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Multiple CD4\(^+\) T Cell Subsets Produce Immunomodulatory IL-10 During Respiratory Syncytial Virus Infection

Kayla A. Weiss,* Allison F. Christiaansen, † Ross B. Fulton, †,1 David K. Meyerholz, ‡ and Steven M. Varga*†,‡

The host immune response is believed to contribute to the severity of pulmonary disease induced by acute respiratory syncytial virus (RSV) infection. Because RSV-induced pulmonary disease is associated with immunopathology, we evaluated the role of IL-10 in modulating the RSV-specific immune response. We found that IL-10 protein levels in the lung were increased following acute RSV infection, with maximum production corresponding to the peak of the virus-specific T cell response. The majority of IL-10–producing cells in the lung during acute RSV infection were CD4\(^+\) T cells. The IL-10–producing CD4\(^+\) T cells included Foxp3\(^+\) regulatory T cells, Foxp3\(^-\) CD4\(^+\) T cells that coproduce IFN-\(\gamma\), and Foxp3\(^-\) CD4\(^+\) T cells that do not coproduce IFN-\(\gamma\). RSV infection of IL-10–deficient mice resulted in more severe disease, as measured by increased weight loss and airway resistance, as compared with control mice. We also observed an increase in the magnitude of the RSV-induced CD8\(^+\) and CD4\(^+\) T cell response that correlated with increased disease severity in the absence of IL-10 or following IL-10R blockade. Interestingly, IL-10R blockade during acute RSV infection altered CD4\(^+\) T cell subset distribution, resulting in a significant increase in IL-17A–producing CD4\(^+\) T cells and a concomitant decrease in Foxp3\(^+\) regulatory T cells. These results demonstrate that IL-10 plays a critical role in modulating the adaptive immune response to RSV by limiting T-cell–mediated pulmonary inflammation and injury. The Journal of Immunology, 2011, 187: 000–000.

The lung environment, with its constant exposure to inert foreign Ags, must preserve a fine balance between tolerance and inflammation to effectively clear respiratory pathogens while, at the same time, maintaining tissue homeostasis. Improper regulation of a pathogen-induced immune response can result in excessive immune-mediated injury to the lung tissue, termed immunopathology. Respiratory syncytial virus (RSV) is a leading cause of lower respiratory tract infections in young children, the elderly, and immunocompromised individuals (1–6). The host immune response is thought to contribute to the severity of RSV-induced respiratory disease (7). In addition to RSV, other respiratory viral pathogens, such as pandemic influenza strains and human coronaviruses that cause severe acute respiratory syndrome (SARS), are believed to induce a poorly regulated immune response resulting in immune-mediated disease (8, 9).

The lung environment contains a number of mechanisms that regulate the immune response (10), including the production of immunosuppressive cytokines such as IL-10. Both innate and adaptive immune cells have the capacity to produce IL-10, including mast cells, NK cells, neutrophils, eosinophils, macrophages, dendritic cells (DCs), CD4\(^+\) T cells, CD8\(^+\) T cells, and B cells (11). In addition, almost all immune cells express the IL-10 receptor (IL-10R), and are thus capable of responding to IL-10. Due to the broad expression of IL-10 and the potential responsiveness of most immune cells, it is clear that IL-10 can influence both innate and adaptive immunity.

IL-10 mediates pleiotropic effects on the immune response, including suppressing the activation and/or effector functions of immune cells. In macrophages, IL-10–induced signaling results in reduced production of proinflammatory cytokines and chemokines that are required for the recruitment of T cells, monocytes, DCs, and neutrophils to the site of infection (15). In addition, IL-10 suppresses the surface expression of MHC class II and costimulatory molecules on macrophages and immature DCs, resulting in the decreased activation and proliferation of T cells (16–18). IL-10 can also directly inhibit T cell cytokine production (e.g., IL-2, IL-5, and TNF) and induce T cell anergy (19–21).

The critical role of IL-10 during persistent infections has been well established (15, 22); however, its capacity to modulate the immune response during an acute viral infection resulting in immunopathology remains unknown. In the murine RSV infection model, it is evident that RSV-induced disease is immune mediated because depletion of T cells ameliorates disease even in the presence of prolonged viral replication (23, 24). This makes RSV an ideal model for studying dysregulation of an immune response in the lung mucosal environment. To dissect the role of IL-10...
during a respiratory viral infection resulting in immunopathology, we first identified the IL-10–producing cells during acute RSV infection and examined the effects of IL-10 deficiency on the adaptive immune response. We demonstrate that CD4+ T cells account for the majority of IL-10 production during the peak of the virus-specific T cell response. Furthermore, we show that multiple CD4+ T cell subsets produce IL-10. In the absence of IL-10 or following IL-10R blockade, we observed a significant increase in RSV-induced disease severity correlating with the kinetics of the RSV-specific adaptive immune response. Finally, we show that IL-10R blockade results in an increase in the number of Th1 and Th17 cells with a concomitant decrease in the number of T regulatory cells (Tregs). Taken together, our data suggest that IL-10 plays a critical role in modulating the adaptive immune response to acute RSV infection and in preventing immune-mediated pulmonary injury.

Materials and Methods

Mice, infection, and Ab treatment

Female BALB/cAnNCr mice 6–8 wk old were purchased from the National Cancer Institute (Frederick, MD). BALB/c IL-10–deficient mice (IL-10 knockout [KO]) were obtained from Dr. Daniel Berg (University of Iowa, Iowa City, IA) (25, 26). VERT-x mice, expressing an IL-10–eGFP reporter, were obtained from Dr. Christopher Karp (Cincinnati Children’s Hospital Research Foundation and University of Cincinnati College of Medicine, Cincinnati, OH) (27). The A2 strain of RSV was a gift from Dr. Barney Graham (National Institutes of Health, Bethesda, MD) and was propagated on HEp-2 cells (American Type Culture Collections, Manassas, VA). For infections, mice were anesthetized with isoflurane and infected intranasally with 1–3 × 10^6 PFU of RSV. In some experiments, mice were administered either 0.5 mg rat IgG (MP Biomedicals, Aurora, OH) or 0.5 mg anti–IL-10R mAb (clone 1B1.3) via i.p. injections starting 1 d prior to infection and every other day thereafter. CD4+ and/or CD8+ T cells were depleted via i.p. injection of 0.25 mg of anti-CD4 (clone 18H10.1) (all mAbs obtained from eBioscience)—for 30 min at 4°C. Cells were washed twice with FACS buffer containing saponin, once with FACS buffer, and resuspended in FACS buffer prior to analysis on a BD FACSCanto.

Disease measurements

Disease was assessed daily by monitoring weight loss as well as pulmonary function using an unrestrained whole-body plethysmograph (Buxco Electronics, Wilmington, NC), as previously described (28, 29). Pressure and volume differences in the chamber caused by respiration were monitored for 5 min and recorded to obtain a daily average to calculate the dimensionless parameter-enhanced pause (Penh), as previously described (30). Penh positively correlates to airway hyperreactivity (30), which is narrowing of the small airways in response to stimuli.

Tissue processing

Lungs and bronchoalveolar lavage (BAL) fluid from mice were harvested as previously described (31). Lung tissue and lymph nodes were digested in 4 ml or 1 ml, respectively, of HBSS with CaCl2 and MgCl2 (Life Technologies, Grand Island, NY) supplemented with 60 U/ml DNase I (Sigma-Aldrich, St. Louis, MO) and 125 U/ml collagenase (Sigma-Aldrich) and 1250 pg/ml IFN-γ (clone XM1G1.2), TNF-α (clone MP6-XT22), IL-17A (clone TC11-18H10.1) (all mAbs obtained from ebioScience)—for 30 min at 4°C. Cells were washed twice with FACS buffer containing saponin, once with FACS buffer, and resuspended in FACS buffer prior to analysis on a BD FACSCanto.

Ex vivo cell stimulations

Cells were incubated at 1–2 × 10^6 cells/well in a 96-well round-bottom plate (Corning). For peptide stimulations, cells were incubated with or without 1 μM M2a2.90 peptide (Biosynthesis, Lewisville, TX) and 10 μg/ml brefeldin A (Sigma-Aldrich) in RPMI 1640 supplemented with 10% FCS (Atlanta Biologicals, Lawrenceville, GA), 5 mM 2-ME (Sigma), 2 mM L-glutamine, 10 U/ml penicillin, 10 μg/ml streptomycin sulfate, 10 mM HEPES, 1 mM sodium pyruvate, and 0.1 mM MEM nonessential amino acids (all obtained from Life Technologies) for 5 h at 37°C (31). Alternatively, cells were stimulated in the presence or absence of 50 ng/ml PMA (Sigma-Aldrich), and 500 ng/ml ionomycin (Sigma-Aldrich) in the presence of 10 μg/ml brefeldin A in 10% FCS complemented RPMI 1640 for 5 h at 37°C.

Intracellular cytokine staining

Briefly after ex vivo stimulation, cells were stained for extracellular proteins and fixed in FACS lysing solution as described above. Cells were subsequently incubated in FACS buffer containing 0.5% saponin (Sigma-Aldrich) and intracellular cytokine Abs—IL-10 (clone JES5-16E3), IFN-γ (clone XM1G1.2), TNF-α (clone MP6-XT22), IL-17A (clone TC11-18H10.1) (all mAbs obtained from ebioScience)—for 30 min at 4°C. Cells were washed twice with FACS buffer containing saponin, once with FACS buffer, and resuspended in FACS buffer prior to analysis on a BD FACSCanto.

Plaque assays

Lungs were harvested on days 4 and 7 postinfection (p.i.) and processed for plaque assays as previously described (32).

ELISAs

Lungs from RSV-infected mice were disrupted using a tissue homogenizer (Ultra-Turrax T25; IKA Works, Wilmington, NC) in RPMI 1640 supplemented with 10% FCS and a 1/20 dilution of a protease inhibitor mixture (Sigma). Lung homogenates were centrifuged at 2000 rpm for 10 min. Cytokine protein levels in the supernatant were determined for IL-10 and IFN-γ by ELISA, as previously described (33) (IL-10 and IFN-γ Abs were obtained from ebioScience). Detection limits for cytokine protein levels were 62.5 pg/ml for IL-10 and 1250 pg/ml for IFN-γ.

Multiplex bead assays

Lungs from RSV-infected IL-10 KO and wild-type mice were harvested and supernatants were collected as described above. A Milliplex kit was used to determine the protein levels of 27 different cytokines and chemokines by following the manufacturer’s instructions (Millipore, Billerica, MA). The assay was run on a BioPlex instrument (Bio-Rad, Hercules, CA).

Histological examination

Whole lungs from naive and RSV-infected rfgG and anti–IL-10R mAb–treated mice on day 6 p.i. were fixed in 10% formalin (Thermo Fisher Scientific, Waltham, MA). Fixed lungs were embedded in paraffin, sectioned at 4-μm thickness, and stained by the University of Iowa Comparative Pathology Laboratory. Slides were stained with either H&E or periodic acid-Schiff (PAS) and randomized and blinded for analysis by a board-certified veterinary pathologist (Dr. D. Meyerholz, University of Iowa). Sectioned slides were scored for perivascular aggregates of leucocytes (PV; 1, within normal parameters; 2, small numbers of solitary cells with uncommon aggregates; 3, multifocal small to moderate aggregates; and 4, moderate to high cellularity with multifocal large cellular aggregates that may be expansive into adjacent tissues), interstitial disease (ID; 1, within normal parameters; 2, mild, detectable focal to multifocal
congestion, with uncommon to small numbers of leukocytes and some atelectasis; 3, moderate, multifocal to coalescing congestion, leukocyte cellularity, and atelectasis, with rare luminal leakage of cellular and fluid debris; and 4, severe, coalescing interstitial congestion, leukocytes, and atelectasis, with admixed extensive loss of airspace and luminal accumulation of cellular and fluid debris), edema (1, none; 2, rare, with alveoli with wispy to partial pools of eosinophilic seroproteinaceous fluid; 3, focal microscopic fields with contiguous alveoli partially filled by pools of fluid; and 4, multiple fields having coalescing alveoli filled by pools of fluid), and mucus (1, none; 2, epithelial mucinous hyperplasia with none to rare luminal mucus; 3, epithelial mucinous hyperplasia with luminal mucus accumulation in airways; and 4, severe mucinous change with some airways completely obstructed by mucus). The total score is the sum of the PVA, ID, edema, and mucus scores.

Statistical analysis

Data were graphically compiled using Prism software (Graphpad Software, San Diego, CA), with bars representing the SEM. Statistical analysis was completed using InStat software (Graphpad Software); unpaired, two-tailed Student t tests and ANOVAs with a Dunnett’s post test were calculated to determine statistical significance at a level of $\alpha = 0.05$.

Results

CD4+ T cells account for the majority of IL-10–producing cells during the peak of the virus-specific T cell response

To determine the role of IL-10 during acute RSV infection, we first examined the kinetics and cellular source of IL-10 production. Cells from the lung, medLNs, and lung airways (i.e., BAL) were stimulated in vitro with PMA and ionomycin and stained for intracellular cytokine staining for IL-10. The total number of IL-10+ cells (A), IL-10+ CD4+ T cells (B), and IL-10+ CD8+ T cells (C) in the lung, medLNs, and BAL. To determine statistical significance, an ANOVA was performed with a Dunnett’s post test making comparisons between each time point (days 4, 6, 8, and 10) and baseline (day 0). Cumulative data from two independent experiments are shown ($n = 8$). $^* p < 0.05$, $^** p < 0.01$.

FIGURE 1. The kinetics of IL-10–producing cells during acute RSV infection. At the indicated time points, cells from the lung, medLNs, and BAL of RSV-infected mice were isolated and stimulated with PMA and ionomycin, followed by intracellular cytokine staining for IL-10. The total number of IL-10+ cells (A), IL-10+ CD4+ T cells (B), and IL-10+ CD8+ T cells (C) in the lung, medLNs, and BAL. To determine statistical significance, an ANOVA was performed with a Dunnett’s post test making comparisons between each time point (days 4, 6, 8, and 10) and baseline (day 0). Cumulative data from two independent experiments are shown ($n = 8$). $^* p < 0.05$, $^** p < 0.01$.

FIGURE 2. CD4+ T cells are the primary in vivo source of IL-10 during acute RSV infection. Wild-type mice were treated with depleting anti-CD4 and/or anti-CD8 mAbs or control rat IgG and subsequently infected with RSV. On day 6 p.i., lungs were analyzed for IL-10 (A) and IFN-γ protein (B) via ELISA. Cumulative data from two independent experiments are shown ($n = 8$). $^* p < 0.05$, $^** p < 0.01$, $^*** p < 0.001$. n.d., none detected.

The Journal of Immunology 3

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IL-10–eGFP* cells on days 6 and 8 p.i. were T cells, as identified by Thy1.2 staining in the lung and BAL (Supplemental Fig. 1). Within the Thy1.2+ population, CD4+ T cells represented the majority of IL-10–eGFP+ T cells in lungs and BAL at all time points examined. Taken together, these data demonstrate that CD4+ T cells account for the majority of IL-10–producing cells during the peak of the virus-specific T cell response.

To confirm that CD4+ T cells produce the majority of IL-10 in vivo during RSV infection, we quantified IL-10 protein levels in the lungs of mice depleted of CD4+ and/or CD8+ T cells. Our results indicate that CD4+ T cells were responsible for the majority of IL-10 protein produced in the lung during an acute RSV infection (Fig. 2A). Elimination of CD8+ T cells resulted in increased IL-10 production, suggesting that CD8+ T cells may suppress IL-10 production by CD4+ T cells. IFN-γ production was also measured using the above approach, and, as expected, CD8+ T cells produced significantly more IFN-γ than did the CD4+ T cells in response to RSV infection (Fig. 2B). Taken together, these data demonstrate that CD4+ T cells are the major producers of IL-10 in vivo during an acute RSV infection. The capacity of CD4+ T cells to produce IL-10 suggests their potential to significantly modulate the immune response via anti-inflammatory cytokine production.

**Multiple CD4+ T cell subsets produce IL-10 in response to RSV infection**

Because CD4+ T cells accounted for the majority of RSV-induced IL-10–producing cells, we next determined the CD4+ T cell subsets responsible for the IL-10 production. On day 6 p.i., lung, medLN, and BAL cells were stimulated in vitro with PMA and ionomycin and stained for intracellular IL-10, IFN-γ, and Foxp3. In all tissues analyzed, we observed equal proportions of IL-10+ CD4+ T cells that produced only IL-10, coproduced IFN-γ (i.e., Th1 cells), or coexpressed Foxp3 (i.e., Tregs) (Fig. 3). Interestingly, in the BAL we observed a small population of Foxp3+ Tregs that coproduced IL-10 and IFN-γ. These data demonstrate that multiple CD4+ T cell subsets contribute to IL-10 production during an acute RSV infection.

In addition, we determined the frequency of IL-10–producing cells within these CD4+ T cell subsets (Table I). On day 6 p.i., we observed the highest frequency of IL-10–only producing conventional and regulatory CD4+ T cells (5.1 ± 1.6 and 36.7 ± 6.0%, respectively) in the BAL. Furthermore, in the BAL we observed the highest proportion of conventional and regulatory CD4+ T cells that coproduce IL-10 and IFN-γ (11.1 ± 4.4 and 8.8 ± 4.1%, respectively) when compared with other tissues. In all tissues analyzed, IL-10–producing conventional and regulatory CD4+ T cells were observed as summarized in Table I. These data demonstrate the distribution of IL-10–producing cells in the conventional and regulatory CD4+ T cell compartments following acute RSV infection.

**Increased RSV disease severity in the absence of IL-10 or following IL-10R blockade**

To examine the role of IL-10 during acute RSV infection, we used either IL-10–deficient (IL-10 KO) mice or anti–IL-10R mAb to determine disease severity in the absence of IL-10 or following IL-10R blockade. IL-10 KO mice, compared with control mice, exhibited greater weight loss and increased Penh following acute RSV infection (Fig. 4A). Similar results were obtained from mice treated with anti–IL-10R mAb prior to and during RSV infection, compared with IgG-treated controls (Fig. 4B). These data indicate that IL-10 plays a critical role in limiting disease severity at the peak of the adaptive immune response during RSV infection.

To determine if IL-10 had an effect on viral titers, we performed plaque assays using whole lung homogenates to measure viral load at the peak of RSV replication (i.e., day 4 p.i.) and following viral clearance (i.e., day 7 p.i.). No significant differences were noted in viral titer at either time point in IL-10 KO mice (Fig. 5A) or anti–IL-10R mAb-treated mice (Fig. 5B). These results show that the effect of IL-10 deficiency on disease severity is not due to either

![Table I. Frequencies of cytokine-producing CD4+ T cells on day 6 following RSV infection](http://www.jimmunol.org/)

The table shows the frequencies of cytokine-producing CD4+ T cells on day 6 following RSV infection.

<table>
<thead>
<tr>
<th>CD4+ T cell</th>
<th>IFN-γ+ IL-10−</th>
<th>IFN-γ+ IL-10+</th>
<th>IL-10+ IFN-γ−</th>
<th>IL-10− IFN-γ−</th>
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</thead>
<tbody>
<tr>
<td>Conventional (Foxp3−)</td>
<td></td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>Lung</td>
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<td>2.8 ± 0.9</td>
<td>2.5 ± 0.4</td>
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<tr>
<td>MedLN</td>
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<td>0.6 ± 0.2</td>
<td>0.5 ± 0.2</td>
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<tr>
<td>BAL</td>
<td>26.9 ± 6.2</td>
<td>11.1 ± 4.4</td>
<td>5.1 ± 1.6</td>
<td></td>
</tr>
<tr>
<td>Regulatory (Foxp3+)</td>
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<td></td>
<td></td>
</tr>
<tr>
<td>Lung</td>
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<td>2.0 ± 1.7</td>
<td>15.9 ± 1.4</td>
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</tr>
<tr>
<td>MedLN</td>
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<td>0.6 ± 0.2</td>
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</tr>
<tr>
<td>BAL</td>
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<td>8.8 ± 4.1</td>
<td>36.7 ± 6.0</td>
<td></td>
</tr>
</tbody>
</table>

*Percent ± SD

### References

1. Figure 3: Multiple CD4+ T cell subsets produce IL-10 during acute RSV infection. At day 6 p.i., cells from the lung, medLNs, and BAL were stimulated with PMA and ionomycin. Cells were subsequently stained for CD4, Thy1.2, IL-10, Foxp3, and IFN-γ. Gated IL-10+ CD4+ T cells depicting isotype control staining (A) or Foxp3 expression (x-axis) and IFN-γ production (y-axis) (B) are shown. Plots are concatenated using Flowjo software from one of two independent experiments displaying equal representation from four individual mice (n = 4).

2. Figure 4: IL-10 influences RSV-induced disease severity. Weight loss depicted as the percent of initial weight and airway function (Penh) were assessed daily following RSV infection of IL-10–deficient (A) or anti–IL-10R mAb-treated mice (B). Cumulative data from two independent experiments are shown (n = 8). *p < 0.05, **p < 0.01, ***p < 0.001.
altered viral replication or delayed viral clearance but instead may be the result of alterations of the RSV-specific immune response. Increased inflammatory environment in the absence of IL-10

Given the increased disease severity observed during RSV infection in the absence of IL-10, we next examined the overall inflammatory environment in the lung following acute RSV infection. Using a multiplex bead assay, we quantified cytokine and chemokine protein levels from whole-lung homogenates at days 0, 4, 6, 8, and 10 p.i. from control and IL-10 KO mice infected with RSV. In the absence of IL-10, we observed a marked increase in the levels of proinflammatory cytokines and chemokines following RSV infection (Fig. 6). In the lungs of RSV-infected IL-10 KO mice, we observed a significant increase in IL-6 (p < 0.01) and TNF-α (p < 0.001) on day 6 p.i., and a significant increase in IFN-γ on days 6 (p < 0.001) and 8 p.i. (p < 0.05) as compared with control mice. In addition, CXCL1 was significantly increased in the lungs of IL-10 KO mice at days 4 (p < 0.05) and 6 p.i. (p < 0.01) and slightly increased on day 8 p.i., when compared with control mice. CXCL9 was increased and CCL2 was significantly increased (p < 0.001) on day 6 p.i. in the absence of IL-10 following RSV infection. Overall, in the absence of IL-10, we observed increased levels of proinflammatory cytokines and chemokines that can enhance inflammation and the infiltration of immune cells responsible for mediating immunopathology. Taken together, these data support the hypothesis that IL-10 plays a critical role in regulating the severity of RSV-induced pulmonary disease by regulating inflammation and limiting immunopathology.

IL-10R blockade results in an increase in the magnitude of the RSV-specific CD8⁺ T cell response

Because the increase in RSV-induced disease severity (days 6–8 p.i.) in the absence of IL-10 or following IL-10R blockade occurred during the rise of the RSV-specific adaptive immune response, we next investigated the impact of IL-10 deficiency on the magnitude of the RSV-specific CD8⁺ T cell response. In mice treated with anti–IL-10R mAb, the total number of CD8⁺ T cells significantly increased (p < 0.01) in the medLNs on day 6 p.i. (Fig. 7A). Furthermore, the total number of CD11a⁺CD8⁺ T cells significantly increased (p < 0.001) in the medLNs of mice treated with anti–IL-10R mAb on day 6 p.i. (Fig. 7B), indicating the presence of Ag-experienced cells (34).

Using the M282–90 peptide to stimulate RSV-specific CD8⁺ T cells ex vivo, we observed a significant increase in the magnitude of the M282–90 RSV-specific CD8⁺ T cell response in the medLNs of mice treated with anti–IL-10R mAb on day 6 p.i. (p < 0.001) and in the lung on day 8 p.i. (p < 0.01) (Fig. 7C, 7D). In the medLNs of mice treated with anti–IL-10R, we observed a significant increase in the total number of M282–90-specific CD8⁺ T cells producing IFN-γ (p < 0.01) or coproducing IFN-γ and TNF-α (p < 0.0001) on day 6 p.i. In the lungs of mice treated with anti–IL-10R mAb, there was a significant increase in the total number of M282–90-specific CD8⁺ T cells that produce IFN-γ (p < 0.001) or coproduce IFN-γ and TNF-α (p < 0.01) on day 8 p.i. These data indicate an increase in both the magnitude and the effector function of RSV-specific T cells.

The Journal of Immunology
CD8+ T cells following IL-10R blockade, demonstrating a role for IL-10 in limiting the RSV-specific CD8+ T cell response.

**IL-10R blockade results in an increased Th1 CD4+ T cell response following acute RSV infection**

Because the RSV-specific CD8+ T cell response was increased following IL-10R blockade, we next evaluated the impact of IL-10 on the magnitude of the effector CD4+ T cell response during acute RSV infection. IL-10R blockade resulted in a significant increase in the total number of CD4+ T cells in the lung (p < 0.01) and BAL (p < 0.05) on day 8 p.i. (Fig. 8A). Intracellular cytokine staining for IFN-γ and TNF-α production by CD4+ T cells from the lung and mediLNs stimulated with PMA and ionomycin revealed a significant increase (p < 0.01) in the total number of effector CD4+ T cells in the lung on day 8 p.i. in mice administered anti–IL-10R mAb (Fig. 8B). Furthermore, on day 8 p.i. in the lung, mice treated with anti–IL-10R mAb exhibited a significant increase in the number of CD4+ T cells that produce IFN-γ (p < 0.001) or coproduce IFN-γ and TNF-α (p < 0.05). Thus, similar to the increase in the number of CD8+ T cells described above, we observed an increase in the magnitude of the Th1 CD4+ T cell response in the lungs of mice administered anti–IL-10R mAb during acute RSV infection. This increase in the number of virus-specific IFN-γ–producing CD8+ and CD4+ T cells following IL-10R blockade during a RSV infection suggests a critical role for IL-10 in limiting the magnitude of the RSV-specific adaptive immune response.

**Increased lung pathology following IL-10R blockade**

To determine if the increased disease severity and effector CD8+ and CD4+ T cell response result in increased lung pathology following IL-10R blockade, histological analysis of sections from whole lungs were stained with either H&E or PAS and analyzed. On day 6 p.i., a significant increase (p < 0.001) was observed in lung pathology when scoring the PVA, ID, eosinophilic seroproteinaceous fluid in the airspaces (edema), and luminal mucus accumulation in the airways (Fig. 9A). The histology of rIgG-treated mice following RSV infection revealed tissue inflammation with minimal mucus in the luminal space of the lung airways (Fig. 9B). The histological sections from mice treated with anti–IL-10R mAb demonstrate the increased pathology with increased areas of edema, tissue inflammation, and increased mucus in the luminal space of the lung airways (Fig 9C). Together these data indicate that IL-10R blockade results in increased lung pathology that is...
With the increased Th1 CD4+ T cell response during RSV blockade, we identified natural Tregs by their expression of the transcription factor Helios (35, 36). On day 8 p.i., a significant decrease occurs in the total number of Foxp3+Helios+ CD4+ T cells. These data suggest that IL-10, directly or indirectly, supports the maintenance or induction of natural Tregs during an acute RSV infection.

Coincident with the decrease in the total number of natural Tregs, we observed a significant increase in the total number of IL-17A+CD4+ T cells in mice treated with anti–IL-10R mAb (Fig. 10B). On day 8 p.i., IL-17A+ CD4+ T cells are significantly increased following IL-10R blockade in the lung (p < 0.01) and BAL (p < 0.05) during acute RSV infection. Similar results were obtained using IL-10 KO mice (data not shown). We did not observe IL-17A+ producing CD4+ T cells in the medLN or IL-17F+producing cells in the lung, medLNs, and BAL of experimental or control mice (data not shown). Thus, IL-10R blockade leads to decreased numbers of Tregs in the lung with a concomitant increase in the number of Th17 cells.

### Discussion

In this article, we examined the kinetics of IL-10 production during acute RSV infection and found that the highest frequency of IL-10–producing cells occurred between days 6 through 10 p.i., a time frame that overlaps with the virus-specific adaptive immune response (Fig. 1; Supplemental Fig. 1) (37, 38). We found that three major subsets of CD4+ T cells were responsible for the majority of IL-10 production at the peak of the immune response following RSV infection (Fig. 3). Approximately one-third of the IL-10–producing CD4+ T cells expressed Foxp3, representing a portion of the Treg population, whereas the remainder were conventional Foxp3+CD4+ T cells that produced either IL-10 alone or coproduced IFN-γ. In addition, we observed a small frequency of IL-10+IFN-γ regulatory (Foxp3+) CD4+ T cells only in the lung airways following RSV infection. We hypothesize that the environment in the lung airways is unique, causing these CD4+ T cells to alter their phenotype. We are currently investigating the plasticity of these IL-10+IFN-γ regulatory (Foxp3+) CD4+ T cells and their role during acute RSV infection.

Th1 cells have been previously reported to be the major producers of IL-10 in other pathogenic models, including influenza A virus (IAV), Leishmania species, mycobacterial, and Borrelia burgdorferi infections (39–43). The observation that other CD4+ T cell subsets are capable of producing IL-10, and that IL-10 can regulate T cell function, has led to the hypothesis that IL-10 acts in either an autocrine manner to directly suppress T cell activation or a paracrine manner through DCs to control immunopathology (11, 44–46). Our results support the conclusion that in the absence of IL-10 and its regulation of T cells, increased RSV-induced disease severity is a consequence of increased immunopathology. However, the molecular mechanisms that induce IL-10 production by effector T cells during RSV infection remain to be determined.

We demonstrate that CD4+ T cells produce the majority of IL-10 in vivo during an acute RSV infection. In contrast, we observe only a small number of IL-10–producing CD8+ T cells (Fig. 1). IL-10 production by T cells has been examined in a number of other respiratory virus infections. Consistent with our results, McKinstry et al. (43) recently reported that CD4+ T cells make the majority of IL-10 during a high-dose IAV infection. Virus-specific CD8+ T cells that coproduce IL-10 and IFN-γ in the lung have been reported during acute SV5 infection (47). However, these studies did not assess IL-10 production by CD4+ T cells during SV5 infection. Therefore it is unclear if either CD4+ or CD8+ T cells represent the dominant cell source of IL-10 during acute SV5 infection. In contrast to our findings with RSV, Sun et al. (48)
report that CD8\(^+\) T cells are responsible for the majority of IL-10 production during an acute low-dose IAV infection. It is currently unclear what accounts for the reported discrepancies in the proportion of CD4\(^+\) and CD8\(^+\) T cells that make IL-10 during respiratory viral infections. A more recent study by Sun et al. (49) indicates that IL-10 production by CD8\(^+\) T cells during an acute IAV infection requires the expression of Blimp-1 and is synergistically promoted by IL-2 produced by helper CD4\(^+\) T cells as well as IL-27 produced by innate immune cells. It remains to be determined if the reduced number of IL-10–producing CD8\(^+\) T cells during an acute RSV infection is the result of a lack of CD8\(^+\) T cell help and/or decreased IL-27 production.

Exacerbated disease severity and increased levels of proinflammatory cytokines and chemokines were observed in the absence of IL-10 following RSV infection (Figs. 4, 6). However, we did not observe any changes in peak viral titers (day 4 p.i.) or viral clearance (day 7 p.i.) between IL-10 KO or anti–IL-10R mAb-treated mice compared with their controls (Fig. 5). These data support the idea that increased disease severity is independent of RSV replication and clearance kinetics (23). Instead, alterations in the RSV immune response (i.e., increased inflammatory conditions) result in increased immunopathology. Similarly, Haeberle et al. (50) reported that treatment with rIL-10 significantly decreased the production of chemokines, such as CCL2, CCL3, and CCL5, in the lung airways when analyzed 24 h after RSV infection. CCL5 has been positively associated with RSV-induced disease severity. In recent studies, depletion of natural Tregs selectively identify natural Tregs (35, 36). Thus, the decreased number of Th17 CD4\(^+\) T cells during an acute RSV infection requires the expression of Blimp-1 and is synergistically promoted by IL-2 produced by helper CD4\(^+\) T cells as well as IL-27 produced by innate immune cells. It remains to be determined if the reduced number of IL-10–producing CD8\(^+\) T cells during an acute RSV infection is the result of a lack of CD8\(^+\) T cell help and/or decreased IL-27 production.

In contrast to the increased number of Th17 cells discussed above, we observed a decrease in the total number of Helios\(^+\) Foxp3\(^+\) Tregs in the lung following IL-10R blockade in RSV-infected mice. Recent work suggests that Helios expression may be necessary to maintain Foxp3 expression of CD4\(^+\) T cells (60). Murai et al. demonstrated that the suppressive function of Tregs was maintained by IL-10R expression and the capacity to directly respond to IL-10. However, the environment in the intestinal mucosa is unique, with different regulatory factors than those in the lung, and the findings of Murai et al. may not necessarily translate to lung mucosal tissue. Therefore, the role of IL-10 in the regulation of Foxp3 expression will need to be further evaluated in the lung environment. We are currently examining the role of IL-10 in the induction and/or maintenance of pulmonary Tregs during acute RSV infection.

IL-10 polymorphisms have been associated with RSV-induced disease severity in humans (61, 62). In this article, we demonstrate that IL-10 plays a critical role in regulating the immune response and preventing exacerbated disease due to immune-mediated pulmonary injury during acute RSV infection. Multiple
respiratory viruses, such as pandemic IAV strains and the human coronavirus that causes SARS, are believed to induce immunopathology resulting in severe morbidity and mortality (8, 9, 56–58). Therefore, identifying novel approaches to modulate virus-induced immunopathology would be beneficial in treating acute respiratory viral infections. Exogenous IL-10 treatment has been shown to be safe and well tolerated in clinical trials for several chronic inflammatory diseases, including Crohn’s disease, psoriasis, and chronic hepatitis C (63–68). In patients with chronic hepatitis C who were nonresponsive to IFN-based therapy, IL-10 treatment did not affect viral titers but reduced chronic liver inflammation and fibrosis (66). Using IL-10 as an immunosuppressive treatment during a respiratory viral infection resulting in immunopathology may be feasible to prevent severe morbidity and mortality. Our data indicate that IL-10 is an attractive target based on its ability to regulate RSV-induced innate and adaptive immune responses without affecting viral clearance.

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