition of 10 ml of lysis buffer at 22°C × 10 min. The cells were washed again; the final solution was resuspended in 50 ml PBS, and total cells were counted. CD14⁺ cells were negatively selected using magnetic MACS small-cell separation column system (Miltenyi Biotec) according to the manufacturer’s instructions.

**Isolation and culture of peritoneal macrophages**

 Resident PMs were collected from uninfected mice. Mice were euthanized by CO₂ inhalation, and peritoneal cavities were washed twice with 5 ml cold RPMI 1640 supplemented with 1% antibiotic-antimycotic in the absence of serum (RPMI+/-). Collected fluid was placed in conical tubes on ice and centrifuged at 1500 rpm for 10 min. Cell counts were performed immediately, and 2 × 10⁶ cells per well were added to culture in 384-well plate (Costar), followed by incubation at 37°C, 5% CO₂ for 2 h. After this period, the media were changed to RPMI 1640, and adherent cells were cultured by 18 h incubation at 37°C, 5% CO₂, as published before (42, 43).

**Fluorometric phagocytosis assay with FITC-labeled GAS**

The phagocytosis of unopsonized FITC-labeled HK GAS was assessed using PMA-treated THP-1 cells, PBMCs, DMs, PMs, and resident PMs. All cell types were plated onto 384-well tissue culture–treated plates with 2 × 10⁵ cells per well, and the phagocytosis was performed as previously reported (20, 38, 44). After an overnight rest, media were removed, and RPMI 1640 containing vehicle or different stimuli were added as indicated in the figure legends. Specific treatments included LTB₄ (15 min pre-treatment, 1–1000 nM), PGE₂ (15 min pre-treatment, 1 μM), 12-HHT (15 min pre-treatment, 0.001–1 μM), U75302 (15 min pre-treatment, 10 μM), and PTX (18-h pre-treatment, 3 μg/ml). After appropriate incubations, GAS were added at multiplicity of infection of 150:1 and incubated at 37°C with 5% CO₂ for 3 h. Extracellular bacterial fluorescence was quenched with trypan blue (500 μg/ml in 0.09 M citrate buffer) for 15 min in the dark before the fluorescence was determined on a microplate fluorometer (485₅/535₅). SPECTRMax GEMINI EM (Molecular Devices, Sunnyvale, CA). A minimum of eight replicates were done for each condition in each experiment. Fluorometric data were expressed in arbitrary relative fluorescence emitted from intracellular phagocytosed bacteria (relative fluorescence unit [RFUₜₜ] and was determined by subtracting the fluorescence from unquenched, extracellular bacteria (RFUₜₜ) from the total fluorescence of the experimental well (RFUₜₜ). The RFUₜₜ was determined using the fluorescence of cells exposed for 30 min to the phagocytosis inhibitor cytochalasin D (20 μg/ml). Thus, the PI = RFUₜₜ = RFUₜₜ - RFUₜₜ (44, 45).

**Bacterial killing assay**

PMA-treated THP-1 cells were adjusted to a suspension of 2 × 10⁵ cells per well in 100 μl RPMI 1640 in replicates of 8 per condition in a 96-well plate and was centrifuged at 500 rpm for 30 s to settle all cells into a monolayer. Cells adhered for 1 h before starting with the treatments. One set of replicates conditions within each plate was treated with cytochalasin D for 30 min. Cells were pretreated with the NADPHox inhibitor apocynin 100 μM and/or LTB₄ 10 nM for 30 and 15 min respectively before infecting cells with a 1:10 multiplicity of infection of live GAS. The plates were centrifuged again to synchronize bacterial contact with the monolayer and incubated for 30 min. After this time, all media were removed and plates washed gently 3 times with RPMI 1640 to remove nonphagocytosed bacteria. Then 100 μl of 0.5% saponin in PBS was added and incubated for 10 min. Serial dilutions were made and the lysates were plated onto TSA–blood plates; CFUs were counted 24 h later (adapted from Refs. 14 and 22).

**Measurement of intracellular reactive oxygen intermediates**

Reactive oxygen intermediate (ROI) generation was performed with 1.2 × 10⁶ PMA-treated THP-1 cells per well, using 384-well plates, according to our previously reported protocol (14, 22, 46). Medium was replaced the following morning, and cells were rested for 2 d. The cells were incubated with the ROI-sensitive fluorochrome carboxy-H2DCFDA at 10 μM for 30 min, followed by two washes with HBSS. Then LTB₄ (10 nM), LTB₄-unopsonized GAS 10:1, or both were added and incubated for 30 min. The fluorescence intensity was measured at an excitation wavelength of 493 nm and emission wavelength of 522 nm, using the fluorescent plate reader (above).

**Measurement of intracellular cAMP**

For intracellular cAMP measurement, 3 × 10⁶ PMA-treated THP-1 per well were used. The medium was changed to RPMI 1640, and cells were incubated for 0, 15, 60, 300, 600, and 900 s with 10 nM LTB₄ for some experiments or 60 s with 10 nM LTB₄ plus 900 s followed by 1 μM of PGE₂, for others. Culture supernatants were aspirated, and the cells were lysed by incubation for 10 min with 0.1 M HCl at room temperature, followed by disruption using a cell scraper (20). Intracellular cAMP levels were determined by ELISA according to the manufacturer (Enzo Life Sciences).

**Measurement of LTB₄ and cytokines**

Uteri were removed 24 h postinfection to measure LTB₄ and cytokines. Briefly, tissue was homogenized (Tissuemiser; Fisher Scientific, Suwanee, GA) in 1 ml PBS, centrifuged, and stored at −80°C until assayed. The OD of samples was determined at 405 nm on a microplate reader (VERSA-MAX; Molecular Devices, Sunnyvale, CA), and concentrations of eicosanoids were calculated based on a standard curve. The sensitivity for LTB₄ was 5.63 pg/ml. A specific enzyme immunoassay was used to quantify LTB₄ (Enzo Life Sciences) according to the manufacturer’s instructions. The tissue was recovered and stored at −80°C until assays were performed to determine the levels of IL-17A, IL-6, IL-1β, TNF-α, IL-10, IFN-γ, and MCP-1 by ELISA (R&D Duoset; R&D Systems). ELISAs were performed by the University of Michigan Cancer Center Cellular Immunology Core (14).

**Analysis of calcium mobilization**

PMA-treated THP-1 cells were used at a density of 4 × 10⁵ cells per well, using 96-well, white-bottom plates (BD Biosciences). The cells were incubated at 37°C plus 5% CO2 for 1 h to rest, and then the media were removed from the adherent cell culture, to eliminate sources of baseline fluorescence, particularly esterase activity. A volume of 100 μl of the dye-loading solution was added to each well immediately. The plate was incubated for 30–45 min at 37°C, and after this, LTB₄ (10 nM) and our positive control A23187 (1 μM) were added in the specific wells at the moment of the reading. Fluorescence was measured with excitation at 494 nm and emission at 516 nm. The assay was performed using a Fluor-4-NM Calcium Kit (Invitrogen) according to the manufacturer’s instructions and adapted according to previous works (47, 48).

**Statistical analysis**

Data are presented as the mean ± SEM and were analyzed with the Prism 5.0 statistical program (GraphPad Software). Comparisons among three or more experimental groups were performed with one-way ANOVA, followed by a Bonferroni correction, and between two experimental groups a Student t test was used, unless otherwise stated. Differences were
Results

5LO<sup>−/−</sup> mice have enhanced susceptibility to intrauterine GAS infection

Leukotrienes, including LTB<sub>4</sub>, are important endogenous regulators of immune defense against bacterial pathogens (21, 25, 49, 50), suggesting that they might be involved in supporting host defense against GAS. To test this, wild-type C57BL/6 mice or 5LO<sup>−/−</sup> mice were subjected to intrauterine GAS infections and monitored for survival, uterine bacterial clearance, and bacterial dissemination. As illustrated in Fig. 1A, 5LO<sup>−/−</sup> mice were significantly more susceptible to overwhelming infection caused by GAS than were wild-type mice. It was found that 80% of the wild-type mice survived after 7 d, whereas only 30% of 5LO<sup>−/−</sup> mice survived ($p < 0.0001$). The enhanced susceptibility of 5LO<sup>−/−</sup> mice to intrauterine GAS infection was correlated with an impaired ability of these genetically deficient mice to contain the infection within the uterus (Fig. 1B). At 24 h postinfection, the number of bacteria that disseminated to the spleen in 5LO<sup>−/−</sup> mice was 1.48 times greater than that observed for wild-type mice. Levels of LTB<sub>4</sub> were measured in uterine homogenates from wild-type and 5LO<sup>−/−</sup> mice following GAS infection. As shown in Fig. 1C, intrauterine infection with GAS significantly increased LTB<sub>4</sub> production in the wild-type uterus. However, uterine tissues obtained from infected 5LO<sup>−/−</sup> mice did not reflect an increase in LTB<sub>4</sub> generation, compared with uninfected tissues. Interestingly, as evidenced in Fig. 1D, inflammatory mediator generation, including IL-1β, IL-6, IL-17, and MCP-1, was markedly increased in the uterine tissues of infected 5LO<sup>−/−</sup> mice compared with wild-type animals.

**LTB<sub>4</sub> enhances the phagocytosis of unopsonized GAS**

The importance of LTs in supporting clearance of GAS in vivo and the role of macrophages in host defense against GAS (10, 11) suggested that LTB<sub>4</sub> might regulate macrophage interactions with GAS in vitro. Thus, primary human macrophages, mouse macrophages, or THP-1 cells were pretreated with various concentrations of LTB<sub>4</sub> (1–100 nM) that were found to enhance macrophage phagocytosis in other model systems (20, 50, 51). As outlined in Fig. 2A–E, LTB<sub>4</sub> significantly increased the phagocytosis of unopsonized GAS by THP-1 cells, human PBMCs, human DMs, human PMs, and mouse PMs. Because the actions of LTB<sub>4</sub> on THP-1 cells were similar to primary cells, we used this human macrophage–like cell line for mechanistic studies to build a model of how LTB<sub>4</sub> regulates phagocyte function during GAS infection.

The high-affinity, Ga<sub>q</sub>-coupled BLT1 receptor for LTB<sub>4</sub> modulates phagocytosis in THP-1 cells

LTB<sub>4</sub> has two major G protein–coupled receptors: the high-affinity BLT1 receptor and the low-affinity BLT2 receptor (28). The BLT1 receptor has been implicated in macrophage immunoregulation by LTB<sub>4</sub> (52) through a PTX-sensitive suppression of intracellular cAMP (20). We confirmed that THP-1 cells express the BLT1 receptor (39) by Western immunoblot analysis (not shown) and demonstrated that the LTB<sub>4</sub>-mediated enhancement of GAS phagocytosis by THP-1 cells was completely blocked by the BLT1 receptor antagonist U75302 (Fig. 3A). Further, 12-HHT, an agonist for the BLT2 receptor that is more potent than LTB<sub>4</sub> (28), had no significant effect on THP-1 internalization of unopsonized bacteria (Fig. 3B). The capacity of LTB<sub>4</sub>–BLT1 signaling to enhance GAS phagocytosis suggested that LTB<sub>4</sub> reduced cAMP in these cells. We measured intracellular cAMP in macrophages after exposure to

**FIGURE 1.** Leukotrienes are important determinants of survival and containment of intrauterine GAS infection in mice. (A) Wild-type C57BL/6 or 5LO<sup>−/−</sup> mice ($n = 10$ per group) were inoculated with $10^4$ CFU of GAS in the right uterine horn on day 0, and survival was recorded over time. Vehicle (PBS)–treated mice did not die ($n = 10$; data not shown). Data are expressed as mean ± SEM followed by log-rank (Mantel–Cox) test. **p < 0.001. (B) Nonpregnant mice C57BL/6 or 5LO<sup>−/−</sup> ($n = 10$ per group) were inoculated with $10^7$ CFU of GAS in the right uterine horn and then euthanized 24 h postinfection. Bacterial loads from blood, spleen, and uterus were determined as described in Materials and Methods. Data shown are from one representative experiment from two independent experiments with similar results. Data are expressed as mean ± SEM followed by Student t test. *p < 0.05. (C) After 24 h of infection, enzyme immunoassay quantification of LTB<sub>4</sub> concentrations was performed on uterine homogenates obtained from mice that had received either an intrauterine PBS injection (uninfected) or GAS. Data are presented as the mean ± SEM followed by one-way ANOVA and are representative of two independent experiments with similar results ($n = 10$ mice per group). **p < 0.001 compared with wild-type uninfected control. (D) Inflammatory mediators in uterine homogenates of C57BL/6 or 5LO<sup>−/−</sup> mice 24 h after inoculation with $10^7$ CFU of GAS. At 24 h after intrauterine inoculation with GAS, cytokines and chemokines were measured by ELISA. **p < 0.01, ***p < 0.001 by Student t test ($n = 10$ mice per group from two combined experiments with similar results).
Notably, cAMP levels fell acutely. Within 15 s of exposure, the intracellular levels were undetectable (Fig. 4A). Phagocytosis experiments with PTX (Fig. 4B) confirmed that the capacity for LTB4 to stimulate THP-1 cell ingestion of GAS was Gαi-dependent. Because the BLT1 receptor can signal through a Gαq-dependent release of intracellular calcium (53), we tested whether LTB4 treatment could increase Ca2+ in THP-1 cells. As opposed to the positive control compound, the calcium ionophore A23187, LTB4 failed to significantly alter basal calcium levels (Fig. 4C), consistent with results in lung macrophages (20). This finding was verified over a range of LTB4 concentrations and incubation times (data not shown).

**LTB4 enhances bacterial killing of GAS**

Along with phagocytosis, intracellular microbicidal activity is an important step in the control of infection. Previous work in other model systems has revealed that PGE2 can suppress the capacity of macrophages to kill bacteria (13, 18, 54), whereas endogenous and exogenous LTs enhance this action (20, 55). We determined the importance of LTB4 to macrophage killing of GAS using THP-1 cells. As noted in Fig. 5A, the number of intracellular bacteria present in THP-1 cells following 30 min of incubation was significantly reduced by treating cells with LTB4 (10 nM). ROIs generated by NADPHox are important for macrophage elimination of GAS (56), and it is known that LTB4 enhances NADPHox activation and ROI generation in macrophages (25, 55, 57–59). The extent to which the LTB4 stimulation of ROI production enhanced macrophage GAS killing was evaluated using the selective NADPHox inhibitor apocynin (60) Treatment with apocynin prevented the acute killing of intracellular bacteria during the 30 min of incubation of GAS with THP-1 cells (Fig. 5A). In addition, LTB4 failed to reduce the number of intracellular bacteria in macrophages pretreated with apocynin, suggesting that the actions of LTB4 depend upon NAPDHox activation. We then tested whether LTB4 enhanced intracellular ROI generation in THP-1 cells infected with GAS (Fig. 5B). Notably, live GAS suppressed cellular ROI concentrations, an effect not associated with cell death (data not shown). However, LTB4 alone significantly increased ROI generation, and this overcame the suppressive action of GAS (Fig. 5B).

If the actions of LTB4 are mediated through suppression of intracellular cAMP, then it can be hypothesized that this lipid would antagonize the capacity for PGE2 (which stimulates cAMP) to suppress macrophage host defense functions. Indeed, PGE2 raised intracellular cAMP concentrations and inhibited the phagocytosis of GAS by THP-1 cells (Fig. 6A, Fig. 6B). As noted in Fig. 6A, PGE2 reduced cAMP, LTB4 increased it, and the effect of a combination of both lipids was intermediate (Fig. 6A). In addition (Fig. 6B), 10 μM of PGE2 significantly inhibited GAS phagocytosis by 56.5 ± 3.0%, whereas 100 nM of LTB4 enhanced it by 44.4 ± 7.3%. However, when cells were exposed to both com-

**FIGURE 2.** LTB4 increases leukocyte phagocytosis of GAS. Macrophages were exposed for 15 min to LTB4 before challenge with FITC-labeled HK GAS, and phagocytosis was determined as described in Materials and Methods. Data shown are from (A) PMA-treated human THP-1 cells, (B) human PBMCs, (C) human DMs, (D) human PMs, and (E) C57BL/6 mouse peritoneal macrophages. All data are expressed as a percentage of the phagocytic activity of untreated cells. *p < 0.05, **p < 0.01 versus untreated, as determined by one-way ANOVA for (A)–(D) and t test for (E).

**FIGURE 3.** LTB4 enhances GAS phagocytosis in THP-1 cells via the BLT-1 receptor. (A) PMA-treated THP-1 cells were incubated with the BLT1 receptor antagonist U75302 (10 μM) or LTB4 (10 nM) for 15 min before challenge with FITC-labeled HK GAS, as described in Materials and Methods. Data shown are mean ± SEM of four independent experiments. (B) PMA-treated THP-1 cells were incubated with 0.001–1 μM of BLT2 agonist (12-HHT) for 15 min before challenge with HK FITC-GAS at 150:1. Phagocytosis was quantified after 3 h by fluorometry, as detailed in Materials and Methods. *p < 0.05 versus untreated, #p < 0.05 versus LTB4, as determined by one-way ANOVA.
pounds, the net effect was a lack of significant change in phagocytic activity relative to untreated cells (Fig. 6B).

Discussion

*S. pyogenes* is a leading cause of severe postpartum sepsis worldwide (1, 61). A better understanding of why pregnancy and the postpartum state increase the risk for GAS is needed to develop better preventive or therapeutic strategies. In this article, we report the novel finding that LTB4 is an endogenous lipid mediator that augments innate immune defenses against invasive GAS infection. Macrophages are important sentinels of host defense against GAS (10, 11). These studies suggest that LTB4, a major product of the 5LO metabolic pathway, enhances the phagocytosis and bacterial killing of unopsonized GAS by macrophages, including primary human macrophages from the female reproductive tract. The mechanism for these actions involves signaling via the Gαi-coupled BLT1 receptor with an acute suppression of cAMP.

Earlier studies established the importance of LTs as immunostimulatory mediators during infection with non-GAS pathogens (21, 25, 42, 49, 62, 63). In fact, the capacity of LTB4 to regulate host–streptococcal interactions has been shown in a model of lung infection caused by *S. pneumoniae* (21–24). In addition, the administration of aerosolized LTB4 to mice with pre-existing pneumococcal pneumonia increased mononuclear phagocyte/macrophage accumulation in the lungs, NADPHox subunit expression in pulmonary macrophages, and pulmonary bacterial clearance (25). Although these data provided an important rationale for our hypothesis, little was known about the role of LTB4 in GAS infections or reproductive tract infections.

A significant finding includes the critical importance of the enzyme 5LO in supporting immune containment of intrauterine *S. pyogenes* infection. Unlike wild-type mice, mice lacking the 5LO enzyme did not produce meaningful quantities of LTB4 following infection, they were significantly more susceptible to death from GAS endometritis, they were less capable of preventing dissemination of bacteria from the uterus to distant sites, and they displayed a significantly enhanced inflammatory response to infection. These findings suggest that an overwhelming inflammatory response, due to poor innate immune containment of a local GAS infection, played a role in the mortality of this infection in 5LO−/−.
mice. Establishing a causal relationship between the exaggerated inflammatory response to GAS in SLO−/− mice and their mortality will require further study.

It is important that LTB4 has been shown to augment macrophage immune defenses against several pathogens in vivo (49, 51, 55, 62, 63) and to amplify the ability of leukocytes to phagocytose and kill microbes (43, 49, 51, 62–65). Our results support the generalizability of the previous reports and add evidence that one causal mechanism by which LTB4–BLT1 coupling regulates macrophage behaviors is by reducing cAMP production through a PTX-sensitive signaling cascade. How the acute lowering of basal cAMP levels impairs host defense against GAS requires more exploration, but both phagocytosis and ROI-dependent bacterial killing are certainly involved. cAMP is a critical intracellular immunoregulatory molecule that can rapidly dampen macrophage antimicrobial responses (19). The lipid mediator PGE2, for example, has been shown to suppress macrophage–GAS interactions via this second messenger (18). Our results confirm that report and extend them to demonstrate that a significant interaction is possible when LTB4 and PGE2 costimulate a macrophage at the time of GAS infection (summarized in Fig. 7). Such crosstalk between these lipid mediators has been shown in other systems (17), adding generalizability to this finding.

This work has limitations, which underscore areas requiring investigation in future studies. For example, SLO−/− mice not only failed to generate increased LTB4 during infection but also lacked the capacity to produce the cysteinyl LTs, such as LTC4, D4, and E4. The extent to which such compounds affect survival or bacterial clearance during infection will require further study. Preliminary studies in our laboratory suggest that exogenously added cysteinyl LTs also have positive immunomodulatory actions on macrophage–GAS interactions (data not shown), and their contribution to the in vivo phenotype of infected mice remains to be determined. Another limitation was our use of a human macrophage–like cell line to build a model of LTB4 immunoregulation. We used this THP-1 cell line because it demonstrated responsiveness to LTB4 in the regulation of GAS phagocytosis similar to that of several other primary macrophage types, including human and mouse macrophages. A longer term plan is to validate our complete model in primary cells.

The mouse model of intrauterine infection that we have newly developed for this study involved nonpregnant mice that were also not immediately postpartum. It is possible that different results would be obtained if experiments were conducted in such animals, particularly given the hormonal and immunological differences associated with pregnancy and the puerperium (6, 66). Experiments to assess the impact of pregnancy and the postpartum state on this infection are planned.

In summary, this study newly establishes a role for LTs in positively regulating innate immune defenses against the puerperal sepsis pathogen GAS. These findings confirm those in other model systems and suggest that exploiting this endogenous signaling system pharmacologically might be a robust mechanism for improving therapeutic approaches to this infection. For example, amplifying the synthesis and signaling LTB4, while limiting the production and responsiveness to PGE2, might significantly enhance the efficacy of existing medical treatments of severe GAS infections. Future studies to assess such approaches are in order. The global problem of increasing antimicrobial resistance further highlights the ongoing need for new treatment modalities for life-threatening infections. Reducing maternal mortality requires ongoing studies of fundamental mechanisms of disease, with a goal of identifying novel targets for treatment and prevention.

**TABLE 3.** Summary of immunoregulation of macrophages by LTB4 during S. pyogenes infection. Ligation of BLT1 by LTB4 results in the activation of Goi protein, with a subsequent decrease in intracellular cAMP levels. This, in turn, limits the activation of protein kinase A (PKA), an endogenous suppressor of both phagocytosis and NADPHox-dependent ROI generation. These actions oppose those of the endogenous lipid mediator PGE2, which stimulates cAMP production via EP2 and/or EP4 receptors in macrophages. Compounds used in our studies to modulate signaling are shown. AC, Adenylate cyclase; APY, apocynin.

![FIGURE 6](http://www.jimmunol.org/)

**FIGURE 6.** Effects of exogenous PGE2 and LTB4 on phagocytosis and cAMP production in macrophages. (A) PMA-treated THP-1 cells were exposed for 1 min to LTB4 (10 nM) or vehicle followed by PGE2 (1 μM) for 3 min. Intracellular cAMP was determined as described in the text. Data are expressed as the mean ± SEM from seven independent experiments. **p < 0.01 compared with untreated cells and ***p < 0.001 compared with PGE2 treatment. (B) PMA-treated THP-1 cells were cultured with LTB4 (100 nM) and PGE2 (10 μM) for 15 min before challenge with HK FITC-GAS. Phagocytosis was quantified after 3 h, as detailed in Materials and Methods. Data are expressed as the mean ± SEM relative to untreated cells for four independent experiments. **p < 0.01 compared with untreated cells and ***p < 0.001 compared with LTB4.
References


