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Signal-Transducing Adaptor Protein-2 Modulates Fas-Mediated T Cell Apoptosis by Interacting with Caspase-8

Yuichi Sekine,* Chikako Yamamoto,* Michinori Kakisaka,* Ryuta Muromoto,* Shigeyuki Kon,* Dai Ashitomi,* Natsuko Fujita,† Akihiko Yoshimura,‡ Kenji Oritani,† and Tadashi Matsuda*

We found that an adaptor protein, signal-transducing adaptor protein (STAP)-2, is a new member of the Fas–death-inducing signaling complex and participates in activation-induced cell death in T cells. STAP-2 enhanced Fas-mediated apoptosis and caspase-8 aggregation and activation in Jurkat T cells. Importantly, STAP-2 directly interacted with caspase-8 and Fas, resulting in enhanced interactions between caspase-8 and FADD in the Fas–death-inducing signaling complex. Moreover, STAP-2 protein has a consensus caspase-8 cleavage sequence, VEAD, in its C-terminal domain, and processing of STAP-2 by caspase-8 was crucial for Fas-induced apoptosis. Physiologic roles of STAP-2 were confirmed by observations that STAP-2–deficient mice displayed impaired activation-induced cell death and superantigen-induced T cell depletion. Therefore, STAP-2 is a novel participant in the regulation of T cell apoptosis after stimulation. The Journal of Immunology, 2012, 188: 6194–6204.

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Materials and Methods

Reagents, Abs, and mice

Recombinant human TRAIL, PHA, and dexamethasone (DEX) were obtained from Wako (Osaka, Japan). Recombinant human TNF-α was a gift from Dainippon Sumitomo Pharma (Osaka, Japan). Recombinant murine His-tagged FasL was purchased from R&D Systems (Minneapolis, MN). Propidium iodide, staurosporine, and etoposide were purchased from Sigma-Aldrich (St. Louis, MO). The pan-caspase inhibitor z-IETD-fmk and the caspase-8 specific inhibitor z-VDVAD-fmk were purchased from Calbiochem (Darmstadt, Germany). Expression vector for Halo-tagged human Fas and HaloLink resin were purchased from Promega (Madison, WI). Expression vectors for HA-tagged caspase-8/Mch5 (17) and FLAG-FADD were a gift from Daiichi Sankyo Co., Ltd.

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(18) were provided by T. Miyazaki (Hokkaido University) and S. Akira (Osaka University). Recombinant soluble Fas ligand (sFasL) (19) was provided by S. Nagata (Kyoto University). Epitope-tagged STAP-2, GST-fusion STAP-2 mutants, and Myc-tagged STAP-2 deletion mutants were described previously (14). Myc-tagged STAP-2 D260A (STAP-2 DA), Myc-tagged STAP-2 D260E (STAP-2 DE), and FLAG-tagged-Caspase-8 deletion mutants were generated by PCR methods and sequenced (primer sequences are available on request). Anti-Fas Ab (CH11) was purchased from Ortho Clinical Diagnostics (Raritan, NJ). Anti–caspase-8, anti-caspase-7, anti-cleaved caspase-7, anti-caspase-3, anti-caspase-7, anti-cleaved caspase-7, anti-caspase-8, anti-caspase-3, anti-caspase-8, anti-cleaved PARP, and anti-myc (9B11) Abs were purchased from Cell Signaling Technologies (Beverly, MA). Anti-GST and anti-Fas (M-20) anti-His probe sequences are available on request). Anti-Fas Ab (CH11) was purchased from BD Pharmingen.

The mouse model of in vivo AICD was performed as described previously (23). In brief, anti-CD3 Ab (145-2C11) (5 µg) or isotopic control Ab (5 µg) was injected i.v. into the tail veins of 9–11-wk-old WT or STAP-2 knockout (KO) mice. Then, 20 h later the spleenocytes were isolated from the mice and stained with FITC–anti-TCR-β (BD Pharmingen) and PE–anti-B220 (BD Pharmingen) Abs. The ratio of T cells (TCR-β+) or SEB-unresponsive (Vβ-6+)CD4+ T cells were analyzed by flow cytometry.

**Statistical methods**

The significance of differences between group means was determined by Student t test.

**Results**

**STAP-2 enhances PHA- and DR-mediated apoptosis in Jurkat T cells**

To initially examine the role of STAP-2 in T cell apoptosis, we used Jurkat T cells overexpressing STAP-2 (Jurkat/STAP-2 cells).
We observed significantly enhanced cell death in Jurkat/STAP-2 cells after PHA treatment compared with empty vector-expressing Jurkat T cells (Jurkat/pcDNA3 cells; Fig. 1A). Next, we examined the involvement of STAP-2 in DR-mediated cell death. Fas-mediated cell death was induced in Jurkat T cells by exposure to a Fas agonist mAb. Jurkat/STAP-2 cells displayed higher susceptibility to cell death than Jurkat/pcDNA3 cells. Similarly, Jurkat/STAP-2 cells showed enhanced cell death in response to TNF-α and TRAIL, although their susceptibilities to cell death were lower than that for Fas stimulation. We then examined the involvement of STAP-2 in the Bim-dependent mitochondrial apoptotic pathway in T cells. We used etoposide and staurosporine as Bim-dependent apoptotic stimuli. No significant differences in cell death in response to these drugs were observed between STAP-2-overexpressing and control Jurkat T cells (Supplemental Fig. 1A). Thus, the enhanced cell death displayed in the STAP-2 transfectants is selective for the DR-mediated apoptotic pathway.

FIGURE 1. STAP-2 enhances Fas-mediated apoptosis and caspase activation in Jurkat T cells. (A) Jurkat/pcDNA3 (filled circle), Jurkat/STAP-2#1 (filled triangle) or Jurkat/STAP-2#2 (filled square) cells (2 × 10^4/well) were cultured in 96-well plates in the absence or presence of the increasing amount of PHA (24 h), anti-Fas Ab (24 h), TNF-α (48 h) or TRAIL (48 h). The cell viability was measured using a Cell Counting Kit-8. The data are the means of triplicate experiments, which generally varied by ±10%. Similar results were obtained in three independent experiments. (B) Jurkat/pcDNA3 or Jurkat/STAP-2 cells (1 × 10^6) were stimulated without (−) or with (+) anti-Fas Ab (10 ng/ml). At 12 h after stimulation, apoptotic cells were determined by flow cytometry using TUNEL-staining. Similar results were obtained in three independent experiments. (C) Jurkat/pcDNA3 or Jurkat/STAP-2 cells (1 × 10^6) were stimulated with anti-Fas Ab (10 ng/ml) for the indicated periods. The cells were then lysed, and total cell lysate (TCL) was immunoblotted (IB) with anti–caspase-8, anti-cleaved caspase-8, anti–caspase-3, anti-cleaved caspase-3, anti–caspase-7, anti-cleaved caspase-7, anti-PARP, anti-cleaved PARP, anti-Myc, or anti-actin Ab. (D) Jurkat/pcDNA3 or Jurkat/STAP-2 cells (1 × 10^6) were stimulated with increased amounts of anti-Fas Ab (0, 1, 5, 10 ng/ml). The cells were then assessed for caspase-8 (3 h after stimulation) or caspase-3/7 (5 h after stimulation), and activity was analyzed as described in Materials and Methods. Data are expressed relative to the value of control samples without Fas stimulation. Results are representative of three independent experiments, and the error bars represent the SD of means. (E) Jurkat/STAP-2 cells (2 × 10^4/well) were cultured in 96-well plates without or with anti-Fas Ab (0.5 ng/ml) and increased concentrations of z-VAD-fmk (0.5, 1 μM) or z-IETD-fmk (20, 40 μM) for 24 h. The cell viability was measured using a Cell Counting Kit-8. Similar results were obtained in three independent experiments. Data are the means of triplicate experiments, and the error bars represent the SD of means. *p < 0.05 (Student t test).
Increased caspase activation through Fas in STAP-2-expressing Jurkat T cells

We focused on effects of STAP-2 on Fas-mediated apoptosis, because Fas and FasL have a major role in AICD of T cells. To determine whether STAP-2 affected the susceptibility of Jurkat cells to Fas-mediated cell death, apoptosis of the stable transfectants was evaluated with TUNEL staining (Fig. 1B) as well as annexin V staining (Supplemental Fig. 1B) after the treatment with an anti-Fas mAb. Enhanced induction of apoptosis upon Fas stimulation was detected in Jurkat/STAP-2 cells. We then analyzed the activation of caspases after Fas stimulation. As shown in Fig. 1C, the cleavage of caspase-3, -7 and -8 as well as that of PARP was significantly increased in Jurkat/STAP-2 cells compared with control cells. These findings were confirmed by observations that Fas stimulation markedly increased the caspase enzymatic activities in Jurkat/STAP-2 cells (Fig. 1D), although slightly enhanced basal caspase activation was observed. To confirm the requirement for caspase-8 in the STAP-2–mediated apoptotic process, cells were cultured in the presence of a general caspase inhibitor, Z-VAD, or a caspase-8–specific inhibitor, IETD. Pretreatment with these compounds completely blocked Fas-mediated cell death in Jurkat/STAP-2 cells (Fig. 1E). Therefore, enhanced activation of caspase-8 is a critical step in the increased susceptibility to Fas-mediated cell death exhibited in STAP-2 transfectants.

Caspase-8 is recruited to the multiprotein Fas-DISC that is rapidly formed on the cytoplasmic portion of the Fas receptor after ligand engagement by FADD. Therefore, we examined the effect

FIGURE 2. STAP-2 influences Fas-DISC formation. (A–C) Jurkat/pcDNA3 or STAP-2 cells were stimulated with anti-Fas Ab (20 ng/ml) at the indicated periods. The immunoprecipitate with anti-IgM Ab was separated on SDS-PAGE and immunoblotted with anti-caspase-8 (A), anti-FADD (B), or anti-Myc (C, upper panel). An aliquot of TCL was blotted with the indicated Abs (lower panels). Similar results were obtained in three independent experiments. (D) Jurkat/pcDNA3 or STAP-2 cells (1 × 10⁶) were lysed after stimulation at the indicated times with anti-Fas Ab (20 ng/ml). The cells were lysed, immunoprecipitated (IP) with anti–caspase-8 Ab and immunoblotted with the indicated Abs (upper panels). TCL was also blotted with indicated Abs (lower panels). (E) 293T cells (1 × 10⁶) were transfected with HA-tagged caspase-8 (8 μg) without or with Myc-tagged STAP-2 (10 μg). At 36 h after transfection, the cells were lysed, immunoprecipitated with anti-Myc Ab, and immunoblotted with the indicated Abs (upper panels). TCL was blotted with indicated Abs (lower panels). (F) Human T cell lymphoma, HUT78 cells (1 × 10⁵) were lysed, immunoprecipitated with control IgG or anti–caspase-8 Ab, and immunoblotted with anti–STAP-2 (upper panel) or anti–caspase-8 (lower panel) Ab. (G) 293T cells (1 × 10⁵) were transfected with HA-tagged caspase-8 (15 μg), FLAG-tagged FADD (20 μg) and increasing amount of Myc-tagged STAP-2 (0, 1.5, 5 μg). At 36 h after transfection, the cells were lysed, immunoprecipitated with anti-HA Ab, and immunoblotted with the indicated Abs (upper panels). TCL was blotted with the indicated Abs (lower panels).
of STAP-2 on the Fas-DISC formation. Anti-Fas mAb treatment led to caspase-8 association with Fas in the Fas-DISC, and cleaved caspase-8 was produced (Fig. 2A). Importantly, the amount of caspase-8 in the Fas-DISC was remarkably increased in Jurkat/STAP-2 cells compared with control cells. The amount of FADD in the Fas-DISC was also increased in Jurkat/STAP-2 cells (Fig. 2B). Notably, STAP-2 itself existed in the Fas-DISC in Jurkat/STAP-2 cells (Fig. 2C). Moreover, the association of caspase-8 with FADD was markedly enhanced by overexpression of STAP-2 (Fig. 2D). Thus, STAP-2 is likely to enhance the Fas-DISC formation after Fas stimulation. We then examined effects of STAP-2 on caspase-8 aggregation. Aggregation of caspase-8 in the absence of 2-ME was markedly increased in 293T and Jurkat cells over-expressing STAP-2 (Supplemental Fig. 2A, 2B). These findings indicate that STAP-2 enhances caspase-8 aggregation in the Fas-DISC.

Association of STAP-2 with caspase-8

As shown in Fig. 2E, the immunoprecipitates for STAP-2 contained caspase-8 protein in samples from 293T cells expressing HA–caspase-8 and Myc-STAP2. A similar interaction between STAP-2 and caspase-8 was also observed in Jurkat/STAP-2 cells expressing Myc-tagged STAP-2 (Supplemental Fig. 2C). The immunoprecipitates for STAP-2 contained caspase-8 protein even in the absence of Fas stimulation, suggesting that the interaction between STAP-2 and caspase-8 occurred spontaneously. To confirm the endogenous association of the two proteins, we used a T cell lymphoma cell line, HUT78, which abundantly expressed both endogenous STAP-2 and caspase-8. The immunoprecipitates with the anti–caspase-8 Ab contained endogenous STAP-2 protein, whereas those with the control Ab did not (Fig. 2F). In addition, both endogenous STAP-2 and caspase-8 existed in the Fas-DISC (Supplemental Fig. 2D). Thus, STAP-2 constitutively associates with caspase-8 in cells. Next, we examined whether STAP-2 interacts with FADD, another component of the Fas-DISC. STAP-2 failed to interact with FADD in samples of 293T cells expressing FLAG-FADD and Myc-STAP2 (Supplemental Fig. 2E). Similarly, an association between STAP-2 and FADD was not observed in Jurkat/STAP-2 cells (Supplemental Fig. 2F). However, coexpression of STAP-2 with caspase-8 and FADD caused an enhanced interaction between caspase-8 and FADD, which is consistent with the data shown in Fig. 2G. These findings strongly suggest that STAP-2 has an important role in the complex formation between caspase-8 and FADD.

Next, we determined the caspase-8–interacting domain in STAP-2 using a series of Myc-tagged STAP-2 deletion mutants (Fig. 3A). Deletion mutants of the C-terminal domain or the PH domain of STAP-2 as well as full length of STAP-2 showed interactions with caspase-8, but a deletion mutant lacking the SH2 domain failed to bind to caspase-8 (Fig. 3B). To further confirm these findings, a series of deletion mutants of STAP-2 fused with GST (GST-STAP-2-A, and immunoblotted with the indicated Abs were lysed, immunoprecipitated with anti-FLAG mutants (15) and/or a series of FLAG-tagged caspase-8 deletion mutants (10). At 36 h after transfection, the cells were lysed, immunoprecipitated with anti-Myc Ab, and immunoblotted with the indicated Abs (upper panels). TCL was blotted with indicated Abs (lower panels). (C) 293T cells (1 × 10⁶) were transfected with HA-tagged caspase-8 (8 μg) or a series of Myc-tagged STAP-2 WT or deletion mutants (10 μg). At 36 h after transfection, the cells were lysed, pulled down with glutathione-Sepharose, and immunoblotted with anti-HA or anti-GST Ab (upper panels). TCL was blotted with anti-HA or anti-GST Ab (lower panels). (D) 293T cells (1 × 10⁶) were transfected with Myc-tagged STAP-2 (8 μg) and/or a series of FLAG-tagged caspase-8 deletion mutants (15 μg). At 36 h after transfection, the cells were lysed, immunoprecipitated with anti-FLAG Ab, and immunoblotted with the indicated Abs (upper panels). TCL was blotted with indicated Abs (lower panels). (E) Jurkat transfectants (2 × 10⁶/well) were cultured in 96-well plates with the indicated amounts of anti-Fas Ab for 24 h. The cell viability was measured using a Cell Counting Kit-8. The data are the means of triplicate samples, which generally varied by <10%. Similar results were obtained in three independent experiments. (F) Jurkat transfectants (1 × 10⁶) were stimulated without or with anti-Fas Ab (10 ng/ml) for 3 h. The cells were then assessed for caspase-8 activities. Data are expressed relative to the value of control samples without Fas stimulation. Results are representative of three independent experiments, and the error bars represent the SD of means. *p < 0.05 (Student t test).
PH, GST-STAP-2 SH2, and GST-STAP-2 C) were used (Fig. 3A). The precipitates for the GST–STAP-2 SH2 protein clearly contained caspase-8 protein (Fig. 3C). However, no caspase-8 protein was detected in the precipitates for GST–STAP-2 PH protein or GST–STAP-2 C protein. These findings further indicate that the SH2-like domain of STAP-2 interacts with caspase-8. We also determined the STAP-2–interacting domain on caspase-8 using a series of caspase-8 deletion mutants (Fig. 3A). The two N-terminal death effector domain (DED) domains of caspase-8 showed obvious interactions with STAP-2 as strong as full-length caspase-8 did, whereas the catalytic domain failed to bind to STAP-2 (Fig. 3D). Therefore, caspase-8 interacts with STAP-2 through its N-terminal DED domains.

To assess the physiologic roles of the association between STAP-2 and caspase-8, we used stable Jurkat cell transfectants expressing an empty vector (pcDNA3), WT STAP-2 or STAP-2 ΔSH2, which lacks the SH2-like domain. Significant cell death in response to Fas stimulation was observed in Jurkat/STAP-2 WT cells compared with Jurkat/pcDNA3 cells (Fig. 3E). However, the Fas-induced cell death in Jurkat/STAP-2 ΔSH2 cells was moderate. We also found that the enhanced Fas-induced caspase-8 enzymatic activity was significantly reduced in Jurkat/STAP-2 ΔSH2 cells compared with Jurkat/STAP-2 WT cells (Fig. 3F). Therefore, the association of STAP-2 with caspase-8 through its SH2-like domain enhances Fas-induced cell death and caspase-8 enzymatic activation in Jurkat T cells.

**Proteolytic cleavage of STAP-2 by caspase-8 is required for Fas-mediated signaling**

As seen in Fig. 2A, Fas-mediated proteolytic cleavage of STAP-2 was observed in Jurkat/STAP-2 WT cells after Fas stimulation. Therefore, we focused on the potential role of STAP-2 proteolysis during Fas-mediated signaling. First, we examined the proteolytic cleavage of endogenous STAP-2 after Fas stimulation using HUT78 cells. Cleaved STAP-2 as well as cleaved caspase-8 was observed in HUT78 cells treated with the anti-Fas Ab (Fig. 4A). We then investigated which domains of STAP-2 were cleaved in response to Fas stimulation. Jurkat cells expressing a series of Myc-tagged STAP-2 deletion mutants were treated with an anti-Fas Ab. Mutants lacking the PH or SH2 domain, but not a deletion mutant for the C-terminal domain of STAP-2, successfully generated cleaved STAP-2 within 1 h after Fas stimulation (Fig. 4B), indicating that the proteolytic cleavage site of STAP-2 may exist in the C-terminal domain. To examine the involvement of caspases in the Fas-mediated processing of STAP-2, we transfected specific siRNAs for caspase-3 (nos. 1 and 2) or caspase-8 (nos. 1 and 2) into Jurkat/STAP-2 cells. Reduction in caspase-8 expression resulted in a marked decrease in STAP-2 cleavage after Fas stimulation, whereas the reduction in caspase-3 expression did not (Fig. 4C).

We next tested whether recombinant active caspase-8 can cleave the STAP-2 protein in vitro. As shown in Fig. 4D, recombinant active caspase-8 cleaved the partially purified STAP-2 protein from the Myc-STAP-2–transfected 293T cells. Furthermore, Z-VAD treatment inhibited generation of the cleaved STAP-2.

**FIGURE 4.** Proteolytic cleavage of STAP-2 by caspase-8. (A) HUT78 cells (1 × 10⁷) were stimulated with anti-Fas Ab (100 ng/ml) for the indicated periods. The cells were then lysed, and an aliquot of total cell lysate (TCL) was immunoblotted (IB) with anti–STAP-2, anti–caspase-8 or anti-actin Ab. (B) Jurkat/STAP-2 WT, Jurkat/STAP-2 ΔPH, Jurkat/STAP-2 ΔSH2, and Jurkat/STAP-2 ΔC cells (1 × 10⁶) were stimulated with anti-Fas Ab (10 ng/ml) for the indicated periods. The cells were then lysed, and an aliquot of TCL was immunoblotted with anti-Myc or anti-actin Ab. The asterisks indicate the migration positions of the cleaved STAP-2. (C) Jurkat/STAP-2 cells (5 × 10⁶) were nucleofected with control, caspase-3#1, caspase-3#4, caspase-3#1 or caspase-3#2 siRNA (200 pmol) by nucleofection with cell line Nucleofector Kit V as described in Materials and Methods. At 36 h after nucleofection, the cells were stimulated without (−) or with (+) anti-Fas Ab (20 ng/ml) for 3 h. The cells were then lysed, and an aliquot of TCL was immunoblotted with anti-Myc, anti-caspase-8, anti-caspase-3, or anti-actin Ab. (D) 293T cells (1 × 10⁶) were transfected with Myc-tagged STAP-2 (10 μg). At 36 h after transfection, the cells were lysed and immunoprecipitated with anti-Myc Ab. The immunoprecipitates were soaked with caspase-8 assay buffer and incubated with recombinant active caspase-8 (5 U) at 37°C for 4 h in the absence or presence of Z-VAD. After boiling in the SDS sample buffer, samples were subjected to SDS-PAGE, followed by immunoblotting with anti-Myc or anti-caspase-8 Ab.
Thus, caspase-8 is required for the STAP-2 processing during Fas-mediated signaling. In general, caspases cleave their substrates specifically at the C-terminal side of aspartic acid (the P1 site), which is dictated by arginine and glutamine residues (24). Indeed, the consensus caspase-8 cleavage site was reported to be (I/L/V)EXD (25). Thus, we searched for this consensus sequence in the C-terminal domain of STAP-2, because a deletion mutant of the C-terminal domain failed to produce the cleaved fragment after Fas stimulation. We found a VEAD sequence (residues 257–260) as a potential caspase-8 cleavage site (Fig. 5A). To further evaluate the potential cleavage of STAP-2 at this site, we generated an STAP-2 mutant with substitution of alanine for aspartic acid at

![FIGURE 5. Caspase-8–dependent STAP-2 cleavage occurs at the position of Asp260.](image-url)
Myc Ab (Fas or anti-Myc Ab) and immunoprecipitated with anti-Myc Ab and immunoblotted with anti-Halo-tagged Fas (10 kDa). DISC formation was analyzed by immunoblotting with anti-caspase-8.

followed by cross-linking with anti-His probe mAb for 10 min at 37˚C. The cells were then lysed and immunoprecipitated with anti-Fas Ab, and the Fas-associated protein (Fas-AP) was immunoprecipitated with anti-Fas Ab, and the Fas-AP was immunoprecipitated with anti-Fas Ab and immunoblotted with anti-Myc or anti-HA Ab (upper panels). TCL was blotted with anti-Myc or anti-HA Ab (lower panels).

STAP-2 KO mice were stimulated with sFasL for the indicated periods. The cells were then lysed, and TCL was analyzed by immunoblotting (IB) with anti-Myc or anti-HA Ab (Fig. 6A). 293T cells (1 × 10^7) were transfected with Myc-tagged STAP-2 WT or DA (6 μg) and with or without Halo-tagged Fas (10 μg). At 36 h after transfection, the cells were lysed and immunoprecipitated with anti-Myc Ab and immunoblotted with anti-Fas or anti-Myc Ab (upper panels). TCL was blotted with anti-Fas or anti-Myc Ab (lower panels).

STAP-2 associates with Fas. (A) 293T cells (1 × 10^7) were transfected with Myc-tagged STAP-2 WT or DA (6 μg) and with or without Halo-tagged Fas (10 μg). At 36 h after transfection, the cells were lysed and immunoprecipitated with anti-Myc Ab and immunoblotted with anti-Myc or anti-HA Ab (upper panels). TCL was blotted with anti-Myc or anti-HA Ab (lower panels). (B) 293T cells (1 × 10^7) were transfected with Myc-tagged STAP-2 WT (6 μg) and with or without Halo-tagged Fas (10 μg). At 36 h after transfection, the cells were lysed and immunoprecipitated with anti-Myc Ab and immunoblotted with anti-Fas or anti-Myc Ab (upper panels). TCL was blotted with anti-Fas or anti-Myc Ab (lower panels). (C) 293T cells (1 × 10^7) were transfected with Myc-tagged STAP-2 WT or DA (6 μg) and with or without Halo-tagged Fas (10 μg). At 36 h after transfection, the cells were lysed and pulled down with HaloLink resin and immunoblotted with anti-Myc or anti-Fas Ab (upper panels). TCL was blotted with anti-Myc or anti-Fas Ab (lower panels).

FIGURE 6. STAP-2 associates with Fas. (A) 293T cells (1 × 10^7) were transfected with Myc-tagged STAP-2 WT or DA (6 μg) and with or without Halo-tagged Fas (10 μg). At 36 h after transfection, the cells were lysed and immunoprecipitated with anti-Myc Ab and immunoblotted with anti-Myc or anti-HA Ab (upper panels). TCL was blotted with anti-Myc or anti-HA Ab (lower panels).

upper panels and lower panels.

FIGURE 7. STAP-2 influences FasL-mediated apoptosis and caspase activation. (A and B) Splenocytes (5 × 10^6) from WT or STAP-2 KO mice (9 wk old) were stimulated without (−) or with (+) sFasL. At 18 h after stimulation, cells were stained with CD4 and annexin V (A) or TUNEL (B). Then annexin V-positive or TUNEL-positive apoptotic cells in CD4-positive cells were determined by FACS. Each circle represents data from one mouse; horizontal bars represent the mean; n = 8 (A), n = 6 (B); t value for sFasL (+)/WT versus sFasL (+)/KO, p < 0.05 (Student t test). (C) Splenic T cells (1 × 10^6) from WT or STAP-2 KO mice were stimulated with sFasL for the indicated periods. The cells were then lysed, and TCL was analyzed by immunoblotting (IB) with anti–caspase-8, anti–caspase-3, or anti–actin Ab. (D) Splenic T cells (2 × 10^6) from WT or STAP-2 KO mice were cultured for 72 h in a six-well plate precoated with anti-CD3 mAb (10 μg/ml). The cells were incubated with recombinant mouse His-tagged FasL (0, 2, 4 μg/ml) on ice for 30 min, and followed by cross-linking with anti-His probe mAb for 10 min at 37˚C. The cells were then lysed and immunoprecipitated with anti-Fas Ab, and the Fas-DISC was analyzed by immunoblotting with anti-caspase-8.

residue 260 (STAP-2 DA). We also constructed another STAP-2 mutant by substitution of glutamic acid for aspartic acid at residue 260 (STAP-2 DE) to avoid the effect of alteration in local charge at residue 260. Expression vectors of STAP-2 WT, STAP-2 DA, or STAP-2 DE were transiently expressed into Jurkat cells and cells were stimulated anti-Fas mAb for the indicated periods. Although substitution of Asp260 with a neutral amino acid, Ala, differing in size and polarity, resulted in a slightly faster mobility in the SDS-PAGE gels, Fas-mediated cleavage of STAP-2 was not observed in Jurkat cells expressing either STAP-2 DE or STAP-2 DA (Fig. 5B). Therefore, Fas/caspase-8-mediated processing of STAP-2 occurs at Asp260 in its C-terminal domain.

Activation of caspase-3 and -8 was greatly decreased in Jurkat/STAP-2 DA cells compared with Jurkat/STAP-2 WT cells (Fig. 5C). Furthermore, the amount of caspase-8 or FADD, or both, in the Fas-DISC was also reduced in Jurkat/STAP-2 DA cells compared with Jurkat/STAP-2 WT cells (Fig. 5D, 5E). We further examined whether enhanced enzymatic activation of caspasers by STAP-2 was affected by this mutation. As shown in Fig. 5F, Fas-induced caspase-3 and -8 enzymatic activities in Jurkat/STAP-2 DA cells was markedly reduced compared with Jurkat/STAP-2 WT cells. Moreover, TNF-α–induced caspase-8 enzymatic activity in Jurkat/STAP-2 DA cells was also significantly reduced compared with Jurkat/STAP-2 WT cells (Fig. 5G), suggesting that the cleavage of STAP-2 at Asp260 by caspase-8 is required for other DR-mediated apoptotic pathways. Fas-induced apoptosis in Jurkat/STAP-2 DA cells was weaker than that in Jurkat/STAP-2 WT cells, and at a level similar to that in Jurkat/pcDNA cells (Fig. 5H, 5I). Therefore, proteolytic cleavage of STAP-2 at Asp260 by caspase-8 is required for the enhancement of Fas-induced apoptosis and caspase-8 enzymatic activation by STAP-2 in Jurkat T cells.

We examined the molecular mechanisms related to how the STAP-2 DA mutant affects the Fas-DISC formation. We tested the association of caspase-8 with STAP-2 WT and DA in 293T cells. As shown in Fig. 6A, both STAP-2 WT and DA interacted with caspase-8. Because STAP-2 interacted with Fas (Fig. 6B), we tested the association of Fas with STAP-2 WT and DA in 293T cells.
STAP-2 KO (filled circle) mice (6–8 wk old) were immunized s.c. in the hind footpads with OVA/CFA (100 μg) in CD4+ T cells calculated according to the formula: % apoptosis = (% annexin V+ apoptotic cells at indicated day in CD4+ T cells/% annexin V+ apoptotic cells at day positive and FITC-CD4–positive cells were detected by FACS. (20 h after injection, isolated splenocytes were stained with anti–TCR-α/β Abs and analyzed by FACS. The above findings suggest that absence of STAP-2 results in longer life span of Ag-stimulated CD4+ T cells because of lowered sensitivity to AICD. Notably, the frequencies of apoptotic CD4+ T cells in the draining lymph nodes decreased after immunization in STAP-2–deficient mice (Fig. 8E). However, DEX-induced cell death in splenic T cells was not altered by STAP-2 deficiency (Supplemental Fig. 3C). In addition, DO11.10 T cells over-expressing STAP-2 responded to DEX as strong as control cells (Supplemental Fig. 3D). Therefore, these data support the notion that STAP-2 has a physiologic role in the ability of the immune system to eliminate activated T cells.

Discussion

The domain structure of STAP-2 contains several potential tyrosine phosphorylation sites as well as PH- and SH2-like domains, and STAP-2 shows the characteristics of an adaptor protein that can interact with several other signaling molecules. In this study, we identified new functions of STAP-2, specifically that it is a member of the Fas-DISC components and enhances Fas-mediated apoptosis in T cells. The STAP-2–mediated effect is selective for the DR-

![Graphs and data from the study showing the effects of STAP-2 on AICD in vivo.](http://www.jimmunol.org/)

**FIGURE 8.** STAP-2 influences AICD in vivo. (A) WT or STAP-2 KO mice were injected i.v. with 5 μg anti-CD3 mAb (+) or isotype control Ab (−). At 20 h after injection, isolated splenocytes were stained with anti–TCR-β and anti-CD220 Abs and determined the T cell-to-B cell ratio by FACS. Each circle represents data from one mouse; horizontal bars represent the mean; n = 6–9; t value for CD3/WT versus CD3/KO, p < 0.01 (Student t test). (B) Splenocytes from WT or STAP-2 KO mice injected with anti-CD3 Ab or control Ig (5 μg) for 20 h were stained with FITC-CD4 and TUNEL. TUNEL–positive and FITC-CD4–positive cells were detected by FACS. (C and D) WT or STAP-2 KO mice were injected i.p. with SEB. Percentages of Vβ8+CD4+ (C) and Vβ6+CD4+ (D) subsets of peripheral blood cells taken from the tail vein were analyzed on days 0 and 7 by FACS. Each circle represents data from one mouse; horizontal bars represent the mean; n = 6–8; t value for day 16 WT versus day 16 KO, p < 0.05 (Student t test). (E) WT (open circle) or STAP-2 KO (filled circle) mice (6–8 wk old) were immunized s.c. in the hind footpads with OVA/CFA (100 μg). Percent of apoptotic CD4+ T cells was calculated according to the formula: % apoptosis = (% annexin V+ apoptotic cells at indicated day in CD4+ T cells/% annexin V+ apoptotic cells at day 0 in CD4+ T cells) × 100. Each circle represents data from one mouse; horizontal bars represent the mean; n = 4–8; t value for day 16 WT versus KO, p < 0.01, for day 20, 24 WT versus KO, p < 0.05 (Student t test).
mediated pathway because STAP-2 expression does not alter the sensitivity of Jurkat cells to Bim-dependent apoptotic stimuli. The critical target of the STAP-2-mediated effects is the activation of caspase-8, a key initiator caspase in the Fas signaling pathway. Indeed, stable expression of STAP-2 leads to a marked increase in caspase-8 activation upon Fas engagement, and the addition of a caspase-8 inhibitor blocks Fas-mediated cell death in STAP-2-transfected cells. Splenic T cells from STAP-2-deficient mice exhibit a reduced ability to undergo AICD, as well as defects in the SEB-mediated elimination of V$\beta^8$CD4$^+$ T cells. These findings indicate that STAP-2 can control T cell apoptosis by modulating one of the critical initiating events in the Fas signaling cascade.

In general, caspase-8 activation within the Fas-DISC is believed to occur in two steps. For the first step, recruitment of FADD to Fas promotes dimerization and conformational changes of caspase-8, and allows caspase-8 to gain full enzymatic activity. Next, active caspase-8 undergoes autoproteolytic processing and leaves the Fas-DISC to access its substrates. Among the Fas-DISC components, FLASH associates with procaspase-8 in nonapoptotic cells (27). After Fas signaling is stimulated, FADD and then a FLASH–procaspase-8 complex are recruited into the intracellular domain of the aggregated Fas, resulting in the promotion of caspase-8 activation. Similar to FLASH, STAP-2 recognized the DED domain of caspase-8, even under steady-state conditions, and enhanced the interaction between FADD and caspase-8 as well as the aggregation and activation of caspase-8 during Fas signaling. Although STAP-2 did not bind to FADD, the molecular mechanisms for how FLASH and STAP-2 enhance Fas-induced apoptosis may be similar. For example, STAP-2 may control the local concentration and intracellular distribution of procaspase-8 before Fas stimulation. Alternatively, STAP-2 may promote a conformational change of procaspase-8 to allow dimerization or rapidly expose its autoproteolytic active sites in the Fas-DISC.

STAP-2 has a consensus caspase-8 cleavage sequence, VEAD, in its C-terminal domain, and was clearly cleaved by caspase-8 after Fas stimulation. It is noteworthy that processing of STAP-2 was crucial for Fas-induced apoptosis. Consequently, STAP-2 may participate in the Fas-DISC as a suicide substrate to enhance Fas-induced apoptosis. For example, activated caspase-8 may easily leave the Fas-DISC after processing of STAP-2. Although FLASH is also proteolytically cleaved during Fas signaling, the requirement for the cleavage is likely to differ between STAP-2 and FLASH. In the case of FLASH, its C-terminal region contains a potential cleavage site for caspase-8 and a nuclear localization signal (27). Thus, the cleaved FLASH C-terminal is thought to translocate into the nucleus. Although not a Fas-DISC component, hematopoietic progenitor kinase 1 (HPK1), a mammalian Ste20-related protein kinase, is a substrate of caspases (28, 29). HPK1 is crucial for TCR-mediated NF-κB activation; therefore, AICD-resistant naive T cells contain full-length HPK1. Following the proliferation of activated T cells, HPK1 is cleaved into N- and C-terminal fragments (HPK1-N and HPK1-C, respectively). The kinase-inactive HPK1-C inhibits NF-κB by binding to IKK-α and IKK-β, and sequestering the IKK complex in its inactive state. Therefore, HPK1-C blocks the induction of prosurvival genes and sensitizes T cells to AICD (8, 28, 29). At the present time, the roles of the cleaved STAP-2 fragments remain unknown. Further detailed studies are required to clarify this issue.

Our data also indicate that both the SH2 domain and a cleaved site of STAP-2 are essential for full activation of caspase-8 after Fas-stimulation. We have proposed possibility that STAP-2 SH2 is involved in recruitment of caspase-8 into the Fas-DISC and that STAP-2 cleavage is required for the activation of caspase-8 itself.

Although our data are reproducible, there are some questions against these molecular events in the Fas-DISC. One question is how STAP-2 ΔSH2, lacking the binding capacity to caspase-8, was cleaved by caspase-8 (Fig. 4B). This might suggest that STAP-2 cleavage is independent on tight binding between STAP-2 and caspase-8, because both proteins may exist closely enough in the Fas-DISC. The other question is why STAP-2 DA inhibited recruitment of caspase-8 into the Fas-DISC (Fig. 5D). For the possible mechanism, the caspase-8–cleaved site might bind to another Fas-DISC component. Indeed, we showed that STAP-2 DA could recognize caspase-8, but not Fas (Fig. 6A, 6C). Because molecular events in the DISC during Fas-stimulation are complicated, further analysis will clarify these strange phenomena regarding to STAP-2.

Notably, expression of FLICE-like inhibitory protein (FLIP), an inhibitory protein of Fas-mediated apoptosis (30), was slightly decreased in Jurkat/STAP-2 cells (Supplemental Fig. 4A). Furthermore, STAP-2–deficient T cells also showed a slightly increased FLIP expression (Supplemental Fig. 4B). Thus, the involvement of STAP-2 in FLIP expression is likely to be another possible mechanism for the DISC formation, although we need more careful examinations in this regard.

Given that STAP-2 interacts with a variety of signaling molecules, an attractive hypothesis is that the presence or absence of specific partners may modulate the ability of STAP-2 to function in several signaling pathways. In this study, we identified caspase-8 as a novel partner of STAP-2 in T cells. Future investigations will clarify when and how STAP-2 acts as a regulator of caspase-8 activation in vivo.

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References


