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Pneumococcal Surface Protein A Inhibits Complement Deposition on the Pneumococcal Surface by Competing with the Binding of C-Reactive Protein to Cell-Surface Phosphocholine

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In the presence of normal serum, complement component C3 is deposited on pneumococci primarily via the classical pathway. Pneumococcal surface protein A (PspA), a major virulence factor of pneumococci, effectively inhibits C3 deposition. PspA’s C terminus has a choline-binding domain that anchors PspA to the phosphocholine (PC) moieties on the pneumococcal surface. C-reactive protein (CRP), another important host defense molecule, also binds to PC, and CRP binding to pneumococci enhances complement C3 deposition through the classical pathway. Using flow cytometry of PspA+ and PspA− strains, we observed that the absence of PspA led to exposure of PC, enhanced the surface binding of CRP, and increased the deposition of C3. Moreover, when the PspA− mutant was incubated with a pneumococcal eluate containing native PspA, there was decreased deposition of CRP and C3 on the pneumococcal surface compared with incubation with an eluate from a PspA mutant. Also, there was much greater C3 deposition onto the PspA− pneumococcus when exposed to normal mouse serum from wild-type mice as compared with that from CRP knockout mice. Furthermore, when CRP knockout mouse serum was replenished with CRP, there was a dose-dependent increase in C3 deposition. The combined data reveal a novel mechanism of complement inhibition by a bacterial protein: inhibition of CRP surface binding and, thus, diminution of CRP-mediated complement deposition.

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Abbreviations used in this article: CBA/N, CBA/CaHN-Btkxid/J; CRP, C-reactive protein; CRPKO, CRP knockout; FL-PspA, full-length native pneumococcal surface protein A; GVB, gelatin veronal buffer; MFI, mean fluorescence intensity; NHS, normal human serum; NMS, mouse serum from nonimmunized mice; PC, phosphocholine; PFA, paraformaldehyde; PspA, pneumococcal surface protein A; PspC, pneumococcal surface protein C; rCRP, recombinant CRP; SAF, serum amyloid P; THY, Todd-Hewitt broth supplemented with 5% yeast extract; WT, wild-type.

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protein A (PspA), which is present on virtually all strains of pneumococci and is highly immunogenic (9–12). There are two major families of PspA, family 1 and family 2, which are further divided into clades (13, 14). Any given strain expresses only a single PspA allele, and thus expresses PspA of only one family. Strains of each major capsular type includes strains that express PspA family 1 and strains that express PspA family 2 (10, 14). Both families of PspA are able to inhibit C3 deposition on the pneumococcal surface and confer virulence on pneumococci (12). Proteins within PspA families are immunologically cross-reactive (15, 16), and immunity to a member of either family is generally protective against members of the same family and frequently cross-protective against strains of the other PspA family (9, 17, 18).

Full-length native PspA (FL-PspA) consists of five distinct domains. From the N to C terminus, these are a signal peptide, a highly charged antiparallel coiled coiled α-helical domain, a proline-rich region, a choline-binding domain, and a C-terminal 17-aa tail (19–21). PspA is a member of a family of choline-binding proteins on the pneumococcus, which are anchored to the pneumococcal surface via their choline-binding domains (22). Choline-binding domains recognize the phosphocholine (PC) on the cell wall and cell-membrane-associated teichoic acids (23). Mutant PspA proteins lacking the choline-binding repeat are largely absent from the bacterial surface (12, 23, 24), and mutant strains lacking surface PspA show reduced virulence and greater complement deposition in normal serum (11, 12, 25). An isogenic strain lacking 6 of its 10 choline-binding repeats showed intermediate surface expression of PspA, reduced virulence, and intermediate deposition of C3 (12). It has been shown using normal serum lacking detectable anti-pneumococcal Ab that complement deposition on pneumococci occurs via the classical pathway (26, 27). In the absence of PspA, the classical pathway–dependent, C1q-dependent deposition of complement on pneumococci in normal serum is greatly increased (12, 27). Accordingly, PspA strains show greater clearance from blood compared with PspA− strains (11, 28). However, the mechanism accounting for PspA’s effect on complement has not been described.

CRP is found in low amounts (generally 1–3 μg/ml) in the serum of healthy humans but can increase ≥500-fold in response to inflammation or infection (29, 30). CRP is evolutionarily conserved, and healthy mice have blood concentrations of CRP comparable with healthy humans (31). Importantly, however, mouse CRP is a minor acute-phase reactant and can only increase in concentration ~2-fold during inflammation (31). Like PspA, CRP can bind PC on the pneumococcal cell surface (32) where it activates the classical pathway in a C1q-dependent manner (33, 34) and can protect against invasive infection (35, 36). Because PspA is the most abundant pneumococcal choline-binding protein (22, 37), its absence should significantly increase: 1) PC exposure on pneumococcal surface, 2) binding by CRP, 3) complement activation, and 4) greater opsonization of pneumococci. In this study, we tested the hypothesis that surface-bound PspA minimizes the binding of CRP, thus decreasing CRP-mediated activation of the complement pathway. PspA+ and PspA− strains were examined for their exposure of PC and their ability to bind both CRP and natural Ab to PC. We also examined the ability of these strains to be targets of complement deposition when exposed to complement from wild-type (WT) versus CRP knockout (CRPKO) mice.

**Materials and Methods**

**Bacterial strains and cultures**

The strains used in this study included capsular serotype 2 pneumococci D39 (PspA+)(38) and its isogenic mutants (JY182) and (TRE108) lacking PspA or pneumococcal surface protein C (PspC), respectively (24, 27, 39).

The PspA of D39 is family 1, clade 2 (40). We confirmed that the respective PspA and PspC proteins were not expressed in these mutants by immunoblot assay. For the PspA mutants, we also examined PspA expression by flow cytometry where JY182 showed only background fluorescence with anti-PspA and D39 showed 7-fold higher mean fluorescence intensity (MFI). PspA expression was not changed in the pspC mutant. Both JY182 and TRE108 showed somewhat more capsule than strain D39; however, this difference was not statistically significant (data not shown).

Capsule quantitation was done using a multiplex immunoassay developed in Dr. Moon Nahm’s laboratory at the University of Alabama at Birmingham (41). Strains D39, JY182, and TRE108 were grown in triplicate in 3-m1 volumes of Todd–Hewitt broth supplemented with 5% yeast extract (THY), with or without erythromycin (0.3 μg/ml), to OD<sub>600</sub> 0.5. Next, 500 μl of each culture was lyzed in 1/4× lysis buffer (41). Ten-fold dilutions ranging from 1/10 to 1/1000 of lysates were used to inhibit the ability of type 2 anti-capsule Abs to bind polysaccharide-coated latex beads. Purified type 2 polysaccharide ATCC 16-X (American Type Culture Collection, Manassas, VA) was used as the standard.

Strain Rx1 (PspA−)(11, 42) is a nonencapsulated derivative of D39. WG44.1 is a pspA-null mutant of Rx1. PspA− WT strains TIGR4, P765, P1547, and P1121 are capsular types 4, 6B, 6A, and 23F, respectively (43–45). A PspA mutant strain was made by double crossover insertion–deletion as described previously (44). The pspA mutation was transferred to the type 4, 6B, and 23F strains by sequential back transformation and selecting with 200 μg/ml spectinomycin (Sigma-Aldrich, St. Louis, MO). A 6B revertant was prepared by transforming the 6B pspA mutant with a PCR product of the 6B pspA gene including ~800 bp flanking regions and screening for PspA protein expression by colony immunoblot with clade-specific anti-PspA sera (46). The revertant was confirmed by loss of spectinomycin resistance and by DNA sequence analysis. Strains P765, P1547, and P1121 are PspA family 1, and are PspA clades 1, 2, and 1, respectively. Capsular type 4 strain TIGR4 is family 2, clade 3 PspA.

Bacteria that were used in the CRP binding and C3 deposition assays were grown directly from frozen stocks on the day of assay and were washed in PBS before use. The cells were grown in THY (with erythromycin added to the mutant strains) to an OD<sub>600</sub> 0.4, diluted back to an OD<sub>600</sub> 0.1, and then regrown in THY to an OD<sub>600</sub> 0.4 for use in the assays. For enumeration of pneumococci, cultures were serially diluted, plated on blood agar with or without erythromycin, and grown in a candle jar overnight.

**Sera and buffers**

Fresh-frozen “normal” mouse serum from nonimmunized mice (NMS) was obtained from CBA/CaBRH−H(1, 2) (CBAN) mice, which have no natural Ab to PC (47); BALB/C mice, which have natural Ab to PC (48); and CRPKO mice (Cpr<sup>−/−</sup> (C57BL/6f background)), which do not express any CRP (49). NMS for use as a complement source was pooled from groups of 8–10 mice of each genotype. The sera from all sources were aliquoted and stored at –80°C. In assays to measure CRP or C3 deposition, all buffers had a pH of 7.3; all dilutions were in gelatin veronal buffer (GVB) containing 0.15 mM calcium, 0.5 mM magnesium (GVB<sup>2+</sup>; CompTech, Tyler, TX); and all the washing steps were performed using PBS with 0.02% NaN<sub>3</sub> or Ca<sup>2+</sup> (Mediatech, Manassas, VA). These studies were conducted in accordance with the principles set forth in the Guide for Care and Use of Laboratory Animals (Institute of Laboratory Animal Resources, National Research Council, revised 1996), and approved by the University of Alabama at Birmingham Institutional Animal Care and Use Committee.

**Isolation of eluates containing FL-PspA**

Rx1 and WG44.1 pneumococci were grown to an OD<sub>600</sub> 0.4 in 200 ml of chemically defined medium containing 0.03% choline chloride. FL-PspA was isolated and dialyzed as previously described (37).

One-fourth serial dilutions of cell eluates were immunoblotted on nitrocellulose membranes, blocked with 1% BSA in PBS, incubated with PspA-specific mAb, Xir278 (50), and developed with biotinylated goat anti-mouse Ig, alkaline phosphatase-conjugated streptavidin (Southern Biotechnology Associates, Birmingham, AL) and nitroblue tetrazolium substrate with 5-bromo-4-chloro-3-indolyl phosphate p-toluidine salt (Fisher Scientific, Norcross, GA), to qualitatively detect PspA. The presence of eluted PspA was further confirmed by a Western immunoblot assay (37, 50). The amount of total protein in these bacterial eluate samples was determined by Bio-Rad molecular weight Standards and the amount of BSA added in the FL-PspA isolation step was subtracted to yield net (total) bacterial eluate protein.

The amount of PspA in the cell eluate from Rx1 was determined using an ELISA. Microtiter 96-well plates (NUNC, Wiesbaden, Germany) were
coated overnight at 4°C with 100 μl of a mixture of mAb XR278 (50) and RX1003 (Genschmer and Briles, manuscript in preparation) at 2 μg/ml PBS total concentration (1 μg/ml each). Plates were washed, blocked with 150 μl 1% BSA/0.2% casein (BSA/casein) for 1 h, and washed; then Rx1 (PspA) and WG44.1 (PspA) cell eluates were titrated out on the plate in one-third dilutions in BSA/casein starting at 100, 10, and 1 μg/ml total protein. Finally, the plates were incubated at room temperature for 1 h with rabbit anti-PspA (1:1000) (14), washed, incubated for 1 h with 1:10,000 biotin-conjugated donkey anti-rabbit Ig antiseraum IgG (H+L, Southern Biotechnology Associates), washed, incubated for 1 h with 1:4000 streptavidin conjugated to alkaline phosphatase (Southern Biotechnology Associates), and developed with p-nitrophenyl phosphate (Sigma-Aldrich), and the absorbance read at 405 nm.

rUAB055 (51) was used as a standard at 60 ng/ml titered 1:3 in BSA/casein. The total PspA concentration in the cell eluate was calculated and expressed as ng/ml PspA.

Detection of exposed PspA and PC on the pneumococcal surface

Pneumococci (D39, JY182, TRE108, Rx1, and WG44.1) were grown as above and resuspended to 1 × 10^9 CFU/ml in GVB. For the PspA expression assay, D39, JY182, and TRE108 were incubated in 100 μl of 1:500 PspA-specific mAb (XR278) at 37°C for 30 min with shaking. Samples were then washed and incubated with 50 μl FITC-labeled goat anti-mouse Ab (Southern Biotechnology Associates) diluted 1:100 in GVB at 37°C for 30 min with shaking. They were then washed and fixed with 500 μl 2% paraformaldehyde (PFA; Electron Microscopy Sciences, Hatfield, PA) for analysis by flow cytometry. The PC exposure assay was carried out the same way as the PspA expression assay except that the IgA hybridoma TEP15 (Sigma-Aldrich) was used to detect PC on strains and mutants for 20,000 gated events.

Ab binding to pneumococcal surface

Types 6B, 4, 6A, and 23F, together with their pspA mutants (and revertant, in the case of 6B), were grown to midlog phase, washed, and resuspended in 1% BSA/PBS containing primary Ab. Ab for primary incubation was as follows: human IgG to PC (1:10) purified from pooled γ-globulin as previously described (52), normal human serum (NHS; 1:100) obtained from healthy adults, and NMS (1:10) pooled from BALB/c mice. Bacteria were incubated with primary Ab at 37°C for 30 min, washed, and bound Ab was detected using the appropriate species-specific anti-IgG or anti-IgM conjugated to FITC (1:200) (Sigma-Aldrich) at room temperature for 30 min. Finally, bacteria were washed and resuspended in 2% PFA for analysis by flow cytometry for 20,000 gated events.

CRP binding assay

D39, JY182, and TRE108 strains were grown fresh from frozen stocks. Cultures were centrifuged at 3000 rpm, washed twice, and pneumococci were resuspended in GVB to a concentration of 1 × 10^9 CFU/ml. Two hundred microliters of bacterial suspension was incubated with the indicated concentrations of human recombinant CRP (rCRP, US Biologicals, Swampscott, MA) at 37°C for 30 min. For the assays where cell eluates from Rx1 and WG44.1 were used, bacteria were first incubated with appropriate concentration of eluate for 1 h at 37°C followed by the incubation step with CRP. These were washed twice and then incubated with 50 μl FITC-conjugated mouse anti-human CRP (1:50; HyTest, Turku, Finland) at 37°C for 30 min. Finally, bacteria were then washed and fixed with 500 μl 2% PFA. The controls included bacteria incubated with only GVB and bacteria incubated with only FITC-conjugated secondary Ab. Flow cytometric analysis was carried out using an FACSCalibur instrument (Becton Dickinson, Mountain View, CA) for 40,000 gated events. To make sure that the CRP was not agglutinating pneumococci, we examined bacteria by slide agglutination. Even at concentrations as high as 250 μg/ml, rCRP was not able to agglutinate strains D39, JY182, TRE108, and a nonencapsulated mutant of D39, AM1000 (53) (data not shown). We also failed to see evidence of CRP-induced aggregation in our flow cytometry studies with D39, JY182, and TRE108. These findings are consistent with the fact that all PC binding sites of CRP are on the same face of the CRP pentamer.

CRP binding assay

Pneumococci at 1 × 10^9 CFU/ml in GVB were prepared as described earlier. Fresh-frozen NMS (complement) from CBA/N mice or CRPKO mice were resuspended in GVB to a concentration of 100, 10, and 1 μg/ml total protein. Finally, the plates were incubated at room temperature for 1 h with rabbit anti-PspA (1:1000) (14), washed, incubated for 1 h with 1:10,000 biotin-conjugated donkey anti-rabbit IgG (H+L, Southern Biotechnology Associates), washed, incubated for 1 h with 1:4000 streptavidin conjugated to alkaline phosphatase (Southern Biotechnology Associates), and developed with p-nitrophenyl phosphate (Sigma-Aldrich), and the absorbance read at 405 nm. 

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to PC probably reflects the fact that NHS invariably contains Ab to PspA (57), which would bind more strongly to WT than mutant pneumococci.

There is greater binding of CRP to the surface of the pspA mutant strain than to its WT parent
CRP is known to efficiently deposit C3 on encapsulated pneumococci (32, 34), so it was likely that CRP might contribute to increased complement deposition on PspA−-deficient pneumococci. The WT D39 strain was examined along with its pspA mutant (JY182) and pspC mutant (TRE108) to determine the relative effects of PspA and PspC on the binding of CRP. As predicted, the pspA mutant showed an increase of 2.28-fold in CRP binding as compared with the WT strain (Fig. 2). A measurable increase in CRP binding to the pspC mutant was not observed, consistent with PspC’s smaller effect on PC exposure (Fig. 2).

The CRP present in normal serum is sufficient for C3 deposition on WT pneumococci and pspA mutant pneumococci
CRP is known to efficiently deposit C3 on encapsulated pneumococci (32, 34), so it was likely that increased CRP binding because of the absence of PspA might contribute to increased complement deposition on PspA-deficient pneumococci. Despite low levels of CRP in NMS, we suspected that there would be enough CRP to support the classical pathway-dependent C3 deposition that has been observed on WT and PspA-deficient pneumococci in NMS (26, 27). In the presence of WT NMS, we observed much stronger C3 deposition than in the presence of NMS from CRPKO mice (Fig. 3A). Moreover, in the presence of normal serum, there was more complement deposition on PspA− versus PspA+ D39 pneumococci. However, when CRPKO serum was the complement source, the amount of C3 deposited was essentially the same as when we incubated the bacteria without complement (Fig. 3A). These data confirmed that CRP was necessary for the deposition of C3 on pneumococci.

The deposition of C3 onto the surface of PspA− pneumococci is CRP dose dependent
The interpretation that CRPKO NMS failed to support complement deposition because of its lack of CRP (Fig. 3A) depends, in part, on the assumption that the CRPKO NMS contained sufficient C3 to cause detectable deposition onto pneumococci if CRP had been present. To test this, we incubated JY182 (PspA−) with NMS from CRPKO mice at 37˚C, with and without the addition of human rCRP. C3 binding to the surface of JY182 was measured by flow cytometry. As a negative control, pneumococci received neither CRP nor CRPKO NMS. Compared with pneumococci incubated with CRPKO sera, there was a 1.5-, 2.2-, and 2.8-fold increase in C3 binding after addition of 50, 100, and 150 μg/ml CRP, respectively (Fig. 3B). Thus, these data make it clear that the very low C3 deposition in the presence of CRPKO NMS is due to a lack of CRP and not due to inadequate complement levels.

A pneumococcal eluate containing FL-PspA inhibited CRP binding to the surface of JY182
The earlier data are consistent with the premise that PspA on pneumococci reduces PC exposure leading to impaired binding by CRP and the deposition of C3. Even though independent pspA
mutants were used in some of the experiments and all mutants are well characterized, the possibility exists that differences between the WT and mutant strains may have been caused by unknown changes in non-\textit{pspA} genes. To independently determine whether PspA can inhibit CRP deposition and C3 deposition, we added FL-PspA to JY182 before the addition of CRP or NMS.

FL-PspAs, in contrast with rPspA fragments that lack the choline-binding domain, are quite unstable and cannot be isolated and concentrated without extensive denaturation and loss. However, FL-PspA can be recovered in 2\% choline-chloride eluates of WT pneumococci (37). A cell eluate containing FL-PspA was isolated in this manner from the nonencapsulated strain Rx1 that had been grown in a chemically defined medium. A control lysate was prepared from WG44.1, a \textit{pspA} null mutant of strain Rx1. Rx1 and WG44.1 cell eluates were incubated separately at 5 and 10 \textmu g/ml total eluate protein with JY182 (PspA\textsuperscript{+}), washed to remove unbound FL-PspA, incubated with CRP, and developed to detect CRP binding to the bacteria. The eluates of Rx1 (PspA\textsuperscript{+}) at 5 and 10 \textmu g/ml contained 9.9 and 19.8 ng/ml PspA, respectively, as determined by a PspA-capture assay. This assay detected no PspA (<0.1 ng/ml) in the 5 and 10 \textmu g/ml concentrations of the eluate of WG44.1 (PspA\textsuperscript{−}). The eluate from Rx1, but not WG44.1, inhibited CRP deposition onto JY182 (Fig. 4A). Based on MFI in the presence of 5 and 10 \textmu g/ml total protein from the two eluates, there was 1.13- and 1.37-fold more CRP deposition, respectively, in the presence of the WG44.1 versus the Rx1 extract. Because the only difference between the two eluates was the presence of PspA, the results indicate that FL-PspA can inhibit CRP deposition on JY182.

A FL-PspA–containing pneumococcal eluate inhibited C3 deposition onto the surface of JY182

The FL-PspA–containing eluate of Rx1 and the control eluate from WG44.1 lacking FL-PspA were added to JY182 before the addition of NMS to measure its blocking effect on C3 deposition (Fig. 4B). The PspA-containing Rx1 cell eluate reduced C3 deposition compared with the WG44.1 eluate. At 20 and 40 \textmu g/ml protein added, there was a 1.32- and 1.28-fold greater deposition of C3 on bacteria treated with WG44.1 cell eluate compared with those treated with Rx1 (Fig. 4B). Although these differences were small, they were significant at \textit{p}=0.002 (see legend for Fig. 4B). The fact that the effect was not larger with the higher dose may mean that the lower dose had already saturated the binding sites for PspA. These results show that treatment of PspA\textsuperscript{−} pneumococci with an extract containing FL-PspA was able to inhibit subsequent C3 deposition. Importantly, when the same experiment was repeated using CRPKO NMS as the complement source, there was \textasciitilde1/15 as much C3 deposited on the surface of JY182. Moreover, in the presence of CRPKO NMS, no significant difference was found.

![FIGURE 2](image_url) Relative effects of \textit{pspA} and \textit{pspC} mutations on CRP binding. WT (D39), PspA\textsuperscript{−} mutant (JY182), and PspC\textsuperscript{−} mutant (TRE108) strains were examined for their ability to bind CRP in the presence of 100 \textmu g/ml human rCRP using flow cytometry as described. (A) The average MFI of anti-CRP is shown in the bar plots. Error bars represent the range of duplicate samples. Bacteria that were not exposed to added CRP showed <10 on the MFI scale (not shown). (B) Flow cytometry histogram of the number of events at different fluorescent intensities is shown for D39, JY182, and TRE108. This experiment was conducted twice with duplicates. A representative experiment is shown. Gray peak represents D39 in the absence of CRP, but with FITC-labeled Ab to CRP.

![FIGURE 3](image_url) Effect of CRP on complement C3 deposition on pneumococci. (A) D39 (PspA\textsuperscript{+}) and JY182 (PspA\textsuperscript{−}) were incubated with 10\% NMS from either CBA/N or CRPKO mice as the complement source for 30 min at 37˚C with shaking. The cells were then incubated with 1:100 FITC-labeled Ab to C3 and fixed for flow cytometric analysis to detect bound C3. (B) Effect of exogenously added CRP on C3 deposition on the surface of JY182 (PspA\textsuperscript{−}) using CRPKO NMS as complement source. Ten percent CRPKO NMS was added to all the samples except for the negative control. Human rCRP was added at concentrations of 0, 50, 100, or 150 \textmu g/ml. The negative control had no CRP added. The mixtures were incubated at 37˚C for 1 h. C3 was detected by the addition of 1:100 anti-mouse C3 and 1:100 FITC-labeled goat Ab in separate 30-min incubations. Cells were fixed and analyzed by flow cytometry. Both experiments (A, B) were conducted with duplicates only once. Error bars represent the range of duplicate samples.
between the C3 deposited in the presence of the PspA+ and PspA− eluates (data not shown). These results confirm a role for CRP in complement activation and deposition on the surface of PspA− pneumococci. Furthermore, CRP-dependent activity could be inhibited by the addition of PspA expressing an intact choline-binding domain.

To test the hypothesis that the choline-binding domain was essential to inhibit the deposition of CRP and C3, we repeated the inhibition studies using rPspA, UAB055 (51), that contains the N-terminal α-helical region of PspA and 14 aa of the proline-rich domain, but completely lacks PspA’s choline-binding domain. There was no blocking effect with either 5 or 10 μg/ml rPspA on the binding of CRP to pneumococci or the deposition of C3 on pneumococci (Fig. 5). This lack of inhibition is especially striking because the concentrations of rPspA used were up to 500 times the concentration of FL-PspA used in the prior experiments. This observation is consistent with our hypothesis that because native PspA binds to PC, it is able to inhibit CRP binding and thus indirectly blocks CRP-dependent activation of C3 and deposition of C3b onto the pneumococcal surface.

Discussion

Both the classical and alternative complement pathways are known to be important for host defense against pneumococci (12, 26, 27, 58). Total complement deposition is greatly affected by defects in the alternative pathway because it acts to amplify any complement deposited on the pneumococcus via the classical or lectin pathways. However, the classical pathway appears to be the primary trigger for almost all complement deposition on highly virulent strains of pneumococci. This is true even in the presence of nonimmune sera where complement deposition on pneumococci is dependent on the initial complement activation by the classical pathway (26, 27, 59). PspA is a choline-binding protein of pneumococci that binds to the PC epitopes on teichoic acids (23), and its virulence role of PspA in bacteremia and sepsis has been attributed largely to its ability to interfere with Ab-independent activation of the complement system (12, 27, 60).

CRP, when bound to the PC of teichoic acids, is able to bind C1q and activate complement via the classical pathway (34). CRP can activate complement when bound to S. pneumoniae and Haemophilus influenzae (61, 62). CRP is present at a few micrograms per
milliliter in all human and mouse sera (29, 31). It can protect mice from pneumococcal infection (35, 36), and this protection requires complement (61). PspA can significantly reduce complement deposition on pneumococci in normal serum, and this additional complement deposition is all C1q dependent (27). In this study, we hypothesized that the ability of PspA to reduce C1q-dependent complement activation (27) by pneumococci in normal serum was by inhibiting the binding of CRP to pneumococci, thus reducing CRP-mediated, C1q-dependent activation of the classical complement pathway (26, 34).

Our prior studies have shown that, in addition to exhibiting greater complement deposition in normal serum, PspA- pneumococci exhibit greater in vitro phagocytosis and faster clearance in nonimmune mice as compared with PspA- WT strains (11, 12, 25, 63). In this study, we showed that mutant pneumococci lacking PspA had significantly higher exposure of PC, greater deposition of rCRP, and greater deposition of C3 from NMS than was observed with WT pneumococci that expressed PspA. Moreover, enhanced C3 deposition on the strain lacking PspA was not observed in the presence of nonimmune mouse serum from CRPKO mice. However, when human rCRP was added to CRPKO serum, the deposition of C3 was restored. The role of PspA’s choline-binding activity in inhibiting C3 deposition was strengthened by the observation that FL-PspA, but not rPspA lacking a choline binding domain, could inhibit CRP binding and C3 deposition. These results support our hypothesized mechanism of classical pathway inhibition by PspA. The results also indicate that in normal serum, complement activation via the classical pathway can be largely attributed to CRP. The importance of this inhibitory effect by PspA on complement deposition is probably magnified during a pneumococcal infection in human where CRP is an acute-phase protein, which can increase in concentration >500-fold during an infection (30).

Virtually all mice, except for Xid mice, produce Ab to PC in their NMS (47, 64), and these Abs can have a protective effect against vivo infection (47, 63). However, the fact that so little C3 deposition was observed with the CRPKO sera indicates that it is the CRP, and not the Ab to PC, that accounts for the bulk of the C3 deposition on WT pneumococci in NMS. In fact, in Fig. 3A, we observed much higher C3 deposition using CBA/N NMS, which lack Ab to PC but have CRP, than we did with CRPKO NMS, which lacks CRP but should contain IgM Ab to PC.

The absence of another choline-binding protein, PspC, resulted in only a small exposure of PC, did not increase CRP binding, and does not increase classical pathway complement deposition (27). It is likely that the reason PspA, but not PspC, significantly inhibited CRP binding was because PspA is present in much higher concentrations on the pneumococcal surface than PspC, and is thus able to cover much more PC. Although strict comparisons have not been made between the numbers of molecules of PspA and PspC expressed, it is noted that the PspC (CbpA) protein, and other choline-binding proteins, could not be originally detected in pneumococcal surface extracts without using pneumococci with a pspA mutation to remove the coisolated PspA from the extracts (22). It was also noted in early studies where PspA was isolated by elution with choline chloride from the pneumococcal surface that the major product was PspA (23, 37).

The observation that PspA’s effect on complement deposition resided in its choline-binding domain is in contrast with the observation that all known protective mAb to PspA are directed against the a-helical or proline-rich domains of PspA (50, 65–67). Thus, the protective effects of Ab to PspA are not because they interfere with PspA’s ability to inhibit complement. Rather, the ability of Ab to PspA to enhance complement deposition on pneumococci and be protective (59, 68) probably depends on its ability to bind C1q and activate the classical pathway in the same manner as Abs to other Abs.

The a-helical and proline-rich regions of PspA are highly exposed to Ab at the pneumococcal surface (66, 69) where they can be targeted by protective Ab (50, 65–67). To evolutionarily justify the surface exposure of PspA, there must be a function mediated by the exposed portions of the molecule. The a-helical end of PspA blocks the bactericidal activity of cationic bactericidal peptides released by the degradation of apolactoferrin by binding strongly to the bactericidal peptides (21, 70, 71). PspA may block the bactericidal function of cationic peptides in general because it has been observed to bind and inhibit killing by other bactericidal peptides (S. Mirza, B. Hatcher, and D. Briles, unpublished observations).

Like CRP, serum amyloid P (SAP) is another PC-binding pentraxin that has been shown to opsonize pneumococci with C3b via the classical pathway and reduce infection in vivo in mice (33). SAP, however, has a different fine specificity from CRP in that it binds better to phosphorylcholine at (72, 73), which is not normally expressed on pneumococci (74). Few direct in vivo comparisons between the two pentraxins have been conducted, but in early studies performed in mice, SAP was less protective against pneumococcal infection than CRP (36). Even so, it is anticipated that SAP and CRP each contribute to complement deposition on pneumococci and subsequent bacterial clearance.

Finally, many mucosal pathogens, including S. pneumoniae, H. influenzae, Pseudomonas aeruginosa, Neisseria meningitidis (62, 75), Salmonella enterica Typhimurium (76), Proteus mirabilis (77), and the nonmucosal parasitic worms Brugia malayi (78) express PC on their surface. In most cases, the PC can be detected with the same T15 idiotype Abs to PC that are optimally protective against pneumococci (79). The expression of PC may aid these pathogens in the mucosal environment by facilitating adherence and colonization (62, 80). The frequent expression of PC by such pathogens may explain, in part, why mammalian hosts have evolved choline-binding triggers of innate immunity such as CRP and SAP. Mice and men also make strong (in the case of mice, germline) (81) anti-PC Ab responses. Both mouse (79) and human (52, 63) Abs to PC can protect against pneumococcal infection. Not only have pneumococci developed a means to mask their PC with PspA (this study), they (43) and some Gram-negative mucosal pathogens (62, 75) modify PC exposure by phase variation. In pneumococci, the invariant PC epitope has been masked by PspA, which, unlike PC, is not detected by the innate immune system and is antigenically variable.

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References
PsP inhibits CRP binding to pneumococcal phosphocholine


