GM-CSF Modulates Autoantibody Production and Skin Blistering in Experimental Epidermolysis Bullosa Acquisita

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GM-CSF Modulates Autoantibody Production and Skin Blistering in Experimental Epidermolysis Bullosa Acquisita

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GM-CSF activates hematopoietic cells and recruits neutrophils and macrophages to sites of inflammation. Inhibition of GM-CSF attenuates disease activity in models of chronic inflammatory disease. Effects of GM-CSF blockade were linked to modulation of the effector phase, whereas effects on early pathogenic events, for example, Ab production, have not been identified. To evaluate yet uncharacterized effects of GM-CSF on early pathogenic events in chronic inflammation, we employed immunization-induced epidermolysis bullosa acquisita (EBA), an autoimmune bullous disease caused by autoantibodies to type VII collagen. Compared to wild-type mice, upon immunization, GM-CSF−/− mice produced lower serum autoantibody titers, which were associated with reduced neutrophil numbers in draining lymph nodes. The same effect was observed in neutrophil-depleted wild-type mice. Neutrophil depletion in GM-CSF−/− mice led to a stronger inhibition, indicating that GM-CSF and neutrophils have additive functions. To characterize the contribution of GM-CSF specifically in the effector phase of EBA, disease was induced by transfer of anti–type VII collagen IgG into mice. We observed an increased GM-CSF expression, and GM-CSF blockade reduced skin blistering. Additionally, GM-CSF enhanced reactive oxygen species release and neutrophil migration in vitro. In immunization-induced murine EBA, treatment with anti–GM-CSF had a beneficial effect on established disease. We demonstrate that GM-CSF modulates both autoantibody production and skin blistering in a prototypal organ-specific autoimmune disease. The Journal of Immunology, 2014, 192: 559–571.

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The online version of this article contains supplemental material.

Abbreviations used in this article: AUC, area under the curve; COL7, type VII collagen; DEJ, dermal-epidermal junction; EBA, epidermolysis bullosa acquisita; IgC, immune complex; IF, immunofluorescence; mCOL7, murine type VII collagen; NR-IgG, normal rabbit IgG; PMN, polymorphonuclear cell; ROS, reactive oxygen species; RT, room temperature.

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leading to both neutrophil extravasation and activation. Reactive oxygen species and proteolytic enzymes released from neutrophils then lead to subepidermal blister formation (29–31).

In animal models of EBA, we noted increased levels of GM-CSF in both serum and skin. Based on this observation, we first evaluated the effects of GM-CSF blockade in a model of autoantibody transfer-induced EBA and documented a profound proinflammatory activity of this cytokine mediated by innate immune mechanisms. Subsequently, we investigated the contribution of GM-CSF in modulating adaptive immune functions in immunization-induced EBA and identified a significant contribution of GM-CSF to the formation of autoantibodies.

**Materials and Methods**

**Experiments with human blood samples**

Blood for isolation of polymorphonuclear cells (PMNs) was obtained from EDTA anticoagulated blood drawn from healthy blood donors. Sera from EBA patients and controls were obtained by venipuncture. All experiments using human samples were approved by the Ethics Committee of the University of Lübeck and were performed according to the Declaration of Helsinki. Blood donors gave their written informed consent prior to study participation.

**Mice**

C57BL/6, B6.SJL-H2s, and BALB/c mice were initially obtained from Charles River Laboratories (Sulzfeld, Germany). GM-CSF−/− mice were provided by Prof. Jeff Whitsett (Division of Pulmonary Biology, Cincinnati Children’s Hospital, Cincinnati, OH) (32). Mice included in experiments were from the offspring of all the strains. For experiments using human samples were approved by the Ethics Committee of the University of Luxembourg and were performed according to the Declaration of Helsinki. Blood donors gave their written informed consent prior to study participation.

**Generation of anti-mouse COL7 IgG**

Rabbit anti-mouse COL7 IgG was generated as described elsewhere (24). In brief, recombinant fragments of murine COL7 (termed mCOL7A, mCOL7B, and mCOL7C) tagged with GST were produced using a prokaryotic expression system and purified by glutathione affinity chromatography. IgG from rabbits immunized with these recombinant fragments of mCOL7 and from nonimmunized rabbits was purified by affinity chromatography using protein G-Sepharose affinity chromatography (Amersham Biosciences, Heidelberg, Germany). Reactivity of IgG fractions was analyzed by immunofluorescence (IF) microscopy on murine skin.

**Induction of experimental EBA by autoantibody transfer**

C57BL/6 and GM-CSF−/− mice aged 6–10 wk were used for induction of experimental EBA by autoantibody transfer. In brief, experimental EBA was induced in mice by transfer of rabbit anti-mouse COL7 IgG as described (24). Clinical disease manifestation, expressed as percentage of body surface area covered by EBA skin lesions, was determined before and described (24). Clinical disease manifestation, expressed as percentage of body surface area covered by EBA skin lesions, was determined before and the area under the curve (AUC) was used to calculate the overall EBA severity. Injection of normal rabbit IgG (NR-IgG) served as negative control.

**Induction of immunization induced EBA**

C57BL/6, B6.SJL-H2s, and GM-CSF−/− mice aged 6–10 wk were used for experiments with immunization-induced EBA. All clinical examinations, biopsies, and bleedings were performed under anesthesia with i.p. administration of a mixture of ketamine (100 μg/g) and xylazine (15 μg/g). Animal experiments were approved by local authorities of the Animal Care and Use Committee (Kiel, Germany) and performed by certified personnel.

Biopsies of nonlesional skin were obtained 2 d after last injection of IgG (day 12), and IgG and C3 deposits were detected by direct IF microscopy and/or detection of Abs against COL7 by Western blotting. Serum from healthy blood donors (n = 52) served as a reference. Analysis of GM-CSF serum concentrations was performed by a commercial supplier using the Bio-Plex system.

**IF microscopy**

Measurement of reactive oxygen species and GM-CSF release from human leukocyte populations

Blood for isolation of PMNs was obtained from EDTA anticoagulated blood collected from healthy blood donors. Neutrophil activation was assessed by determination of immune complex (IC)–induced reactive oxygen species
FIGURE 1. Expression patterns of GM-CSF in experimental and human EBA. (A) Increased serum concentrations of GM-CSF in Ab transfer-induced EBA. GM-CSF levels were found to be significantly increased in sera from mice injected with mCOL7-IgG (plG) starting at day 4 of the experiment compared with NR-IgG–injected mice sera (*p = 0.01, one-way ANOVA). (B) Increased serum concentrations of GM-CSF in immunization-induced EBA. GM-CSF concentrations were significantly increased starting 2 wk after immunization compared with control sera (*p = 0.01, one-way ANOVA). (C) Correlation of GM-CSF serum concentrations with clinical disease severity assessed in C57BL/6 mice before and 3, 6, and 12 d after the initial anti-COL7 IgG injection. Levels of GM-CSF in serum increases exponentially with affected area (r = 0.9675, p ≤ 0.001, nonlinear regression). (D) Representative staining from skin specimens obtained from a total of 12 mice per group injected with NR-IgG or anti-COL7 IgG 12 d after the initial IgG injection. GM-CSF was stained in blue (insert, isotype control staining of anti-COL7–injected mouse). (E) Expression of GM-CSF in serum is increased in human EBA patients (n = 26) compared with healthy controls (n = 52; p < 0.001, Mann–Whitney rank sum test). Owing to the nonparametric distribution, data are presented as median (black line), 25th/75th percentiles (boxes), and 5th/95th percentiles (error bars), and green/red dots represent actual results for each sample.
(ROS) release using a slightly modified protocol (37). In brief, human PMNs were isolated using Polymorphprep (Axis-Shield, Heidelberg, Germany) following the protocol provided by the manufacturer. Isolated PMNs were incubated on a 96-well plate (Greiner Bio-One, Frickenhausen, Germany) coated with IC formed by monoclonal anti-COL7 Abs and recombinant fragments located within the human NC1 domain of COL7 (38). ROS release with GM-CSF at different concentrations was analyzed using luminol (Sigma-Aldrich Biochemie, Hamburg, Germany). For positive controls, FMLP (Sigma-Aldrich Biochemie) was used. Each plate was analyzed for 60 repeats using a plate reader (PerkinElmer, San Jose, CA) and the values are expressed as relative fluorescence units. To analyze GM-CSF release from neutrophil and monocyte cell populations, EDTA anticoagulated blood was obtained from healthy blood donors. Both cell populations were isolated using Polymorphprep (Axis-Shield) coated with immune complexes formed by monoclonal anti-COL7 Abs and recombinant fragments located within the human NC1 domain of COL7 and incubated at 37°C on a shaker. At different time points (start and after 1, 2, and 3 h), cell supernatants were collected from different wells. GM-CSF release was analyzed by using a Legend Max human GM-CSF ELISA kit (BioLegend, Fell, Germany) following the protocol provided by the manufacturer. Supernatants from unstimulated cells were used as controls.

**Detection of circulating vWFA2-specific Abs**

To detect circulating vWFA2-specific Abs, ELISA was performed. Briefly, 96-well microtiter plates (Maxisorb; Nunc, Roskilde, Denmark) were coated with 250 ng murine vWFA2. Nonspecific binding was reduced by blocking plates with 1% BSA in PBS-T at room temperature (RT) for 1 h. Subsequently, plates were incubated at 37°C on a shaker. At different time points (start and after 1, 2, and 3 h), cell supernatants were collected from different wells. GM-CSF release was analyzed by using a Legend Max human GM-CSF ELISA kit (BioLegend, Fell, Germany) following the protocol provided by the manufacturer. Supernatants from unstimulated cells were used as controls.

**Statistical analysis**

Unless otherwise noted, data are presented as means ± SE. For comparisons of two groups, a t test or Mann–Whitney rank sum test was used as appropriate. For comparison of more than two groups, ANOVA was used. For equally distributed data, a one-way ANOVA, followed by a Bonferroni t test for multiple comparisons, was used; when data were nonparametric, an ANOVA by ranks (Kruskal–Wallis) was applied, followed by a Bonferroni t test for multiple comparisons. Pearson product moment correlation was used to test for correlations. In all tests, a p < 0.05 was considered significant. All statistical analysis was performed using SigmaPlot 12.0 (Systat Software, Erkrath, Germany).
Results

Increased expression of GM-CSF, but not G-CSF, in experimental and human EBA

Repellent injections of anti-COL7 IgG into either C57BL/6 or BALB/c mice induced experimental EBA as described (24). In both strains, comparable and equally distributed serum concentrations of G-CSF and GM-CSF were noted in control mice. In contrast, induction of experimental EBA lead to a sharp, 8- to 38-fold increase of both CSFs in the tested mouse strains. Additionally, a great variation in the serum levels of both G-CSF and GM-CSF concentrations in mice with experimental EBA was noted. Overall, a significant increase was evident for GM-CSF, but not G-CSF, serum concentrations in C57BL/6 as reported (40), as well as BALB/c (23-fold) mice after EBA induction (Supplemental Fig. 1A). In C57BL/6 mice, serum was also obtained before the initial anti-COL7 IgG injection and 3, 6, and 12 d thereafter. In this experiment, GM-CSF serum concentrations strongly and exponentially correlated with the clinical disease activity, expressed as the body surface area affected by EBA skin lesions (Fig. 1A–C). These GM-CSF serum concentrations strongly and exponentially correlated with the clinical disease activity, expressed as the body surface area affected by EBA skin lesions (Fig. 1A–C). These GM-CSF serum concentrations continuously increased throughout the observation period (Fig. 1A–C). Expression of GM-CSF in the skin of C57BL/6 mice 12 d after the injection of NR-IgG was not observed. In contrast, cutaneous GM-CSF expression was constantly detected 12 d after the administration of anti-COL7 IgG (Fig. 1D). We also observed an increased GM-CSF serum concentration in immunization-induced EBA, which was first noticed 2 wk after immunization and increased until week 4. Thereafter, a plateau was maintained until the end of the 6-wk observation period (Fig. 1B).

In line with these findings, we also observed increased GM-CSF serum concentrations in EBA patients, compared with healthy controls. In detail, in healthy controls (n = 52), GM-CSF serum levels were 30.3 pg/ml (median; 25th percentile, 27.4–31.33 pg/ml; Fig. 1E), whereas in EBA patients levels were 60.6 pg/ml (median; range, 54.7–67.43 pg/ml; p < 0.001, Mann–Whitney rank sum test). Ex vivo stimulation of isolated human neutrophils and monocytes with EBA-IC induced a time-dependent release of GM-CSF by these two leukocyte populations (Supplemental Fig. 1B).

Induction of skin blistering in experimental EBA is hindered in GM-CSF–deficient mice

To test whether the increased serum and cutaneous GM-CSF expression in experimental EBA is of functional relevance, anti-COL7 IgG was injected into mice lacking GM-CSF expression (n = 12) and wild-type control C57BL/6 mice (n = 12). Skin lesions were evident in 11 of 12 C57BL/6 mice with a median of 0.2% (range, 0–0.6%) of affected body surface area as early as 4 d after the initial anti-COL7 IgG injection. Similar disease incidence and severity were observed in GM-CSF−/− mice. In detail, 11 of 12 mice presented with skin lesions with a median of

Table I. EBA-associated neutrophilia is absent in GM-CSF–deficient mice

<table>
<thead>
<tr>
<th>Strain/IgG</th>
<th>NR-IgG</th>
<th>Anti-COL7 IgG</th>
</tr>
</thead>
<tbody>
<tr>
<td>C57BL/6</td>
<td>16.6 ± 2.2</td>
<td>31.5 ± 2.3</td>
</tr>
<tr>
<td>GM-CSF−/−</td>
<td>13.4 ± 1.4</td>
<td>17.6 ± 4.3</td>
</tr>
</tbody>
</table>

Numbers in the table correspond to the percentage of circulating neutrophils, determined by differential blood count 12 days after the injection of the respective IgG preparation (mean ± SEM). Significant differences are observed comparing NR-IgG- and anti-COL7 IgG-injected C57BL/6 mice, as well as in comparison of anti-COL7 IgG-injected C57BL/6 mice with GM-CSF-deficient mice. Data are from 5–6 mice per group.
0.4% (range, 0–1.4%) affected body surface area. This difference in median disease severity was not statistically significant. Eight days after the initial anti-COL7 IgG injection, blister formation affected all mice. Although not statistically significant ($p = 0.053$), a trend toward a lower disease scores in GM-CSF$^{−/−}$ mice was observed. Mean affected body surface area in C57BL/6 mice was 2.9% (0.6–10.3%), as opposed to 1.3% (0.3–2.6%) in mice lacking GM-CSF expression. This trend was validated 12 d after the initial anti-COL7 IgG injection. Whereas in C57BL/6 mice 8.4% (1.6–14.1%) of the body surface area was affected by blistering, this was significantly lower ($p < 0.001$) in GM-CSF$^{−/−}$ mice, which had a mean of 3.0% (0.6–6.1%) affected body surface area (Fig. 2A). Calculation of the overall disease activity, taking onset and maximum severity into account and expressed as the AUC in

**FIGURE 4.** Decreased neutrophil infiltration into blood and skin in GM-CSF$^{−/−}$ mice after injection of anti-COL7 IgG. (A) Injection of anti-COL7 IgG significantly increased neutrophil numbers in peripheral blood of C57BL/6 mice compared with GM-CSF$^{−/−}$ mice starting from 8 d after initial injection ($p = 0.01$, two-way ANOVA). (B) Likewise, injection of anti-COL7 IgG induces a significant increase in dermal leukocyte infiltration in C57BL/6 compared with GM-CSF$^{−/−}$ mice ($p < 0.001$, two-way ANOVA). (C) Correlation of neutrophil number in blood with those in dermal infiltrate in both strains after anti-COL7 IgG injection ($r = 0.895$, $p ≤ 0.001$, Pearson product-moment correlation). (D) Representative H&E-stained sections from both strains injected with anti-COL7 IgG and NR-IgG (original magnification ×200).
C57BL/6 mice, reached a median of 29 (range, 4–97). In GM-CSF−/− mice the AUC was reduced to 26% of wild-type mice (median, 8; range, 3–20; Fig. 2B). These clinical changes were accompanied by unaltered deposition of IgG and C3 at the DEJ (Fig. 2C). Injection of NR-IgG did not lead to the development of skin lesions in both strains (n = 5/strain) throughout the entire observation period.

Decreased skin blistering in experimental EBA after anti–GM-CSF injection

We further tested the impact of GM-CSF in experimental EBA by using neutralizing Abs against GM-CSF. Anti-COL7 IgG was injected into C57BL/6 mice (n = 5) and anti–GM-CSF Abs (50 μg/mouse) were simultaneously administered. Control mice were injected with anti-COL7 IgG and appropriate isotype control Ab (n = 5). Skin lesions were evident in control mice with a median of 0.3% (range, 0–0.6%) of affected body surface area as early as after 4 d of the initial anti-COL7 IgG injection. In anti–GM-CSF–treated mice, no blisters were observed at the same time point (Fig. 3A). Overall clinical disease severity expressed as the AUC reached a median of 36.2 (range, 26.2–45.3) in isotype Ab–injected mice, whereas extent of blister formation was >2-fold reduced in mice treated with anti–GM-CSF, reaching a median AUC of 13.1 (range, 10.2–18.4; p = 0.003, Mann–Whitney rank sum test; Fig. 3B). Representative clinical images are shown in Fig. 3C. Differences in clinical disease activity were independent of IgG or C3 deposition at the DEJ, which was similar between both treatment groups (data not shown).

Absence of EBA-associated neutrophilia in GM-CSF−/− mice

Given the pivotal role of neutrophils in the development of autoantibody-induced tissue injury in this EBA model (41) and the effects of GM-CSF on neutrophil activation (5, 42), levels of circulating neutrophils were determined in both mouse strains 12 d after the injection of either anti-COL7 or NR-IgG. In NR-IgG–injected C57BL/6 and GM-CSF−/− mice the proportion of circulating neutrophils was comparable, that is, 16.6 ± 2.2 and 13.4 ± 1.4%, respectively. Compared to NR-IgG, injection of anti-COL7 IgG into C57BL/6 mice was associated with a marked and statistically significant (p < 0.001, Mann–Whitney rank sum test) increase in circulating neutrophils (31.5 ± 2.3%). This was not observed in GM-CSF−/− mice, which showed similar neutrophil counts after injection of either NR-IgG or anti-COL7 IgG (Table I). In line with this observation, in the differential blood count, the proportion of circulating neutrophils was significantly decreased in GM-CSF−/− mice after injection of anti-COL7 IgG compared with wild-type controls (p = 0.015, Mann–Whitney rank sum test). Similar observations were made evaluating the absolute neutrophil number in the circulation (Fig. 4A).

Decreased dermal infiltration of neutrophils in GM-CSF−/− mice after injection of anti-COL7 IgG

Parallel to the effects of GM-CSF deficiency on circulating neutrophil counts after the injection of anti-COL7 IgG, the impact of lacking GM-CSF expression on the neutrophil-rich dermal leukocyte infiltration in experimental EBA (30) was analyzed. In-
duction of experimental EBA in C57BL/6 mice led to a pronounced increase in dermal leukocyte infiltration (p = 0.01, two-way ANOVA). This phenomenon was also observed in GM-CSF−/− mice (p = 0.03, two-way ANOVA). However, after injection of anti-COL7 IgG, compared with C57BL/6 mice, dermal leukocyte infiltration was significantly lower in GM-CSF−/− mice (p ≤ 0.001, two-way ANOVA; Fig. 4B). Furthermore, a close correlation between circulating and skin-infiltrating neutrophils was observed (r = 0.895, p ≤ 0.001, Pearson product-moment correlation; Fig. 4C). Representative H&E staining images are shown in Fig. 4D.

**GM-CSF enhances IC-induced neutrophil activation**

To test whether IC-induced neutrophil activation is modulated by GM-CSF, ROS release from IC-activated neutrophils was measured in the absence and presence of GM-CSF. Incubation of neutrophils with GM-CSF in the presence of either monoclonal anti-COL7 Abs, or corresponding proteins located within the NC1 domain, had no effect on neutrophil ROS release. In contrast, ROS was readily released from neutrophils after incubation with ICs of anti-COL7 Ab and COL7 protein. This IC-induced neutrophil activation was further enhanced by addition of GM-CSF in a dose-dependent manner (Fig. 5A). A representative ROS release curve shown in Fig. 5B.

**Anti–GM-CSF treatment delays disease progression in already established immunization-induced EBA**

To test whether modulation of GM-CSF function could be potentially used as a treatment, experimental EBA was induced in B6.SJL-H2s mice (n = 16) by immunization with an immunodominant protein located within murine COL7. Thereafter, mice were scored every week and allocated into treatment and isotype groups when 2% or more of the body surface area showed skin lesions in individual mice. Mice were randomly allocated to anti–GM-CSF (50 μg three times per week, n = 8) or appropriate isotype control Ab (n = 8). Disease progression was monitored weekly and is expressed in relationship to the score at allocation to treatment. No difference in disease severity was detected 1 wk after treatment had been initiated (Fig. 6A). At later time points, clinical disease manifestation in isotype-injected mice increased close to 7-fold (Fig. 6A). In contrast, mice treated with anti–GM-CSF showed a significantly lower disease progression at these time points (Fig. 6A). In line with this observation, overall disease activity expressed as AUC reached a median of 47.6 (range, 38.7–54.4) in isotype-treated mice, whereas disease severity was significantly decreased in mice treated with anti–GM-CSF presented with a median AUC of 33.2 (range, 26.1–37.4; p ≤ 0.001, Mann–Whitney rank sum test; Fig. 6B). Representative clinical images are shown in Fig. 6C. Differences in clinical disease activity were independent of circulating Ag-specific IgG (subclasses) Abs, IgG, or C3 deposition at the DEJ, which were similar between both treatment groups (data not shown).

**Lower levels of Ag-specific autoantibody concentrations after immunization of GM-CSF–deficient mice with recombinant murine COL7: correlation with numbers of neutrophils in draining lymph nodes**

Owing to the long half-life of autoreactive anti-COL7–producing B cells (43), as well as the long persistence of circulating and tissue-bound COL7 autoantibodies (44), we investigated whether GM-CSF is required for autoantibody production. To address this, C57BL/6 and GM-CSF−/− mice (n = 8/group) were immunized with an immunodominant protein located within murine COL7 and levels of circulating Abs were determined by ELISA until 6 wk after immunization. Autoantibodies against COL7 were detected at 2 wk after immunization at equal titers in both strains.
of mice (Fig. 7A). After this time point, C57BL/6 mice showed significantly higher autoantibody serum levels compared with GM-CSF<sup>−/−</sup> mice (Fig. 7A). This effect of GM-CSF deficiency on specific autoantibody production affected all Ab subclasses (Fig. 7B).

Recently, GM-CSF has been shown to contribute to formation of anti-influenza IgG (21). Furthermore, Ab generation in humans has been demonstrated to be modulated by B cell helper neutrophils (22). Owing to the long persistence of circulating and tissue-bound anti-COL7 Abs (44), we investigated whether neutrophil homing to draining lymph nodes is changed in GM-CSF<sup>−/−</sup> mice after immunization. Interestingly, at the end of the observation period, we found significantly lower neutrophil numbers in the draining lymph nodes of GM-CSF<sup>−/−</sup> mice compared with C57BL/6 mice (Fig. 7C). We also observed a colocalization of neutrophils with BAFF in draining lymph nodes 6 wk after immunization (Supplemental Fig. 2). The ratio of neutrophils versus total number of cells (Ly6G/DAPI) showed significant reduction in GM-CSF<sup>−/−</sup> mice compared with C57BL/6 mice (n = 5/group; p = 0.001, Student t test; Fig. 7D). Staining of draining lymph node sections with labeled Ag, reflecting the total number of COL7-specific B cells (36), was identical in both groups (Fig. 7E). Furthermore, the number of Ly6G<sup>+</sup> cells in lymph nodes significantly correlated with the circulating anti–mCOL7-IgG at the end of the experiment (Fig. 7F; r = 0.917; p ≤ 0.0001, Pearson correlation test).

**FIGURE 7.** Reduced autoantibody serum levels in GM-CSF-deficient mice after immunization: correlation with reduced neutrophil numbers in draining lymph nodes. COL7-specific Ab serum levels in C57BL/6 or GM-CSF<sup>−/−</sup> mice were analyzed weekly after immunization using ELISA. (A) GM-CSF<sup>−/−</sup> mice mounted a significantly lower autoantibody response compared with C57BL/6 mice from week 4 until the end of the experiment (*p ≤ 0.001, Student t test). (B) Comparison of autoantibody IgG subclasses in mice showed a significantly decreased production at week 6. (C) Representative sections of draining lymph nodes stained with DAPI (blue), Ly6G (red), and COL7 (green) (original magnification ×200). (D) Percentage of neutrophil numbers in lymph nodes (Ly6G/DAPI) of C57BL/6 and GM-CSF<sup>−/−</sup> mice at the end of the experiment showed a significant reduction of the Ly6G/DAPI ratio in GM-CSF<sup>−/−</sup> mice (n = 5; p = 0.001, t test). (E) Percentage of COL7-specific B cells in lymph nodes (COL7/DAPI) of C57BL/6 and GM-CSF<sup>−/−</sup> mice (n = 5) showed no significant difference. (F) Neutrophil numbers in lymph nodes correlate with anti-mCOL7 IgG serum levels after 6 wk of immunization in C57BL/6 and GM-CSF<sup>−/−</sup> mice (r = 0.917; p ≤ 0.0001, Pearson correlation test).
Lower serum levels of Ag-specific autoantibodies in neutrophil-depleted mice

To evaluate the impact of neutrophils on production of specific autoantibodies, we depleted neutrophils in C57BL/6 mice and GM-CSF−/− mice (n = 5 mice/group). In these mice, EBA was induced by immunization with COL7. As observed before (Fig. 7A), autoantibodies were detected 2 wk after immunization (Fig. 8A). At this time point, equal serum levels of anti-COL7 Abs were observed in neutrophil-depleted and isotype control mice. After this time point, isotype Ab-injected C57BL/6 mice mounted a significantly higher immune response compared with neutrophil-depleted mice (p ≤ 0.001, one-way ANOVA). Three weeks after immunization, GM-CSF−/− mice (both isotype- and 1A8-injected) showed a similar Ab response, which was significantly lower than that observed in C57BL/6 mice (both isotype- and 1A8-injected; p ≤ 0.001, one-way ANOVA). The difference in mounting a specific immune response was even more pronounced 6 wk after immunization. At this time point, GM-CSF−/− mice injected with 1A8 showed a very low specific autoantibody response (Fig. 8A). Furthermore, C57BL/6 mice (1A8-injected) and GM-CSF−/− mice (isotype-injected) had serum levels of anti-COL7 IgG significantly lower compared with isotype-injected C57BL/6 mice (p = 0.001, one-way ANOVA; Fig. 8A) and significantly higher levels compared with 1A8-injected GM-CSF−/− mice (p = 0.01 and p = 0.02, respectively, one-way ANOVA; Fig. 8A).

Analysis of the total amount of anti-COL7 IgG produced during the 6-wk observation period (expressed as AUC, Fig. 8B) revealed significant inhibition of autoantibody production in neutrophil-depleted C57BL/6 mice (1A8-injected; mean AUC, 3.4 ± 0.3) compared with isotype-injected C57BL/6 mice (mean AUC, 4.5 ± 0.2; p ≤ 0.001, two-way ANOVA). Furthermore, neutrophil depletion using 1A8 in GM-CSF−/− mice (mean, 2.5 ± 0.1) showed significantly lower cumulative autoantibody levels in sera compared with 1A8-injected C57BL/6 mice (mean, 3.4 ± 0.3; p = 0.003, two-way ANOVA), isotype-injected C57BL/6 mice (mean, 4.5 ± 0.2; p ≤ 0.001, two-way ANOVA), or compared with isotype-injected GM-CSF−/− mice (mean, 3.2 ± 0.2; p = 0.002, two-way ANOVA; Fig. 8B). The effect of neutrophil depletion in GM-CSF−/− mice on autoantibody production is cumulative but not synergistic, as interaction analysis of genotype (C56BL/6 or GM-CSF−/−) and treatment (isotype or 1A8) showed no significance (p = 0.236, two-way ANOVA; Fig. 8B).

Discussion

The data presented in this study provide strong evidence for proinflammatory functions of GM-CSF in an experimental model of a prototypic organ-specific autoantibody-mediated disease. Both genetic and pharmacological blockade of GM-CSF led to significant decrease in clinical disease severity in two different models of EBA. Interestingly, blockade of GM-CSF not only impaired neutrophil functions, but also significantly contributed to autoantibody production.

More specifically, induction of experimental EBA led to increased levels of GM-CSF both locally (skin) and systemically (serum). These increased GM-CSF levels are of functional relevance, as in GM-CSF−/− mice induction of skin blistering by Ab transfer is significantly impaired. Furthermore, in already established EBA, blockade of GM-CSF significantly attenuated clinical disease progression. Our findings are in line with observations in experimental models of arthritis (10–12), but in contrast to data obtained in murine models of contact hypersensitivity, where an increased GM-CSF expression was found in skin and serum samples (17); however, GM-CSF deficiency alone has no effect on disease manifestation (18). These observations underscore the
importance of validating morphologic findings or expression data in appropriate experimental systems to fully understand their contribution to disease pathogenesis. For the effector phase of experimental EBA, that is, autoantibody-induced tissue injury, neutrophil depletion has been shown to protect mice from induction of skin blisters (41). In line with this observation, CD18-deficient mice with impaired leukocyte extravasation into the skin (45) are also completely resistant to blister induction (41). Additionally, impaired release of ROS (41) or proteolytic enzymes (46) from neutrophils has similar protective effects. In this context, our findings provide novel insights into EBA pathogenesis: GM-CSF contributes to EBA-associated tissue injury by modulating several neutrophil functions (Fig. 9). First, GM-CSF is required to induce autoantibody-induced neutrophilia. It is tempting to speculate that the increase in circulating neutrophils allows a higher number of neutrophils to migrate into the skin. In general, both G-CSF and GM-CSF are capable of recruiting neutrophils from the bone marrow into the blood (47, 48). Interestingly, in experimental EBA, this recruitment appears to be mainly mediated by GM-CSF, as, after injection with Ab to COL7, neutrophil numbers were significantly lower in GM-CSF−/− mice. Second, GM-CSF is required for neutrophil extravasation from blood into skin, as after EBA induction, the dermal leukocyte infiltrate was significantly reduced in GM-CSF−/− compared with C57BL/6 mice. Based on the observation of a significant increase in dermal leukocyte infiltration in GM-CSF−/− mice, neutrophil extravasation into the skin, in contrast to recruitment into the blood, is not predominantly mediated by GM-CSF. Furthermore, given that GM-CSF deficiency led to impaired blistering, the effect of GM-CSF on neutrophil recruitment into the skin may be indirect rather than a direct enhancement of neutrophil migration (49). More specifically, we hypothesize that GM-CSF activates neutrophils in the skin, resulting in the release of proinflammatory mediators by neutrophils, which leads to a continued and increased neutrophil migration into the skin (50). Third, IC-mediated activation of neutrophils is also enhanced by GM-CSF, similar to that described for many, but not all, known neutrophil agonists (42). This enhancement of IC-induced neutrophil activation contributes to reactive oxygen release by neutrophils, which is required for autoantibody-induced tissue injury in different models of EBA (41). Additionally, GM-CSF may also enhance other effector functions of neutrophils, such as release of proinflammatory mediators or proteolytic enzymes (Fig. 9).

Our findings confirm and extend previous reports on GM-CSF inhibition in the effector phase of arthritis. However, in these models, GM-CSF inhibition had no effect on autoantibody production (11, 12). In contrast, in the context of infection, GM-CSF (together with IL-4) was shown to stimulate an Ab response to avian influenza in immunodeficient mice engrafted with human hematopoietic stem cells by promoting an Ab response (21). Furthermore, neutrophils, which are among the main targets of GM-CSF (5), have profound effects on B cell functions. More specifically, neutrophils induced Ig class switching, somatic hypermutation, and Ab production by activating B cells through mechanisms involving BAFF, APRIL, and IL-21 (22). The observed colocalization of neutrophils with BAFF in draining lymph nodes after immunization with COL7 in draining lymph nodes suggests that neutrophils are also involved in shaping the process of anti-COL7 IgG in immunization-induced EBA, presumably by mediating the influx of (I) APCs and (II) neutrophils (Fig. 7). This enhancement of IC-induced neutrophil activation contributes to reactive oxygen release by neutrophils, which is required for autoantibody-induced tissue injury in different models of EBA (41). Additionally, GM-CSF may also enhance other effector functions of neutrophils, such as release of proinflammatory mediators or proteolytic enzymes (Fig. 9).

FIGURE 9. Contribution of GM-CSF to pathogenesis of experimental EBA. Schematic diagram summarizing the observed effects of GM-CSF in experimental models of EBA. (A) Mechanisms leading to the loss of tolerance to COL7 and anti-COL7 autoantibody production (path 1). In experimental EBA, a complex interplay with APCs and CD4+ T cells leads to a COL7-specific B cell response (33). In path 2, this results in production of circulating autoantibodies in peripheral lymph nodes, which are released into the circulation (path 3). As shown in this study (Fig. 8), both GM-CSF and neutrophils contribute to formation of anti-COL7 IgG in immunization-induced EBA, presumably by mediating the influx of (I) APCs and (II) neutrophils (Fig. 7) into the peripheral lymph nodes. (B) Subsequently, autoantibodies locate from the circulation to the DEJ in the skin (path 4). Furthermore, neutrophils are recruited from bone marrow (BM) into the circulation. (III) As shown in Fig. 4, GM-CSF modulates neutrophil recruitment from the bone marrow into the circulation. (IV) After autoantibody binding, a proinflammatory milieu is generated in the skin (62), leading to neutrophil recruitment into the skin (path 6). (V) This process also depends on the presence of GM-CSF (Fig. 4). Thereafter, neutrophils bind to the immune complexes located at the DEJ in an FcγR-dependent fashion, leading to neutrophil activation and ultimately blister formation (path 7). (V) As shown in Fig. 5, IC-induced neutrophil activation is enhanced in the presence of GM-CSF.
The authors have no financial conflicts of interest.

CSF through which neutrophils modulate the Ab response is unclear. In T cell–dependent Ab production. However, the precise mechanism of a few case reports, such innovative strategies have not yet been implemented in patients with autoantibody bullous dermatoses (62).

In the present study we demonstrate that GM-CSF not only modulates the effector phase of EBA by blocking neutrophil functions, but it is also required for mounting an autoantigen-specific B cell response. A similar inhibitory effect on autoantibody production was observed in neutrophil-depleted mice. Combining GM-CSF inhibition and neutrophil depletion showed additive effects. Our findings extend the potential therapeutic spectrum of GM-CSF inhibition to autoantibody-mediated diseases.

Disclosures

The authors have no financial conflicts of interest.

References


A

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B

Supplement figure 1: Expression and sources of GM-CSF in EBA

A. G- and GM-CSF serum concentrations (pg/ml) in different mouse strains after repetitive injections of either normal rabbit IgG (NR-IgG) or anti-type VII collagen IgG (anti-COL7 IgG). Data has been obtained from at least 3 mice per group, *indicates statistical significance (student’s t-test). §Data on B6 mice has already been published elsewhere (Samavedam UK et al, 2013), but has been included here for completeness.

B. Ex vivo stimulation of isolated human neutrophils and monocytes with EBA-IC, induced a time-dependent release of GM-CSF (*/**/*** p=0.01, Mann-Whitney Rank Sum Test)
Supplement figure 2: Reduced BAFF expressing neutrophils in draining lymphnodes

A. Representative sections of draining lymph nodes stained with DAPI (blue), Ly6G (red) and BAFF (green). B. Average number of Ly6G & BAFF co-localized cells in lymph nodes of C57Bl/6 and GM-CSF^-/- mice at the end of the experiment showed a significant reduction of Ly6G and BAFF cells in GM-CSF^-/- mice (n=5 (10 fields), *p<0.001, t-test).