Cutting Edge: Structural Basis for the Recognition of β-Linked Glycolipid Antigens by Invariant NKT Cells

Esther Dawen Yu, Enrico Girardi, Jing Wang and Dirk M. Zajonc

*J Immunol* published online 1 August 2011
http://www.jimmunol.org/content/early/2011/07/31/jimmunol.1101636

**Supplementary Material**
http://www.jimmunol.org/content/suppl/2011/08/02/jimmunol.1101636.6.DC1

**Subscription**
Information about subscribing to *The Journal of Immunology* is online at:
http://jimmunol.org/subscription

**Permissions**
Submit copyright permission requests at:
http://www.aai.org/About/Publications/JI/copyright.html

**Email Alerts**
Receive free email-alerts when new articles cite this article. Sign up at:
http://jimmunol.org/alerts
Cutting Edge: Structural Basis for the Recognition of β-Linked Glycolipid Antigens by Invariant NKT Cells

Esther Dawen Yu, Enrico Girardi, Jing Wang, and Dirk M. Zajonc

Invariant NKT (iNKT) cells expressing a semi-invariant Vα14 TCR recognize self and foreign lipid Ags when presented by the nonclassical MHCI homolog CD1d. Whereas the majority of known iNKT cell Ags are characterized by the presence of a single α-linked sugar, mammalian self Ags are β-linked glycosphingolipids, posing the interesting question of how the semi-invariant TCR can bind to such structurally distinct ligands. In this study, we show that the mouse iNKT TCR recognizes the complex β-linked Ag isoglobotrihexosylceramide (iGb3; Galα1-3-Galβ1-4-Glcβ1-1Cer) by forcing the proximal β-linked sugar of the trisaccharide head group to adopt the typical binding orientation of α-linked glycolipids. The squashed iGb3 orientation is stabilized by several interactions between the trisaccharide and CD1d residues. Finally, the formation of novel contacts between the proximal and second sugar of iGb3 and CDR2α residues of the TCR suggests an expanded recognition logic that can possibly distinguish foreign Ags from self Ags.

Invariant NKT (iNKT) cells constitute a T lymphocyte population that is able to modulate the functions of several different cell types such as NK cells, B cells, and T cells, therefore bridging the innate and adaptive immune response to self and foreign Ags. Critical to this role is the ability of iNKT cells to rapidly secrete, upon stimulation, large amounts of both Th1 and Th2 cytokines, such as IFN-γ, IL-4, and IL-13 (1). iNKT cells express a semi-invariant TCR recognize a range of highly diverse Ags. The squashed iGb3 orientation of the iNKT TCR. The crystal structure of the mouse CD1d-iGb3-iNKT TCR complex described in this study illustrates how the iNKT TCR is able to recognize complex β-linked glycolipids by flattening the trihexosyl sugar of iGb3 between the CD1d and TCR surface to allow for the conserved TCR-binding mode on CD1d that has previously been observed for the α-linked Ags (4, 5, 8–11), therefore explaining how this conserved TCR can recognize a range of highly diverse Ags.

Materials and Methods

Lipids

β-lactosylceramide (β-LacCer) and β-glucosylceramide (β-GlcCer) were obtained from Avanti Polar Lipids. iGb3 was purchased from Enzo Life Sciences International. Lipids were dissolved at 0.4 mg/ml in 0.5% Tween 20 obtained from Avanti Polar Lipids. iGb3 was purchased from Enzo Life Sciences International. Lipids were dissolved at 0.4 mg/ml in 0.5% Tween 20 and 0.9% NaCl and stored at 4°C.

Cell lines and culture conditions

iNKT hybridoma cell lines 1.2, 1.4, and 2C12 (12, 13) were cultured in RPMI 1640 medium supplemented with 2 mM l-glutamine, 100 mg/ml each of penicillin and streptomycin, 50 mM 2-ME, and 10% FBS. The cell lines were maintained in an incubator with a humidified atmosphere containing 5% CO₂ at 37°C.

Cell-free Ag presentation assay

The cell-free Ag presentation assay for stimulation of mouse iNKT cell hybridomas by soluble mCD1d was carried out following published protocols.

Received for publication June 9, 2011. Accepted for publication July 6, 2011.

This work was supported by a Cancer Research Institute Investigator award and by National Institutes of Health Grant RO1 AI074952 (to D.M.Z.).

Structure factors and coordinates have been deposited in the Protein Data Bank under accession code 3RZC.

Division of Cell Biology, La Jolla Institute for Allergy and Immunology, La Jolla, CA 92037

1E.D.Y. and E.G. contributed equally to this work.

© 2011 by The American Association of Immunologists, Inc. 0022-1767/11/$16.00

Address correspondence and reprint requests to Prof. Dirk M. Zajonc, Division of Cell Biology, La Jolla Institute for Allergy and Immunology, 9420 Athena Circle, La Jolla, CA 92037. E-mail address: dza jonc@lji.org

The online version of this article contains supplemental material.

Abbreviations used in this article: α-GalCer, α-galactosylceramide; β-GlcCer, β-glucosylceramide; iGb3, isoglobotrihexosylceramide; iNKT, invariant NKT; β-LacCer, β-lactosylceramide; β-m, β-microglobulin; PI, phosphatidylinositol; SPR, surface plasmon resonance.

Copyright © 2011 by The American Association of Immunologists, Inc. 0022-1767/11/$16.00

www.jimmunol.org/cgi/doi/10.4049/jimmunol.1101636
with the modifications described below. Briefly, 1 μg soluble mCD1d, G155W, and hCD1d protein was coated in 96-well flat-bottom plate at 4°C overnight. The plates were blocked by PBS and 10% FBS for 1 h after washing, and then the glycolipid of interest at various concentrations (shown in Fig. 1B and Supplemental Fig. 1) was added to each well and incubated for 24 h at 37°C. After washing, 5 × 10^4 hybridoma cells in 200 μl complete media were added to each well and incubated at 37°C for 16 h in a CO2 incubator. IL-2 release in the supernatant was measured after 16 h of culture in a sandwich ELISA, as previously described (12, 13).

**Mouse CD1dβγm expression and purification and Vα14-Vβ8.2 TCR refolding**

The expression and purification methods of fully glycosylated mouse and human CD1d/βγm heterodimer proteins were reported previously (11). Mouse TCR refolding was performed according to previously reported protocols, using the identical construct (11, 14). A representative human Vα24-Vβ11 iNKT TCR (CDR3β: CASSDPNEQFF) was cloned in pET22b (Vα24 chain) and pET30a (Vβ11 chain) Escherichia coli expression vectors. An additional interchain disulfide bond was engineered by mutating to cysteine Thr248 (TRBC 75) on the Cβ domain and Ser57 on the Cα domain. Moreover, a cysteine residue (TRBC 75) on the Cβ domain was mutated to serine to avoid incorrect oligomerization. The two chains were expressed, refolded, and purified, as described for the mouse Vα14-Vβ8.2 iNKT TCR.

**Glycolipid loading and ternary complex formation**

Mouse CD1d was loaded overnight with 4–6 molar excess of iGb3 (0.4 mg/ml in 0.5% Tween 20 and 0.9% NaCl). iGb3-loaded CD1d was purified by size-exclusion chromatography and incubated with an equimolar amount of TCR for 1 h at room temperature. The mixture was concentrated to 4.8 mg/ml in 10 mM HEPES (pH 7.5), 30 mM NaCl before being used for crystallization.

**Surface plasmon resonance studies**

Surface plasmon resonance (SPR) studies were performed using a Biacore 3000 (GE Healthcare), according to the methods described previously (14). Human or mouse CD1d was expressed, purified, and biotinylated, as previously reported, and loaded with iGb3 (8). CD1d–iGb3 complexes and empty CD1d as a control were immobilized on a streptavidin sensor chip (CAPture chip; GE Healthcare), whereas increasing concentrations (0.02–20 μM) of the mouse or human TCR were passed over the chip at 25°C with a flow rate of 30 μl/min. Experiments were repeated twice. Kinetic parameters were calculated in the BIA evaluation software (version 4.1) after background subtraction.

**Crystallization and structure determination**

Crystals of mCD1d–iGb3–TCR complexes were grown at 27.6°C by sitting drop vapor diffusion, mixing 0.5 μl protein with 0.5 μl precipitant (16% polyethylene glycol 3350, 8% v/v Tacsimate [pH 5.0]). Crystals were flash cooled at 100 K in mother liquor containing 20% glycerol. Diffraction data were collected at the Stanford Synchrotron Radiation Lightsource beamline 7-1. Data processing and structure determination by molecular replacement using the CD1d and TCR coordinates from PDB ID 3QUY were carried out as reported for other CD1d-glycolipid complexes (11). Model building and refinement were performed analogous to other CD1d-glycolipid–TCR structures (11). Data collection and refinement statistics are presented in Supplemental Table I.

**Results and Discussion**

Mouse NKT cells recognize iGb3 presented by mouse or human CD1d

We tested the ability of iGb3 and its biosynthetic precursors β-LacCer and β-GlcCer, which lack one or two distal sugar rings compared with iGb3 (Fig. 1A), to activate three Vα14 NKT hybridomas in a cell-free Ag presentation assay (Fig. 1B, Supplemental Fig. 1). Consistent with other studies (7, 15), the truncated ligands β-LacCer and β-GlcCer did not activate either hybridoma. However, iGb3 was able to induce significant IL-2 release from all three hybridomas. These results confirm the importance of the distal α1-3-linked sugar of iGb3 for its antigenicity. Interestingly, iGb3 was able to stimulate mouse iNKT cell hybridomas when presented by human CD1d, whereas presentation of iGb3 by the humanized mCD1d mutant G155W (in hCD1d, Trp153 is at the equivalent position compared with Gly155 in mCD1d) did not lead to mouse iNKT cell activation. Recognition of α-GalCer was unimpaired when presented by the G155W
mutant, indicating that this mutant was still able to bind and properly present glycolipids. Most residues lining the entrance to the mCD1d-binding groove are conserved in hCD1d (except for the G155W exchange), but differences are observed in the area above the A’ pocket (i.e., Met69 in mCD1d to Ile, Gly155 to Trp, Met162 to Trp). The results obtained with the G155W mutant therefore suggest that the observed cross-reactivity of mouse iNKT cells toward human CD1d-presented iGb3 is not dependent on a single residue; it is in fact abrogated by the single amino acid substitution, and it is rather the result of several contacts unique to the human CD1d protein surface that compensate for the G155W substitution, suggesting an alternative mode of iGb3 presentation by mouse and human CD1d.

**iGb3 has a submicromolar affinity for the mouse iNKT TCR**

Previous SPR experiments carried out using a short-chain version of iGb3 and an iNKT TCR with a truncated CDR3α loop reported a low-affinity ($K_D \sim 50 \mu M$) interaction (16). In this study, we measured the affinity of both the mouse (Vα14/β8.2 derived from the 2C12 hybridoma (14) and human (Vα24β11) iNKT cell TCR toward the full-length iGb3 presented by either mouse or human CD1d (Fig. 1, Supplemental Fig. 1B). The mouse TCR binds to mCD1d-iGb3 with submicromolar affinity ($K_D$ of 718 nM). Binding was characterized by a relatively fast on-rate ($k_2 1.95 \times 10^8 \text{M}^{-1}\text{s}^{-1}$) and a slow off-rate ($k_1 1.19 \times 10^{-2} \text{s}^{-1}$), not dissimilarly from other previously characterized α-GalCer analogs, but in contrast to microbial Ags that are characterized by much faster TCR dissociation rates (GalA-GSL from *Sphingomonas spp.*, $k_2 1.43 \times 10^9 \text{M}^{-1}\text{s}^{-1}$, $k_1 9.4 \times 10^{-2} \text{s}^{-1}$; BbGlc from *Borrelia burgdorferi*, $k_2 2.16 \times 10^4 \text{M}^{-1}\text{s}^{-1}$, $k_1 0.165 \text{s}^{-1}$) (14). SPR measurements for β-LacCer and β-GlcCer did not result in measurable affinities, consistent with their failure to activate iNKT cells (data not shown).

Human CD1d-iGb3 is bound with weaker affinities by both mouse and human iNKT TCRs ($K_D$ of 5.1 and 6.8 μM, respectively), compared with mCD1d-iGb3, whereas no measurable binding was observed for the interaction of mCD1d-iGb3 with the human iNKT TCR. Taken together, these data confirm the pattern observed in the Ag presentation assay (Fig. 1B, Supplemental Fig. 1A) and further suggest a limited cross-reactivity between human and mouse iNKT cells. Finally, our data suggest that in vitro iGb3 is a surprisingly more potent iNKT Ag than previously appreciated.

**Crystal structure of the mCD1d–iGb3–iNKT TCR complex**

We determined the crystal structure of the mouse CD1d–iGb3–iNKT TCR complex at 2.8Å resolution (Fig. 2, Supplemental Table I). The overall docking orientation of the iNKT TCR on mCD1d is highly similar to other previously determined ternary complexes with bound α-GalCer (5), its various analogs (9, 11), as well as microbial Ags (8). In the mCD1d–iGb3–TCR complex, the TCR adopts a parallel orientation on mCD1d characterized by a footprint centered above the F’ pocket. Key residues in the CDR3α, CDR2β, and CDR3β loops (Supplemental Fig. 1C) mediate polar contacts with the α1 and α2 helices of mCD1d, whereas the TCR α-chain exclusively mediates contacts with the Ag.

**FIGURE 2.** Structure of the mouse CD1d–iGb3–TCR complex. A. Crystal structures of the mTCR–iGb3–mCD1d complex. iGb3 in yellow; mCD1d H chain and β2m in gray; TCR α-chain in cyan; TCR β-chain in orange. Final 2Fo-Fc electron density map for iGb3. The 2FoFc map at 1σ is shown as a blue mesh in side view (top panel) and top view (bottom panel). C. Superposition of α-GalCer (purple) and iGb3 (yellow) in the mouse ternary complex illustrates the similar binding orientation of the β-linked glucose and the α-linked galactose sugars after TCR binding.
Consistent with all the sphingolipid Ags analyzed to date (3), the iGb3 molecule binds to mCD1d with its sphingosine chain in the F′ pocket and the C26 acyl chain in the A′ pocket. Surprisingly, upon TCR binding, the trisaccharide head group of iGb3 gets squashed over the α2 helix of CD1d, with each sugar making polar and VdW contacts with CD1d residues (Fig. 3A). In particular, the proximal β-Glc residue contacts Asp153 through its 2′-OH, whereas the second β-Gal sugar contacts Thr159 through its 2′-OH. In contrast, the terminal α-Gal residue nestles in a relatively hydrophobic pocket formed between Ala158, Thr159, and Met162, while packing against the side chain of Met162 (Fig. 3A). The flat binding orientation of the trisaccharide molds the proximal β-linked glucose of iGb3 to bind into a highly similar position compared with α-linked glycolipids such as α-GalCer, with the 2′-OH forming a H-bond with Asp153 of CD1d (Fig. 2C). The same residues on mCD1d (Asp158) and the TCR (Asn90α, Gly96α) are involved in recognizing the iGb3 β-linked proximal sugar and the sugar of other α-linked Ags, therefore suggesting remarkably conserved recognition logic for both α- and β-anomeric glycosphingolipids. Surprisingly, considering the essential role of the terminal sugar of iGb3 in determining its antigenicity, contacts between the Ag and the TCR α-chain involve only the β-Glc and β-Gal sugars, but not the terminal α-Gal (Fig. 3B). In addition, unlike any previously characterized exogenous iNKT cell Ag, all three CDR loops from the Vα14 chain are involved in contacting the iGb3 molecule. In particular, Gly96 and Asn90 on the CDR3α and CDR1α loops, respectively, form H-bonds with the 2′ and 3′ hydroxyl groups of β-Glc. At the same time, the carbonyl group of both Asn90 on CDR1α and Val50 on CDR2α, as well as the framework residue Lys68, is interacting with the 6′-OH of the β-Gal residue (Fig. 3B). Interestingly, a similar interaction pattern with the iNKT TCR was recently reported for the self Ag phosphatidylinositol (PI) (10), which is not a β-linked glycosphingolipid self Ag, but instead a phosphoglycerolipid and, as such, presented differently by the relatively slow TCR dissociation rate and the well-defined electron density observed. Consistent with this binding mode, the shorter β-GlcCer and β-LacCer ligands that lack the terminal sugar are not antigenic. Moreover, the closely related compounds iGb4 (GalNAcβ1-3Galα1-3LacCer) and Gb3 (Galα1-4LacCer) are not antigenic because of the additional sugar moiety or the suboptimal orientation of their distal sugar that cannot fit or reach the anchoring pocket on the mCD1d surface. In conclusion, we hypothesize that complex β-linked glycolipids that cannot anchor the sugar moiety to the CD1d surface after TCR flattening require more binding energy to stabilize the CD1d–TCR complex and, as such, have significantly weaker potency.

Finally, as described for other iNKT Ags, this TCR is also able to force mCD1d in a conserved binding mode by inducing the formation of a hydrophobic roof above the F′ pocket (Supplemental Fig. 1D). The F′ roof formation involves the repositioning of the side chains of residues Leu84 and Val149 and Leu′50 of mCD1d and was previously shown to affect the stability of the mCD1d–iNKT TCR complex (8). Although TCR-induced conformational changes for MHC-peptide and CD1d-glycolipid have been reported (8, 18), the extent of reorientation observed for iGb3 is unprecedented among TCR Ags, highlighting once again the unique nature of the iNKT TCR.

Acknowledgments
We thank Stanford Synchrotron Radiation Lightsource BL 7-1 for remote data collection and Dr. Kronenberg (La Jolla Institute for Allergy and Immunology) for providing the Vα24β11 TCR cDNA.

Disclosures
The authors have no financial conflicts of interest.

References


