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*J Immunol* published online 2 April 2010
http://www.jimmunol.org/content/early/2010/04/02/jimmunol.0902819

**Supplementary Material**
http://www.jimmunol.org/content/suppl/2010/04/02/jimmunol.0902819.DC1

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Selective Inhibition of JAK1 and JAK2 Is Efficacious in Rodent Models of Arthritis: Preclinical Characterization of INCB028050

Jordan S. Fridman,1 Peggy A. Scherle,1 Robert Collins,1 Timothy C. Burn, Yanlong Li, Jun Li, Maryanne B. Covington, Beth Thomas, Paul Collier, Margaret F. Favata, Xiaoming Wen, Jack Shi, Ryan McGee, Patrick J. Haley, Stacey Shepard, James D. Rodgers, Swamy Yeleswaram, Greg Hollis, Robert C. Newton, Brian Metcalf, Steven M. Friedman, and Kris Vaddi

Inhibiting signal transduction induced by inflammatory cytokines offers a new approach for the treatment of autoimmune diseases such as rheumatoid arthritis. Kinase inhibitors have shown promising oral disease-modifying antirheumatic drug potential with efficacy similar to anti-TNF biologics. Direct and indirect inhibition of the JAKs, with small molecule inhibitors like CP-690,550 and INCB018424 or neutralizing Abs, such as the anti-IL6 receptor Ab tocilizumab, have demonstrated rapid and sustained improvement in clinical measures of disease, consistent with their respective preclinical experiments. Therefore, it is of interest to identify optimized JAK inhibitors with unique profiles to maximize therapeutic opportunities. INCB028050 is a selective orally bioavailable JAK1/JAK2 inhibitor with nanomolar potency against JAK1 (5.9 nM) and JAK2 (5.7 nM). INCB028050 inhibits intracellular signaling of multiple proinflammatory cytokines including IL-6 and IL-23 at concentrations <50 nM. Significant efficacy, as assessed by improvements in clinical, histologic and radiographic signs of disease, was achieved in the rat adjuvant arthritis model with doses of INCB028050 providing partial and/or periodic inhibition of JAK1/JAK2 and no inhibition of JAK3. Diminution of inflammatory Th1 and Th17 associated cytokine mRNA levels was observed in the draining lymph nodes of treated rats. INCB028050 was also effective in multiple murine models of arthritis, with no evidence of suppression of humoral immunity or adverse hematologic effects. These data suggest that fractional inhibition of JAK1 and JAK2 is sufficient for significant activity in autoimmune disease models. Clinical evaluation of INCB028050 in RA is ongoing. The Journal of Immunology, 2010, 184: 000–000.

The pathogenic role of inflammatory cytokines in rheumatoid arthritis (RA) is well established (1). Multiple strategies have been pursued to affect their function using molecularly targeted biotherapeutics, such as those neutralizing TNF-α. More recently, Ab-mediated antagonism of IL-6 with tocilizumab has proven effective and bolsters the hypothesis that mitigating the presence and/or activity of cytokines can be efficacious (2). Preliminary clinical data with Abs targeted against IL-15 or IL-17 in RA are consistent with this concept (3, 4). Nonetheless, because of the complex etiology of RA, the potential redundancy of ILs, the inconvenience and risk of the dosing route, and limiting toxicities of current therapeutics, an unmet medical need clearly persists.

JAKs associate with various cytokine receptors and translate signals triggered by cytokine binding into intracellular responses (5). Cytokines that signal through JAKs include those associated with both adaptive and innate immune responses as well as inflammation (6). Multiple inflammatory cytokines signal through receptors that recruit two of the four JAK family members (JAK1, JAK2, JAK3, and Tyk2) into a signaling complex, and it has been proposed that inhibiting either or both JAKs in the signaling complex would effectively inhibit cytokine signaling (7). Theoretically, it is not necessary to inhibit all four JAK kinases to affect signaling from those cytokines associated with the pathogenesis of RA. In fact, doing so may be detrimental because of the fundamental roles that JAKs play in the immune system. JAK3 is expressed primarily in lymphocytes where it associates with essentially one cytokine receptor subunit, the common γ-subunit (γc), and conveys signals from six known cytokines, including IL-2 and IL-7 (8). Interestingly, mutations in γc, JAK3, or the IL-7 receptor (IL-7R) account for the majority of cases of SCID in humans. For the treatment of chronic inflammatory diseases, in which immune suppression is undesirable, we hypothesized that an attractive alternative to pan-JAK inhibition would be selective inhibition of JAK1 and JAK2. Conceptually, this strategy can antagonize the effects of inflammatory cytokines such as IL-6 while minimizing immune suppression.

The realization of therapeutic benefit from JAK inhibition in RA has now been demonstrated with the use of CP-690,550, a pan-JAK inhibitor originally believed to be an immune suppressive JAK3 inhibitor for use in organ transplant recipients (9). However, it is now clear that in addition to its effects on JAK3, this compound inhibits JAK1 and JAK2 at similar concentrations (10, 11). Moreover, tocilizumab has exhibited impressive clinical activity by antagonizing...
IL-6 signaling, a JAK1/2-dependent process (2, 12). Therefore, we proposed to test the therapeutic hypothesis that selective inhibition of JAK1/2 could be efficacious using a number of cellular and animal models relevant to RA and other autoimmune disorders. INCBO28050, a potent and selective inhibitor of JAK1/2, is shown to inhibit signaling from pathogenic cytokines such as IL-6 and IL-23. Inhibition of JAK signaling prevents the production of chemokines and cytokines associated with uncontrolled RA (13). Finally, in three complementary rodent models of arthritis, we show that selective and fractional inhibition of JAK1/2 is both sufficient for efficacy and well tolerated.

Materials andMethods

Biochemical assays

Enzyme assays were performed using a homogeneous time-resolved fluorescence assay with recombinant epitope tagged kinase domains (JAK1, 837–1142; JAK2, 828–1132; JAK3, 718–1124; Tyk2, 873–1187) or full-length enzyme (cMET and Chk2) and peptide substrate. Each enzyme reaction was performed with or without test compound (1-point dilution), JAK, cMET, or Chk2 enzyme, 500 nM (100 nM for Chk2) peptide, ATP (at the 4-fold concentration for each kinase or 1 nM), and 2.0% DMSO in assay buffer. The calculated IC50 value is the compound concentration required for inhibition of 50% of the fluorescent signal. Additional kinase assays were performed at Cerep (Redmond, WA) using standard conditions at 200 nM. Enzymes tested included: Abi, Akt1, AurA, AurB, CDK2, CDK4, Chk2, c-kit, EGFR, EphB4, ERK1, ERK2, FLT-1, HER2, IGFIR, IKKα, IKKβ, JNK, Lck, MEK1, p38α, p70S6K, PKA, PKCo, Src, and ZAP70.

Cellular assays

Human PBMCs were isolated by leukapheresis followed by Ficol-Hypaque centrifugation. For the determination of IL-6–mediated MCP-1 production, PBMCs were plated at 3.3 × 105 cells per well in RPMI 1640 + 10% FCS in the presence or absence of various concentrations of INCBO28050. Following preincubation with compound for 10 min at room temperature, cells were incubated by adding 10 ng/ml human recombinant IL-6 to each well. Cells were incubated for 48 h at 37°C. 5% CO2. Supernatants were harvested and analyzed by ELISA for levels of human MCP-1. The ability of INCBO28050 to inhibit IL-6–induced secretion of MCP-1 is reported as the concentration required for 50% inhibition (IC50). Proliferation of Ba/F3-TEL-JAK3 cells (a gift from Ross Levine, Memorial Sloan-Kettering Cancer Center, New York, NY) was performed over 3 d using Cell-Titer Glo (Promega, Madison, WI) following standard assay conditions.

For the determination of IL-23–induced IL-17 and IL-22, PBMCs were maintained in RPMI 1640 medium supplemented with 10% FBS, 2 mM l-glutamine, 100 μg/ml streptomycin, and 100 U/ml penicillin. T cells were activated by culturing with anti-CD3 and anti-CD28 Abs. After 2 d, the cells were washed and recultured with IL-23 (100 ng/ml), IL-2 (10 ng/ml) and various concentrations of INCBO28050. Cells were incubated for an additional 4 d at 37°C, then supernatants were collected, and secretion of IL-17 and IL-22 were measured by ELISA. The ability of INCBO28050 to inhibit IL-23–induced secretion of IL-17 and IL-22 is reported as the concentration required for 50% inhibition (IC50).

Phospho-STAT3 analysis

Isolated cells. For analysis of phospho-STAT3 in human PBMCs or PHA-stimulated T cells, cell extracts were prepared after 10–15 min preincubation with different concentrations of INCBO28050 and stimulation of cells for 15 min with IL-6 (100 ng/ml), IL-12 (20 ng/ml), or IL-23 (100 ng/ml). The extracts were then analyzed for phosphorylated STAT3 by using a phospho-STAT3–specific ELISA.

Whole blood. Blood drawn from rats was collected into heparinized tubes and then aliquoted into microfuge tubes (0.3 ml per sample). In stimulation experiments, INCBO28050 at various concentrations was added for 10 min prior to stimulation with human IL-6 (100 ng/ml) for 15 min at 37°C. RBCs were lysed using hypotonic conditions. WBCs were then quickly pelleted and lysed to make total cellular extracts; the extracts were analyzed for phosphorylated STAT3 by using a phospho-STAT3–specific ELISA. Blood from animals that were dosed with INCBO28050 was drawn at various times after INCBO28050 administration and processed as described above.

In vivo experiments

Animals were housed in a barrier facility accredited by the Association for Assessment and Accreditation of Laboratory Animal Care International. All of the procedures were conducted in accordance with the U.S. Public Health Service Policy on Humane Care and Use of Laboratory Animals and with Incyte Animal Care and Use Committee guidelines. Animals were fed standard rodent chow and provided with water ad libitum.

Pharmacokinetics. Female rats (n = 6 per gender per group) were given a dose of 10 mg/kg INCBO28050 suspended in 0.5% methylcellulose and given by oral gavage at 10 ml/kg. The first three rats were bled at 0 (pre-dose), 2, 8, and 24 h, and the second three rats were bled 1, 4, and 12 h after dosing. EDTA was used as the anticoagulant, and samples were centrifuged to obtain plasma. An analytical method for the quantification of INCBO28050 has been developed and used to analyze samples from toxicology studies. The method combines a protein precipitation extraction with 10% methanol in acetonitrile and LC/MS/MS analysis. The method has demonstrated a linear assay range 1–5000 nM using 0.1 ml of study samples. Data were processed using Analyst 1.3.1 (Applied Biosystems, Foster City, CA). A standard curve was determined from peak area ratio versus concentration using a weighted linear regression (1/x2).

Rat adjuvant-induced arthritis. Adjuvant-induced arthritis was elicited in rats according to established methods. Lewis rats (150–200 g, female; Charles River Laboratories, Wilmington, MA) are injected at the base of the tail with 100 μl of an emulsion of CFA (10 mg/ml Mycobacterium butyricum in incomplete Freund’s adjuvant). Rats exhibited signs of inflammation within 2 wk of the injection of CFA. Each rat paw was scored following visual observation using a rating of 0–3 (0 = normal: 1 = redness and minimal swelling of digits; 2 = moderate swelling of the digits and/or paw; 3 = severe swelling of digits and/or paw). Individual animal paw scores are combined and recorded as a sum of all four paws and groups means of these totals are reported. Percent inhibition in clinical score/severity is calculated using the following formula:

\[
\text{Vehicle – Treated)} / \text{Vehicle} \times 100
\]

In addition, a plethysmometer (Stoelting, Wood Dale, IL) was used to measure paw volumes taken at baseline and study termination. At the termination of the experiment, paws were removed from euthanized rats for histologic analyses. Treatment was initiated when significant signs of disease were noted, and the earliest signs of inflammation in individual paws would be equivalent—usually occurring 2 wk after adjuvant injection. Graphs reflect endpoints collected only immediately prior to and after therapy was initiated (treatment day 0). Groups consisted of six animals, and statistical differences between treatment and vehicle controls were assessed using two-tailed Student t tests or ANOVA with a Dunnett’s test when appropriate.

Collagen-induced arthritis. DBA/1j mice (4–5-wk old males) were purchased from The Jackson Laboratory (Bar Harbor, ME). The model was established as described with minor modifications (14). Mice are immunized intradermally with 100 μl bovine type II collagen solution (Chondrex, Redmond, WA; IMB11) in CFA (100 μl) in the base of the tail with 100 μl l-arthogenic anti-collagen Ab (ArthroMAB; Millipore, Bedford, MA). Rats exhibited signs of inflammation within 2 wk after adjuvant injection. The model was terminated when all animals had at least one affected paw and groups randomized to contain similar mean scores. Each group contained six animals. Anti-type II collagen Ab titers were determined using the Rheumera ELISA platform (no. 3000; Astarte Biologics, Foster City, CA). A standard curve was determined from peak area ratio versus concentration using a weighted linear regression (1/x2).
Histology and microcomputed tomography imaging. Ankles from rats and mice were fixed in 10% neutral buffered formalin and sent to Bolder BioPATH (Boulder, Colorado) for analysis. Following incubation in 5% formic acid decalifier (2–3 d for mice, 5–7 d for rats), tissues were processed for paraffin embedding and then stained with H&E (rats) or toluidine blue (mice). All tissues from all animals were examined microscopically by a board certified veterinary pathologist. Rat joints were given scores of 0–7 for inflammation and bone resorption. Measurements were taken from the dorsal skin surface (in flexion angle) to ventral skin surface (across the tarsal joints) to semiquantitate the inflammatory edema. Corrections were made for any artificial tissue separation. Scoring of mice was on a scale of 0–5 for inflammation, pannus, cartilage damage, and bone resorption. A four-joint mean score was determined for each parameter as were summed parameters for each animal, and these values were used to determine group means. Statistical analysis of histopathologic parameters were done by comparing group data using the Student t test with significance set at 5%.

Rat samples for microcomputed tomography (micro-CT) imaging were fixed in 10% neutral buffered formalin and sent to MB Preclinical Services (Ann Arbor, MI). Micro-CT images were obtained of each sample using a GE RS-100 (General Electric, Waukesha, WI) small animal micro-CT scanner. After each acquisition, the images were isosurface rendered, and images of each sample were generated using EVS Beam and MicroView 2.1.2 software (General Electric).

Real-time PCR. Transcript levels of cytokines were evaluated using real-time PCR. Primers and probes were synthesized and purified by Biosearch Technologies (Novato, CA) with the 18S rRNA levels and relative abundance levels calculated based on the median of JAKs (Table I) in a biologically relevant manner by performing these assays at ATP concentrations approximating those within cells. INCBO28050 was a potent inhibitor of JAK1 and JAK2 with IC50 values of 5.9 and 5.7 nM, respectively. In an attempt to mitigate the potentially immunosuppressive effects of JAK3 inhibition, INCBO28050 was designed using previously established structure-activity relationships to be JAK3 sparing (IC50 < 560 nM), as demonstrated by ∼100-fold selectivity over the enzymatic IC50 values for JAK1 and JAK2. INCBO28050 also demonstrated moderate (∼10-fold) selectivity against Tyk2 (IC50 = 53 nM) and marked selectivity over the unrelated c-Met (IC50 > 10,000 nM) and Chk2 (IC50 > 1,000 nM) kinases. Moreover, when tested at a concentration ∼100-fold the IC50 of JAK1 and JAK2 against a diverse panel of 28 kinases (see Materials and Methods for list), no significant inhibition was observed (17). Two structurally related compounds (INCBO27753 and INCBO29843) were found to have JAK1/2 potencies exceeding 200 nM (the highest concentration tested; Supplemental Table I) and were used to support the conclusion that the cellular activities of INCBO28050 described below were due to JAK1/2 inhibition.

In cell-based assays, INCBO28050 proved to be a potent inhibitor of JAK signaling and function. In PBMCs, INCBO28050 inhibited IL-6-stimulated phosphorylation of the canonical substrate STAT3 (pSTAT3) and subsequent production of the chemokin MCP-1 with

<table>
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<th>JAK1</th>
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<td>5.9 ± 0.9</td>
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<sup>a</sup>Highest concentration tested.

### Results

**Biochemical and cellular activity of INCBO28050**

To characterize the biochemical potency and selectivity of INCBO28050 within the JAK family of kinases, we assessed the ability of this compound to inhibit the enzymatic activity of the four JAK family members. Because INCBO28050 is an ATP competitive kinase inhibitor, we were able to characterize the relative selectivity between JAKs (Table I) in a biologically relevant manner by performing these assays at ATP concentrations approximating those within cells. INCBO28050 was a potent inhibitor of JAK1 and JAK2 with IC50 values of 5.9 and 5.7 nM, respectively. In an attempt to mitigate the potentially immunosuppressive effects of JAK3 inhibition, INCBO28050 was designed using previously established structure-activity relationships to be JAK3 sparing (IC50 < 560 nM), as demonstrated by ∼100-fold selectivity over the enzymatic IC50 values for JAK1 and JAK2. INCBO28050 also demonstrated moderate (∼10-fold) selectivity against Tyk2 (IC50 = 53 nM) and marked selectivity over the unrelated c-Met (IC50 > 10,000 nM) and Chk2 (IC50 > 1,000 nM) kinases. Moreover, when tested at a concentration ∼100-fold the IC50 of JAK1 and JAK2 against a diverse panel of 28 kinases (see Materials and Methods for list), no significant inhibition was observed (17). Two structurally related compounds (INCBO27753 and INCBO29843) were found to have JAK1/2 potencies exceeding 200 nM (the highest concentration tested; Supplemental Table I) and were used to support the conclusion that the cellular activities of INCBO28050 described below were due to JAK1/2 inhibition.

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lated by IL-23 (IC\textsubscript{50} = 20 nM; Fig. 1)

isolated naive T-cells, INCB028050 also inhibited pSTAT3 stimulation
with IC\textsubscript{50} values of 44 nM and 40 nM, respectively (Fig. 1)

Oral pharmacokinetics of INCB028050 in rats. Female rats

**FIGURE 2.** Oral pharmacokinetics of INCB028050 in rats. Female rats

were dosed orally with 10 mg/kg INCB028050 in methocellose (0.5%),

and anticoagulated blood samples were harvested prior to and 1, 2, 4, 8, 12,

and 24 h after dosing. Six animals were dosed, using three animals for each
time point. Blood samples were centrifuged and individual plasma samples
were analyzed for compound content using LC/MS/MS with average con-
centration shown on the graph. Error bars represent SE. For reference,
a hashed horizontal line was inserted at the whole blood IC\textsubscript{50} for inhibition of
IL-6–stimulated STAT3 phosphorylation.

IC\textsubscript{50} values of 44 nM and 40 nM, respectively (Fig. 1A, 1B). In
isolated naive T-cells, INCB028050 also inhibited pSTAT3 stimulation
by IL-23 (IC\textsubscript{50} = 20 nM; Fig. 1C). Importantly, this inhibition
prevented the production of two pathogenic cytokines (IL-17 and
IL-22) produced by Th17 cells—a subtype of helper T cells with

**Pharmacodynamic and pharmacokinetic characterization of
INCB028050**

Because plasma protein binding of kinase inhibitors can sometimes
be significant and may therefore impact their in vivo effectiveness,
we developed a whole blood assay (WBA) to better characterize
what might be considered pharmacodynamically active plasma
levels of INCB028050. Recombinant IL-6 was added to rat
whole blood following preincubation with INCB028050 at various
concentrations. The effects of selective JAK1/2 inhibition on IL-6
signaling were determined by analyzing the extent of phosphorylation
of STAT3 by ELISA. INCB028050 inhibited IL-6–stimulated phos-
phorylation of STAT3 in whole blood with an IC\textsubscript{50} of 128 nM (Fig.
1D). Compared with results obtained in PBMCs (IC\textsubscript{50} = 44 nM), these
data suggest that constituents in blood (e.g., plasma proteins) se-
quester nearly two thirds of the otherwise active drug.

Because of the essential role JAKs play in a normal physiology,
we were interested to determine whether modest or intermittent
inhibition of JAK1/2 would be effective. By integrating the WBA data
with the favorable rat pharmacokinetics (Fig. 2), it is possible to
calculate the approximate extent and duration of JAK1/2 inhibition
in these experiments, recognizing there are limitations to such
extrapolations. Making use of this modeling, an oral dose of 10 mg/kg is
expected to inhibit JAK1/2 signaling (by ≥50%) in rats for ∼8 h.
To determine whether inhibition of JAK3 was likely to occur at the
dose levels of INCB028050 used in these studies, we determined
the effect of INCB028050 on the proliferation of the cell line
shown elsewhere to depend on JAK3 activity—Ba/F3-TEL-JAK3
cells (18). We did so because some cytokines can signal through
interactions between JAK1 and JAK3, and this may be dependent on context.
INCB028050, at concentrations up to 10 µM, had no effect on
the proliferation of Ba/F3-TEL-JAK3 cells (Supplemental
Fig. 1). Because a dose of 10 mg/kg does not approach 10 µM, even
at its peak (Fig. 2), the doses used in this study are not expected to
inhibit JAK3 signaling.

**In vivo characterization of INCB028050 in a rat model of arthritis**

To assess the potential therapeutic utility of systemic JAK1/2
inhibition for the treatment of autoimmune arthritides, we used
multiple rodent models of arthritis. Although related data have been
published on a distinct compound (19), we evaluated the effects of a
novel JAK3-sparing inhibitor in animals with established disease,
a more strenuous measure of activity. Furthermore, we as-

cessed the effects of INCB028050 on both cellular and humoral
immunity. Initially, we used the rat adjuvant-induced model of arthritis (raIA)
beginning treatment in the therapeutic mode—that is, when rodents have established disease (see Materials and
Methods). This model results in T cell–dependent inflammatory
arthritis. A number of JAK-activating cytokines have been im-
planted in the pathobiology of this model, including IL-6 and
multiple cytokines associated with Th17 cells (20). Although the
IL-17 produced by pathogenic T cells does not necessarily signal
through JAKs, the differentiation to and maintenance of IL-17–
producing Th17 cells requires IL-6 and IL-23, both of which use
JAK1 and/or JAK2 (21, 22).

We assessed the efficacy following daily oral administration of
INCB028050 at doses of 1, 3, or 10 mg/kg based on its pharma-
ecokinetic profile in this species. Disease severity was assessed pe-

dioidically, scoring clinical signs of disease (see Materials and
Methods for details). These doses were based on the PK/PD
relationship established with the IL-6 WBA, with the goal of in-
hibiting JAK1/2 signaling by no more than 50% for half of the
day. Relative to vehicle-treated animals, increasing doses of
INCB028050 inhibited disease scores by 24% (p < 0.05), 57% (p <
0.01), and 81% (p < 0.01), respectively (Fig. 3A). Moreover, while the
observed changes in clinical scores at study termination were
impressive, suppression of clinical disease by INCB028050 was
observed as early as 2 d after initiation of treatment. On the last
day of treatment, paw volumes were measured by plethysmography
as an objective endpoint. INCB028050 treatment, compared with
vehicle, inhibited the increase in hind paw volumes during the 2 wk
of treatment by 50% at a dose of 1 mg/kg and >95% at doses of 3 or
10 mg/kg (Fig. 3B). Because baseline paw volume measurements
are taken on treatment day 0—in animals with significant signs of
disease—it is possible to have >100% inhibition in animals
showing marked improvement in swelling.

Blinded histologic analysis of the paws was performed on vehicle-
or INCB028050-treated rats, and two specific parameters were

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**INHIBITION OF JAK1 AND JAK2 IN RA**

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destruction (Fig. 3D, compare upper to middle). Consistent with the observed histologic effects, orally administered INCB028050 (10 mg/kg, daily for 2 wk) resulted in radiographic improvements with restoration of the normal architecture and appearance to the ankle and tarsals (Fig. 3D, compare middle to bottom). These data support the potential for JAK1/2 inhibitors such as INCB028050 to have disease-modifying antirheumatic drug (DMARD) potential, although clinical translation of such findings is by no means assured.

Finally, to determine the extent of JAK1/2 inhibition associated with the efficacy observed above, we determined the effects of the compound on JAK/STAT signaling in peripheral blood samples collected at various times prior to (24 h) or immediately after (1 and 4 h) the last oral dose (treatment day 14). The quantity of pSTAT3 is elevated 2–3-fold in the peripheral blood of arthritic rats, consistent with a robust and ongoing inflammatory reaction (data not shown). Levels of pSTAT3 were reduced by INCB028050 in a dose- and

FIGURE 3. Anti-inflammatory and DMARD activity of once daily INCB028050 in rats with established disease in the adjuvant arthritis model. Rats with demonstrable signs of inflammation were orally administered vehicle or increasing doses of INCB028050 (1, 3, or 10 mg/kg) once daily for 2 wk (n = 6 per group; see Materials and Methods for details). Improvements in clinical signs of disease (A) were noted as early as 2 d after initiating treatment and were maintained in a dose-dependent manner. Plethysmography (B) was used at study termination to measure changes in paw volumes, comparing compound-treated groups to vehicle-treated animals (no inhibition) and naive controls (100% inhibition). Blinded analysis of histologic sections of ankles and paws was consistent with clinical observations illustrating dose-related improvements in joint inflammation (C, left), ankle width (C, middle) and bone resorption (C, right). Data are presented as mean ± SE. Micro-CT imaging (D) of representative ankles from similarly treated animals showed clear evidence that INCB028050 (10 mg/kg/d) normalized the joint architecture and prevented bone destruction (white arrows).
time-dependent manner (Table II). Interestingly, efficacy was achieved in this rAIA experiment at doses that inhibit pSTAT3 levels by <15% 4 h after an oral dose (3 mg/kg). Moreover, even the maximally efficacious dose of 10 mg/kg resulted in no detectable pSTAT3 inhibition 24 h after administration. These data show that INCB028050 possesses significant activity in this aggressive pre-clinical model of arthritis in the absence of complete and continuous pathway inhibition.

In similar experiments, we explored the efficacy of continuously infused INCB028050 to better determine the average amount of JAK inhibition required for efficacy at steady-state in the rAIA model. To do so, we treated arthritic cohorts of rats using miniature osmotic pumps implanted on treatment day 0 when the mean clinical score was greater than 3 (~2 wk after adjuvant injection). Importantly, surgical implantation of pumps has no significant impact on the progression of the disease model (R. Collins and J. S. Fridman, unpublished data). Implanted pumps were set up to deliver vehicle or 0.18, 0.6, or 1.8 mg/kg/day of INCB028050. These doses resulted in mean plasma levels of 13, 53, and 181 nM, respectively. Based on the potency of INCB028050 in inhibiting pSTAT3 levels in IL-6–stimulated rat whole blood (IC50 ~ 130 nM; Fig. 1D), these plasma levels correlate with 14%, 36% and 62% inhibition of JAK/STAT signaling. However, these values are extrapolated and might not accurately reflect variables such as organ site-specific compound and inhibition levels. Administration of INCB028050 at these levels was efficacious as indicated by inhibition in clinical signs of disease of 25% (p < 0.05), 50% (p < 0.05), and 90% (p < 0.01; Table III) at 0.18, 0.6, or 1.8 mg/kg, respectively. Similar to oral dosing, the improvement in clinical signs of disease following continuous administration coincided with a reduction in paw swelling, as determined by plethysmography, by 63%, 96%, or ≥100% with increasing dose levels. A dose-dependent improvement in histologic evidence of disease, measured as inflammation and bone resorption scores, was also noted (Table III; p < 0.05 for middle and high doses; see Materials and Methods for details) in addition to improvements in radiologic endpoints (not shown).

Molecular analysis of draining lymph nodes from a satellite group of similarly diseased animals (treated orally as described above, 10 mg/kg once daily) was conducted to determine the potential contribution of Th1- and Th17-associated cytokines in this model. Whereas the former has been well established, the latter is a more nascent finding and, as mentioned previously, preliminary clinical data with IL-17Abs implies that antagonizing this signaling pathway may be advantageous. Both Th1- and Th17-associated cytokines were found to be elevated during the inflammatory phase of the rAIA model (day 14, Table IV). Observed increases ranged from an ~6-fold increase in expression of the Th1 cytokines IL-12 and IFNγ to a >100-fold increase in expression of the Th17 cytokines IL-17 and IL-22. Treatment with INCB028050 reduced the expression of these pathogenic cytokines by 55 to ~80%. These data are consistent with the in vitro cellular data showing that selective JAK1/2 inhibition mitigates signaling from multiple inflammatory cytokines as well as the production of inflammatory factors in response to cytokine stimulation (Fig. 1).

Effects of INCB028050 in murine models of humoral and cellular autoimmunity

The therapeutic potential of INCB028050 was also explored in mouse models of arthritis, which are largely driven by humoral mechanisms. The collagen-induced arthritis (CIA) model has been extensively used in the evaluation of novel therapeutics and recapitulates many of the clinical and histologic features of human RA. This complex model requires humoral and cellular immune responses for a robust inflammatory response. A number of cytokines have been implicated as major regulators in mouse CIA, including IL-6, IL-23, IL-17, TNF-α, and IL-1β (23–27). Indeed, circulating plasma levels of IL-6 correlate with disease severity in the CIA model, and genetic or biologic antagonism of IL-6 markedly suppresses or eliminates disease incidence and severity, at least in part, by suppressing a robust humoral response (23, 27). Similar experiments have demonstrated an important role for Th17 cells and associated cytokines (e.g., IL-17, IL-21, and IL-23) (20, 24–26).

In the murine CIA model, we assessed the effectiveness of INCB028050 in animals with established disease (i.e., therapeutic mode; Fig. 4A). Doses of 1, 3, or 10 mg/kg were administered twice daily for 15 d in animals with demonstrable signs of inflammation. This dosing schedule was used because the t1/2 of INCB028050 in mice is approximately one third that observed in rats (data not shown). Improvements in clinical signs of disease were noted as early as 4 d after initiation of dosing, and a dose-dependent reduction of clinical scores compared with vehicle controls—19% (p = 0.55), 67% (p < 0.0001, and 61% (p < 0.0001), respectively—was seen at study termination. This finding was not surprising based on the potent ability of INCB028050 to inhibit IL-6 and IL-23 signaling and function (Fig. 1), both of which are essential in the development of disease in the CIA model. However, because of the supportive effects of some JAK-activating cytokines on B cell proliferation and function, it was possible that the observed efficacy of INCB028050 in this model was due to suppression of a humoral response to collagen. To assess this experimentally, we determined the levels of anti-collagen Abs in serum samples from a separate cohort of control and treated mice. INCB028050 treatment administered at the doses used in this study in a therapeutic model did not suppress the production of anti-collagen Abs in these experiments (Fig. 4B; p = 0.4), indicating that the effects are not due to direct suppression of Ab responses. However, because INCB028050 was administered in therapeutic mode in these studies, we cannot rule out the possibility that earlier administration (preventative mode)

<table>
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<tr>
<th>Dose (mg/kg/d)</th>
<th>INCB028050 Plasma Concentration (nM)</th>
<th>pSTAT3 (% Inhibition)</th>
<th>Clinical Score (% Inhibition)</th>
<th>Paw Swelling (% Inhibition)</th>
<th>Histologic Improvement (%)</th>
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<tbody>
<tr>
<td>0.18</td>
<td>13 ± 2</td>
<td>14</td>
<td>25 ± 42</td>
<td>63 ± 15</td>
<td>29 ± 15</td>
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<tr>
<td>0.6</td>
<td>53 ± 8</td>
<td>36</td>
<td>50 ± 34</td>
<td>96 ± 36</td>
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<tr>
<td>1.8</td>
<td>181 ± 19</td>
<td>62</td>
<td>90 ± 18</td>
<td>≥100</td>
<td>60 ± 10</td>
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*Based on a body weight of 200 g.

*Extrapolated from Fig. 1D.
might have affected anti-collagen Ab titers. Paws from the vehicle control group and the 10 mg/kg INCB028050 group were analyzed for histologic signs of damage. INCB028050 treatment improved a composite score of joint damage by 47% (Fig. 4C; n = 6; p < 0.01). In addition, each individual parameter was significantly improved (p < 0.01, t test): inflammation (43%), pannus (53%), cartilage damage (41%), and bone damage (53%). Representative histlogic sections are shown (Fig. 4D) with arrows pointing to affected joints.

We also used an alternative model of arthritis that is dependent on Abs, but circumvents the need for the generation of humoral immune response—the collagen Ab-induced arthritis (CAIA) model (15). Arthritis in CAIA is induced by direct administration of a mixture of mAbs against various epitopes of type II collagen. Localization of collagen-specific Abs in the joints and subsequent local immune activation results in significant infiltration of polymorphonuclear and mononuclear cells leading to synovitis, pannus formation, and cartilage and bone destruction. Therefore, the CAIA model is suitable for measuring the effectiveness of therapeutic intervention subsequent to immune priming to auto-Ag. In this rapidly progressing model, we assessed the effects of twice daily oral INCB028050 administered beginning 3 d after inoculation of anti-collagen Abs. Doses of 1 and 10 mg/kg, twice daily resulted in a 13% (p > 0.05) and 56% (p < 0.05) improvement in clinical signs of disease, respectively (Fig. 5A; n = 5 per group). Histologic changes in vehicle and INCB028050 treated mice were also compared (Fig. 5B). Mice treated with vehicle all had mild to marked inflammation, minimal to moderate pannus, and damage to the cartilage and bone. Treatment with INCB028050 (10 mg/kg) reduced pannus (74%; p < 0.05) and bone damage (78%; p < 0.05) and improved cartilage damage (43%; p > 0.05) and signs of inflammation (33%; p > 0.05), resulting in a 53% improvement in an aggregate score of disease (Fig. 5C; p = 0.059). These data, with those from the CIA model, illustrate that JAK1/2 inhibition can be efficacious in preclinical RA models without affecting humoral immunity, implying an ability to affect cellular immune function.

To directly assess the effects of INCB028050 on cellular immune function, we performed a delayed-type hypersensitivity (DTH) experiment. DTH is a widely accepted measure of cellular immunity in which Ag-specific memory T cells become activated and migrate to subsequent sites of hapten challenge where they initiate an inflammatory response (reviewed in Ref. 28). In our studies, we used dinitrofluorobenzene as the hapten, applying it ventrally during the sensitization phase and to the ears (pinna) during the challenge phase. Because a twice-daily dose of 10 mg/kg INCB028050 proved efficacious in both the CIA and CAIA models, we investigated the effects of this dose on inflammation and immune activation in mouse ears (Fig. 6). INCB028050, compared with vehicle, inhibited the DTH response by 48% (p < 0.00001, t test). These data demonstrate that INCB028050 modulates cellular immunity, likely accounting for at least some of the observed in vivo activity.

**Hematological impact of selective JAK1/2 inhibition**

Whereas INCB028050 was clearly efficacious, it should also be recognized that JAK signaling is central to a number of fundamental processes, including the generation of RBCs (29, 30). As such, continuous absolute inhibition of JAK signaling is expected to be deleterious, as suggested by various knockout studies in mice and the reduced hemoglobin levels observed in nonhuman primates receiving the broad spectrum JAK inhibitor CP-690,550 (6, 9). Therefore, at the termination of the CAIA study, blood samples were analyzed to determine whether efficacious levels of JAK1/2 inhibition adversely affected hemoglobin, RBCs, total WBCs, or absolute neutrophil or lymphocyte counts (Fig. 7). No significant

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**FIGURE 4.** INCB028050 improves clinical and histologic signs of disease in the murine CIA model. DBA/1 male mice with clinical signs of joint inflammation were randomized to receive twice daily oral doses of vehicle or INCB028050 (1, 3, or 10 mg/kg; n = 8 per group). Treatment with the selective JAK1/2 inhibitor resulted in marked improvement in clinical signs of disease (A, p < 0.0001 for 3 and 10 mg/kg doses) without suppression of humoral immunity (p > 0.05), as demonstrated by a lack of suppression of anti-collagen Ab titers from serum (B, n = 5). Paws from the vehicle and 10 mg/kg dose groups were processed for histologic assessment of joint inflammation, pannus formation, and bone and cartilage destruction, and a composite score was determined (C, n = 6). INCB028050 inhibited histologic signs of disease by 47% (p < 0.01). Examples of front and hind paws from both groups with representative signs of damage are shown (D). Arrows point to affected joints and W identifies the wrist. Original magnification ×16.
differences were noted in any parameter, which is consistent with the hypothesis that periodic incomplete inhibition of JAK1 and JAK2 can be efficacious in the absence of suppressing normal bone marrow function \( (p > 0.05, t \text{ test}) \). Recognizing that any effect on hemoglobin and/or RBCs can be underestimated owing to the duration of treatment, we also analyzed reticulocytes, which are temporally more responsive, and found a similar lack of effect \( (p > 0.05, \text{t test}) \), consistent with a lack of myelosuppression at these otherwise efficacious and well-tolerated doses.

**Discussion**

INCB028050 is a potent, selective, orally bioavailable inhibitor of JAK1 and JAK2 with demonstrable selectivity against JAK3 and a broad panel of unrelated kinases. In cell-based assays relevant to autoimmune diseases such as RA and psoriasis, INCB028050 inhibited JAK signaling and function initiated by two clinically validated therapeutic targets (IL-6 and IL-23) at concentrations \( \leq 60 \text{nM} \) and did not significantly affect JAK3-dependent proliferation at concentrations up to 10 \( \mu \text{M} \) (Supplemental Fig. 1). Structurally similar compounds lacking JAK1/2 inhibitory activity were inactive in these cellular cytokine assays (Supplemental Table I) consistent with a central role for JAK1/2 in these models systems and the selectivity of INCB028050. However, we recognize that the absence of observable off-target activity with INCB028050 does not exclude the possibility that this compound impacts an untested enzyme, receptor, channel or transporter.

In vivo, INCB028050 was efficacious in the rAIA model when administered orally or by continuous infusion. In this aggressive, T cell-driven arthritis model periodic and/or fractional inhibition of JAK1/2 signaling by INCB028050 appears sufficient to achieve remarkable efficacy in animals with active signs of disease regardless of what endpoint is used, including radiographic improvements associated with DMARD activity. This conclusion is based on the oral efficacy of INCB028050 (Fig. 3) and the associated pharmacodynamic data (pSTAT3 inhibition; Table II), which takes into account any potential accumulation of drug during the experiment as well as any potentially active metabolites of INCB028050 produced during its elimination, although no data for either exist at this time (data not shown). Similarly, continuous infusion of INCB028050 is efficacious at steady-state plasma drug levels below the whole blood assay IC50 for JAK1/2 signaling (Table III; Fig. 1D). Plasma levels achieved by either route of administration were well below those expected to inhibit JAK3 based both on enzyme and cellular potency (Table I; Supplemental Fig. 1) supporting the hypothesis that JAK1/2 inhibition is sufficient for efficacy in this destructive preclinical model of arthritis. Moreover, these efficacy data were at least as impressive as those associated with selective inhibition of IL-17, using a receptor IgG1 Fc fusion protein, consistent with the ability of INCB028050 to affect signaling from multiple inflammatory pathways.
cytokines that signal through JAK1 and/or JAK2 (31). However, because a direct comparison of these modalities was not performed, quantitative comparisons are difficult.

In the murine CIA model, dependent on both B and T cells, oral INCB028050 was also efficacious. Interestingly, although IL-6 is reported to be important in both murine CIA and in RA patients, the effectiveness of INCB028050 in this setting may be due, at least in part, to its ability to affect IL-6 signaling to such a degree that it is anti-inflammatory without being immune suppressive, as assessed by quashing an established humoral responses. This hypothesis is based on reports that anti-collagen Ab titers were suppressed in both the IL-6 knockout mice and those treated with anti-IL-6R Abs (23, 27).

This implies that some degree of IL-6 signaling is central to mounting and/or maintaining a robust humoral response and that complete ablation of IL-6 signaling, whether through genetic recombination or Ab sequestration, significantly antagonizes humoral immunity. In contrast, INCB028050 did not suppress an ongoing humoral response to collagen, but was efficacious when administered in the therapeutic mode at doses affecting but not eradicating IL-6 signaling. Conceivably, the ability of INCB028050 to affect the IL-17/23 axis may have also contributed to the impressive responses in the absence of humoral suppression (Fig 1; Table IV). Mice with genetic disruption of IL-23p19, but not IL-12p35, are resistant to CIA, and this correlates with reduced production of IL-17, presumably from Th17 cells (25). In agreement with this finding, mice lacking IL-17 are also resistant to CIA but, similar to the IL-6 knockout mice, have a suppressed humoral response to Ag (26). Nonetheless, neutralization of IL-17 in the therapeutic mode also suppresses the incidence and severity of disease, but does not affect anti-collagen Ab titers (24). The differences in effects on humoral responses likely reflect the ability of IL-17 to contribute to a humoral response during the immune education phase, although this effect is not necessary for IL-17 antagonism to be effective. More exhaustive studies with JAK inhibitors during the various phases of humoral immunity will be required to definitively describe the effects of these kinases and INCB028050 on this biology. However, the ability of INCB028050 to inhibit a DTH response clearly demonstrates its capacity to modulate cellular immunity in a hypersensitivity setting and suggests that at least part of the observed efficacy with INCB028050 is due to this immune modulating capability.

The ability of INCB028050 to affect joint inflammation independently of suppressing an established humoral response was also addressed in the CAIA model, in which joint inflammation is induced by systemic administration of a mixture of anti-collagen II Abs (15). This model induces a polymorphonuclear and monocellular cell infiltrate and a robust inflammatory response, resulting in pannus formation and joint destruction. Interestingly, Kagari et al. (32) showed that IL-6 knockout mice were susceptible to CAIA, which is consistent with a dominant role for IL-6 in supporting a humoral response to autoantigens. In this model, INCB028050 suppressed both clinical and histologic signs of disease consistent with its ability to affect JAK activating pathogenic cytokines including, but not limited to, IL-6.

Because genetic data indicate that complete ablation of JAK1 or JAK2 in mice is lethal, and because decades of research define a fundamental role for these kinases in the signaling pathways for numerous important cytokines, it is unlikely that unremitting inhibition of their activity would be tolerated (reviewed in Ref. 6). In this study, we have demonstrated that a selective JAK1/2 inhibitor (INCB028050) is efficacious in three rodent models of arthritis without complete or constant inhibition of JAK signaling. Effective treatment paradigms in these models have been well tolerated as exemplified by the lack of effects of INCB028050 on a number of hematologic parameters and body weight (Fig. 3E and data not shown). Moreover, a related compound, INCB018424, has been well tolerated in RA and cancer patients suggesting that this mechanism is a viable therapeutic option for patients suffering from RA or other diseases in which JAK signaling is dysregulated (33, 34). These results warrant further clinical evaluation of selective JAK1/2 inhibitors such as INCB028050.
cytokine IL-23 is composed of IL-12Rbeta1 and a novel cytokine receptor subunit, IL-23R.


