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Antibodies Generated against Streptococci Protect in a Mouse Model of Disseminated Aspergillosis

Rebekah E. Wharton,*† Emily K. Stefanov,* R. Glenn King,* and John F. Kearney*

Invasive aspergillosis (IA) resulting from infection by Aspergillus fumigatus is a leading cause of death in immunosuppressed populations. There are limited therapeutic options for this disease and currently no vaccine. There is evidence that some anti–A. fumigatus mAbs can provide protection against IA. However, vaccine development has been impeded by a paucity of immunological targets on this organism demonstrated to provide protective responses. Sialylated oligosaccharide epitopes found on a variety of pathogens, including fungi and group B streptococci (GBS), are thought to be major virulence factors of these organisms facilitating pathogen attachment to host cells and modulating complement activation and phagocytosis. Because some of these oligosaccharide structures are conserved across kingdoms, we screened a panel of mAbs raised against GBS serotypes for reactivity to A. fumigatus. This approach revealed that SMB19, a GBSib type-specific mAb, reacts with A. fumigatus conidia and hyphae. The presence of this Ab in mice, as a result of passive or active immunization, or by enforced expression of the SMB19 H chain as a transgene, results in significant protection in both i.v. and airway-induced models of IA. This study demonstrates that some Abs generated against bacterial polysaccharides engage fungal pathogens and promote their clearance in vivo and thus provide rationale of alternative strategies for the development of vaccines or therapeutic mAbs against these organisms. The Journal of Immunology, 2015, 194: 4387–4396.

Fungal infections involving opportunistic pathogens have increased dramatically in the last 20 years. Although normally harmless, infection by these organisms results in severe diseases in immunocompromised individuals, including AIDS patients as well as those subjected to severe immunosuppressive regimens involved in transplantation or myeloablative chemotherapy. Aspergillus fumigatus, the causative agent of invasive aspergillosis (IA), is the most prevalent airborne, opportunistic fungal pathogen that causes life-threatening disease among immunosuppressed populations in medical centers worldwide. IA results in mortality rates ranging from 40 to 80%, and this disease, already a significant health problem, is likely to become more prevalent owing to the lack of effective therapies or vaccines (1). Compounding the serious nature of these infections are increasing rates of immunodeficiencies, overuse of antibiotics, and the emergence of fungicide-resistant strains.

Thus far, most new therapeutic efforts have been directed toward development of vaccines to induce T cell activation or the production of cytokines, which are thought to be helpful in clearing fungal infections (2, 3). However, active vaccination is problematic in the case of immunosuppressed individuals, particularly those with compromised T cell immunity. Although many fungal cell wall components elicit Ab responses, few of these induced Abs provide protection in fungal infection models (4, 5). Additionally, the observation that serum anti–A. fumigatus Ab does not correlate with clinical improvement and that that μMT mice are resistant to A. fumigatus infections (6) have had a negating effect on efforts to generate vaccine strategies to induce protective Ab responses. Although mAbs directed against β-glucans, components of fungal cell walls (7, 8), and to an undefined glycoprotein (9) have been shown to provide protection in A. fumigatus infection models, to our knowledge protection elicited by other Ab-associated A. fumigatus epitopes has not been reported. Additionally, passive Ab treatment alone or in combination with cell-mediated immunotherapy or antifungal reagents has the potential to provide effective therapy in those with impaired immunity or those about to undergo immunocompromising treatments. Despite the few studies that show certain Abs to fungal cell wall components, especially polysaccharides (PS), can provide protection (10), the lack of knowledge of the nature of crucial fungal targets and host effector mechanisms involved in protection by anti–A. fumigatus Abs has hampered the development of an effective anti–A. fumigatus vaccine. Previous attempts to develop vaccines against fungal infections have concentrated on the products made or released by the fungi themselves; however, some but not all of these components have low intrinsic antigenicity or the ability to dampen host responses (4, 5).

In this study we show that a mouse mAb to group B streptococcus (GBS) type Ib (GBSib), SMB19 (IgMκ), reactive with the oligosaccharide sialyl-lacto-N-tetraose (s-LNT) epitope, also binds to A. fumigatus conidia and hyphae and is protective in inhalation and i.v. models of aspergillosis. Because PS–tetanus toxoid conjugate vaccines, which provide protective Ab responses against infection of multiple GBS, have been developed and used successfully in human clinical trials for some years (11), the results we have obtained in these mouse models suggest that...
GBS-PS conjugate vaccines may be repurposed to develop successful vaccines against A. fumigatus and other clinically important human fungal pathogens.

Materials and Methods

Mice

Eight- to 12-wk-old C57BL/6J, B6,129P2-Tcrbeta10Mouse Tcrdelta10Mouse (TCRβ/β−/−), B6,129P2-Tg-VDJ3 Antibody (mAb), and Nεf/11,12 (p4p4/c5/c5−/−) mice were purchased from The Jackson Laboratory (Bar Harbor, ME) and bred and housed in pathogen-free animal facilities. VH J558 transgenic mice were generated using the rearranged VDJ from a J558 IgG3 bred and housed in pathogen-free animal facilities. VH SMB19 Tg mice were generated using the rearranged genomic DNA VDJ from the SMB19 hybridoma (13), National Center for Biotechnology Information accession sequence BN000872. Genomic DNA was amplified using the primers 5'-CGGCTTCAGTTATTCTAG-3' and 5'-GCTTCTAATTTCAG-CAACCCAC-3'. The PCR product was cloned into the previously described pCRII-CT vector (14) at NcoI and BamHI restriction sites and VH SMB19 Tg mice were generated as described previously for the VH81.1 Tg mouse (14).

Microorganisms and mAbs

A. fumigatus (strain 13073) and isolates of Aspergillus niger (strain IMI 31274) and Aspergillus flavus (strain CBC B5107), all purchased from the American Type Culture Collection, were cultured on potato dextrose agar (Fisher) plates at 37°C for 5 d and then harvested as described (15) in PBS, counted on a hemocytometer, and stored at 4°C. GBS (strain names in parenthesis), including GBS1b (H36B), GBS1a, GBSII (18RS21), and GBSIII (COH1), used in flow cytometry experiments and vaccinations were gifts from Dr. David Pritchard (University of Alabama at Birmingham). GBS used for vaccines were grown to log phase, washed three times, and fixed overnight in 1% paraformaldehyde at 4°C. mAbs specific for these GBS serotypes, α-1,3 glucan, and the anti-idiotypic Ab SMB26 against SMB29 are described, together with methods of purification and quality control, in Supplemental Table I.

Biocore analyses

Biocore T200 (University of Alabama at Birmingham Multidisciplinary Molecular Interaction Core) was used to assess the binding of SMB19 to GBS1b capsular PS. Purified biotinylated GBS1b capsular PS was captured on a streptavidin sensor chip for the Biocore T200. After washing in a running buffer containing 3 mM EDTA to chelate Ca2+, SMB19 was injected over the streptavidin-captured GBS1b capsular PS chip surface in the presence of 0.9 mM calcium chloride (red) or 3 mM EDTA (green) (see Fig. 2) in the running buffer at concentrations ranging from 0.68 to 50 μg/ml. Sensorgrams (binding curves) were obtained and displayed simultaneously.

Treatment of A. fumigatus with phospholipase C

A. fumigatus conidia were germinated for 18 h in RPMI 1640 in a petri dish at 37°C, harvested, and washed twice in PBS (20 min, 2000 rpm) and once in phosphatidylinositol phospholipase C (PI-PLC) enzyme buffer (50 mM Tris-HCl, 150 mM NaCl, 5 mM EDTA in dH2O [pH 7.4]). A. fumigatus were then resuspended in 1 ml enzyme buffer containing 0.4 U PI-PLC (MP Biomedicals, Solon, OH) and incubated for 60 min at 37°C. The A. fumigatus preparation was then washed twice in PBS and stained with 0.5 μg mAb for 30 min, followed by goat anti-mouse IgM FITC (SouthernBiotech, Birmingham, AL) and analyzed on a FACSCalibur (BD Biosciences, Mountain View, CA). The no PI-PLC group was treated the same except for the addition of the enzyme. Cleavage of GPI-linked Thy1 on mouse T cells was used as a control for the enzyme function.

Flow cytometry

For A. fumigatus flow cytometry staining, A. fumigatus conidia were cultured in 50 ml conical tubes for 12 h at 37°C in RPMI 1640 to form hyphae. Then, paraformaldehyde-fixed GBS or live resting A. fumigatus conidia or hyphae were blocked at 4°C in 1% BSA and then incubated with purified anti-GBS-type specific IgM mAbs. Polyclonal goat anti-mouse IgM coupled to Cy5 (SouthernBiotech) was used to detect binding by flow cytometry. All GBS type-specific mAbs were gifts from Dr. David Pritchard (University of Alabama at Birmingham). IgM binding to A. fumigatus was analyzed on a FACSCalibur (BD Biosciences) and FlowJo software. For SMB19 Tg mouse characterization flow cytometry, spleen, bone marrow, mesenteric lymph nodes, and peritoneal cavity lavage cells were removed from SMB19 Tg or C57BL/6J mice. Cells were counted, blocked with anti-FcγR (Ab93), and stained with the Abs listed below. PE anti-B220 (RA3-6B2), PE-Cy7 anti-B220 (RA3-6B2), PE anti-AA4.1, FITC anti-CD23 (B3B4), Pacific Blue goat anti-IgM, goat anti-CD23 (B3B4), allophycocyanin-Cy7 anti-CD19 (1D3), FITC anti-CD5 (53-7.3), PE anti-CD11b (M1/70), and Alexa Fluor 488 anti-CD21 (7G6) were purchased from BD Pharmingen (San Diego, CA). SMIB26, rat anti-SMB19 idiotypic hybridoma, was generated using previously described methods (16). Cell frequencies were analyzed on a BD LSR II flow cytometer.

Vaccinations

Eight- to 10-wk-old C57BL/6J mice were vaccinated i.v. with either 1 × 108 paraformaldehyde-fixed GBS1b, or GBSII or PBS vehicle. Six days after vaccination, mice were infected with a lethal dose of A. fumigatus conidia i.v. and monitored for survival. Mice were euthanized with CO2 when they appeared moribund according to University of Alabama at Birmingham Animal Resources Program provisions.

Intravenous infections with A. fumigatus

For experiments involving passive Ab transfer, C57BL/6J, TCRβ/β−/−, C3−/−, or C5-deficient mice were given 200 μg SMB19, SIBD2, or Ab1 and i.p. and then immediately infected i.v. with A. fumigatus conidia and monitored for survival. In experiments involving a neutropenic model of IA in wild-type mice, mice were given 200 μg anti-Ly6G (IA8 clone hybridoma; a gift from Dr. Thomas Malek) i.p. 1 d prior to infection with A. fumigatus, and in some experiments another 2 d post-infection with A. fumigatus. On day 0, mice were passively administered 200 μg SMB19 or SIBD2 i.p. and then immediately infected i.v. with A. fumigatus and monitored for survival. Endpoints for these experiments were gauged as mouse unresponsiveness to touch for > 2 h (this includes uncontrolled rolling over in the cage, but not responding when touched). Mice were euthanized with CO2 when they appeared moribund. In experiments involving IA in Tg mice, all SMB19 Tg, J558 Tg, or C57BL/6J mice were infected with 2 × 104 A. fumigatus conidia i.v. and monitored for survival. Mice were euthanized with CO2 when they appeared moribund.

Intratracheal infections with A. fumigatus

In experiments involving a neutropenic model of intratracheal (i.t.) A. fumigatus infection, C57BL/6J, SMB19 Tg, and J558 Tg mice were given 200 μg anti-Ly6G (IA8 clone hybridoma; a gift from Dr. Thomas Malek) i.p. 2 h prior to i.t. infection with 2 × 104 A. fumigatus. Nεf/11,12 (p4p4/c5−/−) mice were infected i.t. with 104 A. fumigatus and 50 μg SMB19 or control (No Ab) i.t. Intratracheal infections were performed as described in Kin et al. (15). Mice were monitored for survival. Mice were euthanized with CO2 when they appeared moribund.

Inhibition ELISA

A. fumigatus conidia were germinated in a Nunc 96-well MaxiSorp plate in DMEM supplemented with 10% FCS for 8 h at 37°C in a CO2 incubator, then dried overnight at 37°C. Wells were washed with PBS and blocked with 1% BSA. SMB19 (250 ng/ml) in 1% BSA was then added to each well along with 1 mg/ml s-LNT or LNT (Carbohydr, Berkshire, U.K.), 0.5 mg/ml GBS1b PS (gift from Dr. Dennis Kasper, Harvard University), or α-1,3 dextran (a gift from Dr. A. Janes) in Dulbecco’s PBS. After 2 h incubation at 37°C, the wells were washed with PBS and incubated with goat anti-mouse IgM–alkaline phosphatase (AP; SouthernBiotech) for 1 h. Plates were then washed with PBS, developed with AP substrate buffer (pH 9.8), stopped with 5 N NaOH, and the OD was measured at 405 nm on a microplate reader from MBG Labtech (Cary, NC).

GBS1b type–specific PS and GBSII type–specific PS ELISAs

Nunc 96-well MaxiSorp plates were coated with poly-L-lysine for 30 min at room temperature. Purified capsular PS from GBS1b or GBSII was diluted in Dulbecco’s PBS and added to each well and dried overnight at 37°C. Wells were washed three times with PBS and blocked with 1% BSA. Mouse anti-sera were diluted and added to each well, then incubated for 2 h at 37°C. AP goat anti-mouse IgM (SouthernBiotech) was used as the secondary Ab and plates were developed with AP buffer (pH 9.8). OD was read at 405 nm on a MBG Labtech microplate reader.
ELISA assays for detection of the SMB19 idiotype, α-1,3 dextran, and IgM

Costar 96-well plates were coated with 1 μg/ml (SMBi26) anti-idiotype Ab to SMB19, α-1,3 dextran, or anti-IgM* (RS3.1) in PBS overnight at 4°C. Plates were then washed with PBS three times and blocked with 1% BSA. Mouse sera were diluted in PBS, added to each well, and plates were incubated for 2 h at 37°C. AP goat anti-mouse IgM (SouthernBiotech) was used as the secondary Ab and plates were developed with AP buffer (pH 9.8). OD was read at 405 nm on a BMG Labtech microplate reader.

Immunofluorescence tissue section analysis

Spleens were collected from mice, frozen in OCT, and cryostat sections were fixed in acetone, frozen, and stained with Alexa Fluor 350 anti-MOMA-1, Alexa Fluor 647 anti-IgM* (RS3.1), Alexa Fluor 488 anti-SMB19 idiotype (SMBi26), and PE anti-CD4 (RM4-5, BD Pharmingen) and PE anti-CD8 (53-6.7, BD Pharmingen). Brain and kidney sections from SMB19 Tg, J558 Tg, μMT, and C57BL/6 mice were frozen and stained with Alexa Fluor 647 anti-Ly6G (1A8), Alexa Fluor 555 anti-CD11b (M1/70), Alexa Fluor 488 anti-β-1,3 glucan (a gift from Dr. M. Feldmesser), and biotin rabbit anti-laminin (NB300-144) (Novus Biologicals, Littleton, CO). Biotinylated anti-laminin was visualized with Alexa Fluor 350 streptavidin (BD Pharmingen).

Quantitative PCR analysis

Brain and kidney DNA was extracted from SMB19 Tg, J558 Tg, μMT, and WT C57BL/6 mice using the DNeasy blood and tissue kit (Qiagen, Hilden, Germany). Samples were processed and primers used as described previously (17) prior to analysis on a Bio-Rad iQ5 RT-PCR machine. Results are expressed as A. fumigatus conidial equivalents per gram tissue.

A. fumigatus microscopy analysis

A. fumigatus conidia were diluted in RPMI 1640 and seeded on glass slides. Slides contained resting A. fumigatus conidia or A. fumigatus that was germinated for 8 h and fixed in 95% ethanol at 4°C. Slides were washed with PBS, blocked with 1% BSA, stained with Alexa Fluor 555 SMB19 and Alexa Fluor 488 anti-β-1,3 glucan (clone 744, IgM, a gift from Dr. Marta Feldmesser) (1), and mounted with DAPI fluoromount. Slides were analyzed using a Leitz DMRB fluorescent microscope.

Cytokine analysis

Sera from SMB19 Tg, J558 Tg, μMT, and C57BL/6J mice were collected at various time points before and after infection with A. fumigatus. Sera were diluted 3-fold and processed using the Milliplex mouse cytokine/chemokine kit (Millipore). Prepared serum samples were run on the Luminex Magpix system and results analyzed on Milliplex Analyst software.

Statistical analysis

Statistical comparisons were performed using Prism 4.0 software (GraphPad Software). Data with three or more groups were analyzed by a one-way ANOVA test followed by post hoc analysis, whereas data with two groups were analyzed by a two-tailed unpaired t test to determine whether overall statistically significant differences existed. For survival curves, statistical analysis was performed using the Mantel–Cox Log-rank test. A p value <0.05 was considered statistically significant.

Results

mAbs to the GBSIb capsular PS bind A. fumigatus hyphae and conidia

We found in recent surveys that certain mAbs, which recognize components from a variety of bacterial species, also bind to A. fumigatus at multiple stages of its life cycle (15). In this analysis, we observed that a previously described mouse mAb, SMB19, bound to A. fumigatus conidia (0 h) and hyphae (12 h) of A. fumigatus as detected by flow cytometry (Fig. 1A) and fluorescence microscopy (15). This mAb has been shown previously to bind to a sialylated oligosaccharide, s-LNT, associated with the capsular PS of GBSIb (13). Other mAbs to members of this streptococcal group, including GBSIa (SIaE7), GBSII (SIIF5C4), GBSIII, (SIIV18), and desialylated GBSIb (Sldb2) (described in Supplemental Table 1), do not bind either A. fumigatus conidia or hyphae by flow cytometry (Fig. 1A), although all mAbs bind to their respective type-specific PS expressed by the cognate serotype of GBS bacteria (Fig. 1B). The binding of SMB19 was notably brightest at the tip of the germination tube and on outgrown hyphae (Fig. 1C). Similar SMB19 reactivity was observed with A. flavus and A. niger (Supplemental Fig. 1A, 1B).

Fine specificity and Ca2+-dependent binding of SMB19 to the target oligosaccharide

The terminal oligosaccharides that define the type-specific capsular PS of GBS are known and are shown in Fig. 2A for types Ia, II, and III. The terminal oligosaccharides that define the type-specific capsular PS of GBS are known and are shown in Fig. 2A for types Ia,
Ib, II, and III. SMB19 binding to its target is highly specific because the capsular PS expressed by GBSIb differs from that of GBSIa and the others by having a $\beta_1\rightarrow\beta_3$ instead of the more common $\beta_1\rightarrow\beta_4$ galactose–N-acetylglucosamine linkage (Fig. 2A). Additionally, SMB19 does not react with the desialylated form of GBSIb, which is associated with the oligosaccharide LNT (13). Inhibition of SMB19 binding to A. fumigatus with either s-LNT– or GBSIb-purified capsular PS, but not LNT or $\alpha$-1,3 dextran, confirms that SMB19 binds a similar epitope to that found on GBSIb (Fig. 2B, 2C). Additionally, surface plasmon resonance analysis was used to confirm previous observations (13) that the binding of SMB19 to the purified GBSIb capsular PS is highly dependent on the presence of calcium (Fig. 2E). Treatment of A. fumigatus hyphae with PI-PLC to cleave GPI-linked proteins ablated SMB19 binding whereas 1-21 (anti–$\alpha$-1,3 glucan) binding was not affected (Fig. 2D).

Passive transfer of purified SMB19 mAb and vaccination with GBSIb improves survival in a mouse model of disseminated IA

Because SMB19 binds to A. fumigatus conidia and hyphae, we determined whether this Ab would protect in a mouse model of disseminated IA. Purified SMB19 (anti-GBSIb), SibD2 (anti-desialylated GBSIb), or A16 (anti–$\alpha$-1,3 glucan) was passively administered i.p. to naive C57BL/6J mice immediately before infection with 23 LD50 A. fumigatus conidia i.v. Approximately 75% of mice that received SMB19, but only 10–30% of mice that received either SibD2 or A16, survived 60 d postinfection (Fig. 3A). There were no significant differences in protection between mice that received SibD2 or A16. Injection i.p. of SMB19 revealed that this IgM mAb has an in vivo half-life of $\sim$2 d (Fig. 3B), similar to that found for other IgM Abs. These results indicate that a single treatment with GBSIb PS-specific SMB19, but not SibD2 specific for the desialylated GBSIb PS.

Figure 2. SMB19 is highly specific for s-LNT, is calcium-dependent, and recognizes a PI-PLC–sensitive epitope expressed by A. fumigatus. (A) The SMB19-reactive oligosaccharide expressed by GBSIb differs from other GBS capsular PS by a $\beta_1\rightarrow\beta_3$ instead of the more common $\beta_1\rightarrow\beta_4$ linkage. Binding of SMB19 to A. fumigatus visualized by ELISA assay. A. fumigatus–coated plates were incubated with SMB19 (black), SMB19 plus s-LNT (green), or SMB19 plus LNT (red) followed by an AP-labeled secondary Ab. Data were analyzed by one-way ANOVA with a Dunnett posttest; overall ANOVA, $p = 0.0097$. **$p < 0.01$ by post hoc, pairwise comparisons with a Dunnett test. (B) ELISA plates were coated with A. fumigatus and incubated with SMB19 (black), SMB19 plus purified GBSIb PS (Ib PS, green), or SMB19 plus $\alpha$-1,3 dextran (red) and then with AP-labeled secondary Ab. SMB19 binding was analyzed by measuring the OD at 405 nm following development. Inhibition ELISAs were performed in triplicate and repeated twice with similar results. Data were analyzed by one-way ANOVA; overall ANOVA, $p = 0.0524$, $*p < 0.05$ by post hoc, pairwise comparisons with a Dunnett test. (D) Flow cytometric histograms of A. fumigatus hyphae treated with PI-PLC (shaded histograms) or buffer treated (open histograms) were stained with SMB19 (green), SibD2 (red), or the IgM anti–$\alpha$-1,3 glucan mAb 1-21 (blue), followed by secondary anti-mouse IgM-Cy5. (E) Surface plasmon resonance Biacore analysis of SMB19 binding in the presence or absence of Ca$^{2+}$. The vertical axis (RU) indicates mAb bound at multiple concentrations during 200 s of flow. Overlaid sensograms obtained with multiple concentration of SMB19 in the presence or absence of calcium are shown. DEX, $\alpha$-1,3 dextran; Ib PS, GBSIb-purified capsular PS.
or A16 specific for *A. fumigatus*–expressed α-1,3 gluclans, improves survival against disseminated IA.

The significant protection provided by passive administration of SMB19 suggested that anti-GBSIb responses, as a result of immunization with GBS-PS, would protect mice from *A. fumigatus* challenge. To determine whether the Ab response following vaccination is sufficient to replicate the protection observed in passive transfer experiments, and to determine whether a protective response is specific to the GBSIb-PS vaccine, C57BL/6J mice were vaccinated i.v. with GBSIb, GBSII, or administered PBS. Six days following vaccination, at the predetermined peak of the anti-GBS Ab response, mice were infected i.v. with a lethal dose of *A. fumigatus* conidia. PBS-treated and GBSII-immunized mice died within ∼15 d, whereas ∼50% of GBSIb-vaccinated mice survived 60 d (at which point the experiment was ended) (Fig. 3C). Thus, GBSIb vaccination significantly prolonged survival compared with PBS sham or GBSII-immunized controls, indicating that the protection induced is specifically associated with GBSIb vaccination. In GBSIb-immunized mice, there was an increase in GBSIb type–specific IgM Ab, in addition to that of the SMB19 idiotype. In contrast, mice vaccinated with GBSII had ∼100 μg/ml GBSII-reactive serum Ab, but very little GBSIb-specific or SMB19 idiotype+ Ab (Fig. 3D-F). These results indicate that immunization of mice with GBSIb, but not with GBSII, promotes survival in ∼50% of mice for 60 d after *A. fumigatus* infection. Furthermore, this protection is associated with increased levels of anti-GBSIb Abs and specifically SMB19 idiotype+ Ab.

### SMB19 Tg mice are significantly protected against disseminated IA

We have previously observed that passively administered IgM Ab has a limited half-life, and levels of Ab to GBSIb peaks at ∼7 d after immunization and then subsides. To circumvent the transient nature of the anti-PS response in mice, we used a Tg mouse expressing the SMB19 H chain (characterized in Supplemental Fig. 2). In this model, SMB19 idiotype+ Ab is constitutively maintained at ∼10-fold higher levels than in C57BL/6J mice. Additionally, previous studies using μMT mice, which lack functional B cells, suggested that B cells and Abs may increase the susceptibility of mice to *A. fumigatus* infection (6). To address these issues and further investigate the protective role of SMB19 in IA, we infected SMB19 IgH Tg (SMB19 Tg), μMT, J558 IgH Tg (J558 Tg), or C57BL/6J mice with a lethal dose of *A. fumigatus* conidia i.v. SMB19 Tg mice were highly protected compared with C57BL/6J, μMT, or J558 Tg mice (Fig. 4A). Interestingly, in contrast to previous reports, μMT mice rapidly succumbed to *A. fumigatus* infections at similar frequencies to C57BL/6J mice. Additionally, J558 Tg mice, which maintain high levels of Abs specific for α-1,3 glucan (Fig. 4D), a major component of the *A. fumigatus* cell wall, were not protected compared with C57BL/6J mice. Although C57BL/6J mice and J558 Tg mice both have similar levels of GBSIb PS-binding serum IgM as do SMB19 Tg mice, SMB19 idiotype Abs are significantly lower in these mice. As observed in the GBS vaccination studies described above, the increased levels of SMB19 idiotype-bearing IgM was significantly correlated with protection in this model of IA (Fig. 4B, 4C). In J558 Tg mice, the observed anti-GBSIb Abs likely arose from B cells bearing endogenous Ig gene rearrangements. As expected, μMT mice did not have detectable levels of serum Igs. These results, in conjunction with the protection afforded by purified SMB19 mAb, further indicate that not all polyclonal GBSIb-reactive Abs provide protection in this model of IA (Fig. 4B). Instead, a particular subset of Abs, including those of the SMB19 idiotype, is responsible for the protection observed. Importantly, J558 Tg mice, which express Abs to a major *A. fumigatus* cell wall...
component, are as susceptible as C57BL/6J and μMT mice to A. fumigatus infection.

Because cytokines have been associated with both enhanced susceptibility or resistance to IA, we analyzed cytokine levels in the sera of each of these groups of mice immediately before and at 4 and 7 d after infection with A. fumigatus. SMB19 Tg mice had significantly lower levels of granulocyte proliferation and survival cytokines G-CSF and GM-CSF, which have been previously associated with protection against IA. Additionally, none of the groups of mice analyzed had significantly different levels of Th1 or Th2 cytokines previously associated with protection or susceptibility to IA, including IL-4, IL-5, IL-10, IL-2, and IFN-γ (Supplemental Fig. 3). Taken together, these data show that Ab-mediated protection in this model of IA differs mechanistically from protection reported in other systems.

**SMB19 Tg mice survival correlates with decreased A. fumigatus fungal burden and neutrophil infiltrates in the brain and kidneys**

Because SMB19 Tg mice demonstrated significantly higher survival rates following challenge with A. fumigatus than did WT C57BL/6J, μMT, or J558 Tg mice, we sought to determine whether this protection correlated with decreased fungal burden in the tissues affected during A. fumigatus pathogenesis. It has previously been reported that brain and kidneys are the major tissues damaged during this A. fumigatus infection model (17); therefore, we analyzed both tissues by immunofluorescence and quantitative PCR following A. fumigatus infection. Immunofluorescence analysis revealed that SMB19 Tg mice had significantly fewer A. fumigatus lesions in brain (Fig. 5A–E) and kidneys (Fig. 5G–K) compared with C57BL/6J, μMT, or J558 Tg mice 4 d after i.v. infection. There were no detectable A. fumigatus–containing lesions in brain or kidney tissue sections from mice that survived 60 d post-infection. Furthermore, immunofluorescence analysis showed that A. fumigatus–containing lesions in the brains of C57BL/6J and μMT mice were filled with leukocyte infiltrates, including CD11b+ and Ly6G+ neutrophils. In contrast, A. fumigatus lesions were undetectable in tissues from most SMB19 Tg mice 4 d postinfection. The few lesions observed were surrounded by cells weakly expressing CD11b but negative for Ly6G (Fig. 5D, 5J). None of the SMB19 Tg mice had visible lesions of A. fumigatus in liver or spleen (data not shown). To more accurately access the global fungal burden within these mice, quantitative PCR for A. fumigatus genomic DNA was performed. This analysis confirmed that SMB19 Tg mice had less fungal burden in their brain (Fig. 5F) and kidney (Fig. 5L) than did WT C57BL/6J, μMT, and J558 Tg mice. These data show that the survival of SMB19 Tg mice infected with A. fumigatus correlates with a decreased fungal burden and fewer neutrophil infiltrates in tissues known to be damaged as a result of fungal outgrowth following i.v. infection of mice with A. fumigatus.

**Neutrophil-depleted SMB19 Tg mice and Ncf1m1J/J mice administered SMB19 i.t. are protected after i.t. A. fumigatus infection**

Although i.v. infection with A. fumigatus conidia may rarely occur, the most common route of disseminated infection in immunodeficient patients is by the inhalation of conidia. Because neutrophil insufficiency as a result of myeloablation is a major risk factor for susceptibility to IA, we used a model of infection that renders
mice transiently neutropenic. SMB19 Tg, J558 Tg, and WT C57BL/6J mice were given one i.p. injection of neutrophil-depleting anti-Ly6G mAb 2 h before i.t. infection with *A. fumigatus* This dose of anti-Ly6G is sufficient to maintain neutropenia during the course of 3–4 d after injection. After mice were confirmed to be neutropenic (Fig. 6A), these three groups of mice were infected i.t. with 2.5 × 10⁷ *A. fumigatus* conidia and their survival was monitored. As seen in Fig. 6B, SMB19 Tg mice were highly protected against infection compared with the C57BL/6J and J558 Tg mice. This reduced susceptibility of SMB19 Tg mice to IA is similar to that observed in the i.v. model (Fig. 4). In a separate model we used *Ncf1<sup>m<sup>i</sup>/J* mice, which harbor a NADPH oxidase mutation resulting in an impaired intracellular killing mechanism rendering these mice highly susceptible to *A. fumigatus* infection by inhalation. *Ncf1<sup>m<sup>i</sup>/J* mice were challenged i.t. with 1 × 10⁵ *A. fumigatus* conidia alone (black) or in combination with 50 μg SMB19 (green) and monitored for survival. As shown in Fig. 6C, 80% of mice treated with SMB19 survived 11 d postinfection, a time when all control mice had succumbed to infection. Additionally, 20% of Ab-treated mice survived through day 20. The results of these experiments differ from the i.v. model of IA in which passive SMB19 administration failed to significantly protect transiently neutropenic mice. In this case, mice passively immunized with SMB19 died at the same rate as unimmunized C57BL/6J mice within the first 7 d. These observations indicate that SMB19’s mechanism of protection in the i.v. IA model is at least partially dependent on neutrophil activity (Supplemental Fig. 1C). Protection evident after 7 d may be a result of the combined activity of SMB19 Ab with recovering neutrophils because there was still ∼25 μg/ml SMB19 present in sera of passively transferred mice at this time (Fig. 3B). To determine the role of complement components in the protection
observed by passive immunization with SMB19, we passively transferred SMB19 or isotype control mAbs i.p. to C3−/− mice then immediately infected them i.v. with a lethal dose of *A. fumigatus* conidia. There were no differences in survival of C3−/− mice that received SMB19 or isotype control mAbs (Supplemental Fig. 1D). C5−/− mice treated with SMB19 were as resistant to *A. fumigatus* infection as were SMB19-treated C57BL/6J mice (data not shown). These data suggest that early complement components are involved in SMB19-mediated protection in mice, likely by promoting opsonophagocytosis of *A. fumigatus* and not via formation of membrane attack complexes. Because large volumes of data suggest that T cells are necessary for long-term survival of *A. fumigatus*-infected mice, we next analyzed the role of T cells in SMB19-mediated protection against i.v infection. SMB19 or isotype control Ab was administered i.p. to TCRβ−/− mice immediately before i.v. infection with 2 × 10^6 *A. fumigatus* conidia. TCRβ−/− mice that received SMB19 mAb are more resistant to infection than are the mice that received isotype control mAb (Supplemental Fig. 1E). The degree of protection observed in these animals is similar to that of C57BL/6J mice that received SMB19 mAb (Fig. 3A). Collectively, these results show that SMB19, when maintained at relatively high levels, endogenously in the Tg mice or passively introduced, provides substantial protection against infection following inhalation of *A. fumigatus* conidia. These findings contrast with those in the i.v model, which indicate that SMB19-mediated protection is dependent on neutrophil and complement activity, but not the activity of T cells.

**Discussion**

Abs and B cells historically have not been associated with protection in IA due to the lack of correlation between *A. fumigatus* Ab titers and IA protection, and partially due to a study demonstrating that μMT mice, which do not make Abs, were less susceptible in comparison with IA than were WT mice (6). However, recent studies on the role of mAbs in protection against *A. fumigatus*, especially those against β-1,3 glucans (7), have generated new interest in this field. Few studies, however, have identified new epitopes on *A. fumigatus* that could be targeted therapeutically with mAbs. Our results show that SMB19, an mAb generated against GBSIb, also bound resting *A. fumigatus* hyphae and, as such, is the target site for certain antifungal agents. The observation that binding of SMB19 to its oligosaccharide Ag is calcium-dependent may also have some bearing on SMB19 observation that binding of SMB19 to its oligosaccharide Ag disrupts tip morphology. It is possible that SMB19 preferentially binding to *A. fumigatus* hyphae tips is facilitated by high levels of calcium flux and contributes to the unique protective properties of this Ab.

Passive administration of SMB19 results in significant protection in the i.v. model of infection, in contrast to treatment with A16, *closely linked* to the nascent cell wall of growing hyphae is especially vulnerable to external influences and, as such, is the target site for certain antifungal agents. The observation that binding of SMB19 to its oligosaccharide Ag is calcium-dependent may also have some bearing on SMB19 binding to *A. fumigatus*, because proper outgrowth of fungal hyphal tips relies on high cytoplasmic calcium gradients (18). Dysregulation of these gradients inhibits hyphal growth and disrupts tip morphology. It is possible that SMB19 preferentially binding to *A. fumigatus* hyphae tips is facilitated by high levels of calcium flux and contributes to the unique protective properties of this Ab.

This mAb binds an epitope distinct from the sialic acid–containing capsular PS found on other GBS serotypes by its preference for the GBSIb type–specific oligosaccharide containing a β1→3 instead of the more common β1→4 galactose–N-acetylglucosamine linkage characteristic of the other subtypes. The fact that SMB19 bound preferentially to the tips of *A. fumigatus* hyphae is particularly interesting because the nascent cell wall of growing hyphae is especially vulnerable to external influences and, as such, is the target site for certain antifungal agents. The observation that binding of SMB19 to its oligosaccharide Ag is calcium-dependent may also have some bearing on SMB19 binding to *A. fumigatus*, because proper outgrowth of fungal hyphal tips relies on high cytoplasmic calcium gradients (18). Dysregulation of these gradients inhibits hyphal growth and disrupts tip morphology. It is possible that SMB19 preferentially binding to *A. fumigatus* hyphae tips is facilitated by high levels of calcium flux and contributes to the unique protective properties of this Ab.

**Abs PROTECT AGAINST ASPERGILLOSIS**
which binds α-1,3 glucan, a major cell wall component of *A. fumigatus*. Furthermore, C57BL/6J mice vaccinated with GBSb, which induces moderate increases of SMB19 idiotype Ab, were protected to a greater extent than were the mice that received passive Ab. Finally, SMB19 Tg mice, which maintain constitutively high levels of SMB19 idiotype Ab, are nearly as protected in longed. SMB19 coadministration with conidia was nearly as protective in co-infected J558 Tg mice, this protection was highly effective and prolonged. In the case of the J558 Tg mice, which maintain high titers of Abs against α-1,3 glucans, a major component of the *A. fumigatus* cell wall, are not protected in this model of *A. fumigatus*. This observation further highlights the unique role of SMB19 idiotype-bearing Abs and its target Ag in mediating the observed protection.

Survival of i.v. infected SMB19 Tg mice correlates with decreased fungal load, neutrophil infiltrates, and abscesses containing actively growing hyphae in the brain and kidneys when compared with infected J558 Tg, C57BL/6J, and μMT mice. Rarely, small areas in the brain of SMB19 Tg mice 4 d postinfection contained what appeared to be *A. fumigatus* remnants surrounded by MAC1+ and Ly6G+ cells. These cells could be resident active brain cells such as microglia. This observation suggests that some *A. fumigatus* conidia penetrated the blood brain barrier and germinated into the hyphal form in SMB19 Tg mice, but their growth was eventually halted by SMB19-mediated killing leaving only nonmetabolizing hyphal remnants as shown in Fig. 5. Additionally, C3-/- mice, in contrast to C57BL/6J mice, are not protected from i.v. induced IA after passive administration of SMB19. This indicates that complement activation is, at least in part, responsible for the therapeutic effect of SMB19 Ab. Also of interest is that, contrary to previous studies on C5 in IA, our C5-deficient mice were no more susceptible to IA than were WT mice. In this study, we used C5-deficient mice on the C57BL/6J background compared with WT C57BL/6J mice, whereas in the previous study DBA/2N mice, which are deficient in C5, were compared with CFW mice (19). Differences in these mouse strains could possibly contribute to the differences in survival of *A. fumigatus*-infected mice in these studies, which were attributed to the loss of complement components. Many studies have implicated T cell immunity as being particularly important in clearing *A. fumigatus* following infection. We have shown in the present study that SMB19 Abs do not depend on T cells for the immediate protection observed in this model of acute *A. fumigatus* infection.

Our studies of *A. fumigatus* infections during transiently induced neutropenia indicate that neutrophils are important for protection against IA afforded by SMB19 Abs in mice challenged i.v. Interestingly, however, in two models where neutrophils were transiently depleted or were severely impaired in their bactericidal capacity, SMB19 Abs were highly protective against infection when conidia were administered into the lung. In the case of the SMB19 Tg mice, this protection was highly effective and prolonged. SMB19 coadministration with conidia was nearly as protective in Ncf1m1J/J mice, which harbor a defect in NADPH oxidase activity, during the first 11 d following infection. Although many of the SMB19-treated mice eventually succumbed to infection, 20% of these mice survived at least 20 d. These results suggest that if the level of SMB19-like Abs is maintained, it will protect against lethal infection in mice that have severely impaired neutrophil numbers or function, which is commonly associated with aspergillosis in humans. These results also suggest that the protective effects of SMB19-like Abs may be provided by different mechanisms in response to airway or parenteral *A. fumigatus* infection.

There are several additional strategies to pursue regarding the development of SMB19-like Abs as a potential active or passive therapeutic for patients at risk for IA. Passive Ab could be combined with other antifungal therapies such as amphoterin B, voriconazole, and caspofungin, which are administered as standard treatments for IA patients. Amphoterin B and voriconazole target ergosterol synthesis on the *A. fumigatus* membrane, whereas caspofungin targets β-1,3 β-glucan synthase enzymatic activity, which also ultimately affects *A. fumigatus* membrane synthesis. A recent study showed that mAbs against β-1,3 glucans had a modest inhibitory effect on fungal growth in vitro. In limited studies, we found that SMB19 did not appear to have a direct effect on in vitro growth of *A. fumigatus*.

In patients with the capacity to mount an Ab response, active immunization is feasible. Additionally, GBS-PS conjugate vaccines against GBS serotypes I–V have already been developed and validated through phase II clinical trials in humans (11). In preliminary studies of sera from humans immunized with these GBSb conjugate vaccines, we have found that there are both IgM and IgG Abs that bind in a similar pattern to *A. fumigatus* conidia and hyphal tips as does SMB19. Furthermore, sera from individuals who received the GBSb vaccine inhibited SMB19 binding to *A. fumigatus*. These results indicate that GBS conjugates could be repurposed for use as antifungal vaccines in at-risk individuals. Our preliminary studies show that SMB19 Ab and these human GBS immune sera also show intense staining of *Candida albicans* and *Rhizopus oryzae* during their hyphal growth stages, as well as germinating *Fusarium* spp. macroconidia, indicating that Abs derived from GBS immunization recognize a common epitope on multiple fungi. Future research will determine whether the induction of SMB19-like Abs of IgM and IgG isotypes induced by GBS conjugate vaccines are “globally” protective against *A. fumigatus* and other clinically relevant fungi (20, 21).

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**Disclosures**

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