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Pathogenic Natural Antibodies Propagate Cerebral Injury Following Ischemic Stroke in Mice

Andrew Elvington,*1 Carl Atkinson,*1 Liudmila Kulik,† Hong Zhu,‡ Jin Yu,‡ Mark S. Kindy,‡§ V. Michael Holers,† and Stephen Tomlinson*§

Self-reactive natural Abs initiate injury following ischemia and reperfusion of certain tissues, but their role in ischemic stroke is unknown. We investigated neoeptiope expression in the postischemic brain and the role of natural Abs in recognizing these epitopes and mediating complement-dependent injury. A novel IgM mAb recognizing a subset of phospholipids (C2) and a previously characterized anti-annexin IV mAb (B4) were used to reconstitute and characterize injury in Ab-deficient Rag1−/− mice after 60 min of middle cerebral artery occlusion and reperfusion. Reconstitution with C2 or B4 mAb in otherwise protected Rag1−/− mice restored injury to that seen in wild-type (wt) mice, as demonstrated by infarct volume, demyelination, and neurologic scoring. IgM deposition was demonstrated in both wt mice and reconstituted Rag1−/− mice, and IgM colocalized with the complement activation fragment C3d following B4 mAb reconstitution. Further, recombinant annexin IV significantly reduced infarct volumes in wt mice and in Rag1−/− mice administered normal mouse serum, demonstrating that a single Ab reactivity is sufficient to develop cerebral ischemia reperfusion injury in the context of an entire natural Ab repertoire. Finally, C2 and B4 mAbs bound to hypoxic, but not normoxic, human endothelial cells in vitro. Thus, the binding of pathogenic natural IgM to postischemic neoeptiopes initiates complement-dependent injury following murine cerebral ischemia and reperfusion, and, based also on previous data investigating IgM reactivity in human serum, there appears to be a similar recognition system in both mouse and man. The Journal of Immunology, 2012, 188: 1460–1468.

A number of events occur during tissue ischemia, and the pathophysiology of ischemia reperfusion injury (IRI) is complex. Clinical and animal studies have established a causal role for complement in IRI of various organs and tissues (1), including the brain following ischemic stroke (2). Cleavage of complement component C3 is a central event in complement activation, and studies with C3-deficient (3) and inhibited (4) mice have revealed a key role for complement in murine ischemic stroke. Complement can be activated via the classical, lectin, and alternative pathways, and recent data indicate a central role for the lectin pathway in ischemic stroke (5, 6).

In models of intestinal (7, 8), myocardial (9), and skeletal muscle (10) IRI, complement activation is triggered by natural circulating IgM that recognizes cellular neoeptiopes that become exposed as a result of ischemia. These pathogenic natural Abs recognize a restricted Ag repertoire and are mainly the product of B-1 lymphocytes in mice and humans (11) and are constitutively expressed throughout life. Although IgM bound to reperfused tissue recruits both C1q and mannose-binding lectin (MBL) (classical and lectin pathway recognition molecules, respectively) (12, 13), IRI has been shown to be dependent on MBL binding to IgM, at least with regard to intestinal and myocardial IRI (14–16).

Seminal studies by Zhang et al. (7) showed that Ab-deficient Rag1−/− mice were resistant to intestinal IRI and that natural self-reactive IgM restored IRI. Subsequently, the target of a clonally specific mAb that reconstituted intestinal IRI in Rag1−/− mice was identified as nonmuscle myosin (17), and the same Ab has since been shown to also restore myocardial and skeletal muscle IRI in Rag1−/− mice (9, 18). However, even though a peptide mimic of nonmuscle myosin can block IRI in wild-type (wt) mice (9, 19), it is clear that other targets for self-reactive Abs exist, at least in the postischemic intestine. In this context, intestinal IRI in Rag1−/− mice can also be restored by administration of an anti-annexin IV IgM mAb (8) or the combined administration of anti-phospholipid and anti-β2-gp1 mAbs (20).

Thus, whereas current evidence indicates that multiple cellular neoeptiopes are exposed postischemia, the predominance of their expression in different tissues, as well as the relative contributions of different targets and self-reactive Abs in driving IRI in different tissues, is not known. In this study, we identify a novel IgM mAb that recognizes a subset of phospholipids and show that this mAb, as well as a previously characterized anti-annexin IV mAb, recognizes postischemic neoeptiopes in the brain, activates complement, and propagates cerebral IRI.

Materials and Methods

C2 mAb isolation and purification

The C2 mAb hybridoma was isolated following the fusion of spleen cells from unmanipulated wt C57BL/6 mice as described previously (8). Briefly, splenocytes from healthy C57BL/6 mice were fused with the SP2/0-AG14 myeloma cell line by standard protocol to establish...
hybridomas. The hybridomas were then screened by both Western blot analysis using intact epithelial cell lysates and by flow cytometric analysis of isolated intestine epithelial cells. Positive wells were further subcloned until a monoclonal population was obtained. To purify mAbs, Ab from the exhausted supernatants of cultured hybridomas was affinity purified on a column of agarose beads with goat anti-human IgM (Sigma-Aldrich, St. Louis, MO). Bound mAb was eluted with a buffer containing 0.1 M glycine (pH 2.3) and collected into a buffer containing 1.5 M Tris (pH 8.8). Eluted mAb was dialyzed against PBS (pH 7.4) for 48 h and concentrated using centrifugal filtration on Centricon Plus-20 (Millipore, Billerica, MA). Ab concentration was determined by measuring the absorbance at 280 nm of the sample and purity was confirmed by analysis on 10% SDS-PAGE gel.

Characterization of C2 mAb and anti-phospholipid Abs in mouse serum

ELISAs to determine reactivity of Abs to various phospholipids were performed using microtiter plates (Immulon 1B; Dynatech Laboratories, Chantilly, VA) coated with 100 μlwell 50 μg/ml phospholipid in methanol. The plates were dried under blowing air to allow the organic solvent to evaporate, and the wells were then washed with PBS and blocked with 1% BSA. Supernatant from the mAb hybridoma cell line was added to wells and bound Ab detected by alkaline phosphatase-conjugated goat anti-mouse IgM (Jackson ImmunoResearch Laboratories, West Grove, PA). For detection of Abs, serial dilutions of mouse serum samples prepared in RPMI 1640 containing 10% FBS were added to wells coated with phospholipids and bound Ab detected with AP-conjugated anti-mouse IgG or anti-mouse IgM Ab, followed by p-nitrophenylphosphate (Sigma-Aldrich) at 1 mg/ml. Relative units of Ab were calculated by comparing OD at 405 nm for individual titrated serum with standard curve of OD measurements of titrated standard high-titer polyclonal antiphospholipid hybridoma supernatants. High-titer polyclonal supernatants were generated during the isolation of the C2 mAb hybridoma; they represent polyclonal hybridoma supernatants. High-titer polyclonal supernatants were generated during the isolation of the C2 mAb hybridoma; they represent polyclonal hybridoma supernatants. Phospholipids assayed: phosphatidylethanolamine (PE)-1,2-diacyl-sn-glycero-3-phosphoethanolamine, phosphatidylserine (PS)-1,2-distearoyl-sn-glycerol-3-[phospho-L-serine] (Avanti). Abs were detected using the goat-IMMpress (Vector Laboratories), and negative controls omitted primary Abs. Slides were coverslipped with Cytoseal-60 (Richard-Allan Scientific, Kalamazoo, MI) and imaged by light microscopy.

Immunohistochemistry

Cryosections were cut at 8 μm, fixed in cold acetone, washed in running water, and equilibrated in PBS. Double staining for C3d and IgM deposition was performed. Goat anti-mouse C3d (1:20; R&D Systems) was applied and detected with rat anti-goat IgG Alexa Fluor-555 conjugate (1:200; Invitrogen, Carlsbad, CA). After washing with PBS, anti-mouse IgM FITC conjugate (1:50; Sigma-Aldrich) was applied, followed by ToPro-3 (1:5000; Invitrogen) as a nuclear marker. Slides were coverslipped with Vecta fluorescent hard mount (Vector Laboratories) and imaged on a Leica TCS-SP2 confocal microscope (Leica Microsystems).

Annexin IV purification and treatment

Recombinant annexin IV was generated and purified as previously described (8). In one experiment, annexin IV was administered i.v. to C57BL/ 6 mice (100 μg in 100 μl PBS) 5 min prior to reperfusion. In a second experiment, annexin IV (100 μg in 100 μl PBS) was administered i.v. to Rag1−/− mice 5 min prior to infusion of 300 μl freshly isolated normal mouse serum, which was administered just prior to reperfusion. Briefly, recombinant protein was expressed in transformed Escherichia coli by 0.3 mM isopropyl-D-thiogalactoside. Bacteria were collected and lysed, and following centrifugation, the supernatant was adjusted to pH 7.6 and run on a TALON resin column (BD Clontech, Mountain View, CA). A discontinuous urea gradient was used to refold the protein, and an imidazole gradient was used to elute the protein. Coomassie staining confirmed recombinant protein purity.

In vitro binding of IgM

Mouse brain endothelial cells (bEnd.3; American Type Culture Collection, Manassas, VA) and human endothelial cells (HUVEC; Lonza, Walkersville, MD) were grown to at least 80% confluence in Endothelial Growth Media (Lonza) on chambered microscope slides (Nunc, Rochester, NY). Slides were washed with PBS and incubated in DMEM serum-free medium for 3 h in a Coy Anaerobic Chamber (Coy Laboratory products, Grass Lake, MI) with O2 monitoring in not >0.1% oxygen. The medium was then supplemented with IgM mAb (15 μg/ml) and incubation continued under normoxic cell-culture conditions for 3 h. The slides were fixed in cold acetone, and IgM was detected with an IgM anti-mouse IgM-FITC (1:100; Sigma-Aldrich). Slides were coverslipped with Vecta fluorescent hard mount (Vector Laboratories) and imaged on a Leica TCS-SP2 confocal microscope (Leica Microsystems). Differences in detecting viable cells after hypoxic culture precluded analysis by flow cytometry.

Statistical analysis

Statistical analysis was done using Prism 4 (Graphpad). Infarct volumes were compared using ANOVA or Student t test, and neurologic deficits were compared using the Kruskal–Wallis or Mann–Whitney U test, as applicable. The p values <0.05 were considered significant.

Neurological deficit

Neurological deficit was determined, independent, and blinded, as described (26). Scoring was assigned as follows: 0, normal motor function; 1, torso and contralateral forelimb flexion when lifted by tail; 2, contralateral circling when held by tail on flat surface, though normal at rest; 3, contralateral leaning when at rest; and 4, no spontaneous motor activity.
Results

C2 mAb recognizes a subset of phospholipids

Kulik et al. (8) previously reported a strategy to isolate and identify mAbs that bind to neoepitopes exposed on ischemic cells. The strategy involved the use of isolated intestinal epithelial cells expressing ischemia or apoptosis-associated neoepitopes to screen and identify hybridomas created from B cells of wt C57BL/6 mice (8). In this study, we characterize three of the IgM mAbs isolated by this technique (D5, B4, and C2) in a model of murine ischemic stroke. The specificity of two of these mAbs has been previously determined (8); B4 mAb recognizes annexin IV, and D5 mAb, used in this study as an isotype control Ab, recognizes cytokeratin 19. C2 mAb did not react in Western blots of intestinal epithelial cells (not shown), and because altered phospholipid exposure on apoptotic/ischemic cells has been suggested as a target for pathogenic natural Abs (20, 27), we investigated C2 mAb specificity in antiphospholipid ELISA. C2 was shown to recognize a subset of phospholipids that included phosphatidylcholine, PE, and cardiolipin (CL), but not PG or PS (Fig. 1). B4 mAb and D5 mAb did not recognize any phospholipid tested (Fig. 1).

mAbs B4 and C2 restore injury in Rag1−/− mice following ischemic stroke

C57BL/6 wt and Ab-deficient Rag1−/− mice were subjected to 1 h MCAO-induced cerebral ischemia followed by 24 h reperfusion. Rag1−/− mice showed significantly improved survival at 24 h compared with wt controls, with 100% (18 out of 18) and 59% (10 out of 16) of Rag1−/− and wt mice surviving, respectively (p = 0.006). Infarct size was also significantly reduced 24 h postreperfusion in Rag1−/− mice compared with surviving wt mice (Fig. 2B), and this is in agreement with previous data (21, 22). Rag1−/− mice also displayed an improved neurologic function (Fig. 2C), correlating with the reduction of infarct volume.

It was shown previously that B4 mAb restores intestine IRI in otherwise protected Rag1−/− mice, identifying annexin IV as a postischemic neoepitope expressed in the intestine (8). B4 mAb, but not D5 mAb, also restored cerebral IRI in Rag1−/− mice in terms of infarct size and neurologic outcome, demonstrating that annexin IV is also expressed postischemically in the brain (Fig. 2). C2 mAb also restored cerebral IRI in Rag1−/− mice. To investigate whether there may be a quantitative difference in the postischemic exposure of annexin IV and a subset of phospholipids in terms of IgM-dependent cerebral IRI, we performed mAb dose-response reconstitution experiments. There was no significant difference between the ability of B4 mAb or C2 mAb to restore post-IR infarct volume or neurologic deficit in Rag1−/−/− mice (Fig. 2). In a separate experiment, we also demonstrated that C2 mAb restored intestinal IRI in Rag1−/−/− mice (data not shown).

The effect of Rag1 deficiency and Ab reconstitution on cerebral IRI was further investigated by analysis of demyelination at 24 h postreperfusion. Luxol Fast Blue and Nissl staining of brain sections revealed less myelin loss in the ipsilateral brain of Rag1−/−/− mice compared with wt mice, with restoration of myelin loss in Rag1−/−/− mice reconstituted with B4 and C2 mAbs (Fig. 3). The contralateral brain of all groups was unaffected following MCAO or mAb administration (Fig. 3).

As expected, there was no damage observed in the contralateral hemisphere in any group (not shown). Also, to confirm that cerebral blood flow was interrupted by the MCAO procedure, blood flow was measured by laser Doppler before ischemia, during ischemia, and 10 min postischemia (4). There were no significant differences in cerebral blood flow between any of the groups (not shown). Changes in blood pressure and body temperature can significantly influence the outcome post stroke, and we therefore measured blood pressure, heart rate, and temperature before, during, and after ischemia in Rag1−/−/− and wt mice. There were no differences between the groups (Supplemental Table I).

Analysis of IgM binding and C3 deposition following ischemic stroke

The deposition of C3 in postischemic mouse brains has been shown previously, and C3 deficiency or inhibition protects against ischemic stroke (3, 4). To investigate the relationship between postischemic IgM binding and complement activation, we investigated IgM and C3 deposition in infarcted areas of the brain at 24 h postreperfusion (C3 is a cleaved activation fragment of C3). As would be expected, there was no detectable IgM in postischemic brain sections from Rag1−/−/− mice, and only very low levels of C3 deposition were detected (Fig. 4A). In contrast, postischemic brain sections from wt mice and Rag1−/−/− mice reconstituted with either B4 or C2 mAb showed high levels of deposited IgM and C3. IgM deposition was not detected in brains of Rag1−/−/− mice reconstituted with mAb D5 (Fig. 4A), and no IgM or C3 deposition was detected in contralateral brain tissue (not shown). Additional confocal studies revealed colocalization of IgM and C3 on vessel endothelium of postischemic brains from Rag1−/−/− mice reconstituted with B4 (Fig. 4B). Together, these results indicate specific binding of B4 and C2 mAbs to neo-
epitopes in the postischemic brain with subsequent IgM-mediated activation of complement.

Recombinant annexin IV blocks cerebral IRI in wt mice
Reconstitution of cerebral IRI in Rag1−/− mice with either B4 or C2, but not D5, shows that specific IgM is sufficient to mediate cerebral injury. Previous data from Carroll’s group (9, 17) has shown that an IgM mAb recognizing nonmuscle myosin on postischemic tissue is also capable of reconstituting IRI, at least in the intestine, heart, and hind limb. Thus, multiple neoepitopes are expressed on postischemic tissue, and to investigate whether a single Ab reactivity is sufficient to develop cerebral IRI in the context of an entire natural Ab repertoire, we investigated the effect of recombinant annexin IV on cerebral IRI in wt mice. Recombinant annexin IV (100 μg) or vehicle control (PBS) was injected into wt mice 5 min before reperfusion. Recombinant annexin IV significantly reduced infarct volumes, with percentage infarct similar to that seen in Ab-deficient Rag1−/− mice (Fig. 5A). There was also a strong trend toward reduced neurologic deficit in annexin IV-treated mice (Fig. 5B), although this did not reach significance.

Normal mouse serum restores injury in Rag1−/− mice following ischemic stroke
To address the physiological relevance of the above mAb reconstitution experiments in the context of cerebral IRI, we determined whether normal mouse serum was capable of restoring injury in Rag1−/− mice. After 1 h of MCAO and upon reperfusion, Rag1−/− mice were administered 300 μl of freshly isolated pooled C57BL/6 mouse serum via tail vein injection. At 24 h following ischemia, Rag1−/− mice that were reconstituted with mouse serum had significantly larger infarct volumes (Fig. 6A) compared with Rag1−/− controls (injected with 300 μl saline). Infarct data were functionally relevant in that the increased infarct resulted in significantly worsened neurologic deficits (Fig. 6B). Normal mouse serum restored injury levels to those seen in wt mice (refer to Fig. 2). Data above showing that recombinant annexin IV protected wt mice from cerebral IRI (Fig. 5) indicate that a single Ab reactivity is sufficient to drive injury in the context of an entire natural Ab repertoire. To further strengthen this conclusion, we also administered recombinant annexin IV together with normal mouse serum to Rag1−/− mice. The cerebral injury seen in Rag1−/− mice after administration of normal mouse serum was reversed with the coadministration of recombinant annexin IV in terms of infarct volume (Fig. 6A). There was also a strong trend toward improved neurologic deficit (p = 0.058; Fig. 6B). In control experiments, we also infused wt mouse serum into wt mice prior to either MCAO or a sham procedure; there was no deleterious effect of the serum infusion itself (Fig. 6A).

Natural Abs recognizing similar phospholipids as C2 mAb are present in wt mice
It has been shown previously that natural IgM Abs to annexin IV are present in wt C57BL/6 mice (8). We used an ELISA to investigate specificities and relative levels of anti-phospholipid Abs in wt C57BL/6 mouse serum. IgM reactivity to phosphatidylcholine and PE, albeit variable, was found in all sera analyzed (Fig. 7A), and these two phospholipids are recognized by C2 mAb (refer to Fig. 1). IgG reactivity to certain phospholipids was also seen in some sera (Fig. 7B), although the overall proportion of sera with negative reactivity was much higher than for IgM.

B4 and C2 mAbs bind to hypoxic but not normoxic human endothelial cells
Previous work has demonstrated the presence of anti-annexin IV and anti-phospholipid IgM in the natural human Ab repertoire (8, 28), and postischemic blockade of annexin IV or phospholipid neoepitopes represent a potential therapeutic strategy to inhibit IRI. Also, direct translation development of B4 and C2 mAb derivatives may be feasible. We therefore determined whether human endothelial cells express B4 and C2 epitopes following exposure to hypoxia. HUVEC were exposed to a period of hypoxia followed by a period of reoxygenation and then incubated with B4, C2, or control D5 mAb. Immunofluorescence detection
showed strong binding of both B4 and C2 mAbs to hypoxic but not normoxic, cells (bEnd.3). As with HUVEC, both mAbs bound strongly to hypoxic but not normoxic, cells (Fig. 8B).

Discussion

In this study, we identified a novel IgM mAb (C2) that recognizes a subset of phospholipids and that reconstitutes cerebral IRI in Rag1−/− mice. The mAb was identified from a panel of mAbs recognizing intestine epithelial cells, a major cellular target of intestine IRI, using the same approach for the previous identification of anti-annexin IV B4 mAb (8). Previous studies have shown that anti-phospholipid and anti-annexin IV Abs are present in normal mouse and human serum (8, 20), and because both C2 and B4 mAbs restored cerebral IRI in Rag1−/− mice, the data indicate that distinct neoepitopes and their recognition by natural Abs are involved in the propagation of acute cerebral IRI. To put these findings in a more physiological context, we demonstrated that IgM Abs that recognize a similar phospholipid subset as C2 mAb were present in normal mouse serum and that normal mouse serum restored cerebral IRI in Rag1−/− mice. We further showed that IgM is deposited in infarcted areas of wt mouse brains following ischemic stroke, and recombinant annexin IV significantly reduced infarct volumes in wt mice and Rag1−/− mice reconstituted with normal mouse serum, demonstrating that a single Ab reactivity is sufficient to develop injury in the context of an entire natural Ab repertoire. Also, an IgM mAb isolated from the same panel that contained C2 and B4 mAbs [D5 mAb, anti-cytokeratin 19 (8)] did not restore cerebral IRI in Rag1−/− mice, demonstrating that injury was dependent on specific recognition of self-Ag and not passive deposition of IgM. Postischemic binding of IgM activates complement as indicated by the colocalization of IgM with C3d in wt mice and Rag1−/− mice reconstituted with B4 mAb, and previous data have shown that complement activation following cerebral IR results in a proinflammatory and prothrombogenic phenotype within the cerebral microvasculature (4).
Previous studies have shown that T cells traffic to the postischemic brain within 24 h of reperfusion and play a role in the pathophysiology of ischemic stroke. Indeed, reconstitution of Rag1−/− mice with either CD4+ or CD8+ T cells restores injury after cerebral IRI in the same model of transient ischemic stroke used in this study (22). The mechanism of T cell-dependent injury is not known, but it is not dependent on Ag recognition, TCR used in this study (22). The mechanism of T cell-dependent injury in the same model of transient ischemic stroke used in this study does not restore cerebral IRI (31), whereas mice completely deficient in mature B cells sustained more severe renal IRI compared with wt mice. The dual role of B cells was ascribed to the production of pathogenic natural IgM by B-1 cells that bound to postischemic mesangium on the one hand and the production of protective IL-10 by mature B cells on the other (31). Also of interest, B cells and pathogenic Abs have been shown to play an important role in posttraumatic spinal cord injury, a condition involving ischemia and reperfusion of the spinal cord (32). Although it was shown that pathogenic Abs mediated spinal cord pathology, the primary source and specificity of Abs was not determined, and natural Abs produced by B-1 cells are a potential source.

Natural Abs contribute to host defense against infection and serve homeostatic functions in the immune system, as well as contribute to pathogenic processes, as described in this study. Anti-

![Figure 5](http://www.jimmunol.org/Downloadedfrom/1465)

**FIGURE 5.** Effect of recombinant annexin IV on postischemic infarct size and neurologic deficit in C57BL/6 wt mice. Mice were subjected to 60 min MCAO and 24 h reperfusion with injection of 100 μg recombinant annexin IV immediately after reperfusion. A, Infarct volume was significantly reduced in mice treated with 100 μg Annexin IV (p < 0.003 versus control, n = 4). Results are expressed as mean ± SD. B, Neurological deficit was not significantly different between groups at 24 h (p = 0.058, n = 4). Horizontal line represents median score.

Reconstitution of Rag1−/− mice with B cells prior to ischemia in the same model we use in this study does not restore cerebral IRI (21), and although this finding appears to rule out a direct role for B cells in acute murine ischemic stroke, it does not exclude a role for Abs (which must be synthesized by transferred B cells) and, in particular, natural Abs because they are mainly the product of peritoneal B-1 cells and which were not specifically reconstituted in the previous studies. Nevertheless, it has also been shown that B cell-deficient mice are not protected from ischemic stroke (22), which does bring into question the role of both B cells and Abs in cerebral IRI and appears to be in contradiction to the current findings. Although the T cell population is not affected in B cell-deficient mice, the apparent discrepancy may be due to the fact that B cells can have both protective immune suppressive function as well as pathogenic proinflammatory functions. Indeed, a previous study showed that specific depletion of peritoneal B-1 cells did not alter overall circulating levels of IgM, but did reduce renal IgM binding and protected kidney function following renal IR (31), whereas mice completely deficient in mature B cells sustained more severe renal IRI compared with wt mice. The dual role of B cells was ascribed to the production of pathogenic natural IgM by B-1 cells that bound to postischemic mesangium on the one hand and the production of protective IL-10 by mature B cells on the other (31). Also of interest, B cells and pathogenic Abs have been shown to play an important role in posttraumatic spinal cord injury, a condition involving ischemia and reperfusion of the spinal cord (32). Although it was shown that pathogenic Abs mediated spinal cord pathology, the primary source and specificity of Abs was not determined, and natural Abs produced by B-1 cells are a potential source.

Natural Abs contribute to host defense against infection and serve homeostatic functions in the immune system, as well as contribute to pathogenic processes, as described in this study. Anti-
phospholipid Abs have long been recognized as a significant component of the natural Ab repertoire, and Abs to PC (a component of phosphatidylcholine) appear to be a dominant specificity (33). Further, natural Abs for annexin IV, a soluble cytosolic Ca$^{2+}$-dependent membrane binding protein family that has been identified in extracellular fluids and bound to cellular surfaces (34), represent a potential ischemic injury catalyst. Annexin IV is expressed on early apoptotic cells (35) and elevated in the brain in ethanol-induced injury (36, 37) and ischemic injury (36) and, as previously shown in an intestinal murine IRI model (8), can be bound by specific natural Abs to elicit injury. We demonstrated that the C2 mAb binds to PC, as well as certain other phospholipids, such as CL and PE. Thus, the C2 mAb binding epitope is present on various phospholipids, and accessibility of the epitope depends on the polar head because negatively charged phospholipids are not bound by C2 mAb. Our data indicate that the C2 mAb is only exposed on injured or stressed cells. Further, of the antiphospholipid IgM specificities analyzed, anti-PC IgM was present in C57BL/6 sera at the highest relative levels. It is important to note that anti-PC natural Abs do not bind nonoxidized phosphatidylcholine, which explains why they do not interact with healthy cells. The PC headgroup of phosphatidylcholine is exposed following oxidative damage to the polyunsaturated fatty acid side chain in position 2 of the glycerol backbone (38). In addition, apoptotic cell death can also lead to caspase-3 activation of the calcium-independent phospholipase A$_2$ that can remove the fatty acid at the sn-2 position of phosphatidylcholine to generate lyso-phosphatidylcholine, which is also recognized by PC-specific Abs (27, 39). Thus, the maintenance of a high level of anti-PC Ab can be expected to promote the efficient clearance of injured and apoptotic cells. Significantly, it has been shown that reduction in the level of anti-PC Ab is observed in many pathological conditions, such as Alzheimer’s disease, rheumatoid arthritis, multiple sclerosis, and others (40–44).

IgM can activate both the classical and lectin pathways, and previous studies indicate that IgM-mediated activation of the lectin pathway drives injury after intestinal and myocardial ischemia and reperfusion (14, 16). With regard to ischemic stroke, it has been shown that MBL-deficient mice have reduced infarct following cerebral IRI (5, 6) and that genetically defined MBL deficiency is associated with improved outcome after acute stroke in humans (5, 38). Additionally, a recent study by Ducruet et al. (45) found that IgM and C3 localized together in the ischemic cerebral vasculature immediately following reperfusion and that MBL is the

**FIGURE 7.** The wt sera from C57BL/6 mice show presence of Ab binding to phospholipids. Ab binding was detected by ELISA as described in the text. Normal serum samples were tested for levels of IgM Ab (A) and IgG Ab (B) recognizing the following panel of phospholipids: PC-BSA, CL, PG, PE, and PS. Each dot represents a single animal.

**FIGURE 8.** Recognition of hypoxic mouse and human endothelial cells by monoclonal Abs in vitro. HUVEC (A) and mouse brain endothelial cells, bend.3 (B), were subjected to 3 h hypoxia followed by 3 h reoxygenation and C2, B4, and D5 mAb binding determined by immunofluorescence (D5 mAb shown only for HUVEC). Normal culture conditions were used as normoxic control. C2 and B4 mAbs bound to hypoxic, but not normoxic human and mouse endothelial cells. D5 control mAb did not bind to either normal or hypoxic HUVEC. Representative images from three experiments. Original magnification ×63.
first complement protein involved in this ischemic complex. The classical pathway does not appear to play a role in murine ischemic stroke (3), and studies using C1-inhibitor (39), which also inhibits the lectin pathway (40), implicate the lectin pathway. Interestingly, Ducruet et al. (45) also observed that the protective effect afforded by genetic MBL deficiency was lost in the subacute phase of stroke. Although further work is needed, these data support a putative role for complement in neurogenesis subsequent to ischemic stroke (46, 47). Thus, together with the data presented in this study, it is likely that complement-dependent injury after cerebral ischemia and reperfusion is mediated by natural Ab-mediated activation of the lectin pathway, but the role of IgM and the lectin pathway in neurogenesis and long-term outcomes after stroke remains unclear.

In summary, natural IgM has been shown to play an important role in the pathogenesis of IRI in certain organs via complement activation, but its role in cerebral IRI was unknown. In this study, we show that pathogenic IgM plays an important role in activating complement and driving cerebral injury after ischemic stroke, and we identify the involvement of two distinct self-reactive Ab specificities. Previous studies have shown that normal human sera contain IgM reactivity to phospholipids recognized by C2 mAb (28) and to annexin IV (8), in this study, we show that hypoxic, but not normoxic, human endothelial cells also bind C2 and B4 mAb, indicating that similar recognition processes occur in mouse and man.

Disclosures

The authors have no financial interests of conflict.

References


Table I: Physiologic measurements of *Rag1*−/− and wildtype mice before, during, and following middle cerebral artery occlusion \(^a\)

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<th>MAP</th>
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<th>Temp</th>
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<tr>
<td><em>Rag1</em>−/−</td>
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<td><strong>During Ischemia</strong></td>
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\(^a\) MAP, mean arterial (blood) pressure; HR, heart rate; Temp, temperature (in degrees Celsius). Results are shown as mean ± SD, n=10.