Early Responding Dendritic Cells Direct the Local NK Response To Control Herpes Simplex Virus 1 Infection within the Cornea


*J Immunol* published online 30 December 2011
http://www.jimmunol.org/content/early/2011/12/30/jimmunol.1101968

**Supplementary Material**
http://www.jimmunol.org/content/suppl/2011/12/30/jimmunol.1101968.DC1

**Subscription**
Information about subscribing to *The Journal of Immunology* is online at:
http://jimmunol.org/subscription

**Permissions**
Submit copyright permission requests at:
http://www.aai.org/About/Publications/JI/copyright.html

**Email Alerts**
Receive free email-alerts when new articles cite this article. Sign up at:
http://jimmunol.org/alerts

*The Journal of Immunology* is published twice each month by
The American Association of Immunologists, Inc.,
1451 Rockville Pike, Suite 650, Rockville, MD 20852
Copyright © 2011 by The American Association of Immunologists, Inc. All rights reserved.
Print ISSN: 0022-1767 Online ISSN: 1550-6606.
Early Responding Dendritic Cells Direct the Local NK Response To Control Herpes Simplex Virus 1 Infection within the Cornea


Dendritic cells (DCs) regulate both innate and adaptive immune responses. In this article, we exploit the unique avascularity of the cornea to examine a role for local or very early infiltrating DCs in regulating the migration of blood-derived innate immune cells toward HSV-1 lesions. A single systemic diphtheria toxin treatment 2 d before HSV-1 corneal infection transiently depleted CD11c+ DCs from both the cornea and lymphoid organs of CD11c-DTR bone marrow chimeric mice for up to 24 h postinfection. Transient DC depletion significantly delayed HSV-1 clearance from the cornea through 6 d postinfection. No further compromise of viral clearance was observed when DCs were continuously depleted throughout the first week of infection. DC depletion did not influence extravasation of NK cells, inflammatory monocytes, or neutrophils into the peripheral cornea, but it did significantly reduce migration of NK cells and inflammatory monocytes, but not neutrophils, toward the HSV-1 lesion in the central cornea. Depletion of NK cells resulted in similar loss of viral control to transient DC ablation. Our findings demonstrate that resident corneal DCs and/or those that infiltrate the cornea during the first 24 h after HSV-1 infection contribute to the migration of NK cells and inflammatory monocytes into the central cornea, and are consistent with a role for NK cells and possibly inflammatory monocytes, but not polymorphonuclear neutrophils, in clearing HSV-1 from the infected cornea.  The Journal of Immunology, 2012, 188: 000–000.

The body’s innate immune system is rapidly mobilized to combat infection by a variety of pathogens. Dendritic cells (DCs) are present in skin and mucosal surfaces, where they serve as sentinels that alert the body to the invasion of pathogens. They are widely recognized for their ability to bridge innate and adaptive immunity by sampling Ags during infection, transporting them to the lymphoid organs, and mobilizing the adaptive immune response (1, 2). In recent years, the phenotypic and functional diversity of the DC population has become increasingly apparent. Distinct DC subpopulations occupy different strata of mucosa and skin, and vary in their ability to migrate and cross-present Ags (3–5). DCs residing in lymphoid organs are also functionally and phenotypically diverse. Moreover, recent reports implicate DCs in the orchestration of innate immunity (6, 7). For example, depletion of DCs during HSV-1 infection of the footpad resulted in a diminished NK cell response within the spleen during the first 24 h postinfection (7, 8).

Previous studies have suggested that NK cells and polymorphonuclear neutrophils (PMNs) are primarily responsible for clearing HSV-1 from infected corneas (9–12). However, neutrophils were implicated in HSV-1 clearance in studies involving vivo depletion of neutrophils using the RB6-8C5 (anti-Gr-1) Ab, which was subsequently shown to deplete a variety of cell types in addition to neutrophils (13, 14). In fact, two recent studies demonstrated impaired HSV-1 clearance in mice that were depleted of Gr-1+ cells, but not those specifically depleted of neutrophils, with Ab to Ly6G, suggesting that neutrophils probably do not play a major role in HSV-1 clearance (14, 15).

NK cells provide a rapid response to viral infections through their ability to recognize generic changes that occur in virally infected cells, such as reduced expression of MHC (16, 17). NK cells can inhibit virus replication through generally lethal release of lytic granules or nonlethal release of antiviral cytokines, including IFN-γ (17). Because NK cells are potentially lethal and not activated by specific pathogen-derived Ags, their activity is closely regulated by the immune system. It is now apparent that NK cells require activation, or licensing, to gain full effector functionality (18). DC production of IL-12 and IL-15 has been implicated in NK cell activation (19, 20). In fact, IL-15 presentation by DCs appears to be required for both the activation and the homeostatic proliferation of NK cells (6, 21).

The interactions between DCs and NK cells have been largely studied in lymphoid organs. A separate role for DCs in regulating NK cell function locally within a viral lesion is more difficult to assess. DCs rapidly accumulate at the edges of HSV-1 and HSV-2...
lesions in skin, genital mucosa, and cornea (5, 22–24). Owing to the avascularity of the cornea, infiltrating cells must extravasate from blood vessels in the limbal region abutting the peripheral cornea, and then migrate into the central cornea, where HSV-1 lesions are established in the mouse model of HSV-1 keratitis (25, 26). This model affords a unique opportunity to separately observe tissue extravasation and in situ migration of innate immune responders to sites of viral infection within the central cornea. HSV-1 infection of the corneas of CD11c-DTR mice that express a high-affinity diphtheria toxin receptor from the CD11c-promoter (pCD11c) afforded the opportunity to observe the effect of DC ablation during the early stages of the innate response that clears the virus. We show that DCs are required during the first 24 h after HSV-1 corneal infection to orchestrate an innate immune response that clears virus from the cornea from 2 to 6 days postinfection (dpi). Depleting DCs during the first 24 h postinfection inhibits the migration of NK cells and inflammatory monocytes to the site of the lesion in the central cornea. This study reveals a previously unappreciated local role of dendritic cells in directing the innate immune response to viral infection.

**Materials and Methods**

**Reagents**

The following fluorochrome-labeled Abs were used: PerCP-conjugated anti-CD45 (30-F11), FITC-conjugated anti-CD69 (H1.2F3), PE-conjugated anti-CD3 (clone 17A-2), anti-CD49b (DX-5), and allophycocyanin-conjugated anti-granzyme B (GrB; GB11); anti-CD49b (DX-5) was purchased from BD Pharmingen (San Diego, CA). V450-conjugated anti-CD11b (M1/70), allophycocyanin-conjugated anti-CD11c (N418), allophycocyanin-eFluor-780-conjugated anti-GR-1 (RB6-8CA), and PE-Cy7–conjugated F4/80 (BM-8) were purchased from eBiosciences (San Diego, CA). The appropriate isotype control Abs were purchased from their respective vendors.

**Mice**

Female wild-type (WT) BALB/cJ and CD11c-DTR [C.FVB-Tg(Itgax-DTR/EGFP)57Lan/J] mice, 6–8 wk old, were purchased from The Jackson Laboratory (Bar Harbor, ME).

Bone marrow chimeras were created by transferring bone marrow from CD11c-DTR mice into lethally irradiated WT BALB/cJ mice to avoid lethality associated with multiple DT treatments of CD11c-DTR mice (27). Briefly, BALB/cJ hosts underwent two treatments of 500 rads in an animal γ-iradiator, and 2.5 × 10⁵ bone marrow cells from a CD11c-DTR donor were transferred i.v. The resulting mice (referred to herein as CD11c-DTR...
Chimeras were housed under immunocompromised mouse conditions and treated regularly with 2 mg/ml neomycin from Sigma-Aldrich (St. Louis, MO) in their drinking water. The CD11c-DTR chimeras were fully reconstituted and ready for experimental use after 6 wk. All experimental animal procedures were reviewed and approved by the University of Pittsburgh Institutional Animal Care and Use Committee and adhered to the ARVO Statement for the Use of Animals in Ophthalmic and Vision Research.

In vivo diphtheria toxin and anti–ASGM-1 treatment

Diphtheria toxin (DT) purchased from Sigma–Aldrich was prepared in a sterile solution of PBS at a concentration of 1 mg/ml. Transient DC depletion was effected in CD11c-DTR chimeras by a single i.p. injection of 175 ng DT; continuous DC depletion involved additional i.p. treatments of 100 ng DT per mouse every 3 d. NK cell depletion was accomplished by a single i.p. treatment with 40 \( \mu \)l rabbit anti–ASGM-1 Ab (Wako Pure Chemicals, Osaka, Japan) at 2 dpi.

Ocular HSV-1 infection

HSV-1 strain RE or HSV-1 KOS RFP-K26 was grown in Vero cells, and intact virions were isolated on Optiprep gradients according to the manufacturer’s instructions (Accurate Chemical and Scientific, Westbury, NY). For ocular HSV-1 infection, mice were anesthetized by i.p. injection of 2.0 mg ketamine hydrochloride and 0.04 mg xylazine (Butler Schein, Pittsburgh, PA) in 0.2 ml HBSS (Mediatech, Manassas, VA). The abraded central corneas of anesthetized mice were infected by topical application of 3 \( \mu \)l RPMI 1640 (Lonza, Walkersville, MD) containing 1 \( \times \) 10^5 PFUs of virus. All animal experiments were conducted in accordance with guidelines established by the University of Pittsburgh Institutional Animal Care and Use Committee.

Flow cytometry for phenotypic analysis

At indicated times, draining lymph nodes, spleens, and corneas were harvested from euthanized mice. Draining lymph nodes and spleens were minced and incubated with 500 \( \mu \)l DMEM (Lonza) containing 10% FCS (Atlanta Biologicals, Atlanta, GA) and 420 U/ml collagenase type I (Sigma-Aldrich) for 30 min at 37°C. Dissected corneas were treated with PBS with 2 mM EDTA for 15 min to separate the epithelium from the corneal stroma. Cells were then released from the corneal stroma by incubation for 1 h in 100 \( \mu \)l collagenase type I (Sigma-Aldrich) at a concentration of 840 U/ml, followed by trituration with a micropipette. In some experiments, the conjunctiva, peripheral cornea, and central cornea were examined individually for leukocytic infiltration. The cornea and conjunctiva were excised, the conjunctiva dissected from the cornea based on anatomical differences and the central cornea dissected from the peripheral cornea with a 2-mm trephine. Data were collected on a FACSVerse cytometer and analyzed by FACSVerb software (BD Biosciences, San Diego, CA).

Whole-mount fluorescent microscopy

Whole corneas were excised, flattened by making radial incisions, washed in PBS 4% FBS, fixed in 1% paraformaldehyde for 2 h, washed once more in PBS 4% FBS, and mounted. Images were acquired on an Olympus Fluoview 1000 confocal microscope with a 1.4 NA 60\( \times \) oil objective or a 0.85 NA

![FIGURE 2. DT treatment (Tx) ablates DCs from the lymphoid organs and the cornea. CD11c-DTR chimeras were treated i.p. with 175 ng of DT, or mock-treated with PBS. A. At 2 d postinjection, whole corneas were harvested, mounted, and analyzed via confocal microscopy for pCD11c-EGFP. Serial images of whole mounted corneas were taken starting at the corneal periphery and moving to the central cornea. Data are representative serial image sets of DT-treated or mock-treated corneas visualizing pCD11c-EGFP ablation. B. At 2 d postinjection, spleens and draining lymph nodes were harvested, dispersed, stained with anti-CD11c and -CD11b Abs, and analyzed via flow cytometry. Representative flow plots depicting CD11c+ cells within the cornea, draining lymph nodes, and spleen following DT and PBS treatment. n = 4–6 mice/group, two independent experiments.](http://www.jimmunol.org/Download/)
Detection of infectious virus from corneas

Mouse corneas were swabbed with sterile Weck-Cel surgical spears (Medtronic Solan, Jacksonville, FL) at 2, 4, 6, and 8 dpi; spears were then placed in 0.5 ml RPMI 1640 and frozen at −80˚C until assayed. Samples were added to confluent Vero cells, incubated for 1 h at 37˚C, and overlaid with 0.5% methylcellulose. The cultures were incubated for 72 h, fixed with formalin, stained with crystal violet, and a viral cytopathic effect was detected with the aid of a dissecting microscope.

Gene expression profiles in infected corneas, using GeneChip

Total RNA was isolated from whole corneas, using RNeasy and QIAshredder kits (Qiagen, Valencia, CA), and the cDNA was generated using Affymetrix 3’ IVT Express Kit (Affymetrix, Santa Clara, CA). The resulting cDNA was transcribed in vitro in the presence of biotin-labeled ribonucleotides and fragmented. Hybridization controls were added and hybridized overnight to an MG 430 2.0 GeneChip (Affymetrix). The chips were then washed, developed, and scanned in an Agilent Chip Scanner (Affymetrix). Raw data were processed and analyzed using Affymetrix GeneChip Operating Software (GCOS) v. 1.4 with default statistical settings. GCOS 1.4 was used to assess the presence or absence of the target sequence of each panel, as well as its expression level, and then to make all relevant pairwise statistical comparisons among samples. Processed data were sorted and inspected using Microsoft Excel. The unscaled mean values were 464.6 ± 36.7 (mean ± S.D., n = 6), and were scaled to a target value of 150, using the GCOS default method (2% trimmed mean). For the highly expressed housekeeping gene GAPDH, the coefficient of variation was 6.9% ± 1.3% (mean ± SD for three redundant panels per chip), whereas the housekeeping gene Pcx (pyruvate carboxylase), which has an expression level >20-fold less than that of GAPDH, showed a coefficient of variation of 26.7% ± 5.0% (mean ± SD for three redundant panels per chip that showed substantial presence of transcript). Using Pcx as an exemplar, a requirement for 2-fold change selects values >3.7 S.D. distant from the control mean, that is, at >99th percentile. This microarray data set has been deposited in the National Center for Biotechnology Information’s Gene Expression Omnibus and is accessible through GEO accession number GSE33991 (www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE33991).

Gene expression profiles in infected corneas using quantitative real-time PCR

Quantitative real-time PCR was performed using supplies from Applied Biosystems (Foster City, CA). Between 0.47 and 1.12 μg total RNA was reverse transcribed using a random primer protocol (High Capacity cDNA Kit; Applied Biosystems). The resultant cDNA was diluted to 2 ng/μl with nuclease-free water. Diluted samples were mixed with an equal volume of reaction medium of TaqMan Universal MasterMix (Roche, Indianapolis, IN). Primer probe sets used were Pcx (Mm00442834_m1 or Mm00500992_m1) as the normalizing housekeeping gene and Ifng.
(Mm00801778_m1) to measure the response of the tissue to viral infection. Both these kits target amplicons that span intron-exon boundaries and are essentially free of interference from genomic DNA. Measurements were also made of the viral gene gH, using a custom primer probe kit (26). Because the gH gene is intronless, in virus-infected cultured trigeminal cells ~25% of the Ct signal is due to viral DNA; however, the uncorrected signal is a good general indicator of infection (26).

**Statistical analysis**

All statistical analyses for nonmicroarray experiments were computed with GraphPad Prism software, using unpaired t tests. The p values < 0.05 were considered statistically significant.

**Results**

**DT treatment ablates DCs from both lymphoid tissue and the cornea**

CD11c-DTR chimeras (CD11c-DTR bone marrow into WT mice) were generated as described in Materials and Methods to avoid the lethality of DT treatment in CD11c-DTR mice (27). Both the cornea (Fig. 1A, 1B) and the lymphoid organs (data not shown) were fully reconstituted with DCs that expressed enhanced GFP (EGFP)+ from the pCD11c by 6 wk, which is similar to the kinetics of reconstitution observed previously (28). DT treatment (175 ng, administered i.p.) effectively depleted CD11c+ DCs from the corneas and lymph nodes of reconstituted CD11c-DTR chimeras for 3 d (Fig. 2A, 2B). These CD11c-DTR chimeras were used in all experiments described herein.

**DCs are required for optimal early clearance of HSV-1 from the cornea**

Viral titers drop precipitously from 2 to 4 dpi in BALB/c mice with HSV-1 corneal infections, suggesting deployment of the innate immune response during this period. Mice that were depleted of DCs through 6 dpi by DT treatment at −2, +1, and +4 dpi exhibited markedly impaired viral clearance, harboring a significantly elevated viral load at 4 and 6 dpi (Fig. 3A). A nearly identical increase in viral titers was observed when DCs were transiently depleted through 1 dpi by a single DT treatment at −2 dpi (Fig. 3A). Following this treatment, DCs were found within the viral lesions in the central cornea by 2 dpi as well as in the spleens at 3 dpi (Fig. 3B, 3C). Thus, DCs that are recruited to the viral lesion within the first 24 h after HSV-1 infection appear to play an important role in mobilizing the innate response that clears HSV-1.

**Figure 4**. DC depletion does not alter extravasation of leukocytes into the peripheral cornea. WT and CD11c-DTR mice were infected with HSV-1-RE. CD11c-DTR chimeras were treated with PBS or DT 2 d before infection. At 3 dpi corneas were excised, and the central 2 mm was dissected away from the remaining peripheral cornea with a trephine. Pools of three peripheral corneas were dispersed with collagenase, then stained with Abs to CD45, GL-3, CD49b, F4/80, and GR-1; next, infiltrating leukocytes were identified by flow cytometry. A, Representative histogram overlays depicting the lack of GL-3+ gd T cells within the peripheral cornea. Histograms: Isotype control is gray shaded, CD11c-DTR PBS treatment (Tx) is solid black, and −2 dpi DT Tx is black dotted. B, Representative flow plots gated on CD45+ cells (numbers are for three corneas). C, Bars represent the mean ± SEM absolute number of neutrophils (PMNs), inflammatory monocytes (IM), and NK cells per individual peripheral cornea. n = 4–6 pooled corneas, three independent experiments.
from the cornea. The virus replicates in the corneal epithelium, which in normal corneas is endowed with a population of DCs (4). The rapid involvement of DCs in viral clearance would suggest the participation of these corneal resident DCs.

**DC depletion does not alter extravasation of leukocytes into the infected cornea**

We proposed that DC depletion influenced viral clearance by controlling leukocytic infiltration of the cornea. Normal corneas are devoid of neutrophils, NK cells, Gr-1^int^ monocytes, and γδ T cells (not shown), but these cells reportedly infiltrate the cornea post-infection. At 3 dpi, we did not detect infiltrating γδ T cells (Fig. 4A) but did detect a robust infiltration of Gr-1^high^ F4/80^-^ neutrophils (PMNs), Gr-1^int^ F4/80^-^ inflammatory monocytes, and Gr1^-^ DX5^+^ NK cells (Fig. 4B). To determine if extravasation of these cells into the peripheral cornea is altered by DC depletion, CD11c-DTR chimeras received a single DT or PBS treatment 2 d before infection, infected corneas were excised at 3 dpi, and the central 2 mm of each cornea was separated from the remaining peripheral cornea, using a trephine. Individual peripheral corneas were dispersed with collagenase and analyzed by flow cytometry. As illustrated in Fig. 4C, DT treatment 2 d before infection did not significantly influence the migration of neutrophils, inflammatory monocytes, or NK cells into the peripheral cornea. The activation phenotype of NK cells in the peripheral cornea is not DC dependent

Recent findings suggest an important function for DCs in the migration of activated NK cells into sites of infection (6). Although DC depletion did not influence NK cell numbers in the peripheral cornea, impaired viral clearance could reflect DC regulation of their activation status. CD11c-DTR chimeras were treated with PBS or DT 2 d prior to HSV-1 corneal infection. At 3 dpi, the corneas were excised and the activation phenotype of the DX5^+^ NK cells in the corneal periphery was determined based on expression of CD69 and GrB. As illustrated in Fig. 5A and 5B, ~70% of the NK cells in the infected corneas were activated as indicated by CD69 and GrB expression, and their activation phenotype was not altered by DC depletion. However, as noted above, a single DT treatment at −2 dpi maintains DC depletion only through 1 dpi. It was possible, therefore, that NK cells acquired the activation phenotype during DC reconstitution of the cornea (1–3 dpi). To address this, CD11c-DTR mice received DT treatments at −2 and +1 dpi to maintain DC depletion through 3 dpi. However, even continuous DC depletion through 3 dpi did not influence the activation status of NK cells in the peripheral cornea (Fig. 5B).

**Fewer NK cells and inflammatory monocytes migrate into the central cornea following DC depletion**

In our corneal infection model, HSV-1 lesions are largely restricted to the central area of the cornea (Fig. 6A, 6B). Because the cornea is devoid of blood vessels, leukocytes must extravasate from blood vessels in the peripheral cornea and then migrate to the site of the lesion within the central cornea. We proposed, therefore, that the failure of DC-depleted corneas to control virus replication might reflect an inability of the extravasated leukocytes to traffic to the central cornea. To test this, we treated mice with PBS or DT 2 d before infection, and then quantified NK cells, inflammatory monocytes, and neutrophils in the central corneas at 3 dpi. DC depletion significantly reduced migration of NK cells and inflammatory monocytes into the central cornea, whereas neutrophil migration was not affected (Fig. 7).

**Depletion of NK cells results in increased viral burden within the cornea**

The above findings were consistent with the idea that DCs indirectly contributed to HSV-1 clearance by regulating NK cell migration to the site of the lesion in the central cornea. Therefore, we
predicted that NK cell depletion would have an impact on HSV-1 clearance from the cornea similar to what was seen with DC depletion. To test this prediction, WT mice were depleted of NK cells by treatment with anti–ASGM-1 Ab or mock-depleted with PBS 1 d before corneal infection, and viral clearance was monitored through 8 dpi. Depletion of NK cells delayed clearance of virus from the cornea in a pattern similar to that seen following DC depletion (Fig. 8).

**DC depletion alters gene expression in the infected cornea**

Gene array analyses following DC depletion revealed changes in gene expression that were consistent with the observed changes in NK cell, monocyte, and neutrophil migration to the central cornea. DC depletion from infected corneas resulted in significant (>2-fold) reductions in expression of several genes whose products are involved in attracting and activating NK cells and monocytes (Supplemental Table I). These include the chemokines CCL5 (activates NK cells), CCL7 (activates and activates macrophages), CCL8 (activates and activates monocytes and NK cells), CXCL9 (activates NK cells), CXCL10 (activates macrophages and NK cells), and CXCL11 (activates NK cells). In contrast, CXCL2, the chemokine primarily responsible for neutrophil migration into the cornea, was significantly increased (Fig. 8).
HSV-1–infected cornea, was upregulated in infected corneas but was not reduced in infected corneas of DC-depleted mice (29). We did not observe a consistent reduction of IFN-γ mRNA in DC-depleted corneas (data not shown). However, we did note a consistent decrease in mRNA for several IFN-γ-inducible genes (Supplemental Table I).

Discussion

It is now clear that DCs do not represent a homogeneous cell population, but rather are composed of phenotypically and functionally diverse cells. For instance, distinct populations of Langerin-positive DCs reside in the epidermal and dermal layers of the skin, with the DEC-205+ CD11b+ epidermal Langerhans cells implicated in inhibition of contact hypersensitivity reactions, and the CD103+CD11b+dermal DCs implicated in the stimulation of contact hypersensitivity responses (30, 31). The cornea of the eye represents a mucosal surface that enjoys a degree of immune privilege previously attributed in part to a lack of professional APCs. However, recent studies have demonstrated that the normal mouse cornea contains a small, but clearly detectable, population of CD11c+CD11b−DCs in the draining lymph nodes of HSV-1–infected CD11c-DTR mice (not shown). Moreover, we see very few, if any, CD8+ T cells in the infected corneas of BALB/c mice, and never observe them in the cornea as early as 4 dpi when viral titers are augmented by DT treatment of CD11c-DTR mice. These findings support the view that DT treatment augments viral titers indirectly by depleting the DCs that regulate NK cell infiltration of the cornea.

We considered that DCs could influence NK cell function by controlling their extravasation, directed migration, or activation within the HSV-1–infected cornea. A recent study demonstrated that DCs are required for NK cell activation and their subsequent migration into inflamed tissue (6). However, we observed normal numbers of NK cells in the peripheral corneas of DC-depleted mice, showing that DCs are not needed for the initial extravasation of NK cells into HSV-1–infected corneas. Thus, in this infection model, DC priming of NK cells in lymphoid organs appears unnecessary for their subsequent migration into HSV-1–infected corneas.

The potential lethality of NK cells dictates that their effector functions be closely regulated. Indeed, NK cell effector mechanisms are regulated by a balance of positive and negative signals that are delivered through surface receptors (38). Strong evidence also supports direct activation of NK cells through TLRs. For instance, viral-sensing TLR-2 and TLR-7 can directly activate NK cells, and TLR-2–mediated NK cell activation is critical for control of vaccinia virus in vivo (39, 40). Moreover, DCs have been assigned a critical role in the survival and function of NK cells (8, 19, 21, 41). Cross-presentation of IL-15 appears to be the primary mechanism of DC-mediated activation of NK cell function (13). Thus the relative contribution of direct versus DC-mediated activation to NK cell function at sites of infection remains unclear. We noted that NK cells that infiltrated the peripheral cornea at 3 dpi expressed an activated phenotype (∼70% CD69+ Grb3) regardless of DC depletion. These phenotypic findings suggest that any NK cell functional deficiencies resulting from inadequate DC priming in the lymphoid organs can be at least partially compensated for by stimulation within the infected tissue. Moreover, because the majority of NK cells in the corneas of DC-depleted mice express an activated phenotype, the delayed HSV-1 clearance in these corneas is probably not due to inadequate activation of NK cells.
In vascularized tissue it can be difficult to distinguish extravasation of leukocytes from the blood and their subsequent directed migration to the site of infection. However, following HSV-1 infection of the avascular cornea, leukocytes must extravasate from blood vessels in the limbal region adjacent to the peripheral cornea and then migrate through ~3 mm of avascular tissue to reach the lesion in the central cornea. Thus, one can readily differentiate an effect on extravasation of leukocytes into the peripheral cornea and their directed migration into the central cornea (26, 42). Although both DCs and NK cells are attracted to the site of HSV-1 lesions, it is not clear if they are attracted simultaneously by the same or concurrently produced chemokines, or if the two cell types are sequentially attracted. We show that when DCs are depleted from the cornea, NK cell migration from the peripheral to the central cornea is markedly diminished, and control of virus replication is delayed. Our observations establish an important role for DCs in regulating the directed migration of NK cells to sites of viral infection. Because HSV-1 replication in this model occurs primarily in the central region of the avascular cornea, a lack of directed NK cell migration might have a more dramatic effect on viral clearance in the cornea than would be observed in a more vascularized tissue. For instance, a recent report established that DC depletion influenced NK cell function in lymphoid organs but did not influence HSV-1 clearance from infected footpads of mice (8).

The migration of inflammatory monocytes (F4/80<sup>+</sup> Gr-1<sup>−</sup>) from the peripheral to the central region of infected corneas was also impaired by DC depletion. Inflammatory monocytes produce antiviral compounds, including TNF and NO, that contribute to the control of HSV-1 replication in the trigeminal ganglion (43). Therefore, the reduction in infiltration of inflammatory monocytes into the central cornea might also have contributed to the delayed clearance of HSV-1 from the cornea.

Impaired HSV-1 clearance following DC depletion was not associated with reduced neutrophil infiltration into the peripheral or central cornea. Although a previous study suggested an important role for neutrophils in controlling HSV-1 replication in the cornea, that study employed the Gr-1 Ab for neutrophil depletion, which is now known to deplete a variety of cell populations in addition to neutrophils (13). Although not ruling out a contribution of neutrophils to viral clearance from the cornea, our findings are consistent with recent reports demonstrating a lack of neutrophil involvement in HSV-1 clearance from the lungs and skin (14, 15).

The reduced migration of NK cells and inflammatory monocytes into the central cornea that was observed following DC depletion was associated with a significant reduction in several chemokines known to direct the migration of these cell types. As noted in Supplemental Table I, the chemokines CCL5, CCL7, CCL8, CXCL9, CXCL10, and CXCL11 were upregulated in infected corneas and reduced by >2-fold by DC depletion. It is not clear if these chemokines are produced directly by the DCs, or if DCs induce their production by other cells, such as monocytes. A number of these chemokines are induced by IFN-γ, and expression of several other IFN-γ-inducible genes was reduced in DC-depleted corneas. We saw reduced IFN-γ mRNA in some experiments, but this was not a consistent finding in DC-depleted corneas (data not shown). However, the mRNA was obtained from whole corneas, so it remains possible that local IFN-γ production at the site of the lesion was reduced by DC depletion. This possibility would be consistent with reduced NK cell migration to the lesion, and with the observation that NK cell depletion dramatically reduced IFN-γ mRNA in the infected corneas (Supplemental Fig. 1). DC depletion did not alter neutrophil migration into the central cornea. This observation is consistent with the finding that expression of MIP-2/CXCL2, which is primarily responsible for directing neutrophil migration into the HSV-1–infected cornea (29), was not affected by DC depletion. A previous study showed that early MIP-2 production in HSV-1–infected corneas is regulated by IL-17, detected in corneas within 24 h of infection (44). The source of IL-17 in infected corneas was not established, but our studies suggest that its production is DC independent.

Taken together, our data suggest that the corneal resident or an early responding blood-derived DC population reacts rapidly to HSV-1 infection by directly or indirectly inducing production of several chemokines that are needed to attract NK cells and inflammatory monocytes to the location of the viral lesion within the central cornea. Following a single DT treatment 2 d before HSV-1 corneal infection, DCs begin to recover by 1 dpi and are found in and around the HSV-1 lesion in the central cornea by 2 dpi. However, viral clearance remains impaired at 4 dpi and to a lesser extent at 6 dpi. The impaired viral clearance is associated with a nearly complete exclusion of NK cells and a significant reduction of inflammatory monocytes from the central cornea at 3 dpi. The absence of NK cells in the central cornea as a result of either DC depletion or direct NK cell depletion resulted in similar impairment of viral clearance. This finding strongly suggests a role for NK cells in controlling HSV-1 replication in the cornea. We believe the unique requirements of the avascular cornea for NK cell trafficking allowed us to uncover an important function of DCs in controlling NK cell migration following their extravasation from blood, a function that would be more difficult to detect in a vascularized tissue.

Acknowledgments
We thank Jessica Spehar for technical assistance, Nancy Zuroski for flow cytometry acquisition, and Kira Lathrop for assistance with confocal microscopy.

Disclosures
The authors have no financial conflicts of interest.

References


