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Semaphorin 4A Exerts a Proangiogenic Effect by Enhancing Vascular Endothelial Growth Factor-A Expression in Macrophages

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S emaphorins (Semas) are a large family of extracellular proteins present from viruses to primates and divided into eight classes. Classes 1, 4, 5, 6, and 7 are membrane-bound, whereas classes 2, 3, and V are secreted (1–3), though some

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membrane-associated Semas become soluble after proteolytic cleavage (4). Originally discovered as steering molecules for developing axons, Semas have subsequently been shown to be involved in neuronal function, vascular patterning, heart development, and the immune response (1). The biological effects of Semas impinge on plexins that, alone or in association with other coreceptors endowed with modulatory functions, signal inside cells (5, 6). However, most secreted Semas, including Sema3A and Sema3F, do not bind directly to plexins and use neuropilins as coreceptor ligand-binding subunits (7–9). As reported in axon navigation (10–12), Semas may be both chemorepellent and chemoattractive on endothelial cells (ECs) (1). Class 3 Semas have also been implicated in experimental models of tumor angiogenesis (13). For instance, Sema3E regulates cancer growth, tumor dissemination, and angiogenesis (14). Moreover, recent studies indicated that Sema3A is an endogenous angiogenic inhibitor that regulates the angiogenic switch, inhibits tumor growth, and normalizes tumor vasculature by reducing tumor hypoxia (15).

Besides class 3 Semas, class 4 Semas also regulate angiogenesis as well. Both in vitro and in vivo Sema4D promotes angiogenesis through PlexinB1 (16). More recently, these observations have been extended to tumor angiogenesis, and strong evidence indicated that macrophages recruited in tumors are a major source of Sema4D (17). Another membrane-bound class 4 Semaphorin, namely Sema4A, progressively increases its expression in developing mouse embryos (18) and becomes prominent in the adult brain, lung, kidney, spleen, testis, and mammary gland (19). In contrast to Sema4D, upon binding to PlexinD1, Sema4A inhibits EC migration and in vivo angiogenesis by suppressing the vascular endothelial growth factor (VEGF)-mediated activation of Rac and integrin-dependent cell adhesion (20). Moreover, in the presence of the Rho family GTPase Rnd1, the binding of Sema4A to...
PlexinB1-B2 and B3 induces COS7 cell contraction through the R-Ras GTGase-activating protein enzymatic activity that characterizes all Plexin cytodomains studied so far (21). Besides acting as a chemorepulsive cue via B-type plexins (21, 22), Sema4A has been shown to be constitutively expressed by dendritic cells, where it stimulates T cell activation through Tim-2 receptor, a member of the T cell Ig and mucine domain proteins expressed on activated T cells (19). Moreover, T cells require Sema4A to allow Th cell differentiation. Sema4A-deficient mice displayed defective Th1 responses, indicating that the induction of Sema4A in Th1 cells is necessary for their differentiation either by cognate cellular interactions among Th1 cells or by following an autocrine pathway (23). Recently, Sema4A has been shown to be upregulated by allergen or VEGF in bronchial epithelial cells expressing both PlexinD1 and PlexinB1, thus pointing to Sema4A as a potential player for the allergic airway inflammatory diseases (24).

This evidence highlights the important role played by Sema4A in regulating angiogenesis and the immune system function; however, its specific involvement in controlling both angiogenic and inflammatory response in macrophages is still poorly understood. In this work, we analyzed the expression and the function of Sema4A in macrophages in vitro and in vivo experimental models, including chicken chorioallantoic membrane (CAM) angiogenic assay, thioglycollate-induced peritonitis, and a model of cardiac ischemia/reperfusion (I/R) injury, characterized by increased angiogenesis, tissue remodeling, and recruitment of inflammatory cells (25–28). In this study, we identify a new function for Sema4A as modulator of macrophage functions in the context of both angiogenic and inflammatory processes.

Materials and Methods

Cell culture

Human monocytes were isolated from buffy coats of healthy donors obtained through the courtesy of Transfusion Center AVIS (Torino, Italy). Blood was washed once with PBS 1× at 400 × g to remove plasma and platelets and then centrifuged at 600 × g for 30 min at room temperature in a Picoll gradient (GE Healthcare). Cells at the interface were collected, washed twice with PBS 1×, and monocytes were isolated in a Percoll gradient (Sigma–Aldrich) at 700 × g for 30 min at room temperature. Cells at the interface were collected, washed twice with PBS 1×, and plated at a density of 1.5 × 10⁵ cells/cm² in RPMI 1640 (Lonza). Monocytes were allowed to adhere for 1 h, and then the medium was replaced by RPMI 1640 supplemented with 10% FCS. HUVECs were isolated from umbilical cord veins, characterized, and grown in M199 (Lonza) containing 20% FCS, Insulin (Inovogen), bovine brain extract, heparin (50 μg/ml; Sigma–Aldrich), and penicillin-streptomycin (200 U/ml; Lonza) on gelatin-coated tissue-culture dishes, as previously described (29).

Human macrophage cell culture conditions, differentiation, and polarization

To obtain differentiated macrophages, human monocytes fromuffy coats were cultured for 7 d in RPMI 1640 supplemented with 10% FCS and 100 ng/ml M-CSF (PeproTech) at normoxic oxygen levels (21% O₂) in a humidified incubator at 37°C. For hypoxic experiments, differentiated macrophages were incubated for a further 18 h at an atmosphere of 1% O₂. One week-differentiated macrophages were polarized by replacing the medium for additional 18 h with RPMI 1640 supplemented with 5% FCS. To obtain polarization, 100 ng/ml LPS (Sigma–Aldrich) with 20 ng/ml IFN-γ (PeproTech; M1 polarization) or 20 ng/ml IL-4 (PeproTech; M2 polarization) was added to the medium for 18 h. Differentiated macrophages were exposed to PAM3Cys (2 μg/ml; Calbiochem), polyinosin–poly- cytidylic acid (poly I:C; 100 μg/ml; Invivogen), flagellin (5 μg/ml; Invivogen), R848 (3 μg/ml; Alexo Biochemicals), and CpG ODN 2006 (1 μg/ml; MWG Biotech) for 18 h.

Real-time RT-PCR analysis

RT-PCR analysis was performed as previously described (15). cDNA from I/R tissues and Sema4A-treated macrophages were analyzed by TaqMan Low Density Array based on Applied Biosystems Microfluidic Card (Applied Biosystems) containing 48 probes for I/R tissues and 96 probes for macrophages. cDNA from cells were analyzed by real-time RT-PCR and performed in triplicate using a TaqMan Gene Expression Assay Mix (Applied Biosystems) specific for Sema4A, PlexinD1, PlexinB1, and PlexinB2. cDNAs were run on an ABI Prism 7900HT Fast Real-Time PCR System (Applied Biosystems), and the data were analyzed by SDS and Qq Manager Software to obtain a relative quantification based on the arithmetic equation 2⁻ΔΔCt, in which ΔΔCt is the normalized signal level in a sample relative to the normalized signal level in the corresponding calibrator sample. mRNA from I/R tissues were normalized to the housekeeping 18S gene, whereas the GAPDH gene was used to normalize mRNA from human macrophages. Normalized relative quantification (RQ) fold changes were calculated compared with ΔCt of nontreated mice or control cells.

Western blot analysis

Western blot analysis was performed as previously described (30). Briefly, human macrophages, HUVECs, and mice peritoneal cell proteins were extracted with lysis buffer (50 mmol/l Tris-HCl, 150 mmol/l NaCl, 5 mmol/l Na₂VO₄, 1 mmol/l PMSF, 0.01 mmol/l ZnCl₂, protease inhibitors, and 1% Triton X-100). Proteins were quantified using the BCA assay reagent (Thermo Scientific). Equal amounts of proteins (50 μg) were resolved by SDS-PAGE on a 12% or 8% polyacrylamide gel and transferred to nitrocellulose membranes (Amersham Biosciences). After blocking with 3% BSA or 5% skim milk, membranes were incubated with anti-VEGF and anti-Sema4A (PeproTech), anti-Sema3E (Everest), anti-VEGF receptor (VEGFR)-2, anti- phospho–VEGFR-2, anti–phospho-Akt, anti–phospho-ERK, anti-Akt, anti-ERK (Cell Signaling Technology), anti-phospho–VEGFR-1 (R&D Systems), and anti-β-tubulin (Santa Cruz Biotechnology) primary Abs at 4°C overnight. Membranes were incubated with HRP-conjugated secondary Abs, and immunoreactive proteins were visualized by ECL system (GE Healthcare). The intensity of the signal was quantified by means of the Chemidoc Bio-Rad program (Bio-Rad), and data are reported as ratios of Sema4A/β-actin or β-tubulin densitometry.

ELISA

ELISA has been performed as previously described (15). Briefly, macrophage supernatants were collected and concentrated by the use of centrifugal filter devices (Centricon YM-10; Millipore), whereas macrophage proteins were extracted using lysis buffer. For Sema4A detection, coating was performed using 2 μg/ml protein lysates or 50 μl supernatants dispensed into 96-well polystyrene microtiter plates (Nunc). The plate included duplicates of each sample. After incubation at 4°C overnight, plates were washed once with PBS and blocked with 3% BSA in PBS 1× for 1 h at 37°C. Primary Ab anti-Sema4A (PeproTech) was diluted 1:200 in 3% BSA in PBS 1× and added to the plate. After incubation at 37°C for 2 h, plates were rinsed three times, and the secondary Ab peroxidase-conjugated AffiniPure goat anti-rabbit IgG (Jackson ImmunoResearch Laboratories) was diluted 1:2000 in wash solution and added to the plate. The plate was incubated at 37°C for 1 h and washed again. The 1-step Turbo TMB-Elisa peroxidase substrate (Pierce) was added and incubated at room temperature in the dark. The reaction was stopped by adding 2 M H₂SO₄. The OD was measured at a wavelength of 450 nm using a Synergy HT automated microplate reader (Bio-Tek). For VEGF-A detection, supernatants from macrophages were collected after 24 h, and Sema4A (10 and 100 mmol/l) for 18 h were collected, concentrated, and loaded on a Quantikine 96-well plate for VEGF-A level determination (R&D Systems). To generate a standard curve, we measured the absorbance of increasing concentration of recombinant Sema4A or VEGF-A (10, 25, 50, and 100 ng).

Migration assay

The migration of macrophages was measured with a 48-well chemotaxis Boyden chamber (Neuro Probe) using polyvinylpyrrolidone filter membranes with a pore diameter of 5 μm. Upon 18 h exposure to LPS (100 ng/ml) and IFN-γ (20 ng/ml), human macrophages were harvested with Accutase (PAAB Laboratories) and resuspended in RPMI 1640 supplemented with 1% FCS at a concentration of 4 × 10⁵ cells/ml. Lower wells of the chamber were loaded with 29 μl RPMI 1640 plus 1% FCS alone or supplemented with 50 mmol/l human recombinant Sema4A or Sema3E (R&D Systems). Upper wells were loaded with 50 μl cells alone or in the presence of 50 mmol/l recombinant Sema4A, 20 μg/ml rabbit anti-PlexinD1 Ab (H-70, sc-67145; Santa Cruz Biotechnology), 20 μg/ml mouse anti-PlexinB2 Ab (MAB5329; R&D Systems), and 20 μg/ml IgG or IgG2A controls, respectively (DakoCytomation). After incubation for 4 h at 37°C, the top side of the insert membrane was scrubbed free of cells.
and the bottom side was fixed with methanol and stained with hematoxylin and eosin. Cells that migrated to the bottom side of the membrane were counted under an Olympus BX60F-3 microscope (Olympus) using a 10X original magnification. For ECs migration assay, cells resuspended in M199 medium were seeded on the upper surface of a polycarbonate 8-µm pore Transwell membrane (BD Falcon) and allowed to migrate toward the human macrophage monolayer. After 5 h of incubation, ECs on the upper side of the filters were then mechanically removed. ECs of the filter's lower side were then fixed in 2.5% glutaraldehyde for 30 min and stained with 0.1% crystal violet (15, 30).

Chicken embryo chorioallantoic membrane assay

Fertilized chicken eggs were incubated at 37°C in a humidified incubator, as previously described (31). On day 10 of incubation, CAM were added with sterilized paper disks. The disks were saturated with 100 ng VEGF-A in the presence or not of human recombinant Sema4A (100 nmol/l) or bevacizumab (34 µg/CAM). In addition, the disks were loaded with 10 µl conditioned medium (CM) from human macrophages or 10 µl CM from human recombinant Sema4A in the presence or not of human VEGF-A (30 ng/ml). For EC and macrophage coculture migration assay, a monolayer of human macrophages was incubated for 18 h with Sema4A (10 and 100 nmol/l) with or without bevacizumab (1 µg/ml; Avastin; Roche), anti–IL-8 (1 µg/ml; MAB208; R&D Systems), anti–PlexinD1 (20 µg/ml; Santa Cruz Biotechnology), or anti–PlexinB2 (20 µg/ml; R&D Systems) blocking Abs or isotype control Abs (1 and 20 µg/ml; DakoCyto). ECs suspended in RPMI 1640 medium supplemented with 1% FCS and seeded on the upper surface of a polycarbonate 8-µm porous Transwell membrane (BD Falcon) were allowed to migrate toward the human macrophage monolayer. After 5 h of incubation, ECs on the upper side of the filters were then mechanically removed. ECs of the filter's lower side were then fixed in 2.5% glutaraldehyde for 30 min and stained with 0.1% crystal violet (15, 30).

Thiglycollate-induced peritonitis

Peritonitis was induced by instillation of thiglycollate as previously described (32). Healthy C57/Bl6 mice were subjected to i.p. injection of a sterile 2.95% thioglycollate solution. After 24 and 48 h, mice were anesthetized with isoflurane 1.5% and euthanized, 5 ml sterile PBS 1× and 1.5 ml air were injected in the peritoneal cavity, and peritoneal exudates were collected. For each experimental condition, pools of eight animals were used. Peritoneal cells were pelleted, washed with PBS 1×, and used for immunofluorescence, FACS analysis, and RNA protein extraction.

Flow cytometry analysis

Mouse peritoneal cells were recovered washed twice with PBS 1× and stained for flow cytometry. A total of 2.5 × 10^7 cells was incubated with anti–Sema4A-FTTC (MBL), anti–F4/80-PE (AbD Serotec), anti–Tim-2-647 (BioLegend), and anti–PlexinD1 and PlexinB2 (Santa Cruz Biotechnology) conjugated with allophycocyanin in PBS/1% FCS for 30 min in the dark. Plexins were revealed by incubation with allophycocyanin-secondary Ab (Jackson & Jackson). Samples were washed, resuspended in PBS 1×, and acquired and analyzed on a flow cytometer (CyAn ADP; DakoCyto). Blood from control and thioglycollate-treated C57/Bl6 mice was collected, erythrocytes were lysed with lysis buffer (8.3 g/l NH4Cl, 1 g/l KHCO3, and 0.1 mmol/l EDTA [pH 7.4]) for 10 min in ice, and, after centrifugation, pellets were resuspended in PBS/1% FCS and characterized for anti–Sema4A-FTTC (MBL), anti–CD11b (M170)-APC (BD Pharmingen), anti–CD115 (AFS98)-PE (eBioscience), and biotinylated anti–Ly6c (BD Pharmingen) for 30 min in the dark. Cell samples were fixed and analyzed on a flow cytometer (CyAn ADP; DakoCyto).

Immunofluorescence confocal scanning microscopy

Tissues from C57/Bl6 mice were embedded in OCT, frozen in dry ice, and stored at −80°C. Ten-micrometer-thick sections were cut using a Leica CM1900 cryostat (Leica Microsystems). Mice peritoneal exudates were centrifuged, and cells were resuspended in PBS 1× at the concentration of 10^6/100 µl. A total of 0.1 ml cell suspension was applied to single-well slides using a Cytospin 2 (Shandon). Tissue and cell sections were fixed and permeabilized. After blocking, tissues were incubated and characterized with anti–Sema4A C-20 (Santa Cruz Biotechnology) together with Abs anti–CD11b (BioLegend), -Ly6G (Abcam), -CD45 (Sigma-Aldrich), -F4/80, -CD68, -CD4, -CD8, and -CD11c (AbD Serotec). Secondary Abs were conjugated to Alexa Fluor 555, Alexa Fluor 647, and Alexa Fluor 488 fluorochromes (Invitrogen). Cell nuclei were DAPI counterstained (1:5000; Invitrogen). Immunofluorescence images were captured and analyzed with a Leica TCS SP2 AOBs confocal laser-scanning microscope (Leica Microsystems), as previously shown (15). All immune-localization experiments were repeated three times on multiple tissue sections and included negative and IgG controls for determination of background staining, which was negligible. To quantify the extent of colocalization, we measured the mean fluorescence intensity of red and green channels by means of the Leica Confocal Software Histogram Quantification Tool (Leica Microsystems). In each analyzed picture, we considered five random regions of interest of the same size, and then we calculated the ratio between red (Sema4A) and green (CD11b or CD68) channel mean fluorescence intensity.

In vivo induction of IR

Twelve-week-old C57BL/6j mice (The Jackson Laboratory) were used and maintained in specific pathogen-free organism facilities. All procedures were conducted in conformity with the institutional guidelines in compliance with national and international laws and policies and approved by the Ethical Commission of the University of Turin and by the Italian Ministry of Health. Animals were anesthetized with isoflurane 1.5%, ventilated with a mechanical ventilator (0.2 ml, 120 acts/min; Ugo Basile 28026 mouse ventilator; Ugo Basile) through an endotracheal cannula, and kept at a body temperature of 37°C. The left anterior descending coronary artery was ligated with a 7–0 silk (Ethicon) suture after exteriorization of the heart through a 15-mm opening at the fourth intercostal space. An overhand knot was tied with two pieces of suture to arrest blood flow and then removed after 45 min. Ischemia was confirmed by the appearance of ventricular ectopy and blanching of the myocardium. The chest was closed under negative pressure, and mice were weaned from mechanical ventilation. Post-surgical analgesia was achieved by buprenorphine (0.1 mg/kg s.c. every 12 h for 1 d). Mice were sacrificed after 1, 6, 12, 24, and 48 h and 6 d of IR, and hearts were excised. The right ventricle and the septum and left ventricular free wall (i.e., infarcted myocardium) and sham-operated hearts were separately snap-frozen and used for RNA extraction. Whole hearts were embedded in OCT and stored at −80°C for tissue analysis.

Statistical analysis

The results of all experiments are expressed as mean ± SD. For all statistical analyses, a two-tailed, unpaired Mann–Whitney U test was used. A p value <0.05 was considered significant.

Results

Sema4A is expressed in activated human macrophages

Sema4A enhances T cell activation (19), but its role in the innate immune system and, in particular, on the activation of macrophages, is still unknown. Therefore, we first analyzed Sema4A expression in human macrophages obtained by in vitro differentiation of PBMCs. Significant amount of Sema4A transcript was present in nonstimulated macrophages and significantly increased by LPS, but not by IL-4 (data not shown). IFN-γ alone did not modulate the gene expression profile and slightly decreased the induction by LPS (Fig. 1A). LPS, but not IFN-γ or IFN-γ plus LPS, significantly increased the levels of PlexinD1 and PlexinB2 (Supplemental Fig. 1B, 1C and data not shown), whereas PlexinB1 was poorly expressed both in resting and activated cells (data not shown). LPS activates cellular response mainly by activating TLR4, a member of the TLR family (33) that recognizes pathogen-specific molecular patterns, thus playing a crucial role in both innate and adaptive immunity (34). Therefore, we investigated the expression of Sema4A and its receptors after macrophage activation by other TLR agonists (PAM3Cys, a TLR1-TLR2 agonist; poly I:C, a TLR3 agonist; flagellin, a TLR5 agonist; R848, a TLR7/8 agonist; and CpG ODN 2006, a TLR9 agonist). Transcript levels of Sema4A were strongly increased by long-term exposure to LPS, poly I:C, and, to a lesser extent, CpG ODN 2006 (Supplemental Fig. 1A), whereas no
significant modulation was observed for the other TLR agonists flagellin, R848, and PAM 3Cys (data not shown). Interestingly, poly I:C and LPS treatment also resulted in the upregulation of PlexinD1 and PlexinB2 transcripts, whereas the other TLR agonists did not modulate their expression (Supplemental Fig. 1B, 1C and data not shown). PlexinB1 was not significantly regulated by any TLR agonist (data not shown). All together, these data indicate that Sema4A expression is induced during selective macrophage activation, and indeed, it may play an important role in the regulation of their inflammatory properties.

**Sema4A stimulates the chemotactic activity of macrophages via PlexinD1**

To better understand the role of Sema4A in human macrophages, we evaluated its activity in eliciting their migration. In a chemotactic Boyden’s chamber assay, human recombinant Sema4A enhanced macrophage migration via PlexinD1 (Fig. 1B). Human recombinant Sema4A (from 0 up to 100 nmol/l) increased the migration in Boyden’s chamber of human macrophages treated for 18 h with LPS and IFN-γ in a dose-response curve with a peak at 50 nmol/l of Sema4A. Values are mean ± SD of four independent experiments. (C) Human macrophages exposed for 18 h to LPS and IFN-γ were incubated in the presence or not of 20 μg/ml of PlexinD1 or PlexinB2-blocking Abs or appropriate control Ig and allowed to migrate toward 50 nmol/l of recombinant Sema4A for 4 h. Values are mean ± SD (n = 4 donors) of four independent experiments. (D) Human macrophages exposed for 18 h to LPS and IFN-γ were incubated in the presence or not of 20 μg/ml of PlexinD1-blocking Ab or IgG and stimulated with 50 nmol/l of recombinant Sema3E and Sema4A for 4 h. Values are mean ± SD (n = 4 donors) of four independent experiments. (E and F) Sema4A and Sema3E protein levels were analyzed by Western blot analysis of lysates of control or LPS- and IFN-γ–treated human macrophages. The data shown are representative (n = 4 donors) of four independent experiments. (F) Relative levels of Sema4A and Sema3E protein expression normalized to β-tubulin. (G) Protein levels of Sema4A were measured by ELISA both in the supernatants and cell lysates of human macrophages exposed or not for 18 h to LPS and IFN-γ (see Materials and Methods). Values are mean ± SD (n = 4 donors) of four independent experiments. Ctrl, Control. **p < 0.01, ***p < 0.001.

**FIGURE 1.** Sema4A expression is increased in activated human macrophages. (A) Real-time RT-PCR analysis using RNA from human macrophages exposed to IFN-γ, LPS alone, or combined with IFN-γ for 18 h. RQ values are compared with control untreated macrophages and are mean ± SD of n = 3 donors for each condition. Sema4A enhanced macrophage migration via PlexinD1. (B) Human recombinant Sema4A (from 0 up to 100 nmol/l) increased the migration in Boyden’s chamber of human macrophages treated for 18 h with LPS and IFN-γ in a dose-response curve with a peak at 50 nmol/l of Sema4A. Values are mean ± SD of four independent experiments. (C) Human macrophages exposed for 18 h to LPS and IFN-γ were incubated in the presence or not of 20 μg/ml of PlexinD1 or PlexinB2-blocking Abs or appropriate control Ig and allowed to migrate toward 50 nmol/l of recombinant Sema4A for 4 h. Values are mean ± SD (n = 4 donors) of four independent experiments. (D) Human macrophages exposed for 18 h to LPS and IFN-γ were incubated in the presence or not of 20 μg/ml of PlexinD1-blocking Ab or IgG and stimulated with 50 nmol/l of recombinant Sema3E and Sema4A for 4 h. Values are mean ± SD (n = 4 donors) of four independent experiments. (E and F) Sema4A and Sema3E protein levels were analyzed by Western blot analysis of lysates of control or LPS- and IFN-γ–treated human macrophages. The data shown are representative (n = 4 donors) of four independent experiments. (F) Relative levels of Sema4A and Sema3E protein expression normalized to β-tubulin. (G) Protein levels of Sema4A were measured by ELISA both in the supernatants and cell lysates of human macrophages exposed or not for 18 h to LPS and IFN-γ (see Materials and Methods). Values are mean ± SD (n = 4 donors) of four independent experiments. Ctrl, Control. **p < 0.01, ***p < 0.001.
inhibitory effect on cell motility induced by PlexinD1-blocking Ab. Of note, PlexinD1 neutralization in macrophages triggered by Sema4A associated with Sema3E was not effective in reducing the migration of macrophages under basal levels (Fig. 1D), as observed in the presence of Sema4A and PlexinD1-blocking Ab (Fig. 1C). Based on these results, we can hypothesize that this effect could be mostly due to a competition between Sema3E and Sema4A for the PlexinD1 receptor. By Western blot analysis, we observed that Sema4A, but not Sema3E, expression increased in activated cells compared with control macrophages (Fig. 1E, 1F). Remarkably, we detected significant amounts of both cell-associated and released forms of Sema4A in macrophages (Fig. 1G), as detected by ELISA. The amount of Sema4A protein significantly increased in both supernatants and lysates of activated macrophages, highlighting that Sema4A acts as a soluble factor regulating macrophage functions both in a paracrine and autocrine manner.

**Sema4A is upregulated during peritoneal inflammation**

Next, we extended the above in vitro observations to a murine model of peritonitis induced by thioglycollate and looked at the expression of Sema4A in peritoneal recruited cells. CD11c, CD4, CD8, and Ly6G+ cells did not express Sema4A, neither in controls nor in thioglycollate-treated animals (data not shown). In contrast, whereas CD11b+ and CD68+ cells isolated from control animals did not express significant amounts of Sema4A, thioglycollate treatment induced a dramatic upregulation of the protein in these cells (Fig. 2). Interestingly, after 24 h, Sema4A was mainly expressed by CD11b+ cells with few positive CD68+ macrophages (Fig. 2C, 2D, 2F). Remarkably, 48 h after the treatment, the majority of CD11b+ cells were negative (data not shown), whereas Sema4A accumulated in CD68+ cells (Fig. 2E, 2F).

FACS analysis revealed that peritoneal cells positive for the macrophage F4/80 marker expressed increasing levels of Sema4A upon thioglycollate treatment (Fig. 3A). In fact, the number of Sema4A-expressing peritoneal cells rose from 8% in resting conditions to 48 and 58% 24 and 48 h after thioglycollate injection, respectively. This increased expression of Sema4A in peritoneal cells 24 and 48 h after thioglycollate treatment was confirmed by Western blot analysis (Fig. 3D, 3E). Next, we investigated the expression of Sema4A receptors in these peritoneal cells. Interestingly, both PlexinD1 and PlexinB2, significantly expressed in control cells, were strongly upregulated in Sema4A+ macrophages upon 24 and 48 h of thioglycollate treatment. It has been shown that under inflammatory conditions, various inflammatory cell types including macrophages increased Tim-2 expression, which is one of the Sema4A receptors (2, 24). Therefore, we evaluated the expression of Tim-2 in peritoneal cells upon

![FIGURE 2. Sema4A is upregulated in thioglycollate-induced peritonitis in mice. Cells from control and elicited peritoneal exudates of 24 and 48 h thioglycollate-treated mice were analyzed by means of fluorescent confocal microscopy analysis. (A and B) No significant amount of Sema4A has been detected in macrophages and their precursors isolated from untreated mice, whereas 24 h of thioglycollate treatment increased Sema4A expression in recruited myeloid precursors (C) and, to a lesser extent, in mature macrophages (D) as revealed by colocalization of Sema4A (red) with CD11b and CD68 (green) Abs (C, D). Forty-eight hours after thioglycollate treatment, Sema4A was abundantly present in CD68+ cells (E). No signal was detected using a control goat IgG Ab (not shown). Confocal analysis has been performed on tissue sections from n = 8 mice per time point, and images are representative of five fields observed per mouse. Scale bars, 25 μm. (F) Bar graph shows percentage of Sema4A colocalization with CD11b+ and CD68+ cells. **p < 0.01.](http://www.jimmunol.org/Downloadedfrom/300026535725712485139)
thioglycollate treatment. Of note, whereas we observed a significant Tim-2 expression in both control and elicited Sema4A + macrophages, differently from the other two receptors, we did not detect a significant increase of Tim-2 after the inflammatory stimulus compared with untreated cells (Fig. 3B, 3C).

Circulating monocytes can be grouped in inflammatory and resident subsets depending on the level of expression of the Ly6C marker (36). When cell-surface expression of Sema4A was analyzed by flow cytometry on Ly6Chigh and Ly6Clow peripheral blood monocyte subsets, it was higher on Ly6Chigh than on Ly6Clow monocytes, both in control and in thioglycollate-treated mice. Furthermore, thioglycollate significantly upregulated Sema4A expression on Ly6Chigh monocytes but not in Ly6Clow cells (Fig. 3F). These data suggest that Sema4A belongs to a specific signature for circulating inflammatory monocytes that could migrate to sites of tissue damage.

Sema4A increases VEGF-A expression in human macrophages through PlexinD1

To further study the involvement of Sema4A in macrophage response, we analyzed its effect on the expression of genes encoding for chemokines and angiogenic inducers (Supplemental Tables I, II). Sema4A selectively upregulated VEGF-A and, to a lesser extent, IL-8, whereas it did not modify the expression of other angiogenic factors and chemokines compared with controls (Fig. 4, Supplemental Tables I, II). Consistent with the gene-expression analysis, we detected increased VEGF-A protein expression in lysates and supernatants of macrophages treated with Sema4A (Fig. 4, Supplemental Fig. 1D).

To evaluate the cellular mechanism of the increased expression of VEGF-A induced by Sema4A, we investigated whether this effect was mediated by PlexinD1 or PlexinB2. As detected by ELISA, we observed that PlexinD1, but not PlexinB2-blocking, Abs strongly reduced the enhanced VEGF-A levels induced by both endogenous and exogenous Sema4A in macrophage supernatants (Fig. 4E) and cell lysates (Fig. 4F). Therefore, these data indicate that Sema4A induces VEGF-A expression through a plexin-mediated pathway and that PlexinD1 is critical in mediating the effects of Sema4A.
Because VEGF-A enhances monocyte/macrophage activity and triggers their motility (42–44), we assessed whether Sema4A was able to increase macrophage function through VEGFR-1 or VEGFR-2 activation. Western blot analysis of lysates of macrophages exposed or not to Sema4A revealed barely detectable levels of phospho–VEGFR-2 in both control and Sema4A-stimulated cells. Remarkably, Sema4A induced an increase of VEGFR-1 phosphorylation (Fig. 4G). These data suggest that the increased activation of VEGFR-1 could be involved, in part, in the Sema4A promigratory effect on macrophages.

Sema4A-treated macrophages increase in vitro and in vivo angiogenesis through VEGF-A/VEGFR-2 signal pathway

To study the proangiogenic effect of macrophages mediated by Sema4A, we first assessed whether the VEGF-A expressed by Sema4A-treated macrophages was able to stimulate EC functions. To this aim, we stimulated HUVECs with supernatants of control and Sema4A-stimulated macrophages, and we checked both VEGFR-2 phosphorylation and the VEGF-induced signal pathway. VEGFR-2 phosphorylation was increased in ECs exposed to supernatants of Sema4A-stimulated macrophages, but not in ECs incubated with CM of control cells (Fig. 5B, 5D). These data suggest that Sema4A-treated macrophages exert a proangiogenic effect by activating specifically the PI3K/Akt pathway in ECs.

Because VEGF-A enhances monocyte/macrophage activity and triggers their motility (42–44), we assessed whether Sema4A was able to increase macrophage function through VEGFR-1 or VEGFR-2 activation. Western blot analysis of lysates of macrophages exposed or not to Sema4A revealed barely detectable levels of phospho–VEGFR-2 in both control and Sema4A-stimulated cells. Remarkably, Sema4A induced an increase of VEGFR-1 phosphorylation (Fig. 4G). These data suggest that the increased activation of VEGFR-1 could be involved, in part, in the Sema4A promigratory effect on macrophages.

FIGURE 4. Sema4A induces a proangiogenic activity on human macrophages. (A) Real-time RT-PCR analysis using RNA from human macrophages treated for 18 h with human recombinant Sema4A (10, 50, and 100 nmol/l). RQ values are compared with control macrophages and are mean ± SD of n = 4 donors for each condition. (B and C) VEGF-A protein levels were analyzed by Western blot analysis of lysates of human macrophages upon 18-h treatment with different concentrations of recombinant Sema4A (10, 50, and 100 nmol/l). The data shown are representative (n = 4 donors) of four independent experiments. (C) Relative levels of VEGF-A protein expression normalized to β-tubulin. (D) Protein levels of VEGF-A were measured by ELISA in the supernatants and lysates of human macrophages treated with 50 nmol/l of human recombinant Sema4A in the presence or not of PlexinD1 and PlexinB2-blocking Abs or LPS. Values are mean ± SD (n = 4 donors) of four independent experiments. (G) Phospho–VEGFR-2 and phospho–VEGFR-1 protein levels were analyzed by Western blot analysis of lysates of human macrophages upon 18-h treatment with 50 nmol/l of recombinant Sema4A. The data shown are representative (n = 4 donors) of four independent experiments. **p < 0.01, ***p < 0.001.
genesis induced by Sema4A-elicited macrophages was completely blocked by bevacizumab (Fig. 6F, 6G), indicating that VEGF-A specifically mediates the proangiogenic effect of Sema4A on macrophages in vivo (Fig. 6F, 6G). Taken together, these data demonstrate that Sema4A activates a proangiogenic phenotype in macrophages by specifically inducing the expression of VEGF-A, which, in turn, enhances EC migration and in vivo angiogenesis.

Sema4A is upregulated in the heart after I/R

Based on the data describing the role played by Sema4A during peritoneal inflammation and on its ability to induce a proangiogenic phenotype in macrophages, we next investigated its expression in the cardiac I/R experimental mouse model, which is characterized by activated angiogenesis and inflammatory response. It has been shown that blood reperfusion by eliciting the recruitment of inflammatory cells such as monocytes at the injury site, accentuates the inflammatory cardiac response to ischemia (26–28). To this aim, mice were sacrificed at different time points after heart I/R, and the expression of Sema4A in the infarcted and control sham-operated tissues was evaluated. Sema4A expression was evident after 6 h, peaked at 24 h, and persisted up to 48 h (Fig. 7A). Of note, Sema4D, which was previously implicated in angiogenesis and inflammation (16, 17), was not significantly modulated during cardiac ischemia (Fig. 7A). Moreover, the increased expression of Sema3A, Sema3E, Sema3F, Sema6A, and Sema7A transcripts at 24 h I/R (Supplemental Fig. 2A, 2B) agrees well with their role in regulating angiogenesis, thus suggesting a potential role of these Semas in the early stages of the ischemic process (15, 45, 46). By Western blot analysis, we detected a very similar expression pattern of Sema4A protein at 24 and 48 h after I/R (Fig. 7C, 7D).

Next, we focused our analysis on the differential expression in these experimental conditions of Sema4A receptors, showing that whereas PlexinD1 expression was unchanged, PlexinB1 was significantly downregulated, and PlexinB2 was upregulated (Fig. 7B). These data suggest the involvement of Sema4A and its receptors in the cellular processes occurring after an ischemic event in the heart.

Sema4A is specifically expressed by myeloid precursors and mature macrophages in the infarcted heart

To better understand the involvement of Sema4A in the cardiac ischemia after I/R, we investigated Sema4A protein localization and expression at the different time points after reperfusion by confocal microscope analysis (Fig. 8). Sema4A was not expressed in the noninfarcted heart (i.e., interventricular septum and right ventricle), but its expression was readily detected in the infarcted and necrotic free wall of the left ventricle (Fig. 8A) (47). Notably, Sema4A was not detectable in the heart of sham-operated control mice (Fig. 8B). Next, we examined Sema4A expression pattern in
different cell types that can be found within the infarcted cardiac tissue and noticed that this protein did not colocalize with either α-actinin+ cardiomyocytes or CD31+ vascular ECs (Supplemental Fig. 2C, 2D). Based on the scattered staining of Sema4A, we then evaluated its expression in the different subsets of recruited leukocytes. At 24 h post–I/R, Sema4A did not colocalize with Ly6G+, CD4+, CD8+, or CD11c+ cells (data not shown). Conversely, Sema4A was expressed by CD11b+ and a subset of CD68+ cells infiltrating the tissue at 24 h post-I/R (Fig. 8C, 8D, 8F). At 48 h post–I/R, when most of the CD11b+ cells were replaced by CD68+ mature macrophages, Sema4A was mainly localized in these cells (Fig. 8E, 8F). Moreover, at 48 h, Sema4A colocalized with CD11b+ cells not yet differentiated into macrophages and, to a lesser extent, with Ly6G+ cells. Similar results were obtained using anti-F4/80 Ab at 24 and 48 h post–I/R (data not shown).

Taken together, these data indicate that Sema4A is produced by mature macrophages recruited at injury sites and suggest it may play an important role in regulating their functions during cardiac I/R.

Discussion

In the last years, a growing body of evidence demonstrated an important role of Semas and their receptors plexins in regulating cellular functions related to angiogenesis and immunity (2, 3, 15, 46). Among these, Sema4A regulates the function of dendritic cells and T lymphocytes and inhibits EC migration and experimental angiogenesis (19, 20), but its role in mediating the interplay between inflammatory and angiogenic processes is still unclear. By using in vitro approaches and in vivo models of peritoneal inflammation and cardiac ischemia, we uncovered in this study a new role of Sema4A in regulating macrophage functions in angiogenesis and inflammation.

In this study, we showed that exogenous Sema4A stimulated macrophage migration in a dose-dependent manner. Moreover, we found that PlexinD1 is the most upregulated plexin in human macrophages stimulated with LPS and poly I:C, suggesting its potential involvement in the response of macrophages to Sema4A during the inflammatory process. Consistently, blocking Ab anti-PlexinD1 but not anti-PlexinB2 abrogated for the migratory effect of Sema4A. The observation that Sema4A slightly increased the expression of IL-8, but did not significantly change the expression levels of any inflammatory chemokine tested (Supple-
Sema3E did not enhance macrophage migration and that Sema3E expression was not enhanced in activated cells. Furthermore, our data of the simultaneous treatment of macrophages with Sema4A and Sema3E in presence of PlexinD1-blocking Ab suggest a competition between the two Semas for PlexinD1 receptor. Even though we did not observe a significant increase of Sema3E expression in activated macrophages, we can hypothesize that the low levels of Sema3E produced by macrophages may be sufficient to counteract the effects of Sema4A on basal migration. Our findings unveil a crucial and specific role of Sema4A in enhancing the motility of activated macrophages and suggest that the competition with Sema3E may regulate the activity of macrophages during inflammation or pathological angiogenesis.

While looking for additional mechanisms by which Sema4A could regulate macrophages, we noticed that the stimulation of these cells with Sema4A induced a significant phosphorylation of VEGFR-1, but not of VEGFR-2. Notably, it has been shown that VEGFR-1 is the principal receptor regulating the VEGF-A–triggered motility of monocytes/macrophages. Moreover, VEGFR-1 mediates the activation and recruitment of these cells during chronic inflammation and pathological angiogenesis (42, 43, 49, 50). Based on these findings and our data, we can assume that the increased activation of VEGFR-1 induced by the Sema4A/PlexinD1 pathway and the consequent binding with VEGF-A produced by macrophages or other cell types could be part of the mechanism by which Sema4A exerts its promigratory effect on macrophages. Recently, Sema4A has been reported to bind to all three B-type plexins (PlexinB1, B2, and B3) and plexins found on macrophages. Recently, Sema4A has been reported to bind to all three B-type plexins (PlexinB1, B2, and B3) and plexins found on macrophages. Recently, Sema4A has been reported to bind to all three B-type plexins (PlexinB1, B2, and B3) and plexins found on macrophages. Recently, Sema4A has been reported to bind to all three B-type plexins (PlexinB1, B2, and B3) and plexins found on macrophages.
pathways, leading to overexpression of VEGF-A by NF-κB activation. TGF-β1 has been recently involved in the upregulation of VEGFRs and VEGF-A expression in macrophages and dendritic cells through Smad3/4 (58). Based on our findings describing the activation of VEGFR-1 in macrophages upon stimulation with exogenous Sema4A, we can hypothesize that the activation of the TGF-β1/Smad signaling pathway may represent an alternative mechanism by which Sema4A upregulates VEGF-A expression in these cells.

The increased ECs migration and vessel formation in vivo induced by macrophages treated with recombinant Sema4A is consistent with the induction by Sema4A of a proangiogenic phenotype in macrophages. It has been recently described that Sema4A inhibits ECs functions in experimental angiogenesis (20). In this study, we described that Sema4A can exert an indirect proangiogenic effect mediated by macrophages that trigger EC migration. Interestingly, a similar opposite effect of Sema4A was recently described in the context of cancer metastasis: Sema4A was shown to induce a proangiogenic phenotype in melanoma cells, leading to increased angiogenesis and tumor growth (62). Based on our data, we can speculate that during pathological angiogenesis, Sema4A may promote neovascularization by recruiting VEGF-A–expressing macrophages to activate ECs.

In this article, we show that VEGF-A produced by Sema4A-elicited macrophages activates a specific signal pathway in ECs by increasing the levels of phospho-Akt. It has been widely described that PI3K/Akt pathways can regulate endothelial migration, proliferation, and survival through the effect of its downstream targets such as endothelial NO synthase, p70 ribosomal protein S6 kinase 1, and forhead box factor O1 to regulate tumor angiogenesis (59–61). In addition, endothelial Akt activation causes the appearance of an enlarged and highly permeable vasculature that recapitulates the abnormal structural and functional phenotype of tumor blood vessels (62). Based on our data, it is conceivable to hypothesize that the induction of a proangiogenic phenotype in macrophages by Sema4A causes the activation of the endothelial PI3K/Akt pathway leading to an increased EC migration and in vivo vessel formation that could prove crucial for angiogenesis in cancers and other pathological conditions. Remarkably, we detected a significant amount of Sema4A in the culture medium of differentiated human macrophages, corroborating previous observations demonstrating that a soluble form of Sema4A is cleaved from membrane-bound Sema4A overexpressed in human embryonic kidney 293 (20). These data indicate therefore that Sema4A can be expressed as soluble mediator and exerts its promigratory function on macrophages as locally released protein. Therefore, the final effects of Sema4A on angiogenesis result from a homeostatic balance between its anti- and proangiogenic activities, respectively, mediated by a direct effect on ECs and macrophages.

It has been widely shown that during cardiac I/R, monocytes/macrophages play a crucial role in the phagocytosis of the necrotic myocardium and the inflammatory response necessary for tissue healing (26). We show in this study for the first time, to our knowledge, that Sema4A is specifically expressed by macrophages recruited in the infarcted area and significantly enhanced macrophage migration, suggesting that Sema4A may be involved in recruitment and activation of a subset of macrophages that contribute early to tissue remodeling after I/R.

Interestingly, by performing a wider gene expression analysis of axon guidance cues at the early stages of cardiac ischemia, we revealed a significant modulation of several Semas along the injury development. Interestingly, at the early stages, members of class 3, such as Sema3A, Sema3E, and Sema3F, known to be relevant for pathological angiogenesis (45), were upregulated. This is reminiscent of the observations done in tumor mouse models describing that the angiogenic inhibitors Sema3A and Sema3F were upregulated to balance the prominent production of angiogenic inducers (15). Moreover, Sema6A, another angiogenesis inhibitor (63), and Sema7A, a mediator of T cell–mediated inflammatory response (64), were upregulated as well. Altogether, these results suggest a complex role of Semas in tissue repair of ischemic injury. This concept is further supported by the persistent expression of a high level of Sema4A early on during the I/R process. Further studies will be performed to better understand this new role of Sema4A on angiogenesis in a pathological condition.

In conclusion, our work unveils a new role of Sema4A in inducing a restricted genetic program in macrophages able to sustain angiogenesis. Based on our in vivo and in vitro observations, it is conceivable to hypothesize that at the site of inflammatory injuries or during pathological angiogenesis such as I/R, Sema4A could recruit macrophages to the ischemic/tumor tissue able to regulate the angiogenic response necessary for tissue healing and remodeling. The increased expression of Sema4A may therefore contribute to the activation of the onset of the angiogenesis process by specifically enhancing VEGF-A expression in infiltrating macrophages.

Acknowledgments

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Disclosures

The authors have no financial conflicts of interest.

References


PROANGIOGENIC ROLE of Sema4A ON MACROPHAGES


SUPPLEMENTAL MATERIAL

Sema4A exerts a pro-angiogenic effect by enhancing VEGF-A expression in macrophages
Claudia Meda, Fabiola Molla, Maria De Pizzol, Donatella Regano, Federica Maione, Stefania Capano, Massimo Locati, Alberto Mantovani, Roberto Latini, Federico Bussolino and Enrico Giraudo

Supplemental Figure Legends

Supplemental Figure 1. Sema4A expression is increased in TLR agonist-stimulated human macrophages. (A, B, C) Real-Time RT-PCR analysis using RNA from human macrophages exposed to TLR agonists. Among different TLR agonists, Poly I:C, LPS and, to a lesser extent, CpG strongly increased Sema4A expression (A). Poly I:C and LPS increased PlexinD1 (B) and PlexinB2 (C) expression. Normalized Relative Quantification (RQ) values are compared to control untreated macrophages and are mean ± SD of n=3 donors for each condition.

Expression of Sema4A and VEGF-A under hypoxic conditions. (D) The gene expression of VEGF-A and Sema4A was evaluated by Real-Time RT-PCR of RNA extracted from macrophages exposed or not to human recombinant Sema4A (10 and 100 nM) for 18 h under hypoxic conditions. Hypoxia did not modulate Sema4A (control) transcript, but induced VEGF-A gene expression compared to normoxic conditions. The treatment with Sema4A did not significantly increase VEGF-A gene levels under hypoxic controls. Normalized Relative Quantification (RQ) values are compared to control macrophages in normoxic conditions and are mean ± SD of n=4 donors for each condition.

Sema4A directly inhibits endothelial cells migration. (E, F) The chemotactic activity of ECs (HUVECs) through filter membranes treated with 0.1% gelatine was significantly inhibited when incubated with 10 or 50 nmol/L of human recombinant Sema4A with or without VEGF-A (30
ng/mL), compared to VEGF-A alone (**p<0.01). (C) Similar results were obtained when the chemotaxis was assessed using 50 μg/mL fibronectin-treated filter membranes (**p<0.01).

Supplemental Figure 2. Gene expression profile of Semaphorins and Plexins in the heart after I/R. (A,B) TaqMan low-density array was employed to analyze the gene expression of 48 axon guidance cues. 24 h after I/R Sema3A, Sema3E, Sema3F (A) and Sema6A (B) transcripts were significantly up-regulated. (B) Sema7A mRNA was significantly increased during cardiac ischemia in all time points analyzed. Normalized Relative Quantification (RQ) values represent the gene expression of infarcted compared to sham-operated hearts (Control) and are mean ± SD of n=6 mice/per time point.

Sema4A is not expressed by cardiomyocytes and endothelial cells after I/R. (C, D) Fluorescent confocal microscopy analysis of I/R tissue at 24 h. Sema4A (red) was not expressed by cardiomyocytes (α-actinin, green) (C), and by ECs (CD31+ cells, green) (D). Images are representative of n=6 mice. Confocal analysis has been performed on tissue sections from n=6 mice and images are representative of five fields observed per mouse. Scale bar = 25 μm.
Supplemental Figure 1

A

- Sema4A (RQ)
- Poly I:C
- LPS
- CpG

B

- PlexinD1 (RQ)
- Poly I:C
- LPS
- CpG

C

- PlexinB2 (RQ)
- Poly I:C
- LPS
- CpG

D

- Hypoxia
- VEGF-A
- Sema4A

E

- Cell number
- Sema4A 10nM
- Sema4A 50nM
- VEGF-A

F

- Cell number
- Sema4A 10nM
- Sema4A 50nM
- VEGF-A
Supplemental Figure 2

A

B

C

D

Sema4A

DAPI

/g68

-actinin

Sema4A

DAPI

CD31

Supplemental Figure 2

A

B

C

D

Sema4A

α-actinin

DAPI

Sema4A

CD31

DAPI
### Gene expression of angiogenic factors in Sema4A-treated human macrophages

<table>
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<td>Sema4A 10 nM</td>
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<tr>
<td>Angiopoietin-1</td>
<td>1,08 +/- 0,12</td>
</tr>
<tr>
<td>Angiopoietin-2</td>
<td>0,57 +/- 0,09</td>
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<tr>
<td>GM-CSF</td>
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<tr>
<td>HGF</td>
<td>1,40 +/- 0,25</td>
</tr>
<tr>
<td>IL6</td>
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<tr>
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<tr>
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<td>TNF-α</td>
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The gene expression of angiogenic factors modulated by Sema4A was analyzed by Real-Time RT-PCR using RNAs from macrophages after 16 h of stimulation with human recombinant Sema4A (10 and 100 nM). Normalized Relative Quantification (RQ) values are compared to untreated macrophages and are mean +/- SD of n=4 donors for each condition.
Supplemental Table 2

Gene expression of inflammatory chemokines in Sema4A-treated human macrophages

<table>
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<tr>
<td></td>
<td>Sema4A 10 nM</td>
</tr>
<tr>
<td>CCL2</td>
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<td>CCL3</td>
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<td>CCR1</td>
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The expression levels of inflammatory chemokines was analyzed by using RNAs from macrophages stimulated for 16 h with human recombinant Sema4A Real-Time RT-PCR (10 and 100 nM). Normalized Relative Quantification (RQ) values are compared to untreated macrophages and are mean +/- SD of n=4 donors for each condition.