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A Critical Role for Mast Cells and Mast Cell-Derived IL-6 in TLR2-Mediated Inhibition of Tumor Growth

Sharon A. Oldford,*† Ian D. Haidl,*† Mackenzie A. Howatt,*† Carlos A. Leiva,*† Brent Johnston,*†§ and Jean S. Marshall*†§

Several TLR agonists are effective in tumor immunotherapy, but their early innate mechanisms of action, particularly those of TLR2 agonists, are unclear. Mast cells are abundant surrounding solid tumors where they are often protumorigenic and enhance tumor angiogenesis. However, antitumor roles for mast cells have also been documented. The impact of mast cells may be dependent on their activation status and mediator release in different tumors. Using an orthotopic melanoma model in wild-type C57BL/6 and mast cell-deficient KitW-sh/W-sh mice and a complementary Matrigel–tumor model in C57BL/6 mice, mast cells were shown to be crucial for TLR2 agonist (Pam3CSK4)-induced tumor inhibition. Activation of TLR2 on mast cells reversed their well-documented protumorigenic role. Tumor growth inhibition after peritumoral administration of Pam3CSK4 was restored in KitW-sh/W-sh mice by local reconstitution with wild-type, but not TLR2-deficient, mast cells. Mast cells secrete multiple mediators after Pam3CSK4 activation, and in vivo mast cell reconstitution studies also revealed that tumor growth inhibition required mast cell-derived IL-6, but not TNF. Mast cell-mediated anticancer properties were multifaceted. Direct antitumor effects in vitro and decreased angiogenesis and recruitment of NK and T cells in vivo were observed. TLR2-activated mast cells also inhibited the growth of lung cancer cells in vivo. Unlike other immune cells, mast cells are relatively radioresistant making them attractive candidates for combined treatment modalities. This study has important implications for the design of immunotherapeutic strategies and reveals, to our knowledge, a novel mechanism of action for TLR2 agonists in vivo. The Journal of Immunology, 2010, 185: 000–000.

The use of TLR agonists as immunological adjuvants in the treatment of cancer has been intensively studied. TLR-based immunotherapies have shown promise clinically for the treatment of multiple tumor types. Specifically, TLR2 agonists have been successfully used as potential therapeutics in animal models of pancreatic and brain carcinoma (1, 2) as well as in clinical trials (3). In general, the successful mechanism of action of TLR agonists is considered to involve dendritic cell maturation with enhanced tumor Ag presentation and generation of Th1 immunity and CD8 antitumor effector cells (4). TLR2 agonists, such as Pam3CSK4, often do not share the strong Th1-polarizing effects of other TLR agonists (5, 6). However, they retain antiangiogenesis and recruitment of NK and T cells in vivo were observed. TLR2-activated mast cells also inhibited the growth of lung cancer cells in vivo. Unlike other immune cells, mast cells are relatively radioresistant making them attractive candidates for combined treatment modalities. This study has important implications for the design of immunotherapeutic strategies and reveals, to our knowledge, a novel mechanism of action for TLR2 agonists in vivo. The Journal of Immunology, 2010, 185: 000–000.

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TLR2/1 agonist Pam\(_3\)CSK\(_4\) using an orthotopic model of B16.F10 melanoma, a well used, highly aggressive, poorly immunogenic tumor (24). B16.F10 growth was compared in wild-type and mast cell-deficient Kit\(^{W-sh}\) mice. Although Kit\(^{W-sh}\) mutation impairs long-term endogenous melanocyte survival but does not affect melanoblast development or differentiation or the survival of transplanted c-Kit–expressing melanocytes. Homozygote adult animals consistently retain melanocytes at selected sites including the ear pinna (25, 26). Comparative growth studies in Kit\(^{W-sh/W-sh}\) and control C57BL/6 mice identified that Pam\(_3\)CSK\(_4\) inhibits tumor growth in a mast cell and mast cell-derived IL-6-dependent manner. This study demonstrates that selective activation of mast cells via TLR2 can reverse their tumor-promoting potential and induce them to inhibit tumor growth.

### Materials and Methods

#### Mice

C57BL/6, B6.129S6-Tgf\(_{\beta}^{pm1Nga}\)J, B6.129S2-B6.129\(_{\beta 1}^{tm1Unc}\)J, B6.129P2-Ctnf\(^{tm1Gkl}\)J, and B6.Cg-Ki67\(^{+}\)H2B\(_{\beta}^{G120C}\)J (Kit\(^{W-sh}\)J) were purchased from The Jackson Laboratory (Bar Harbor, ME). C57BL/6 mice were bred in-house from this stock. B6.129-Tgf\(_{\beta}^{pm1Nga}\)J and Kit\(^{W-sh}\)J were identified by Dr. S. Akira (Osaka, Japan) and bred in-house or purchased from The Jackson Laboratory. Matched groups of male or female mice were used at 8–12 wk of age and housed under pathogen-free conditions with food and water provided ad libitum. All experiments were performed according to protocols approved by the animal research ethics board of Dalhousie University (Halifax, Nova Scotia, Canada).

#### Generation of bone marrow-derived mast cells and local reconstitution of mast cell-deficient mice

Bone marrow-derived mast cells (BMMCs) were generated from wild-type C57BL/6 and gene-deficient TLR2\(^{-/-}\), TNF\(^{-/-}\), IL-6\(^{-/-}\), or CCL3L1\(^{-/-}\) mice as previously described (27) and were used at >98% purity. BMMCs derived from selected gene-deficient animals were functionally similar to control C57BL/6 BMMCs (Supplemental Fig. 1). BMMCs were cultured for 16–24 h in the absence of PGE2 prior to in vitro experiments. Eight- to ten-week-old Kit\(^{W-sh/W-sh}\) mice were reconstituted on the dorsal surface over the right and left flanks in 2 × 2 cm\(^2\) areas via a series of injections with BMMCs (1 × 10\(^6\) intradermal and 1 × 10\(^6\) s.c.), which resulted in mast cell levels comparable with those of wild-type C57BL/6 mice (Supplemental Fig. 2).

#### Established tumor growth experiments

B16.F10 cells (1 × 10\(^5\)) (American Type Culture Collection, Manassas, VA) in 100 μl PBS were injected s.c. above the right and left flanks. At days 4 and 9 (and day 14 as indicated), mice were injected s.c. at the tumor site with 3 μg Pam\(_3\)CSK\(_4\) or FSL-1 (EMC Microcollections, Tübingen, Germany) in 50 μl PBS or diluent control. Tumor volume (V) was determined by caliper measurements (V = L × W\(^2\)). After 14 to 19 d, mice were euthanized, and tumors were removed and weighed. Percentage tumor reduction = (tumor + BMMC + PBS control tumor average)/PBS control tumor average. 

#### In vivo Matrigel–tumor growth experiments

Growth factor-reduced Matrigel (Becton Dickinson, San Jose, CA) suspensions containing 1.5 × 10\(^5\) B16.F10 or LLC1 cells (American Type Culture Collection), with or without 10 μg/ml Pam\(_3\)CSK\(_4\) (EMC Microcollections) and/or 3 × 10\(^5\) BMMCs, were prepared. All Matrigel suspensions contained 3 ng/ml murine rIL-3 (PeproTech, Rocky Hill, NJ). C57BL/6 mice (8–12 wk) were injected s.c. in four sites with 0.3 or 0.15 ml suspensions for 4 or 10 d, as indicated. Mice were euthanized, and tumors were excised and weighed. Percentage tumor reduction = (tumor + BMMC + Pam\(_3\)CSK\(_4\) Matrigel average − tumor + BMMC Matrigel average)/tumor + BMMC Matrigel average × 100, following subtraction of baseline Matrigel plug weight determined from 10-d Matrigel plugs containing BMMCs and Pam\(_3\)CSK\(_4\) without tumor cells.

#### Tumor blood volume determination

Tumor-bearing mice were anesthetized and injected via the tail vein with 100 μl 25 mg/ml FITC-labeled dextran (average m.w. 2 × 10\(^6\); Sigma-Aldrich, Oakville, Ontario, Canada) in PBS. After 5-min perfusion, 20 μl blood was collected, tumors were excised and digested in 300 μl collagenase IV (Sigma-Aldrich), and cell-free supernatants were measured fluorometrically (excitation 488 nm, emission 519 nm). Relative fluorescent units (RFU) were normalized to blood plasma levels.

#### Histology

Formalin-fixed, paraffin-embedded Matrigel–tumor sections (3 μm) were stained with H&E. Mature erythrocyte-containing blood vessel lumens were quantified in non-overlapping fields (magnification ×400) of tumor periphery, on coded sections. Mast cells were visualized using Acrilane blue (pH 0.5)–safranin O staining of Carnoy’s fixed, paraffin-embedded tumor sections.

#### Tumor cell cytotoxicity and cell viability

Tumor cell proliferation was evaluated by the MTT assay. Briefly, 1 × 10\(^4\) tumor cells were incubated overnight in 96-well tissue culture plates then treated, in quadruplicate, for 24 h before MTT addition. Treatments included 0, 1, or 10 μg/ml Pam\(_3\)CSK\(_4\), 5 μg/ml of the DNA topoisomerase I inhibitor camptothecin (CPT) or DMSO vehicle control (Sigma-Aldrich), or supernatants from C57BL/6 or IL-6–/− BMMCs incubated for 24 h in BMMC culture medium with or without 10 μg/ml Pam\(_3\)CSK\(_4\). Tumor cell apoptosis in B16.F10 (0.5 × 10\(^6\)/ml)–BMMC (1 × 10\(^6\)/ml) cocultures was assessed using an annexin V–FITC–propidium iodide apoptosis detection kit (BD Biosciences, Mississauga, Ontario, Canada) according to the manufacturer’s instructions. BMMCs were excluded from analysis via biotinylated anti-CD45 (eBioscience, San Diego, CA) and streptavidin (SA)–allophycocyanin (BD Biosciences) staining.

#### Cell counts and flow cytometry

Established tumors were collagenase digested as above. Matrigel tumors were digested in 2.24 U/ml Dispase (Life Technologies, Carlsbad, CA) in PBS. Overall tumor cellularity was assessed by flow cytometry of digested tumors with reference to a known number of 15 μm Polystyrene microspheres (Polysciences, Warrington, PA). Total leukocytes were stained with biotinylated anti-CD45 (eBioscience) and SA–PerCP (BD Biosciences), anti-TRC–FITC (eBioscience) or anti-CD3–FITC (BD Biosciences), anti-CD8–PE (eBioscience), and anti-NK1.1–allophycocyanin (BD Biosciences). Biotinylated rat IgG2a isotype (eBioscience) with SA–PerCP staining did not exceed 0.1% of total. Samples were gated on CD45\(^+\) cells and leukocyte subsets quantified within this gate. Staining using relevant isotype control Abs (eBioscience) did not exceed 0.2% of CD45\(^+\) cells.

#### Chemotaxis assays

C57BL/6 spleens were dispersed by pushing through a 100-μm nylon cell strainer. Erythrocytes were lysed using 0.14 M ammonium chloride plus 20 mM Tris (pH 7.4). Adherent cells were removed via incubation for 20 min at 37°C in 0.1% BSA in RPMI 1640. Chemotaxis of nonadherent splenocytes was performed using 24-well Transwells with a 5-μm pore size (Corning, Lowell, MA). Test supernatant (600 μl) was added to the bottom chamber, and 1 × 10\(^5\) splenocytes in 100 μl RPMI 1640 containing 0.1% BSA was added to the top. Chemotaxis was carried out for 2 h at 37°C. To enumerate total migrating cells, each well received 15-μm polybead polystyrene microspheres (Polysciences) as an internal standard. NK1.1\(^+\), CD11\(^+\), and CD8\(^+\) cells were assessed by flow cytometry, as described earlier. Total migrated subsets are expressed as a percentage of the starting cell subset population and were calculated based on absolute numbers of each cell subset.

#### In vitro cell stimulation, protein array analysis, and ELISA

B16.F10 cells (0.5 × 10\(^6\)/ml) and/or BMMCs (1 × 10\(^6\)/ml) were incubated with or without 10 μg/ml Pam\(_3\)CSK\(_4\), FSL-1, LPS (Sigma-Aldrich), PG\(_N\) (Invivogen), and polyinosinic-polycytidylic acid (Calbiochem, San Diego, CA) or A23187 ionophore (Sigma-Aldrich) positive control at 37°C in RPMI 1640 containing 10% FBS, 3 ng/ml murine rIL-3 (PeproTech), and 100 μg/ml soybean trypsin inhibitor (Sigma-Aldrich). Cell-free supernatants were assessed using the murine Cytokine III Ab array (Raybiotech, Norcross, GA) and ELISA kits to quantify levels of TNF and IL-13 (both eBioscience), IL-6 (PeproTech), and CCL1 and CCL3 (both R&D Systems, Minneapolis, MN).

#### Statistics

Samples were analyzed using unpaired t test or non-parametric Mann-Whitney U test as indicated based on D’Agostino and Pearson omnibus normality test. Matrigel–tumor experiments were assessed with repeated measures Friedman’s test and between-group comparisons assessed by Wilcoxon matched pairs test (10 d) or repeated measures ANOVA with
Results

TLR2 agonist treatment inhibits B16.F10 tumor growth via mast cell activation

The tumor-inhibitory effects of the TLR2/1 agonist Pam3CSK4 were investigated using an orthotopic model of B16.F10 melanoma. When treated with Pam3CSK4, tumors showed a significant reduction in volume (73%; Fig. 1A) and cellularity (74%; Fig. 1B). Mast cells were detected in close proximity to blood vessels at the periphery of both Pam3CSK4-treated and control tumors (Fig. 1C). As the ability of mast cells to exert a protective role in tumorigenesis may be dependent on an intratumoral distribution (28, 29), we assessed the distribution of mast cells in control and Pam3CSK4-treated tumors. Mast cells were predominately found in the stroma as opposed to intratumorally for both PBS-treated (2.3 ± 0.5/mm² versus 0.5 ± 0.1/mm², n = 6) and Pam3CSK4-treated (3.3 ± 0.7/mm² versus 1.7 ± 0.9/mm², n = 9) tumors. The percentage of mast cells that were found intratumorally did not significantly differ between PBS-treated and Pam3CSK4-treated tumors (21 ± 5% versus 28 ± 11%). The protumorigenic role of mast cells has been well characterized in several models (14, 15). Consistent with these findings, baseline tumor growth rate, in the absence of TLR2 agonist treatment, was significantly decreased in mast cell-deficient KitW-sh/W-sh animals compared with that in mast cell-containing C57BL/6 mice (67%; Fig. 1D).

To investigate a possible role for mast cells in Pam3CSK4-mediated tumor growth inhibition, B16.F10 tumors were treated in mast cell-containing C57BL/6 mice or mast cell-deficient KitW-sh/W-sh mice. In KitW-sh/W-sh mice, Pam3CSK4 did not inhibit tumor growth (Fig. 2A), whereas tumor growth was decreased in parallel groups of mast cell-containing mice (59%; Fig. 2B), suggesting a mast cell-dependent mechanism of Pam3CSK4 action. Decreased end-point tumor weight (65%; Fig. 2C) and tumor blood volume (69%; Fig. 2D) were also mast cell dependent. Enhanced growth of diluent-treated control tumors was observed in C57BL/6 compared with KitW-sh/W-sh, further confirming the protumorigenic role of mast cells in the absence of TLR2 activation.

Whereas KitW-sh/W-sh mice have been documented to be similar to their wild-type littermates with respect to myeloid and lymphoid cell subsets in multiple anatomical sites (30), others have demonstrated myeloid differences compared with wild-type C57BL/6 mice (31). To overcome any inherent defects of the mast cell-deficient mouse model and assess the physiological role of mast cells in Pam3CSK4-mediated effects, KitW-sh/W-sh mice were locally reconstituted with BMMCs, which were allowed to mature in situ prior to performing tumor growth experiments. We also extended the time course of the tumor growth experiment to ensure the lack of effect of Pam3CSK4 in KitW-sh/W-sh mice was not attributable to delayed tumor growth kinetics (Fig. 2A). Even with tumor size reaching levels comparable with those of C57BL/6 mice, the antitumor activity of Pam3CSK4 was absent in mast cell-deficient mice (Fig. 3A). Local reconstitution with C57BL/6, but not TLR2−/− BMMCs, restored the Pam3CSK4-mediated decrease in tumor volume (61%, Fig. 3A) and tumor weight (63%, Fig. 3B).

TLR1 and TLR2 are expressed on both murine BMMCs (32) and B16 melanoma (33) (see also Supplemental Fig. 3). To confirm the specific involvement of mast cell-associated TLR2 in the antitumor effects of Pam3CSK4, the ability of wild-type and TLR2−/− BMMCs to inhibit tumor growth were compared using a Matrigel–tumor model. Even within the context of the tumor growth-promoting Matrigel environment (34), TLR2-expressing BMMCs significantly inhibited melanoma growth in the presence of Pam3CSK4 (49% inhibition), whereas Pam3CSK4-activated TLR2-deficient BMMCs did not (Fig. 3C). Pam3CSK4 or BMMCs alone had no inhibitory effect on tumor growth as weight of Matrigels containing B16.F10 plus BMMCs (0.295 ± 0.046 g) did not differ from that of B16.F10 alone (0.244 ± 0.028 g) or B16.F10 plus Pam3CSK4 (0.250 ± 0.016 g).

The Matrigel model was also used to assess the frequency of mature blood vessels at the tumor periphery because the decreased tumor blood volume of Pam3CSK4-treated tumors (Fig. 2D) suggested mast cells could inhibit tumor angiogenesis. Matrigel plugs in which B16.F10 was grown with Pam3CSK4-activated BMMCs had 60% fewer vessels than those that contained melanoma cells together with BMMCs alone (Fig. 3D).

**FIGURE 1.** Pam3CSK4 treatment decreases B16.F10 tumor growth. C57BL/6 mice were injected s.c. with B16.F10 melanoma cells. Tumors were treated s.c. on days 4 and 9 post-tumor injection (arrows) with Pam3CSK4 or PBS control (n = 10 tumors per group). A, Tumor volume was monitored daily. **p = 0.004. B, After 14-d growth, overall cellularity of collagenase-disrupted tumors was assessed. Right panel depicts representative diluent PBS- or Pam3CSK4-injected tumors. **p = 0.038. C, Tumor-associated mast cells were examined in tissue sections stained with Alcian blue–safranin O (original magnification ×400). Adjacent tumor cell areas are indicated by black arrows. D, Mast cell-deficient KitW-sh/W-sh and C57BL/6 mice were injected s.c. with B16.F10 melanoma cells (n = 10 per group), and tumor volume was followed for 16 d. Pooled data from two independent experiments: *p = 0.026.
proliferation was unaffected by Pam3CSK4 treatment but was demonstrated TNF from mast cells was not critical for tumor inhibition. In vivo experiments have demonstrated IL-6 can induce melanoma growth inhibition (38–41), we assessed the effect of Pam3CSK4-activated BMMC supernatants on B16.F10 proliferation. In contrast, supernatants from IL-6–deficient mice. In contrast, tumor growth was decreased in a parallel group of wild-type C57BL/6 mice (Fig. 6B). We further confirmed the requirement for mast cell-secreted IL-6 using the Matrigel–B16.F10 model. Tumor growth was not inhibited by Pam3CSK4 treatment in the presence of IL-6–deficient mice. Furthermore, tumor blood volume and the number of mature erythrocyte-containing blood vessels were not decreased in the presence of Pam3CSK4-activated BMMC supernatants (Fig. 6C). The requirement of mast cell IL-6 in tumor growth inhibition (38–41), we assessed the effect of Pam3CSK4-activated IL-6–/−/BMMC supernatants on B16.F10 proliferation. In vitro, supernatants from Pam3CSK4-activated IL-6–/−/BMMCs did not significantly decrease B16.F10 cell proliferation (Fig. 6C).

TLR2-activated mast cells inhibit lung carcinoma growth

To ascertain whether the inhibitory effects of TLR2-activated mast cells are unique to melanoma or can be extended to other tumor types, we investigated the ability of Pam3CSK4-activated mast cells to inhibit the growth of the lung cancer model LLC1, at day 10, using the Matrigel–tumor model system in C57BL/6 mice. TLR2 expressing BMMCs significantly inhibited LLC1 growth in the presence of Pam3CSK4 (58%), whereas Pam3CSK4-activated TLR2-deficient BMMCs did not (Fig. 7A). In contrast, Pam3CSK4 or BMMCs alone had no effect on tumor growth (Fig. 7B) as weight

Elevated chemokine production by Pam3CSK4-activated mast cells suggested mast cell-mediated tumor-inhibitory effects might involve effector cell recruitment. Flow cytometry of disrupted tumors demonstrated that Pam3CSK4 treatment was associated with infiltrating CD45<sup>+</sup> leukocytes, including CD8<sup>+</sup>TCRβ<sup>+</sup> and NK.1.1<sup>+</sup> cell subsets. B16.F10 tumors grown in Matrigel also had a marked mononuclear cell infiltration, at day 10, in the presence of Pam3CSK4 and BMMCs (Supplemental Fig. 5). To determine the involvement of TLR2-activated mast cells in early NK and T cell recruitment, we performed flow cytometry of B16.F10 tumors in Matrigel, on day 4, in the presence or absence of Pam3CSK4 and/or BMMCs. Tumor growth was significantly decreased in the presence of BMMCs and Pam3CSK4 and was associated with significant local increases in both NK and T cells (Fig. 5B).

CCL3 induces recruitment of both mature NK cells (35) and activated CD8<sup>+</sup> T cells (36, 37). To determine whether TLR2-activated mast cells directly induce the migration of NK and CD8<sup>+</sup> T cells and specifically to assess the role of mast cell-derived CCL3, we performed in vitro chemotaxis assays of C57BL/6 splenocytes using supernatants generated from wild-type and CCL3<sup>−/−</sup> BMMCs (Fig. 5C). Pam3CSK4-activated BMMC supernatants induced significant chemotaxis of NK.1.1<sup>+</sup> NK cells and both CD3<sup>+</sup>CD8<sup>+</sup> and CD3<sup>+</sup>CD8<sup>+</sup> T cell subsets (all p < 0.05). In contrast, supernatants from mast cells that could not produce CCL3 did not induce migration of NK or T cell subsets, demonstrating a role for CCL3 in mediating mast cell-dependent effector cell recruitment.
of Matrigels containing LLC1 plus BMMCs (0.236 ± 0.028 g) did not differ from that of LLC1 alone (0.257 ± 0.036 g) or LLC1 plus Pam3CSK4 (0.241 ± 0.019 g). In a manner comparable with the B16.F10 melanoma, decreased LLC1 tumor growth in the presence of Pam3CSK4-activated mast cells was also associated with mononuclear cell infiltration (Fig. 7C) and decreased angiogenesis (Fig. 7D).

A TLR2/6 agonist can mediate mast cell-dependent tumor growth inhibition

To assess the broader applicability of mast cell-dependent antitumor responses to TLR-based immunotherapies, we evaluated the ability of mast cells to produce mediators in response to a panel of TLR agonists. These studies demonstrate that at doses similar to that used for Pam3CSK4 in this study (10 μg/ml), the synthetic diacylated lipopeptide FSL-1 (a TLR2/6 agonist) and LPS (a TLR4 agonist) induced mediator production at least 2-fold over media control, similar to that seen with Pam3CSK4. The synthetic dsRNA analogue polyinosinic-polycytidylic acid (a TLR3 agonist) and bacterial peptidoglycan (activates TLR2/6 and other receptors) failed to induce mediator profiles from BMMCs similar to Pam3CSK4 upon 24-h stimulation (Supplemental Table I). To investigate whether additional TLR2 agonists could mediate mast cell-dependent antitumor effects in vivo, we performed similar studies using the TLR2/6 agonist FSL-1. FSL-1 treatment significantly inhibited B16.F10 tumor growth in C57BL/6 mice but not in mast cell-deficient KitW-sh/W-sh mice (Fig. 7E). Mast cell dependence in the antitumor effect of FSL-1 was confirmed via restoration of the antitumor effects of FSL-1 in KitW-sh/W-sh mice locally reconstituted with C57BL/6 BMMCs (day 15 tumor volume: 476 ± 153 mm3 [PBS] versus 55 ± 27 mm3 [Pam3CSK4], n = 4).

Discussion

This study demonstrates a previously unrecognized mechanism for TLR2-based immunotherapy with a critical role for the mast cell. TLR2 expression on mast cells was crucial for mediating the inhibitory effects of Pam3CSK4 on melanoma growth, even though multiple other effector cells, and the melanoma cells themselves, are known to express TLR2. The KitW-sh/W-sh model provides a useful tool to study the role of mast cells as KitW-sh/W-sh mice are profoundly mast cell deficient in multiple anatomical sites, with only rare mast cells detected in the skin of adult animals (30, 42). Restoration of the tumor-inhibitory effect of Pam3CSK4 in KitW-sh/W-sh mice via local reconstitution of mast cells indicates that other potential differences between mast cell-deficient mice and controls (31) were not responsible for the lack of tumor growth inhibition by Pam3CSK4 in KitW-sh/W-sh mice. TLR2-mediated activation completely reversed the normal protumorigenic role of mast cells in this model and instead harnessed them to initiate complex tumor-inhibitory effects that include recruitment of immune effector cells and angiogenesis inhibition.
Although mast cells have been demonstrated to display antitumor activity via TNF-mediated cytotoxicity (17, 18), the direct effects of activated mast cells on tumor growth occurred independently of mast cell-mediated TNF cytotoxicity and apoptosis induction. Pam3CSK4 did not exhibit direct antitumor effects in vitro. However, decreased tumor cell proliferation via Pam3CSK4-activated C57BL/6 but not IL-6−/− mast cell supernatants demonstrates the tumor-inhibitory effects of TLR2-activated mast cells are likely to be, in part, mediated via the direct effects of mast cell-derived IL-6. The role of IL-6 in tumorigenesis is controversial and appears to be dependent on the stage of disease and tumor type (39). However, in vitro studies have demonstrated IL-6 is directly growth inhibitory.

**FIGURE 4.** Pam3CSK4-activated mast cells decrease B16.F10 tumor growth in a TNF-independent manner. A, TNF secretion was measured by ELISA of cell-free supernatants. A23187 ionophore induction of TNF was included as a positive control at the 6-h time point (n = three experiments/group). *p < 0.05; ***p < 0.001 (compared with media control). B, Annexin V–propidium iodide (PI) staining of B16.F10 cells cocultured with Pam3CSK4 and/or C57BL/6 BMMCs (n = 3 experiments/group). CPT was included as a positive control with DMSO vehicle control. ***p < 0.001. C, Mast cell-deficient KitW−/− mice were locally reconstituted with TNF-deficient BMMCs 2 wk prior to tumor injection then treated on days 4 and 9 (arrows) with Pam3CSK4 (n = 12 mice from two independent experiments per group). *p = 0.012. D, B16.F10 proliferation was assessed in the presence of increasing doses of Pam3CSK4 (pooled data from three independent experiments) and cell-free supernatants from C57BL6 BMMCs (pooled data from 12 independent experiments) treated for 24 h with or without Pam3CSK4. CPT was included as a positive control. Data are expressed as fold change over media control for BMMC supernatants and fold change over DMSO vehicle control for CPT. ***p < 0.0001. ns, not significant; UD, undetectable.

**FIGURE 5.** Pam3CSK4-activated mast cells recruit immune effector cells in a CCL3-dependent manner. A, IL-6, IL-13, CCL1, and CCL3 secretion was measured by ELISA of cell-free supernatants (n = 3 experiments). *p < 0.05; **p < 0.01. B, C57BL/6 mice were injected with Matrigel (0.15 ml) containing B16.F10 cells in the presence or absence or Pam3CSK4 and/or BMMCs derived from C57BL/6 mice (n = 6). After 4 d, Matrigels were removed and weighed and assessed by flow cytometry. *p < 0.05; **p < 0.01 (compared with B16.F10 alone). C, C57BL/6 splenocyte chemotaxis was assessed in response to supernatants from C57BL/6 or CCL3−/− BMMCs (n = 3 independent experiments for C57BL/6 BMMC and n = 2 independent experiments for CCL3−/− BMMCs). *p < 0.05.
for several tumor cell types, including melanoma (38, 43–45). The melanoma growth-inhibitory effects of IL-6 have also been confirmed in vivo (40, 41). Studies in melanoma cell lines, including B16.F10.9 cells, have demonstrated IL-6–induced tumor cell growth inhibition is enhanced in the presence of soluble IL-6R and occurs via a STAT3-dependent mechanism that involves upregulation of cyclin-dependent kinase inhibitors and induction of cell cycle arrest (46–48). In addition to direct effects of IL-6 on tumor growth, IL-6 may also function indirectly to inhibit tumor growth by modulating T cell effector function. In other models, elevated tumor IL-6 levels has been shown to enhance both CD4+ and CD8+ effector/regulatory T cell (Treg) ratios and induce significant tumor reduction in vivo (49). In the presence of large amounts of IL-6, activated mast cells may also modify the antitumor immune response by skewing Tregs into Th17 cells (50), which have been shown to mediate enhanced angiogenesis in melanomas (12). Further, our data from in vitro chemotaxis experiments using CCL3 demonstrated to be chemotactic for mature NK cells (35) and activated mast cells secrete large amounts of CCL3, which has been demonstrated with the early recruitment of significant numbers of both NK and T cell subsets. We have also identified multiple other cytokines and chemokines produced from Pam3CSK4-activated mast cells that may enhance effector cell recruitment and activation in vivo. For example, TLR2 ligand-activated BMMCs secrete substantial amounts of CCL1, which can induce migration of NK cells via CCR8 in other systems (55).

The tumor-inhibitory effects of Pam3CSK4 in vivo may also involve activation of other TLR2-expressing immune cells. Indeed, TLR2 engagement via Pam3CSK4 can enhance the proliferation, survival, and effector function of CD8+ T cells and lower their threshold of activation (56). However, the lack of Pam3CSK4-mediated antitumor effects in KitW-sh/W-sh mice and their restoration via wild-type, but not TLR2-deficient, mast cells demonstrates a critical role for mast cell TLR2 expression. Furthermore, the use of the Matrigel–tumor model system, containing only mast cells and tumor cells, demonstrates that although Pam3CSK4 may later act on other TLR2-expressing immune cells, the mast cell is both necessary and sufficient for initiating the antitumor effects after TLR2 immunotherapy. This is in keeping with the role of the mast cell as a sentinel cell in initiating immune responses after pathogen recognition (9).

Our studies of the LLC1 tumor model demonstrate that the mast cell–mediated antitumor effects of TLR2 immunotherapy are
not unique to melanoma and that aspects of this response may be extended to additional tumor subtypes. Mast cells are found in high numbers in lung carcinoma and many other solid tumors, where their presence correlates with increased angiogenesis (57). The marked mononuclear cell infiltration and decreased angiogenic response in Pam3CSK4-treated LLC1 tumors indicate a similar, multifaceted, mast cell-dependent mechanism of tumor growth inhibition to that observed in the context of the melanoma model. The TLR2/6 agonist FSL-1 also induced mast cell-dependent antitumor effects on B16.F10 tumor growth. This demonstrates that mast cell-dependent tumor growth inhibition after TLR2-targeted immunotherapy is not restricted to TLR2 agonists and may have broader implications to other TLR-based immunotherapeutic strategies.

The link between chronic inflammation and tumor growth and progression is well established (58). Mast cells normally play an integral role in promoting tumor growth for many solid tumors. Accordingly, it has been proposed that inhibition of mast cell function may be an attractive therapeutic modality in the treatment of solid tumors (16, 59). However, others have demonstrated mast cell infiltration is an independent marker of improved prognosis (19–21). The distribution of mast cells within solid tumors may be a crucial factor in their ability to provide a protective role against the growth of solid tumors. Indeed, the study by Welsh et al. (28) of non-small cell lung carcinoma and a more recent study by Johansson et al. (29) of prostate cancer have demonstrated that intratumoral but not peritumoral mast cells independently predict improved survival. In contrast, in a large study of 4444 breast tumors, stromal mast cells independently predicted improved survival (21). In a small-scale analysis, we did not observe significant differences in mast cell distribution after Pam3CSK4 treatment. However, an accurate comparison of mast cell distribution in the presence and absence of TLR2 agonist treatment was confounded by the small size of Pam3CSK4-treated tumors.

Overall, the results of the current study suggest TLR2-mediated activation of tumor-resident mast cells may represent a potential applicable immunotherapeutic strategy. The protumorigenic role of mast cells can be reversed by selective activation via TLR2 to effectively turn mast cells into triggers for antitumor effector mechanisms. In support of these findings, it has already been demonstrated that tumor incidence is decreased in the presence of mast cells in a TLR activator-rich intestinal microenvironment (60). Mast cells represent attractive targets for anticancer immunotherapy due to their abundance at the periphery of many solid tumors and ideal location in close proximity to blood vessels. In contrast with other immune cells, mast cells are relatively radioresistant (61), which makes them prime candidates for combined treatment modalities. Mast cell activation via TLR2 therefore provides a novel opportunity to modify the tumor microenvironment as part of a regimen of successful cancer immunotherapy.

To date, TLR2-based immunotherapy has been applied clinically for pancreatic cancer using the TLR2/6 agonist MALP-2 (3). Mast cell numbers are increased in pancreatic cancer and correlate with increased microvessel density (62). Thus, the clinical success observed by Schmidt et al. (3) may, in part, be attributable to the action of mast cells, such as reversal of their proangiogenic properties after TLR2-mediated activation or the recruitment of immune effector cells. Future clinical trials examining TLR2-based immunotherapy should examine the role of tumor-resident mast cells by assessing the correlation between mast cell density, distribution, and activation status and patient outcome. The results of the current study suggest TLR2 targeted immunotherapy may prove particularly effective at mast cell dense tissue sites including the skin and lung. Furthermore, it may be important during TLR2-based

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**FIGURE 7.** Mast cells decrease LLC1 tumor growth in a TLR2-dependent manner and are required for B16.F10 tumor growth inhibition by a TLR2/6 agonist. A, C57BL/6 mice were injected with Matrigel (0.3 ml) containing LLC1 cells in the presence or absence or Pam3CSK4 and/or BMMCs derived from C57BL/6 mouse or TLR2−/− mouse (pooled data from three independent experiments, n = 9 C57BL/6 BMMC, n = 13 TLR2−/− BMMC). Dashed line indicates weight of Matrigel with BMMCs plus Pam3CSK4 in the absence of tumor (0.092 ± 0.008 g, n = 8). **p = 0.008. B, Representative growth of LLC1 in Matrigels containing Pam3CSK4 and/or C57BL/6 BMMCs. C, H&E-stained sections were examined for mononuclear cell infiltrate (black arrows) among tumor cells (arrowheads). Original magnification ×400. D, Mature blood vessels were quantified at tumor periphery of H&E-stained sections. Data are pooled from two separate experiments (n = 7 mice). *p = 0.016. E, C57BL/6 mice or mast cell-deficient KitW−/−/W−/− mice were injected s.c. with B16.F10 cells. Tumors were treated s.c. on days 4 and 9 post-tumor injection (arrows) with FSL-1 or PBS diluent control. Data pooled from four independent experiments (n = 26 to 28 for C57BL/6, n = 16 for KitW−/−/W−/−). ***p < 0.001.

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immunotherapy to ensure that other medications that interfere with mast cell function or survival are not being used concurrently.

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Disclosures
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References


