Preferential Expansion of Human Virus-Specific Multifunctional Central Memory T Cells by Partial Targeting of the IL-2 Receptor Signaling Pathway: The Key Role of CD4+ T Cells

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Primary/secondary T cell deficiency is associated with poor control of viruses, virus-associated tumors, and other intracellular pathogens. Adoptive T cell therapy is a novel therapeutic approach to restore immune competence (1–3), and it might be a particularly efficient tool for accelerating immune reconstitution following hematopoietic stem cell transplantation to control life-threatening viral and fungal infections in those patients (4). Recent proof-of-concept studies demonstrated promising data in EBV-related posttransplant lymphoproliferative disease and human CMV (HCMV) disease in permanently immunosuppressed patients. The infused T cells showed no adverse effects after adoptive transfer and induced rapid and effective control of viruses in almost all SOT patients (7, 8). However, the promising data on adoptive therapy in hematopoietic stem cell transplantation and SOT patients also revealed some limitations: the duration of efficacy is not sufficient in many patients (1, 7, 8).

Our recent data on adoptive therapy of antiviral drug-resistant HCMV disease in SOT patients illustrate the problem: HCMV causes frequent and severe complications in SOT recipients (9). We developed a whole protein-spanning peptide pool-based protocol to generate HCMV-specific T cell lines to almost all CD4/8 epitopes in an HLA-independent manner (7). HCMV-specific T cell lines with effector function can be successfully generated in vitro, even from diseased and chronically immunosuppressed patients. The infused T cells showed no adverse effects after adoptive transfer and induced rapid and effective control of viruses in almost all SOT patients (7, 8). However, the promising data on adoptive therapy in hematopoietic stem cell transplantation and SOT patients also revealed some limitations: the duration of efficacy is not sufficient in many patients (1, 7, 8).

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Abbreviations used in this article: FSC, forward light scatter; HCMV, human CMV; LCL, lymphoblastoid B cell line; LD, low dose; mTOR, mammalian target of rapamycin; rh, recombinant human; nTreg, natural regulatory T cell; SOT, solid organ transplantation; TeM, central memory T cell; TeMR, effector memory T cell; TeMRA, terminally differentiated effector T cell; VLDL, very low dose.

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before/after expansion revealed very few, if any, central memory T cells (T_{CM}a), whereas a variable, but detectable, number of virus-specific T_{CM} contributed to the EBV-specific response (1, 10). According to these and other experimental data, the relapse might be due to the late-differentiation phenotype of the infused T cells and, therefore, their insufficient longevity in vivo (11). Late-differentiated effector memory T cells (T_{EM}) and terminally differentiated effector T cells (T_{EMRA}) express powerful effector function but are not able to establish long-lasting protective memory because of diminished proliferative potential and brief survival following antigenic rechallenge (12). Several preclinical studies indicated the superior lasting engrafment of T_{CM}-derived T cells. The persistence of transferred cells correlates with therapeutic efficacy in animals (13). Balancing engrafment fitness with a strong Ag-specific effector function might lead to the best therapeutic outcome.

The mammalian target of rapamycin (mTOR) has been described as a major regulator of memory CD8+ T cell differentiation (14, 15). Several studies described that mTOR inhibition by rapamycin treatment during the expansion and contraction phases improved both the quality and quantity of memory T cells after primary immunization in preclinical models (14–17). Rapamycin seems to prolong the survival of Ag-specific T cells (18, 19). It paradoxically enhances the Ag-specific CD8+ T cell response and accelerates memory T cell formation in vivo (15). Thus, mTOR inhibition enhances the magnitude and quality of an Ag-specific T cell response (14, 15). However, it remains unclear whether mTOR inhibition might also support the targeted composition of T cell products for improved adoptive T cell therapy in patients, particularly because, in contrast to the preclinical models, the memory response was already established in vivo.

In this article, we demonstrate a novel method that enriches human virus-specific T_{CM} in the T cell product, expanded to clinically useful numbers under GMP-conform conditions by modifying IL-2R signaling. This study reveals a potent strategy to generate Ag-specific T cells with a high functional capacity and, presumably, with a robust long-term persistence and functionality in vivo.

**Materials and Methods**

**Cell culture**
All cell cultures and assays were performed in complete media (RPMI 1640 supplemented with 2 mM l-glutamine, penicillin [100 IU/ml], and streptomycin, all from Biochrom), containing 10% FBS (PAA) in humidified incubators at 37°C and 5% CO2.

**Blood samples**
For all experiments, blood samples were collected from healthy volunteers and PBMC were isolated by lymphoprepation density-gradient centrifugation (PAA). The Charité University Medicine Berlin ethics committee (Institutional Review Board) approved the study protocol, and all blood donors provided written informed consent.

**Enrichment and expansion of Ag-specific T cells**
HCMV-specific T cells were enriched and expanded using a previously described technique (1, 7, 20). Briefly, 4 x 10^7 autologous PBMC were stimulated for 6 d with overlapping HCMV pp65IE-1 peptide pools (JPT Peptide Technologies) at 1 µg/ml each peptide. Pools consisted of 15-mer peptides overlapped by 11 aa and were reconstituted in DMSO. IFN-γ-producing cells underwent positive selection using the IFN-γ-secretion capture MACS system, according to the manufacturer’s instructions (Miltenyi Biotec). Enriched IFN-γ+ cells were cultured for 18 d in 24-well plates with irradiated (30 Gy) autologous feeder cells (derived from capture assay-negative fraction) at a 1:100 ratio in culture media supplemented with 50 IU/ml recombinant human (r)IL-2 (Chiron) and 10 ng/ml rHIL-7 (Cellgenix) or either one alone, when indicated. The medium was changed every 3–4 d during culture. Where indicated, a very low dose (rLD; 2 nM) or a low dose (rLD; 20 nM) of mTOR inhibitor (rapamycin; Sigma-Aldrich) or IL-2R antagonist (20 nM) of mTOR inhibitor (rapamycin; Sigma-Aldrich) or IL-2R antagonist (20 µg/ml; basiliximab, Simulect; Novartis) was added every 2 d, beginning on day 1, together with subsequent media supplementation. Where indicated, CD4+ and CD8+ T cells were MACS purified, according to the manufacturer’s instructions (Miltenyi Biotec).

**Allocation of APCs**
Expanded HCMV-specific T cells were analyzed for effector functions by their ability to recognize and eliminate peptide-loaded or virus-incubated target cells which consisted of autologous lymphoblastoid B cell lines (LCL) transformed with B95-8 virus and autologous monocytes. LCL were generated as described previously (7). CD14+ monocytes were enriched from PBMC by magnetic isolation (MACS; Miltenyi Biotec). Subsequently, CD14+ cells were cultured for 6 d in 1000 IU/ml rhGM-CSF (Cellgenix) and 400 IU/ml rIL-4 (Cellgenix). NEWT or fibroblasts (passaged control cell line) were added to monocyte-differentiation cultures on day 4.

**Assessment of cytotoxic activity**
A modified vital assay was used for cytotoxicity testing, as described previously (21). Briefly, monocytes incubated with HCMV-related laboratory wild-type strain NEWT were used as targets (22). In addition, agarose-embedded splenocytes from noninfected (control) mice were pulsed with 1 µg/ml overlapping HCMV pp65IE-1 peptide pools, whereas unpulsed LCL were used as control targets. Targets were labeled with 10 µM CFDA (Molecular Probes). As controls, unpulsed LCL and monocytes coincubated with fibroblasts were labeled with 5 µM dimethylcyclocyanine oxide-succinimidyl ester (Far Red; Invitrogen). Cells were cocultured for 16 h at a T cell/target cell ratio of 1:1 and 10:1. Probes were analyzed as triplicates using an LSRII flow cytometer. Samples without T cells, containing only APC (pulsed/NEWT or unaspecific) and controls were used as internal control. Analysis was gated on LIVE/DEAD staining to discriminate cells with vital dye (Invitrogen). The mean percentage survival of pulsed or NEWT-incubated targets was calculated relative to Ag-unpulsed controls. Percentage lytic activity of target lysis was calculated as follows: mean percentage survival of targets in cultures containing defined numbers of effector T cells in comparison with control cells without T cells.

**Intracellular cytokine staining**
Following Ag-specific stimulation, cytokine secretion (IFN-γ, TNF-α, and IL-2) and activation marker expression (CD154 and CD137) were determined by intracellular fluorescence staining. All Abs were purchased from Becton Dickinson, except when otherwise indicated. Virus-incubated monocytes and peptide pool-loaded LCL or monocytes (1 µg/ml IE-1/pp65) were added to T cells in a diluting ratio of 1:10. Unpulsed LCL or monocytes (DMSO incubated) were used as unstimulated controls. For effector cytokine detection, cultured T cells were restimulated for 18 h in the presence of 1 µg/ml brefeldin A (Sigma-Aldrich). For CD107a (H4A3) detection, 2 µM monensin (Golgiti Stop; Becton Dickinson) was added together with brefeldin A. Cells were then harvested, and the phenotype was stained with mAbs for surface markers CD3 (UCHT1; ebioscience), CD8 (3B5; Invitrogen), and CD4 (SK3). To define memory phenotypes, T cells were extracellularly stained for CCR7 (#150503; R&D Systems), CD62L (DREG56; Beckman Coulter), CD45RA (2H4LDH11LDB9; Beckman Coulter), and CD127 (hiL-7-R-M21). To exclude dead cells, LIVE/DEAD discrimination staining dye (Invitrogen) was added. Subsequently, cells were permeabilized with Perm2 solution (BD Biosciences) and stained for IFN-γ (4S.B3; ebioscience), TNF-α (MAb11), IL-2 (MQ1-17H12), CD137 (4B4-1), and CD154 (TRAP1). Cells were measured on an LSRII flow cytometer and analyzed using FlowJo Version 8 software (Tree Star). Lymphocytes were gated based on the forward scatter (FSC) versus side scatter profile and subsequently gated on FSC (height) versus FSC to exclude doublets.

**Statistical analysis and calculations**
The Kolmogorov–Smirnov test was used to determine whether the values came from a Gaussian distribution. Normally distributed data were analyzed by the nonparametric Wilcoxon matched-pairs test. Probability (p) values <0.05 were considered statistically significant.

CD4+ and CD8+ T cell frequency was calculated by comparing CD4+/CD8+ T cell frequency (assessed by FACS) with the cell number of the whole culture. Fold expansion was calculated by dividing the cell number on day 18 by the cell number on day 1. T cells exhibiting a T_{CM} phenotype were defined by CCR7 expression on CD4+CD8+ T cells on day 18. Based on
Results

Partial mTOR inhibition results in increased numbers of early memory T cells and leads to superior specific T cell response

HCMV_{pp65/IE1}-specific CD4^{+}/CD8^{+} T cells from PBMC were isolated using an immunomagnetic IFN-γ-secretion assay following stimulation by HCMV_{pp65/IE1} peptide pools (Fig. 1) (23). Based on CCR7 and CD45RA expression, human T memory cells can be segregated into T_{EMRA}, T_{CM}, and T_{EM} (12). HCMV-specific CD4^{+}/CD8^{+} T cells before/after expansion were predominantly enriched for T_{EM} and T_{EMRA}/T_{CM}, respectively (1).

We wondered whether mTOR inhibition during the culture might support T_{CM} expansion. Although an intermediate dose of rapamycin (50–100 nM, commonly used for supporting natural regulatory T cell [nTreg] expansion) attenuated T cell proliferation (data not shown), the expansion was only marginally diminished at VLD and LD concentrations (Fig. 2A). Interestingly, VLD/LD rapamycin treatment resulted in a greater CD4^{+} T cell yield compared with the control cell line after expansion (Fig. 2B, 2C). As expected, CD4^{+} T cells, which exhibited enhanced expansion in the presence of LD/VLD rapamycin, did not express nTreg markers Helios and Foxp3 (data not shown).

Human T_{CM} uniformly express CD62L; however, T_{EM} are heterogeneous for CD62L expression. Moreover, T_{CM} coexpress CCR7 and characteristically lose CD45RA during naïve → effector transition (12). CCR7 expression is rapidly downregulated upon antigen restimulation, along with the differentiation of T_{CM} → T_{EM} (12). Remarkably, rapamycin-treated HCMV-specific T cell lines showed a significantly higher proportion and absolute number of CCR7^{+}/CD62L^{−} (CD45RA−) T_{CM} (Fig. 2D–G). To assess their effector function, we evaluated the response following specific restimulation. Rapamycin-treated HCMV-specific CD8^{+} T cells showed enhanced CD137 expression, whereas CD137 and CD154 expression on CD4^{+} T cells was unchanged. Of note, similarly to rapamycin treatment, IFN-γ and TNF-α effector cytokine secretion was not diminished after treatment, despite a decrease in the T_{EMRA}/T_{EM} proportion (Fig. 3J).

Targeting the IL-2R pathway does not diminish specific cytolytic capacity of T cells

Despite a greater proportion of T_{CM}, rapamycin/anti-CD25 mAb-treated HCMV-specific T cells showed similar specific killing of peptide-loaded target cells (Fig. 3L). The expression of the degranulation marker CD107a on CD8^{+} T cells remained constant; however, CD4^{+} T cells exhibited increased CD107a surface expression (Supplemental Fig. 1B, 1C).

CD4^{+} T cell help improves antiviral CD8^{+} T cell response

Because both approaches of partial IL-2R targeting supported the expansion of CD4^{+} T cells (Figs. 2C, 3C), we wondered whether the maintenance of CD8^{+} effector functions might be related to the enforcement of CD4^{+} T cell help (26). To test the impact of CD4^{+} T cell quantity on CD8^{+} function, we isolated HCMV-specific T cells from PBMC and cultured, in parallel, bulk T cells, sorted CD4^{+} T cells, and CD8^{+} T cells. CD8^{+} T cells alone showed slightly poorer proliferation, whereas CD4^{+} T cell expansion was similar to bulk culture (data not shown). Of note, HCMV-specific restimulation of purified CD8^{+} T cells revealed nearly complete inhibition of cytokine/activation marker responsiveness (Fig. 4A), but neither the memory phenotype nor degranulation was changed (data not shown). Next, we wondered whether CD8^{+} T cell functionality could be enhanced by supplementing CD4^{+} T cells just before specific restimulation but not during the expansion phase. Surprisingly, CD8^{+} T cell effector cytokine secretion and activation marker expression following specific restimulation proportionally increased with the presence of CD4^{+} T cells, up to a CD4/CD8 ratio of 0.2 (Fig. 4A–C). However, CD4^{+} T cell supplementation had no impact on the expression of CD107a on CD8^{+} T cells (Supplemental Fig. 2A).

Next, we separately cultured isolated HCMV-specific CD4^{+} and CD8^{+} T cells with rapamycin/IL-2Ra antagonist. Interestingly, when cultured alone, neither CD4^{+} T cells nor CD8^{+} T cells exhibited any changes with regard to cytokine secretion or acti-

![FIGURE 1. Enrichment of HCMV-specific T cells. Specifically stimulated T cells underwent positive selection by the IFN-γ-secretion capture assay technique. Flow cytometric analysis of IFN-γ^{+} T cells following 6 h of HCMV_{pp65/IE1} peptide pool stimulation (upper panel), before isolation; lower panel, IFN-γ-secretion capture assay positive fraction → enrichment of specific T cells for CD4^{+} and CD8^{+} T cells.](http://www.jimmunol.org/DownloadedFrom)
FIGURE 2. Partial mTOR inhibition by VLD (2 nM) or LD (20 nM) rapamycin during expansion culture for 18 d increased the number of early memory cells and led to superior specific antiviral T cell response. Frequencies were assessed by flow cytometry. Respective inhibitor treatment is indicated. [(A) and (B); summarized in (C)] Rapamycin increases the proportion of CD4+ HCMV-specific T cells within bulk culture without affecting fold expansion, as shown in (A) (absolute cell counts ranged from 3.2 × 10^5 to 6.2 × 10^5 cells before expansion, from 2.6 × 10^7 to 8.3 × 10^7 cells [w/o], from 2 × 10^7 to 7.8 × 10^7 cells [VLD], and from 2 × 10^7 to 7.6 × 10^7 cells [LD] after expansion). [(D); summarized in (E) and (F)] Rapamycin preserves CCR7 and CD62L expression within CD45RA+ T cells [(D); upper panel, TCM = CCR7+/CD45RA−] and CD62L [(D); lower panel]. [(G)] Preferential expansion of TCM (refer to fold expansion of CD4+/CD8+ T cells). CD137 and CD154 activation marker expression [(H); summarized in (I)] and IFN-γ, TNF-α, and IL-2 cytokine secretion [(K); summarized in (J)] were determined by intracellular staining. Cells were stimulated for 18 h with HCMVpp65/IE1 peptide pool-pulsed LCL at a ratio of 10:1. Cells were permeabilized and stained for IFN-γ, TNF-α, IL-2, CD3, CD4, CD8, CD137, and CD154. Summary of six independent experiments showing enhanced polyfunctionality of both CD4+/CD8+ T cells [(J); IFN-γ+, TNF-α+, IL-2+] and activation marker expression of CD8+ T cells [(I): CD137+, CD154+] in response to HCMVpp65/IE1 peptide pool-pulsed LCL. Box plots with median and interquartile range. Mean levels are shown in (A), (C), (E)–(G), (I), and (J). Representative examples are shown in (B), (D), (H), and (K). *p < 0.05, Wilcoxon matched-pairs test.
viation marker expression following specific restimulation (Supplemental Fig. 2B, 2C). Rapamycin/IL-2Rx antagonist-incubated CD8+ T cell bulk cultures, as well as IL-2Rx pathway-targeted CD4+ T cells, showed increased CD62L expression (Fig. 4D). In contrast, CD62L expression on purified CD8+ T cells could not be enhanced. Hence, CD8+ T cell effector function and memory phenotype in Ag-specific T cell cultures can be improved in the presence of CD4+ T cells. Furthermore, CD4+ T cells, when cultured alone, elicited direct antiviral cytolytic function apart from helper mechanisms by means of cytokine secretion (Supplemental Fig. 2H, 2K).

In vitro generation of TCM is growth factor dependent

Our expansion protocol is based on standard IL-2/IL-7 growth factor supplementation. Because both approaches of partial IL-2Rx targeting interfere with cytokine/cytokine receptor interaction,
particularly of IL-2/IL-2R, we wondered whether removal of one or both growth factors would have similar effects on the T cell product composition. T cells expanded with IL-2 alone showed strong proliferation and good cytokine-secretion capacity; however, the majority of the T cells displayed a late differentiated TEM phenotype (Fig. 5). IL-7 alone showed remarkably reduced T cell proliferation but somewhat preserved CCR7/CD62L expression compared with supplementation with IL-2 or IL-2/IL-7. Addition of rapamycin/IL-2R\(\alpha\) antagonist with either IL-2 or IL-7 promotes TCM formation by maintaining cytotoxicity. In the presence of rapamycin/anti-CD25 mAb, IL-2 alone was less effective in preserving the TCM phenotype, but more potent in cell expansion and inducing effector functionality than treated with IL-7 alone (Fig. 5). Most importantly, the best results with regard to the expansion rate and preserving the TCM phenotype with full functionality were seen after cultivation with both IL-2 and IL-7 in the presence of rapamycin/IL-2R\(\alpha\) antagonist. Interestingly, rapamycin/IL-2R\(\alpha\) antagonist generated T cell lines showed prolonged (7 d) expression of IL-7R\(\alpha\) (CD127) compared with controls (Supplemental Fig. 2D, 2E).

**MODIFYING IL-2R SIGNALING ENABLES SUPERIOR PROTECTIVE CD4-MEDIATED CD8\(^+\) T CELL IMMUNITY**

In a prevailing HCMV infection, bone marrow-derived myeloid progenitors are latently HCMV infected (27). Following reactivation, HCMV early Ags can be presented by circulating monocytes and their derivatives. This can be simulated by infecting monocyte-derived immature dendritic cells (iDC) using HCMV wild-type strain NEWT (22). The lower epitope density on infected cells compared with peptide-loaded cells is a challenge requiring highly effective T cell recognition.

Compared with peptide pool-pulsed iDC, the frequency of cytokine-secreting and CD137-expressing conventionally expanded CD8\(^+\) T cells responding to NEWT-infected iDC was remarkably reduced (Fig. 6A–D), whereas specific lysis was unchanged (data not shown). However, CD8\(^+\) T cells derived from IL-2R\(\alpha\) pathway-targeted cultures revealed a much greater IFN-\(\gamma\) response to NEWT-infected iDC, whereas CD137 expression was only marginally upregulated (Fig. 6A–D). Interestingly, CD8\(^+\) T cells secreted substantial amounts of IFN-\(\gamma\), whereas TNF-\(\alpha\)/IFN-\(\gamma\) double positivity was reduced in response to NEWT-infected iDC (Fig. 6A, 6B). Expression of the degranulation marker CD107a at CD8\(^+\) T cells remained constant at a low level compared with peptide-loaded iDC. Yet again, CD8\(^+\) T cells tended to increase CD107a surface expression (Fig. 6E).

To test the impact of CD4\(^+\) T cell quantity on CD8\(^+\) function in this model, we isolated HCMV-specific T cells from PBMC and cultured bulk T cells, sorted CD4\(^+\), and CD8\(^+\) T cells in parallel. Following specific restimulation with NEWT-infected iDC, CD8\(^+\) T cells cultured in bulk showed marginally increased IFN-\(\gamma\) ef-
Adaptive T cell therapy is a promising new option for combating severe infections and tumors. The longevity of adoptively transferred T cells is one challenge for long-lasting success. Conventional in vitro expansion strategies of human Ag-reactive T cells preferentially support T EM/TEMRA cells with high effector potential in vitro expansion strategies of human Ag-reactive T cells; however, the expansion and effector functionality of resulting T cells is strongly reduced compared with expansion in the presence of IL-2, which results in a dominant T EM phenotype. However, in the presence of rapamycin/anti-CD25 mAb, the positive effects of IL-2 on expansion and functionality can be preserved without losing the supportive effects of IL-7 on T CM generation, indicating the usefulness of partial IL-2R signaling (via β/γ-chain). The narrow range of the optimal rapamycin dose supports this view.

Targeting the IL-2R signaling pathway during extensive cell expansion revealed virus-specific CD4+/CD8+ T cells with particular properties: a dominant T CM phenotype, powerful specific effector functions, and enhanced functionality (i.e., multicytokine secretors, including IL-2; enhanced CD137 and CD107a expression on CD8+ and CD4+ T cells, respectively; and effective killing of infected target cells). Gating on T CM and T EM revealed no differences in the cytokine pattern between untreated and IL-2R pathway-targeted cultures, speculating that improved T cell functionality correlates with the quantity of T CM in the T cell product. Remarkably, CD8+ T CM formation was strongly dependent on the increased proportion and functionality of CD4+ T cells within those T cell products. These data might explain why CD4 help is a key factor for establishing long-lasting CD8 memory; obviously, CD4+ T cells support the formation and expansion of long-lived CD8+ T cells by preventing the shift of all Ag-activated early memory CD8+ T cells into the terminally differentiated phenotype. Moreover, our studies show that there is differing sensitivity to mTOR inhibition in distinct T cell subsets, with the least dependency in nTreg, intermediate dependency in T CM, and the greatest dependency on mTOR activity in T EM/T EMRA.

Our data are in line with the findings of Araki et al. (14, 15), who reported that rapamycin increases the quantity and quality of memory T cell differentiation in mice. The positive effects of rapamycin were dissected in nonhuman primates following vaccinia virus vaccination (16). Rapamycin augmented the magnitude, duration, and quality of vaccine-specific T cell responses. However, rapamycin was applied prior to vaccination and was continuously administered for >90 d. Most studies analyzed the impact of rapamycin on the differentiation of memory T cells from naive T cells (14, 28). Rapamycin promotes the generation of memory precursor effector cells and accelerates the effector → memory T cell transition during the contraction phase (15). Rapamycin treatment during both the expansion and contraction phases improves the quality and quantity of memory T cells (15). However, the absolute number of memory T cells was not increased during the contraction phase. Our approach differs from those studies, because we boosted a pre-existing memory response by isolating and expanding virus-specific T cells from seropositive donors. IL-2R targeting in our approach improved the quality, as well as enhanced the growth factor-supported quantity, of early memory T cells.

Studies in mice using mTOR inhibitors described reduced effector cytokine secretion at early stages of differentiation (29, 30). A key finding of our study was that the generated HCMV-specific T CM are not diminished in effector functions compared with conventional T CM cultures, with a significantly higher proportion of the T EM/T EMRA counterpart. Moreover, our T cell products even showed enrichment of polyfunctionality. This contradicts another report, in which early differentiated T CM cultures were incapable of early effector function (31). This might be related to the supplementation by growth factors, particularly IL-2, in our system. Rapamycin mediates several effects, including inhibition of the T cell response to IL-2 (25). We showed in this study that reducing the binding capacity of IL-2 to the IL-2R complex by IL-2Ra an-
FIGURE 6. Modifying IL-2R signaling by targeting IL-2R binding or mTOR pathway enables superior protective CD4-mediated CD8⁺ T cell immunity. HCMV-specific T cell lines were generated, as described. Frequencies were assessed by flow cytometry. Inhibitor treatment is indicated. Assessment of IFN-γ and TNF-α cytokine secretion [(A); summarized in (B)], CD137 and CD154 activation marker expression [(C); summarized in (D)], and CD107a degranulation capacity (E) following specific stimulation with 1 μg/ml HCMVpp65/IE1 peptide pool-pulsed monocytes and NEWT-incubated monocytes for 18 h. (F) CD8⁺ T cell function can be induced by supplementing equal numbers of CD4⁺ T cells just before specific restimulation. Shown is IFN-γ secretion of CD8⁺ T cells within bulk culture or IFN-γ secretion of CD8⁺ T cells that were cultured alone. CD8⁺ T cells were cultured separately with rapamycin (LD) or aIL-2Rα, as indicated. (G) IL-2R pathway-targeted cultures exhibit enhanced T cell recognition. T cells were restimulated for 18 h with peptide-loaded autologous LCL at declining peptide concentrations of 1, 0.1, or 0.01 μg/ml pp65/IE-1 (crescendo bar denotes increasing peptide concentrations). Cells were permeabilized and stained with the respective Abs. In all plots, one representative of three experiments is shown.
tagonist gave similar results. The mode of action of our effects is mediated by modifying IL-2R signaling. Certainly, T_{CM} might be more resistant to survive IL-2 withdrawal. This is consistent with observations in murine models of viral infections, in which rapamycin-treated T cells showed an enhanced ability to survive growth factor withdrawal as the result of a hybrid metabolism induced by rapamycin, upregulating both oxidative phosphorylation and glycosylation (29). Thus, the generated T_{CM} have a greater ability to resist growth factor withdrawal, in contrast to the T_{CM} counterpart. Consequently, inadequate IL-2 signaling preferentially supports less-sensitive T_{CM} expansion.

At culture initiation, the targeted T cell lines showed prolonged IL-7Rα (CD127) expression, which is essential for memory T cell maintenance. However, after 18 d of expansion, a distinct population of CD127+ T cells could not be seen. This could be due to the supplementation of IL-7 during culture and, thereby, down-regulation of the receptor. Interestingly, a previous study showed that in vitro IL-2R antagonistic treatment augmented the T cell responsiveness to IL-7 caused by prolonged binding to the CD127/CD132 complex (32). These results indicate that IL-2R antagonist can favor response to IL-7; however, IL-7 alone was less sufficient than was the combination of IL-2/IL-7 in supporting expansion.

We demonstrated that CD8+ T cell effector function and memory phenotype in Ag-specific T cell cultures proportionally improve with the presence of CD4+ T cells. In agreement, Sun et al. (33) demonstrated that the generation of memory CD8+ T cells in response to an acute infection is independent of CD4 help; however, during memory maintenance, CD4+ T cells provide essential support for memory CD8+ T cell survival. In our studies, CD8+ T cells showed prolonged CD62L expression only when CD4+ T cells were present; this effect could be further augmented by the addition of rapamycin/IL-2R antagonist during culture. This exemplifies the importance of CD4+ T cells for memory CD8+ T cells during a rechallenge response.

Remarkably, we showed that HCMV-specific CD8+ T cells expanded with rapamycin/IL-2R antagonist responded similarly to NEWT-infected autologous monocytes/iDC. Moreover, CD8+ T cells only responded when CD4+ T cells were present. Hence, it is likely that virus-specific CD8 T cell responses are partially Ag specific and are caused through bystander activation (34). Consistent with other studies, our results demonstrated that CD4+ T cells elicited antiviral lytic T cell response apart from helper mechanisms (35) that is essential for maintaining long-term immune control (36, 37). Cytolytic CD4+ T cells are important in many chronic infections, such as EBV, HIV, and HCMV (38). In addition, the CD4+ T cells elicited a profound response to NEWT-infected iDC. Reasonably, HCMV-infected cells downregulate MHC class I expression and, thereby, decrease Ag presentation (39). Potentially, insufficient infection of iDC, resulting in an inadequate display of intracellular viral proteins through MHC class I, might lead to preferential processing of MHC class II.

In comparison with the unlimited MHC loading by incubation of target cells with HCMV-derived peptide pools, the living infection model requires an effective T cell recognition because of reduced specific epitope density on the target cells. By stimulating the differentially generated T cell lines with decreasing peptide concentrations, we were able to show that IL-2R pathway-targeted cultures exhibit enhanced T cell recognition. The advantages of T cells generated by targeting IL-2R signaling compared with control T cells in this clinically relevant target model further underscore their functional superiority.

In summary, this study provided evidence of the functional efficacy of human Ag-specific TCM. These findings identify a novel method of cell expansion that possesses important implications for T_{CM} generation and subsequent selection of adoptive immunotherapy for in vivo clinical use.

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Disclosures

The authors have no financial conflicts of interest.

References


SUPPLEMENTARY FIGURE 1.

(A) Experimental setup is illustrated in Fig. 1. Shown is functionality of CD62L+ CD4+ and CD62L+ CD8+ T cells. IFNγ and IL-2 cytokine secretion were determined by intracellular staining. Cells were stimulated for 18h with 1μg/mL HCMVpp65/IE1 peptide-pool pulsed LCL in ratio 10:1 as described at Fig. 1 K. Cells were permeabilized and stained for IFNγ, IL-2, CD3, CD4, CD8 and CD62L. In all plots, one representative of six experiments is shown. (B / C) Cytometric assessment of the degranulation capacity (CD107a) of CD4+ and CD8+ T cells following specific stimulation. CD107a expression at CD8+ T cells remained constant, CD4+ T cells tended to increase CD107a surface expression (B) Rapamycin and (C) IL-2-receptor antagonistic engagement. Experimental setup is illustrated in Fig. 2 and 3.

SUPPLEMENTARY FIGURE 2.

(A) Cytometric assessment of the degranulation capacity (CD107a) of CD4+ and CD8+ T cells following specific stimulation. Experimental setup is illustrated in Fig. 4A. (B / C) Effector cytokine secretion and activation marker expression of separately cultured CD4+ and CD8+ T cells. Experimental setup is illustrated in Fig. 4D. CD137 and CD154 activation marker expression (B) and IFNγ, TNFα and IL-2 cytokine secretion (C) were determined by intracellular staining. Cells were stimulated for 18h with 1μg/mL HCMVpp65/IE1 peptide-pool pulsed LCL in ratio 10:1. Cells were permeabilized and stained for IFNγ, TNFα, CD3, CD4, CD8, CD137 and CD154. (D / E) CD127 surface expression of CD4+/CD8+ T cells at day 7 of expansion. Experimental setup is illustrated in Fig. 2 and 3. (D) Rapamycin and (E) IL-2-receptor antagonistic engagement preserves CD127 surface expression of CD4+/CD8+ T cells; day 7 of expansion. One representative histoplot of three experiments is shown.