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ADAM17-Mediated Shedding of FcγRIIIA on Human NK Cells: Identification of the Cleavage Site and Relationship with Activation

Laurie Lajoie,*† Nicolas Congy-Jolivet,*† Armelle Bolzec,* Valérie Gouilleux-Gruart,*† Elodie Sicard,* Hsueh Cheng Sung,* Frank Peiretti,‡ Thierry Moreau,§ Henri Vié,¶ Beatrice Clementeau,¶ and Gilles Thibault*,†

FcγRIIIA/CD16A, the low-affinity receptor for the IgG Fc portion expressed on human CD56dim NK cells and involved in Ab-dependent cell cytotoxicity, is shed upon NK cell activation. We found that recombinant a disintegrin and metalloprotease (ADAM) 17 cleaved the ectodomain of FcγRIIIA/CD16A and a peptide for which the sequence encompasses aa 191–201 of the FcγRIIIA/CD16A stalk region but not ADAM10. MALDI-TOF analysis revealed that the peptide was cleaved between Ala195 and Val196 (i.e., 1 aa upstream of the expected position). This location of the cleavage site was confirmed by the finding that ADAM17 failed to cleave a peptide in which Ala and Val were reversed. ADAM17 was found to be expressed on NK cells, and stimulation with PMA or N-ethyl-maleimide resulted in the shedding of FcγRIIIA/CD16A and CD62L, a specific substrate of ADAM17. Selective inhibition of ADAM17 prevented the shedding of both molecules. Moreover, the shedding of FcγRIIIA/CD16A was strongly correlated with degranulation when a wide range of CD56dim NK cell activating receptors were stimulated, whereas both ADAM17-dependent shedding and internalization were involved in FcγRIIIA/CD16A downmodulation when the latter was engaged. Finally, the shedding of FcγRIIIA/CD16A was restricted to activated cells, suggesting that ADAM17 acts mainly, if not exclusively, in cis. Taken together, our results demonstrated for the first time, to our knowledge, at the molecular level that ADAM17 cleaves the stalk region of FcγRIIIA/CD16A and identified its cleavage site. The shedding of FcγRIIIA/CD16A was at least partially ADAM17 dependent, and it may be considered as a marker of FcγRIIIA/CD16A-independent NK cell activation highly correlated with degranulation. The Journal of Immunology, 2014, 192: 741–751.

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Abbreviations used in this article: ADAM, a disintegrin and metalloprotease; AR, activating receptor; Cal, calcium ionophore; DABCYL, 4-(4-dimethylaminophenyl) diazenylbenzoic acid; DNAM, DNAX accessory molecule-1; EDANS, S-(2-aminoethyl) (aminooxidopropionate); CD16/RIIIA, CD16/CD16A, the low-affinity receptor for the IgG Fc portion expressed on human CD56dim NK cells, monocytes, dendritic cells and rare T cells (1) and involved in Ab-dependent cell cytotoxicity, FcγRIIIA/CD16A, is shed upon NK cell activation.

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The Journal of Immunology
ADAM17 and ADAM10 are the main sheddases of FcγRIIIB/CD16B when cells are stimulated by PMA and calcium ionophore (CaI), respectively. The shedding of FcγRIIIA/CD16A on NK cells induced by FcγRIIIA/CD16A cross linking or by stimulation of NK cells with IL-12 and IL-18 has recently been shown to be greatly attenuated in the presence of ADAM17 inhibitor (18). It is of note that FcγRIIIB/CD16B and FcγRIIIA/CD16A are encoded by two highly similar genes (19). However, FcγRIIIA/CD16A is a membrane-spanning protein (19, 20), whereas FcγRIIIB/CD16B is linked to the plasma membrane via a GPI anchor (21, 22). The sequences from Thr210 to Ser220 of FcγRIIIA/CD16A and FcγRIIIB/CD16B are identical and contain the putative cleavage site of FcγRIIIB/CD16B (23) (Fig. 1).

Given the sequence homology of FcγRIIIA/CD16A and FcγRIIIB/CD16B, we were interested to study the roles of ADAM10 and ADAM17 in the shedding of FcγRIIIA/CD16A and to identify the putative cleavage site. Our second aim was to study the relationship between the shedding of FcγRIIIA/CD16A and the functional responses of CD56dim NK cells upon FcγRIIIA/CD16A-independent stimulation.

Materials and Methods

Cells

PBMCs, NK cells, JY cells, NKL cells, and CD16/γ-transduced T cells were isolated and/or cultured as previously described (1, 4, 24, 25).

Abs

The following mAbs were used: FITC- or PC7-conjugated-anti-CD16 (clone 3G8), PE-conjugated-anti-CD62L (clone DREG56), PC7-conjugated-anti-CD19 (clone J3-119), PC5-conjugated-anti-CD107a (clone H4A3) and isotype control (Beckman Coulter); PC5-conjugated-anti-CD56 (clone N901), and isotype control (Beckman Coulter); Abs targeting NKG2D/CD314 (clone 1D11), DNAX accessory molecule-1 (DNAM)/CD226 (clone DX11) and isotype control from BD Biosciences; and FITC-conjugated anti-ADAM17 (clone 111633) and its isotype control from R&D Systems. Unconjugated anti-ADAM17 rabbit polyclonal Ab was from France Biochem, HRP-conjugated anti-rabbit Ab from Cell Signaling Technology, and HRP-conjugated anti-murine IgG from GE Healthcare.

ELISA assay

Soluble rhFcγRIIIA/CD16A protein (0.4 nM) was incubated for 2 h at 37°C in the absence or presence of 0.3 μM HNE or with rhADAM10 or rhADAM17 (0.6–20 nM) in ADAM reaction buffer (25 mM Tris [pH 7.4] containing 2.5 mM ZnCl2 and 0.005% Brij-35 [v/v]). Each sample (100 μl) was applied to the Nickel chelate plate (Fisher Scientific) for 2 h with gentle agitation at room temperature. The uncleaved rhFcγRIIIA/CD16A was detected after incubation for 1 h with anti-CD16 clone Dij30c (1 μg/ml) and then HRP-conjugated anti-murine IgG diluted to 1/5000. Absorbance at 492 and 620 nm was measured with an iEMS reader MF spectrometer (Labsystems).

Fluorogenic assay

The assay was performed with HNE (3 μM), trypsin (3 μM), rhADAM17, or rhADAM10 (10 μg/ml) in 25 μM Tris (pH 9), containing 2.5 mM ZnCl2 and 0.005% Brij-35 (v/v). Cleavage of the fluorogenic substrate (5 μM) was measured at Ex340/Em490nm using a SPECTRAmax

FIGURE 1. Schematic representation of FcγRIIIB/CD16B and FcγRIIIA/CD16A. FcγRIIIB/CD16B (left panel) is linked to the plasma membrane via a GPI anchor, whereas FcγRIIIA/CD16A (right panel) is a membrane-spanning protein. The residue at position 203 (Phe and Ser for FcγRIIIA/CD16A and FcγRIIIB/CD16B, respectively) in the proform of the molecules is critical for this alternative membrane form (48). The proform of FcγRIIIB/CD16B (with Ser203), expressed as a membrane-spanning protein in the endoplasmic reticulum before GPI anchoring is shown as a gray dotted line. The glycation site of the mature form of FcγRIIIB/CD16B is likely to be the Ser at position 201 according to the prediction of the a-site (cleavage site) in the prooprotein sequence (49–51). The sequence from Thr191 to Ser220 of the stalk regions of FcγRIIIB/CD16B and FcγRIIIA/CD16A is indicated (middle panel). The location of the cleavage site (between Val196 and Ser197) of FcγRIIIB/CD16B is putative (23). Black boxes represent ITAMs.
Gemini XS spectrophotometer and SoftMax Pro 5.3 software (Molecular Devices).

**MALDI-TOF–mass spectrometry analysis**
Spectra were carried out on an UltraFlex I mass spectrometer (Bruker Daltonics). Samples were diluted 10-fold in a solution of 4-hydroxy-α-cyano-cinnamic acid saturated in a solution of 33.3% acetonitrile, 66.6% water, and 0.1% trifluoroacetic acid. Matrix-sample solutions were spotted onto a gold-plated sample probe using the ultrathin layer method (26, 27). Spectra were acquired in the reflectron negative mode (1000–1200 laser shots) in the 500–3500 mass-to-charge ratio (m/z) range. Calibration of the instrument was performed externally using a neighboring spot with a peptidic calibration kit consisting of angiotensin I, angiotensin II, substance P, bombesin, adrenocorticotropic hormones (clips 1–17 and clips 18–39), and somatostatin 28 (Bruker Daltonics). MALDI-TOF–mass spectrometry spectra were processed using FlexAnalysis 3.3 software from Bruker Daltonics.

**Western blotting**
Cell extracts were prepared in Nonidet P-40 150 mM NaCl buffer (30 min, 4˚C), centrifuged at 15,000 g for 15 min at 4˚C, and 50 μg protein extracts were loaded on SDS-PAGE under nonreducing conditions and then transferred to nitrocellulose membranes. The nitrocellulose sheet was blocked with PBS containing 5% nonfat dried milk and 0.05% Tween 20 overnight at 4˚C), centrifuged at 15,000 g. The membrane was incubated for 1 h at room temperature and then incubated overnight at 4˚C with anti-ADAM17 rabbit polyclonal Ab (1:250). After three washes, the membrane was incubated for 1 h at room temperature with HRP-conjugated anti-rabbit Ab (1:5000), washed three times, and reacted for 15 min with an ECL Western blotting system (GE Healthcare) according to the manufacturer’s protocol.

**Flow cytometry analysis**
ADAM17 expression was analyzed on PBL after staining with 10 μl anti-CD3, anti-CD19, anti-CD56, and FITC-conjugated anti-ADAM17 mAb or FITC-conjugated isotype control for 30 min at 4˚C. ADAM16 down-modulation and CD107 expression on NK cells were analyzed as previously described (4). CD16 and CD62L comodulation on stimulated NK cells and/or CD16/γ-transduced T cells was analyzed after staining with 10 μl anti-CD16 and anti-CD62L for 30 min at 4˚C. All flow cytometry (FCM) analyses were performed with a minimum of 5000 events using an EPICS XL or a Gallios flow Cytometer as described in Dall’Ozzo et al. (25) and Kaluza version 1.2 (Beckman Coulter).

**Activation of NK cells by PMA/Ca2+ or NEM**
NK cells (1 × 10^5/ml) were laid down on P96 culture plates (BD Biosciences) and then incubated at 37˚C in 5% CO2 humidified air, with indicated concentrations of PMA and/or Ca2+ for 4 h or in the presence of indicated concentrations of NEM for indicated times. In some experiments, cells were preincubated for 1 h in the presence of indicated concentrations of specific pharmacological inhibitor [(2R, 3S)-2-[[4-(2-butynyloxy)phenyl] sulfonyl]amino]-3,4-dihydroxybutanamide (TIMP-2)] or physiological inhibitor (tissue inhibitor of metalloproteinase 3 (TIMP-3)) of ADAM17 before stimulation with 1 ng/ml PMA. TIMP-3 was purchased from R&D Systems, and TIMP-2 was prepared by Dr. J. M. Brunel (Laboratory of Integrative Structural & Chemical Biology, Marseille, France) according to a previously reported procedure (28).

**Activation of NK cells and CD16/γ-transduced T cells by plate-bound mAbs**
Nunc Maxisorp 96-well culture plates (Fisher Scientific) were sensitized overnight at 4˚C as previously described (4) with mAbs targeting CD16, CD3, NK22/CD33, NKP30/CD335, NKP46/CD335, 2B4/CD244, and/or DNAM/CD226. After washing three times with PBS 0.05% Tween, solution plates were saturated for 30 min with 1% BSA (Sigma-Aldrich) and then washed twice with PBS 0.05% Tween. Freshly isolated NK cells preincubated with TMI-2 or TIMP-3 and/or CD16/γ-transduced T cells (1 × 10^5/ml) or not preincubated were laid down on sensitized wells of culture plates and incubated for 4 h at 37˚C with 5% CO2 humidified air in the absence or presence of anti-CD107a.

**Results**
ADAM17 cleaved the stalk region of FcγRIIIA/CD16A between Ala195 and Val196 residues
We first investigated the ability of ADAM10 and ADAM17 to cleave FcγRIIIA/CD16A (Fig. 1). Soluble rhFcγRIIIA/CD16A was incubated in the presence of increasing concentrations of rhADAM17 or rhADAM10 or in the presence of HNE used as positive control. Each sample was then deposited on a nickel plate to capture polyhistidine-tagged rhFcγRIIIA/CD16A. Uncleaved rhFcγRIIIA/CD16A protein was detected by ELISA using the anti-CD16 mAb Dj130c. Detection of rhFcγRIIIA/CD16A was reduced by 67.5% after incubation with HNE, indicating effective cleavage of rhFcγRIIIA/CD16A (Fig. 2A). rhADAM17 was also effective and cleaved rhFcγRIIIA/CD16A in a dose-dependent manner, whereas no cleavage was detected after incubation with a high concentration of rhADAM10.

We then used a fluorogenic peptide, DABCYL–Thr-Gln-Gly-Leu-Ala-Ala-Ser-Thr-Ile-Ser-Ser-Glu(EDANS)-NH2 (designed with a fluorochrome and a quencher at the N- and C-terminal ends of the peptide, respectively), for which the sequence encompasses aa 191–201 of FcγRIIIA/CD16A and FcγRIIIA/CD16B. Fluorescence was measured after incubation of the peptide in the presence of HNE, trypsin (used as a negative control), rhADAM17, or rhADAM10. HNE effectively cleaved the peptide, whereas trypsin had no significant detectable effect (Fig. 2B). rhADAM17 induced substantial cleavage of the peptide after 10 min of exposure, reaching a plateau similar to that of HNE after 45 min. By contrast, no cleavage was observed in the presence of rhADAM10. These results are therefore in accordance with those obtained with the complete ectodomain of FcγRIIIA/CD16A (Fig. 2A) and demonstrate the ability of rhADAM17 to cleave the stalk regions of FcγRIIIA/CD16A and FcγRIIIA/CD16B.

Analyses of the cleavage products by MALDI-TOF–mass spectrometry are shown in Fig. 2C. Two peaks, one corresponding to the intact peptide DABCYL–Thr-Gln-Gly-Leu-Ala-Ala-Ser-Thr-Ile-Ser-Ser-Glu(EDANS)-NH2 (m/z 1688.763) and one corresponding to a peptide containing degraded DABCYL (m/z 1556.696), were detected in the absence of the enzyme (Fig. 2C, top panel). A single peak was observed in the sample containing only rhADAM17 (Fig. 2C, middle panel). Two additional peaks were identified in the cleavage products (Fig. 2C, bottom panel) (i.e., DABCYL–Thr-Gln-Gly-Leu-Ala [m/z 738.274] and Val-Ser-Thr-Ile-Ser-Ser-Glu (EDANS)-NH2 [m/z 967.435]), indicating that ADAM17 cleaves the peptide substrate between Ala195 and Val196 residues.

An Ala residue at P1 position immediately upstream of the cleavage site favors the catalytic action of ADAM17, whereas a Val residue at this position is detrimental (29). We therefore designed a second fluorogenic peptide in which Ala and Val were reversed. As shown in Fig. 2D, the fluorescence observed after incubation of rhADAM17 with the peptide containing the reverse sequence was dramatically decreased compared with that observed with the wild-type (WT) sequence, confirming the specificity of the cleavage site in the peptide corresponding to the stalk regions of FcγRIIIA/CD16A and FcγRIIIA/CD16B. We concluded that rhADAM17 cleaves FcγRIIIA/CD16A and probably FcγRIIIA/CD16B between the Ala195 and Val196 amino acids.

ADAM17 was present on the surface of NK cells and mediated the shedding of FcγRIIIA/CD16A
We then investigated the presence of ADAM17 on NK cells. First, cell extracts from PBMC (used as a positive control), from purified human NK cells, from the JY lymphoblastoid cell line and the NK1 cell line were analyzed by Western blotting. We used a rabbit polyclonal Ab against human ADAM17, allowing simultaneous detection of several bands corresponding to the catalytic form (80 kDa), the precursor (proform 110 kDa), and glycosylated ADAM17 (130 kDa). The results showed the presence of an 80-kDa band (Fig. 3A bottom, arrow) and a 110-kDa band (Fig. 3A top, arrow) in all cell extracts. However, the mature
form of ADAM17 was detected at higher intensity in NK cell extracts than in PBMCs, the JY cell line, or the NKL cell line. Membrane expression of ADAM17 on PBL and purified NK cells was then studied by FCM. As shown in Fig. 3B, ADAM17 was expressed by all PBL populations. Expression was relatively weak, although the staining of CD19+ cells was slightly higher than that of CD3**CD56+ cells and CD3+ cells. ADAM17 expression was detected on almost all purified NK cells.

The shedding of FcγRIIIB/CD16B induced by CaI and PMA on neutrophils is mainly ADAM10 and ADAM17 dependent, respectively (16). We therefore evaluated the shedding of FcγRIIIA/CD16A on NK cells under stimulation with CaI and/or PMA. As shown in Fig. 4A, stimulation with PMA induced dose-dependent downmodulation of CD16. Moreover, this effect was not modified in the presence of HNE, whereas CaI used alone was ineffective. CD62L (L-selectin), a known specific substrate of ADAM17 (10, 11), is expressed on a fraction of NK cells (30). We therefore studied the comodulation of CD16 and CD62L after stimulation of NK cells with NEM, a direct activator of ADAM (31), and after stimulation with PMA. First, stimulation with NEM resulted in dose- and time-dependent downmodulation of CD62L and CD16 (Fig. 4B). Downmodulation of both CD16 and CD62L was detected after 5 to 10 min and was complete after 30 to 45 min, suggesting that the ADAM(s) responsible for the cleavage is (are) expressed on the membrane of NK cells. We did not observe CD62L expression on cells that have downmodulated CD16 (CD62L+CD16low) after NEM stimulation, showing that downmodulation of CD16 and CD62L occurred simultaneously and suggesting that the same ADAM was responsible for the shedding of both molecules. This was confirmed when NK cells were stimulated with PMA (Fig. 4C): CD16 and CD62L were simultaneously and dramatically downregulated. Preincubation of NK cells prior to PMA stimulation with increasing concentrations of TIMP-3 or TIMP-2 [physiologic and selective pharmacologic

![FIGURE 2.](http://www.jimmunol.org/)

(A) Uncleaved recombinant human FcγRIIIA/CD16A was detected by ELISA using anti-CD16 mAb Dj130c, which recognizes an epitope located in the EC1 domain of FcγRIIIA/CD16A (4) after incubation in the absence (black bar) or in the presence of HNE used as positive control (white bar), or in the presence of rhADAM10 (gray hatched bar) or rhADAM17 (gray bars). The activity of rhADAM10 and rhADAM17 was checked using commercial fluorogenic peptide (L. Lajoie, unpublished observations). This experiment was repeated twice, and the results from one representative experiment are shown (as mean ± SD of triplicate). (B) Hydrolysis of the fluorogenic peptide DABCYL–Thr-Gln-Gly-Leu-Ala-Val-Ser-Thr-Ile-Ser-Ser-Glu(EDANS)-NH$_2$ corresponding to the stalk regions of FcγRIIIA/CD16B and FcγRIIIA/CD16A, was analyzed by fluorescence after incubation in the absence or presence of HNE, trypsin used as a negative control, rhADAM17, or rhADAM10. This experiment was repeated four times, and one representative experiment is shown. (C) MALDI-TOF analysis of the fluorogenic peptide DABCYL–Thr-Gln-Gly-Leu-Ala-Val-Ser-Thr-Ile-Ser-Ser-Glu(EDANS)-NH$_2$ (top panel) each amino acid residue is indicated by single letter code), rhADAM17 (middle panel), and the cleavage products obtained after incubation with rhADAM17 (bottom panel). (D) Hydrolysis of the fluorogenic WT peptide DABCYL–Thr-Gln-Gly-Leu-Ala-Val-Ser-Thr-Ile-Ser-Ser-Glu(EDANS)-NH$_2$ (black bars) and of the peptide DABCYL–Thr-Gln-Gly-Leu-Val-Ala-Ser-Thr-Ile-Ser-Ser-Glu(EDANS)-NH$_2$ (white bars) was analyzed by fluorescence after incubation in the absence or presence of HNE, rhADAM17, or rhADAM10. This experiment was repeated twice, and one representative experiment is shown.
induce degranulation (Fig. 5A, five mAbs targeting NKG2D, NKp30, NKp46, 2B4, and DNAM. Cells were incubated in plates sensitized by combinations of one to downmodulation in response to stimulation of NK cell ARs. NK activations requires a combination of activator signals mediated by ac-

The FcRIIIA/CD16A engagement, the triggering of NK cell functions requires a combination of activator signals mediated by activating receptors (ARs) such as NKGD2, NKp30, NKp46, 2B4, and DNAM (34). We therefore studied degranulation and CD16 downmodulation in response to stimulation of NK cell ARs. NK cells were incubated in plates sensitized by combinations of one to five mAbs targeting NKGD2, NKp30, NKp46, 2B4, and DNAM. The coengagement of at least two or more ARs was required to induce degranulation (Fig. 5A, top panel), CD16 downmodulation (Fig. 5A, bottom panel), or IFN-γ production (Supplemental Fig. 1A) in a significant proportion of NK cells. The percentage of degranulating cells depended on the combination used to stimulate the cells, tending to increase with the number of ARs simultaneously engaged (Fig. 5A, top panel). Using the 31 possible combinations of 1–5 mAbs, we obtained a wide range of responses. Importantly, we observed that the profiles of CD16 downmodulation and CD107a expression according to the stimulation combination used were very similar (compare Fig. 5A, top and bottom panels), although the percentages of cells downmodulating CD16 were always substantially higher than the percentages of CD107a+ cells. The calculated coefficient of correlation was therefore very high (\( R^2 = 0.97 \)), whereas a weaker correlation was also observed between CD16 downmodulation and IFN-γ production (Supplemental Fig. 1B). It is unlikely that this weaker correlation resulted from the fact that CD56bright cells, which do not express CD16, might represent a significant proportion of IFN-γ–producing cells. Indeed, IFN-γ+ NK cells observed after stimulation by the effective combination of mAbs targeting NKGD2, NKp30, NKp46, and 2B4 were almost totally confined to CD56dim NK cells (Supplemental Fig. 1C). Finally, we studied the downmodulation of CD16 and CD62L induced by the combination of mAbs targeting NKGD2, NKp30, NKp46, and 2B4. This stimulation combination led to downmodulation of both molecules (Fig. 5B). Preincubation of NK cells with TIMP-3 or TIM-2 led to dose-dependent inhibition of downmodulation of both molecules. We concluded that the ADAM17-dependent shedding of FcγRIIIA/CD16A occurred after FcγRIIIA/CD16A-independent stimulation of NK cells and was strongly correlated with degranulation. It is of note that the FcγRIIIA/CD16A engagement by plate-bound anti-CD16 3G8 led to upregulation of CD107a on 17.7 ± 3.4% of NK cells and to downregulation of FcγRIIIA/CD16A on 95.4 ± 3.9% (n = 6) (i.e., a percentage similar to and substantially higher than that obtained when NK cells were stimulated by the combination of mAbs targeting NKGD2, NKp30, NKp46, and 2B4) (Fig. 5A). Thus, CD16 downmodulation was weakly correlated with upregulation of CD107a when cells were incubated with 3G8 compared with experiments in which other stimuli were used to activate NK cells. We cannot exclude the possibility that the detection of CD16 by fluorescent mAb might be hampered by potential epitope interference (due to cross linking or conformational change) (18) or by a putative masking effect due to detachment of the unconjugated stimulating anti-CD16 from the microplate (4). However, this hypothesis is unlikely because: 1) we observed simultaneous and dramatic downmodulation of CD62L (see below) when NK cells were stimulated by plate-bound 3G8; and 2) we found that ADAM17 inhibitors substantially inhibited the downmodulation of FcγRIIIA induced in this condition (Supplemental Fig. 2A). Another explanation might be that FcγRIIIA/CD16A may be internalized when it is engaged. An internalization method (35) therefore tested in which the cells were incubated with soluble Alexa 488-labeled 3G8, in the presence of an anti–Alexa 488...
Ab, which quenches fluorescence on the cell surface but not from internalized molecules (Supplemental Fig. 2B). Using this approach, we found that FcγRIIIA/CD16A downmodulation induced by soluble 3G8 was partially due to internalization. However, dramatic downmodulation of CD62L on CD56dim NK cells stimulated by soluble 3G8 was observed at the same time, showing that ADAM17-dependent shedding was induced in this condition (Supplemental Fig. 2C, right panel). We concluded that ADAM17-dependent shedding and internalization may both be involved in the loss of CD16 when the latter is engaged and that the relative contribution of each mechanism might depend on the conditions of engagement (for instance, immobilized versus soluble anti-CD16). By contrast, ADAM17-dependent shedding was the main, if not the exclusive, mechanism after FcγRIIIA/CD16A-independent stimulation of NK cells and strongly correlated with degranulation in this situation.

**Shedding of FcγRIIIA/CD16A was restricted to activated cells**

Given that the percentages of cells downmodulating CD16 were substantially higher than the percentages of degranulating cells, we

**FIGURE 4.** ADAM17 involved in the shedding of FcγRIIIA/CD16A. (A) NK cells were stained with anti-CD16 mAb after incubation with indicated concentrations of PMA and/or Ca++ and analyzed by FCM. Results are expressed as the percentage of NK cells downmodulating CD16. This experiment was repeated four times, and one representative experiment is shown. NK cells were stained with anti-CD16 and anti-CD62L mAbs after incubation for 1 h in the absence or presence of indicated concentrations of NEM (top panels) (B) or for the indicated times in the presence of 0.125 μM of NEM (bottom panels) (C) or after initial incubation in the absence or presence of indicated concentrations of specific pharmacological inhibitor (TMI-2) or physiological inhibitor (TIMP-3) of ADAM17 followed by a second incubation in the presence of PMA and analyzed by FCM. Each experiment was repeated at least three times, and one representative experiment is shown.
wondered whether ADAM17 expressed on an activated cell could mediate the shedding of FcγRIIIA/CD16A on the cell surface of a neighboring cell (i.e., in trans). Expression of CD62L and CD16 was therefore evaluated on CD16/γ-transduced T cells (36) and NK cells. As shown in Fig. 6 (top panels), stimulation of NK cells with anti-CD3 mAb did not modify CD16 or CD62L expression, whereas stimulation with mAbs targeting NKG2D, NKp30, NKp44, and 2B4 induced the shedding of both molecules. In contrast, stimulation of CD16/γ-transduced T cells led to the opposite result: CD3 engagement induced substantial shedding of FcγRIIIA/CD16A, whereas coengagement of NKG2D, NKp30, NKp44, and 2B4 was ineffective (Fig. 6, middle panel). When NK cells and CD16/γ-transduced T cells were mixed and stimulated in these conditions, FcγRIIIA/CD16A shedding was restricted to T cells after CD3 stimulation (Fig. 6, bottom middle panel) and to NK cells after NKG2D, NKp30, NKp44, and 2B4 stimulation (Fig. 6, bottom right panel). These results demonstrated that the FcγRIIIA/CD16A shedding was restricted to appropriately stimulated cells and suggest that this is likely to occur mainly, if not exclusively, in cis.

Discussion
This study was undertaken to investigate the roles of ADAM10 and ADAM17 in the shedding of FcγRIIIA/CD16A. We found that only the latter was able to cleave FcγRIIIA/CD16A, and we determined the cleavage site, which was located in the stalk region between Ala195 and Val196 (i.e., 1 aa upstream of the expected position). We also demonstrated that the shedding of FcγRIIIA/CD16A on

FIGURE 5. Shedding of FcγRIIIA/CD16A strongly correlated with degranulation after FcγRIIIA/CD16A-independent stimulation of NK cells. (A) Freshly isolated NK cells were incubated in the presence of fluorescent anti-CD107a mAb in plates sensitized with a combination of one to five mAbs targeting NKG2D, NKp30, NKp46, 2B4, and DNAM. They were then stained with anti-CD16 mAb and analyzed by FCM. Each bar represents the percentage of CD107a+ NK cells (top panel) or of NK cells downmodulating CD16 (bottom panel) according to the mAb combination used (results are the mean ± SD of six separate experiments using NK cells from six different donors). (B) NK cells were treated as described in Fig. 4C except that the second incubation was performed on plates sensitized with a combination of mAbs targeting NKG2D, NKp30, NKp46, and 2B4.
NK cells was at least partially mediated by ADAM17. Moreover, we showed that the ADAM17-dependent shedding of FcγRIIIA/CD16A on CD56dim NK cells was strongly correlated with degranulation when a wide range of NK cell ARs were stimulated. ADAM17-dependent shedding of FcγRIIIA/CD16A was the main, if not the exclusive, mechanism of FcγRIIIA modulation after FcγRIIIA/CD16A-independent stimulation of NK cell, whereas both ADAM17-dependent shedding and internalization may be involved after FcγRIIIA/CD16A-dependent stimulation. Finally, we provided evidence that the ADAM17-dependent shedding of FcγRIIIA/CD16A was restricted to activated cells, suggesting that ADAM17 acts mainly, if not exclusively, in cis. The shedding of FcγRIIIA/CD16A may therefore be considered as a marker of FcγRIIIA-independent NK cell activation correlated to functional responses.

Using specific inhibitors (14, 15, 17) or overexpressing transfected cells (16, 17), previous studies have reported that FcγRIIIA/CD16B may be cleaved by ADAM17 (14–17) and/or ADAM10 (16), depending on the stimulation conditions. The sequences from Thr191 to Ser201 of the stalk regions of FcγRIIIA/CD16A and FcγRIIIIB/CD16B are identical and contain the putative cleavage site of the latter, which was reported to be located between Val196 and Ser197 (23). We therefore evaluated the ability of recombinant ADAM10 and ADAM17 to cleave the complete ectodomain of FcγRIIIA/CD16A or a fluorogenic peptide encompassing the stalk regions of FcγRIIIA/CD16A and FcγRIIIIB/CD16B. We demonstrated that ADAM17 was able to cleave both compounds, whereas ADAM10 was ineffective. To our knowledge, this is the first demonstration at the molecular level that ADAM17 cleaves FcγRIIIA/CD16A. Furthermore, MALDI-TOF analysis of the cleavage products revealed that the cleavage site was located between Ala195 and Val196, which is 1 aa upstream of the cleavage site suggested on the basis of amino acid sequencing of the C-terminal fragment of the purified soluble plasma FcγRIIIA/CD16B (23). However, as stated by the authors, the HPLC chromatogram of the first cycle showing a peak corresponding to Val was difficult to interpret due to high background. By contrast, our MALDI-TOF results were unambiguous. Furthermore, Caescu et al. (29) have already defined the cleavage site selectivity of ADAM17 and ADAM10. They showed that ADAM17 is selective for smaller aliphatic hydrophobic residues at the P1’ position (immediately downstream from the cleavage site), with a Val being the most favored amino acid, whereas ADAM10 can accommodate aromatic amino acids at that position. Moreover, they demonstrated selectivity for small residues such as Ala at P1 (immediately upstream of the cleavage site). They compared their peptide library results to sequences that have been identified as being cleaved by ADAMs and confirmed that an Ala at P1 and a Val at P1’ were frequent at ADAM17 cleavage sites (found for instance in the cleavage site of TNF, TGF-α, and Notch). By contrast, a Val at P1 was detrimental for both enzymes. Our MALDI-TOF results were in line with these findings. Moreover, it was expected from the results of Caescu et al. (29) that reversing the Ala and Val in our peptide (resulting in a Val-Ala-Ser sequence rather than an Ala-Val-Ser sequence) would reduce the cleavage if the cleavage site is located between Ala and Val in the WT peptide but not if it is between Val and Ser. Our finding that the Ala-Val permutation completely abolished cleavage by ADAM17 substantiated the former assumption and confirmed the findings obtained by MALDI-TOF. We concluded that ADAM17 (but not ADAM10) cleaves the stalk region of FcγRIIIA/CD16A and probably FcγRIIIIB/CD16B between Ala195 and Val196. It is of note that the activation of mouse neutrophils, which resulted in substantial mouse l-selectin downregulation and indicated ADAM17 induction, did not lead to a significant downregulation of mouse CD16 or FcγRIV (17). The fact that mouse CD16 and FcγRIV are not regulated by ectodomain shedding may be related to the weak amino acid sequence similarity between the stalk region of human CD16 and the corresponding region of mouse CD16 or FcγRIV (17).

We clearly detected ADAM17 within cells by Western blotting and also at the cell surface by FCM. It is of note that the membrane expression was relatively weak. Interestingly, ADAM17 was also present on the cell surface of both B cells and T cells, as previously reported (37, 38). In accordance with the membrane expression of ADAM17 on NK cells, the shedding of FcγRIIIA/CD16A was very rapidly induced by stimulation with NEM, which interferes with the bond formed between the cysteine proform and the catalytic zinc ion, thus bypassing the necessity to cleave the proform (39, 40). In addition, our results showed that the shedding of FcγRIIIA/CD16A was induced by stimulation with PMA, a known activator of ADAM17 (31, 41), whereas CaI [which activates

**FIGURE 6.** Shedding of FcγRIIIA/CD16A restricted to activated cells. Purified NK cells (top panel), CD16γ-transduced T cells (middle panel), or both (bottom panel) were stained with anti-CD16 and anti-CD62L mAbs after incubation on unsensitized plates (left panel), plates sensitized with anti-CD3 mAb (middle panel), or with a combination of mAbs targeting NKG2D, Nkp30, Nkp46, and 2B4 ARs (right panel) and analyzed by FCM. This experiment was repeated twice, and one representative experiment is shown.
ADAM10 (42) was ineffective. Moreover, CD62L, a well-known specific substrate of ADAM17 (10, 11), was concomitantly shed with FcyRIIIA/CD16A in a time- and concentration-dependent manner after NEM or PMA stimulation of NK cells. Finally, the shedding of both molecules was inhibited when PMA-stimulated NK cells were preincubated with similar concentrations of ADAM17 inhibitors such as TIMP-3 or the more selective pharmacological inhibitor TMI-2 (33). Our results agree with those recently reported by Romee et al. (18) showing that human NK cells express ADAM17 and that NK cell activation by PMA, cytokine exposure, cross linking of activating receptors, or exposure to K562 target cells resulted in decreased expression of FcyRIIIA/CD16A and CD62L, which was blocked by another selective ADAM17 inhibitor (BMS566394). Taken together, these results strongly support the conclusion that ADAM17 is at least partially involved in the shedding of NK cell FcyRIIIA/CD16A.

We and others have previously shown that NK cell FcyRIIIA is downmodulated after incubation with the anti-CD16 mAb 3G8 (3, 4) or the Fc portion of rituximab (4, 6). Such modulation may result from shedding and/or internalization. Several lines of evidence suggest that the former is the main mechanism by which cells are stimulated by immobilized anti-CD16. First, immunohistochemistry and immunoblot analysis revealed little cell-associated CD16 after stimulation of NK cells by 3G8 cross linked with goat anti-mouse–coupled beads or by plate-bound 3G8 (3, 18). Second, incubation of NK cells in the presence of anti-CD16 mAb immobilized on microplates induced a simultaneous dramatic downmodulation of CD62L, a known substrate of ADAM17 (18) (Supplemental Fig. 2C, left panel). Third, FcyRIIIA downmodulation was strongly inhibited in the presence of 1,10-phenanthroline, an inhibitor of Zn2+-dependent metalloprotease (3), and in the presence of different ADAM17 inhibitors (18) (Supplemental Fig. 2A). Finally, the fact that the frequency of NK cells expressing IFN-γ or TNF-α was greater in the presence of the ADAM17 inhibitor BMS566394 after cross linking with CD16 is consistent with the maintenance of CD16 on the cell surface and not with internalization (18). In contrast, when the NK cell responses following engagement of FcyRIIIA and other ARs were compared, we found that FcyRIIIA downmodulation induced by the former was substantially higher than degranulation. It may therefore be assumed that internalization is involved in the FcyRIIIA/CD16A downmodulation induced by its engagement. Indeed, Cecchetti et al. (5) clearly demonstrated internalization of CD16 on NK cells using confocal microscopy and fluorescent soluble 3G8. CD16 is internalized through a very rapid process (10 min), and newly synthesized CD16 is rapidly re-expressed on the membrane (10 min). In our hands, FcyRIIIA downmodulation was not detectable before 1 h of stimulation by plate-bound anti-CD16 mAb or by other stimuli. However, when NK cells were incubated with Alexa 488–labeled 3G8 in the presence of an anti-Alexa 488 Ab, which quenches fluorescence on the cell surface (Supplemental Fig. 2B), we confirmed that internalization actually occurred when FcyRIIIA was engaged by soluble 3G8. However, the total fluorescence (membrane and intracellular) was decreased after incubation at 37˚C for 3 h, showing that the Alexa 488–labeled 3G8 was partially degraded after internalization and/or that shedding was also effective in this condition (Supplemental Fig. 2B, right bottom panel). The fact that CD62L was strongly downregulated in this condition demonstrated that the latter was involved and that it was ADAM17 dependent. These results suggest that both ADAM17-dependent shedding and internalization may be involved in FcyRIIIA downmodulation when the latter is engaged by anti-CD16 mAb and that the relative contribution of each mechanism might depend on the conditions of the engagement (for instance, immobilized versus soluble mAb). Finally, activation of NK cells by rituximab-opsinoned cells leads to downmodulation of CD16 (18, 43), which correlates with ICAM1 up-regulation (43). Romee et al. (18) have shown that when the human CD20-positive Burkitt’s lymphoma cell line Raji precoated with rituximab was incubated with purified NK cells in the presence of the ADAM17 inhibitor BMS566394, modulation of both CD16 and CD62L from NK cells was significantly attenuated, showing that ADAM17-dependent shedding of FcyRIIIA is at least partially involved when FcyRIIIA is engaged by rituximab-opsinoned CD20 positive target cells. Finally, other mechanisms may also be involved when an NK cell interacts directly with a target cell opsonized by an Ab. For instance, it has been shown that both rituximab and CD20 are trogocytosed by NK cells stimulated by rituximab-opsinoned cells (44). It is likely that internalization of CD16 is required in this process. The evaluation of the relative involvement of these different mechanisms in the downmodulation observed after FcyRIIIA engagement requires further investigations.

Modulation of FcyRIIIA/CD16A has also been observed after incubation of NK cells with K562 cells (i.e., independently of FcyRIIIA/CD16A engagement) (7). We therefore simultaneously evaluated the degranulation and the shedding of FcyRIIIA/CD16A after coengagement of different NK cell ARs. Using 31 combinations of plate-bound mAbs targeting NKG2D, Nkp30, Nkp46, 2B4, and DNAM (34), we obtained a wide range of responses. We observed a correlation between IFN-γ production and shedding of FcyRIIIA/CD16A (Supplemental Fig. 1B), as previously reported (18). However, in our hands, the shedding of FcyRIIIA/CD16A was more highly correlated with degranulation according to the combination used. It has been suggested that the major effector function of CD56dim/CD16+ NK cells is cytoxicity, whereas CD56bright/CD16dim−/− NK cells act mainly by secretion of cytokines. However, we observed that IFN-γ-producing NK cells in response to plate-bound mAbs were CD56dim (Supplemental Fig. 1C). This is in agreement with several studies showing that NK cells producing IFN-γ upon stimulation by K562 (7), Ab-coated target (45), Drosophila cells expressing ligands for the NK cell ARs (46) and plate-bound mAbs against ARs (47) belong to the CD56dim subset. In contrast, NK cells that readily respond to monokines such as IL-12 or IL-15 belong to the CD56bright NK cell subset (45, 47). It appears therefore more appropriate to define CD56dim and CD56bright NK cells as target cell responsive and cytokine responsive, respectively (45). It is of note that all degranulating cells downmodulated their FcyRIIIA/CD16A, whereas the converse was not true. The shedding of FcyRIIIA/CD16A and CD62L by engagement of NKG2D, Nkp30, Nkp46, and 2B4 was inhibited in the presence of TIMP-3 and TMI-2, showing that the ADAM17-dependent shedding of FcyRIIIA/CD16A was not restricted to stimulation conditions in which FcyRIIIA/CD16A was engaged, but extended to a wide range of FcyRIIIA/CD16A-independent stimuli. We found that the percentages of cells downmodulating CD16 were substantially higher than the percentages of degranulating cells. This may be explained by the heterogeneous response of NK cells to a given stimulus, as demonstrated by Fauriat et al. (46). Indeed, these authors showed that within stimulated NK cells, CD107a+ cells were confined to a subset expressing MIP-1β, whereas the reverse was not true: the expression of MIP-1β was the sole functional response observed in a very large proportion of cells. In addition, some cells produced two or more responses in this stimulation condition, but CD107a expression did not necessarily correlate with the production of TNF-α or IFN-γ. It is thus likely that the NK cells downmodulating CD16 (CD16−) without degranulating (CD107−)
that we observed in our stimulation conditions had other functional responses such as chemokine or cytokine production. Another possible explanation for this finding was that ADAM17 expressed on an activated cell may shed the FcγRIIIA/CD16A on the cell surface of a neighboring cell (i.e., in trans). However, when CD16γ-transduced T cells and NK cells were mixed, the shedding of FcγRIIIA/CD16A was restricted to T cells after CD3 engagement and to NK cells after NKGR2D, NKp30, NKp46, and 2B4 coengagement. The shedding of FcγRIIIA/CD16A was limited to appropriately stimulated cells and is therefore likely to occur mainly, if not exclusively, in cis.

We conclude that FcγRIIIA/CD16A shedding may be considered as a marker of FcγRIIIA-independent activation of human CD56dim NK cells strongly correlated with degradation. On the basis of our results obtained with CD3-stimulated T cells, it is tempting to speculate that ADAM17-dependent shedding of membrane proteins is a general mechanism taking place during the process of lymphocyte activation. The identification of the cleavage site may help in the designing of inhibitors that could enhance FcγRIIIA-dependent functions of NK cells such as Ab-dependent cell cytotoxicity.

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Disclosures
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