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Regulation of Cytosolic Phospholipase A$_2$ Phosphorylation by Proteolytic Cleavage of Annexin A1 in Activated Mast Cells

Joon Hyun Kwon,* Jea Hwang Lee,* Ki Soon Kim,* Youn Wook Chung, † and Ick Young Kim*

Annexin A1 (ANXA1) is cleaved at the N terminal in some activated cells, such as macrophages, neutrophils, and epithelial cells. We previously observed that ANXA1 was proteolytically cleaved in lung extracts prepared from a murine OVA-induced asthma model. However, the cleavage and regulatory mechanisms of ANXA1 in the allergic response remain unclear. In this study, we found that ANXA1 was cleaved in both Ag-induced activated rat basophilic leukemia 2H3 (RBL-2H3) cells and bone marrow-derived mast cells. This cleavage event was inhibited when intracellular Ca$^{2+}$ signaling was blocked. ANXA1-knockdown RBL-2H3 cells produced a greater amount of eicosanoids with simultaneous upregulation of cytosolic phospholipase A$_2$ (cPLA$_2$) activity. However, there were no changes in degranulation activity or cytokine production in the knockdown cells. We also found that cPLA$_2$ interacted with either full-length or cleaved ANXA1 in activated mast cells. cPLA$_2$ mainly interacted with full-length ANXA1 in the cytosol and cleaved ANXA1 in the membrane fraction. In addition, introduction of a cleavage-resistant ANXA1 mutant had inhibitory effects on both the phosphorylation of cPLA$_2$ and release of eicosanoids during the activation of RBL-2H3 cells and bone marrow-derived mast cells. These data suggest that cleavage of ANXA1 causes proinflammatory reactions by increasing the phosphorylation of cPLA$_2$ and production of eicosanoids during mast-cell activation. The Journal of Immunology, 2012, 188: 5665–5673.

Mast cells are considered central effector cells in allergic diseases, such as asthma, allergic rhinitis, and food allergies, and mast-cell–derived proinflammatory mediators play a key role in these pathologies (1–3). Ag-mediated activation of mast cells is regulated by a complex series of intracellular signaling processes that are initiated by FcεRI aggregation. Activated mast cells show three biological characteristics: degranulation (secretion of mediators stored in intracellular granules), the production of cytokines and chemokines, and the release of newly synthesized lipid mediators (4–7). Most studies on FcεRI-mediated signaling have been carried out using rat basophilic leukemia 2H3 (RBL-2H3) cells and mutated sublines (8). Although a number of studies have described the mast-cell activation pathway, it is still not fully understood.

Annexin A1 (ANXA1) is a member of the ANX family of Ca$^{2+}$ and phospholipid-binding proteins, and plays different roles under various biological conditions, for example, membrane organization, apoptosis, and inflammation (9, 10). ANXA1 consists of two domains: an N-terminal tail and a C-terminal core domain. The C-terminal core domain consists of four repeats of a highly conserved amino acid sequence responsible for binding with Ca$^{2+}$ and phospholipids (9). The N-terminal peptide is known to be cleaved by a number of proteases, including elastase, calpain, plasmin, cathepsin D, and proteinase 3 (11–14), and is also phosphorylated to regulate the activity of the protein (15, 16). N-terminal truncated ANXA1 has a proinflammatory effect and promotes neutrophil transendothelial migration (17). Truncated ANXA1 has also been linked to an epidermal growth factor (EGF)-triggered signaling pathway involving cytosolic phospholipase A$_2$ (cPLA$_2$) in normal and malignant squamous epithelial cells (14). In activated polymorphonuclear (PMN) leukocytes, ANXA1 is cleaved and then translocates to the extracellular surface (13). Mutant ANXA1, which is resistant to cleavage by proteases isolated from PMN cell extracts, was previously shown to have anti-inflammatory activities in the inflamed microcirculation of mice and in a skin trafficking model (18).

We previously demonstrated the proteolytic cleavage of ANXA1 and activation of cPLA$_2$ in a murine OVA-induced asthma model (19). cPLA$_2$ plays a key role in the release of arachidonic acid (AA) for the production of lipid inflammatory mediators, such as leukotrienes and PGs. In mast cells, the aggregation of IgE-loaded FcεRI induced by polyvalent Ag leads to the phosphorylation of ERK and cPLA$_2$, followed by release of AA, which is then further converted to leukotrienes and PGs by 5-lipoxygenase (5-LO) and cyclooxygenase (COX), respectively (20). Cysteinyl leukotrienes (cysLTs) are involved in the pathogenesis of asthma and are determinants of the severity of asthma (21). cPLA$_2$ activity is regulated by intracellular Ca$^{2+}$ concentrations and phosphorylation (22). Increased intracellular Ca$^{2+}$ concentrations promote the translocation of cPLA$_2$ from the cytosol to the membrane (23, 24). The catalytic domain of cPLA$_2$ is phosphorylated by MAPKs (Ser$^{505}$) and MAPK-interaction kinases (Ser$^{767}$) (25–27). Furthermore, cPLA$_2$ activity is inhibited by direct interaction with...
ANXA1 (28, 29). However, the inhibitory mechanisms of ANXA1 remain unclear. In particular, the functional significance of ANXA1 cleavage in cPLA₂ activity during mast-cell activation remains unknown. In this study, we demonstrate that cleavage of ANXA1 activates cPLA₂ by phosphorylation, stimulating a proinflammatory reaction in activated mast cells.

**Materials and Methods**

**Reagents**

Monoclonal anti-DNP Clone SPE-7, A23187 (a calcium ionophore), BAPTA-AM, 2-aminoethoxydiphenyl borate (2-APB), thapsigargin (TG), HEPES, sodium pyruvate, sodium orthovanadate, and PMSF were obtained from Sigma-Aldrich. DNP-BSA and p-nitrophenyl-2-acetyl-β-D-glucosaminide were purchased from Calbiochem. Recombinant mouse IL-3 and stem cell factor were obtained from R&D Systems. Abs against p-ERK1/2, ERK1/2, and p-cPLA₂ (Ser505) were purchased from Cell Signaling Technology. ANXA1 Abs were obtained from Zymed, and cPLA₂ (4-4B-3C) Abs were purchased from Santa Cruz Biotechnology. Abs against β-actin, H−hexosaminidase, cysLTs, and PGE₂ secreted from the activated cells were analyzed using an enzyme immunoassay system. (A, B, C) Data are expressed as the means ± SE of three independent experiments. (B, D–F) Results are representative of at least three independent experiments.

**Mast-cell culture and activation**

Bone marrow-derived mast cells (BMMCs) were isolated from the femurs of C57BL/6 mice (8- to 12wk-old) as described previously (30). The isolated BMMCs were cultured in RPMI 1640 medium (Life Technologies) supplemented with 10% FBS, 2 mM t-glutamine, 1% nonessential amino acids (Invitrogen), 50 μM 2-ME, 1 mM sodium pyruvate, 100 μM penicillin, 100 μg/ml streptomycin, 25 mM HEPES, 10 ng/ml stem cell factor, and 5 ng/ml IL-3 for 4–8 wk at 37°C and 5% CO₂. Purity, measured by flow cytometry analysis using FITC-conjugated rat anti-mouse c-Kit mAbs (BD Biosciences) at 4 wk, was >97%.

**Assay for β-hexosaminidase release**

β-Hexosaminidase release was used as a measure of degranulation activity as described previously (31). In brief, IgE-sensitized and serum-starved RBL-2H3 cells and BMMCs were incubated with Tyroid buffer (135 mM NaCl, 5 mM CaCl₂, 1 mM MgCl₂, 5.6 mM glucose, 1 mg/ml BSA, and 20 mM HEPES [pH 7.4]) and then stimulated with DNP-BSA (1 μg/ml) or A23187 (1 μM) at 37°C in Tyroid buffer for the indicated times. The incubation buffer and cell lysates, prepared by the addition of identical volumes of Tyroid buffer containing 1% Triton X-100, were then recovered for quantification of β-hexosaminidase release.

**Assay for cysLTs and PGE₂ release**

Release of cysLTs and PGE₂ was measured using an enzyme immunoassay system (Amersham), according to the manufacturer’s instructions. IgE-sensitized and serum-starved RBL-2H3 cells and BMMCs were stimulated with DNP-BSA (1 μg/ml), A23187 (1 μM), or TG (1 μM) at 37°C for the indicated times. When necessary, BAPTA-AM (30 μM) and 2-APB (50 μM) were added to the cells for 30 min before activation with DNP-BSA. Then, 100 μl culture medium was concentrated by freeze-drying overnight, followed by reconstitution in assay buffer. Samples and standard cysLTs or PGE₂ were incubated with antiserum in a 96-well plate for 2 or 3 h, followed by cysLT or PGE₂-peroxidase conjugates for 3 or 1 h, respectively, at 4°C. To remove unbound ligand, we washed the wells five times with wash buffer. Then the amount of bound peroxide-labeled cysLTs and PGE₂ was determined by the addition of substrate for 30
in R buffer containing 100 pmol ANXA1 siRNAs for RBL-2H3 cells (siRNA-1, 5′-CCG GAC GTA CAA AGC AUA CUU A-3′; siRNA-2, 5′-CAG AUG AAG ACA CUC UUG AGA U-3′) or 5 μg plasmid DNA for RBL-2H3 cells and BMMCs. Resuspended cells were then transferred into a gold tip and electroporated by 1 pulse at 1600 V for 30 ms, followed by incubation overnight in growth media without antibiotics.

Data analysis and statistics

All data are represented in this study as the means ± SE of the control value. Statistical comparisons from at least three independent experiments were determined using Student t tests. The p values <0.05 were considered significant.

Results

Proteolytic cleavage of ANXA1 in activated mast cells

We previously confirmed the proteolytic cleavage of ANXA1 in lung extracts prepared from a murine asthma model (19). To further
elucidate the regulatory mechanisms of ANXA1 cleavage and their roles in inflammation, we activated both RBL-2H3 cells and BMMCs with DNP-BSA. For activation, IgE-sensitized cells were serum starved and then stimulated with DNP-BSA (1 μg/ml) for the indicated times. RBL-2H3 cell activation was determined by measuring degranulation, cytokine production, and eicosanoid generation. Release of β-hexosaminidase from cytosolic granules to extracellular spaces was used as a measure of degranulation activity. The activity reached a maximum at 30 min after stimulation (Fig. 1A). Production of cytokines was determined by RT-PCR analysis of TNF-α, IL-4, and IL-6 mRNA levels. Expression of both TNF-α and IL-4 was rapidly induced by DNP-BSA treatment, whereas IL-6 production was induced only after ≥30 min of treatment (Fig. 1B). The generation of eicosanoids was represented by the presence of cysLTs and PGE2 in cell-free supernatants (Fig. 1C). Like degranulation activity, the production of eicosanoids also reached a maximum at 30 min. ERK and cPLA2 were differentially phosphorylated on DNP-BSA treatment in RBL-2H3 cells (Fig. 1D). ERK was rapidly phosphorylated, and cPLA2 phosphorylation was detected at later time points. ANXA1 was cleaved without any changes in mRNA expression (Fig. 1E, 1F).

BMMC activation was also determined by measuring degranulation and eicosanoid generation. Release of β-hexosaminidase and production of eicosanoids were similar to those of RBL-2H3 cells (Supplemental Fig. 1A, 1B). Similar to our results in RBL-2H3 cells, cleavage of ANXA1 and phosphorylation of ERK and cPLA2 were also detected in BMMCs (Supplemental Fig. 1C).

Release of eicosanoids and phosphorylation of cPLA2 are increased in ANXA1-knockdown RBL-2H3 cells

To investigate the role of ANXA1 in mast-cell activation, we transfected ANXA1 siRNAs into RBL-2H3 cells (Fig. 2A). There was no difference in either degranulation activity or cytokine production between the NC and ANXA1 siRNA-transfected cells (Fig. 2B, 2C). However, the release of cysLTs and PGE2 was significantly increased in ANXA1-knockdown cells (Fig. 2D). To understand the increased production of eicosanoids, we examined the phosphorylation of ERK and cPLA2. Whereas ANXA1 siRNA transfection did not affect ERK phosphorylation, cPLA2 phosphorylation was significantly increased (Fig. 2E). We also found that expression of COX-2 and translocation of 5-LO were not changed in ANXA1-knockdown RBL-2H3 cells (Supplemental Fig. 2). These results suggest that cPLA2 phosphorylation is associated with ANXA1 during mast-cell activation.

Intracellular Ca2+-dependent translocation of ANXA1 and cPLA2 from the cytosolic to membrane fraction in activated mast cells

ANXA1 and cPLA2 are known to translocate from the cytosol to the membrane on increase in intracellular Ca2+ concentrations (9, 32). Similar to DNP-BSA stimulation, both A23187, a calcium ionophore, and TG, which increases cytosolic Ca2+ levels, activated RBL-2H3 cells and BMMCs (Fig. 3). Thus, ANXA1 cleavage and cPLA2 phosphorylation were measured on treatment with A23187 and TG. CysLTs and PGE2 were also produced by A23187 and TG treatment. When the cells were pretreated with

FIGURE 3. Inhibition of ANXA1 cleavage and eicosanoid production by decreasing intracellular Ca2+ concentrations. IgE-sensitized RBL-2H3 cells and BMMCs were serum starved and stimulated with DNP-BSA (1 μg/ml, 30 min), TG (1 μM, 5 min), or A23187 (1 μM, 10 min). Cells were pretreated with BAPTA-AM (30 μM) or 2-APB (50 μM) for 30 min before stimulation with DNP-BSA. (A) Lysates of activated cells were subjected to immunoblot analysis with Abs against ANXA1, cPLA2, p-cPLA2, and β-actin. Results shown are representative of at least three experiments. (B) The levels of secreted cysLTs and PGE2 were determined by an enzyme immunoassay system. Data are expressed as the means ± SE of three independent experiments. Significant differences were determined by comparison of DNP-BSA + BAPTA-AM or 2-APB with DNP-BSA (*p < 0.05).
BAPTA-AM, a Ca\(^{2+}\) chelator, or 2-APB, a store-operated Ca\(^{2+}\) channel inhibitor, ANXA1 cleavage and the production of cysLTs and PGE\(_2\) significantly decreased (Fig. 3A, 3B). We also observed that full-length ANXA1 was localized to both the cytosolic and membrane fractions of nonactivated RBL-2H3 cells. When the cells were activated by DNP-BSA or A23187, ANXA1 was cleaved, and the amount of full-length ANXA1 in the cytosol decreased. In the membrane fraction, the level of full-length ANXA1 did not differ from that of control cells, although cleavage of ANXA1 increased (Fig. 4A, 4B). Thus, in response to cell activation, cleaved ANXA1 translocated mainly to the membrane. Furthermore, both cPLA\(_2\) and p-cPLA\(_2\) translocated from the cytosol to the membrane in activated RBL-2H3 cells (Fig. 4A, 4C). However, ANXA1 knockdown did not affect the translocation of either cPLA\(_2\) or p-cPLA\(_2\) (Fig. 4D).

**Both full-length and cleaved ANXA1 interact with cPLA\(_2\)**

To elucidate the interactions between ANXA1 and cPLA\(_2\) during mast-cell activation, we performed coimmunoprecipitations of the proteins from whole-cell lysates and subcellular fractions. In the whole-cell lysates, cPLA\(_2\) interacted with either full-length or cleaved ANXA1 (Fig. 5A). However, coimmunoprecipitation of proteins from the subcellular fractions showed that cPLA\(_2\) mainly interacted with full-length ANXA1 in the cytosol and with cleaved ANXA1 in the membrane fraction (Fig. 5B).

**A cleavage-resistant ANXA1 mutant inhibits phosphorylation of cPLA\(_2\)**

It was previously reported that cleavage of ANXA1 occurred at three sites (Ala\(^{11}\), Val\(^{22}\), and Val\(^{36}\)) in the N-terminal region during the activation of neutrophils (13). The ANXA1 mutant (A11R/V22K/V36K), which is resistant to cleavage, controls inflammation in the microvasculature (18). In this study, we investigated whether mutant ANXA1 regulates cPLA\(_2\) activity during RBL-2H3 cell and BMMC activation. GFP-tagged expression vectors containing wild-type ANXA1 (wtANXA1) or triple-mutant ANXA1 (mtANXA1) were constructed; then these vectors were introduced into RBL-2H3 cells and BMMCs as described in the Materials and Methods. We confirmed that mtANXA1 was not cleaved by DNP-BSA stimulation, whereas both endogenous ANXA1 and wtANXA1 were cleaved (Fig. 6A, Supplemental Fig. 3A). We further observed that phosphorylation of cPLA\(_2\) was significantly decreased in both activated RBL-2H3 cells and BMMCs expressing mtANXA1, whereas ERK phosphorylation was not affected (Fig. 6B, Supplemental Fig. 3B). We also found that although both wtANXA1 and mtANXA1 interacted with cPLA\(_2\), p-cPLA\(_2\) was not detected in mtANXA1-expressing RBL-2H3 cells and BMMCs under activation conditions (Fig. 6C, Supplemental Fig. 3C).

**FIGURE 4.** Translocation of ANXA1 and cPLA\(_2\) from the cytosol to the membrane in activated RBL-2H3 cells. (A–C) DNP-BSA or A23187-stimulated RBL-2H3 cells were fractionated into cytosol and membrane fractions. (A) The subcellular fractions were subjected to immunoblot analysis with Abs against ANXA1, cPLA\(_2\), p-cPLA\(_2\), α-tubulin, and calnexin. α-Tubulin and calnexin were used as cytosol and membrane marker proteins, respectively. Results are representative of at least three experiments. Relative band intensities for (B) full-length and cleaved ANXA1 and (C) cPLA\(_2\) are represented as a percentage of the control full-length ANXA1 or control cytosolic cPLA\(_2\), respectively, in each fraction. Data are expressed as the means ± SE of three independent experiments. Significant differences were determined by comparison of DNP-BSA or A23187 with the control (*p < 0.05). (D) NC and ANXA1 siRNA-transfected cells were stimulated with DNP-BSA and then fractionated into cytosolic and membrane fractions. Cytosolic and membrane fractions were subjected to immunoblot analysis with Abs against cPLA\(_2\), p-cPLA\(_2\), α-tubulin, and calnexin. Results are representative of at least three experiments.
ANXA1 cleavage regulates production of eicosanoids in mast-cell activation

Next, we determined the effects of ANXA1 cleavage on the production of proinflammatory mediators in both RBL-2H3 cells and BMMCs expressing wtANXA1 or mtANXA1. Compared with control or wtANXA1, mtANXA1-expressing cells did not show any difference in either degranulation activity or cytokine production (Fig. 7A, 7B). However, release of cysLTs and PGE2 was significantly decreased in mtANXA1-expressing RBL-2H3 cells and BMMCs (Fig. 7C).

Discussion

In this study, we confirmed that ANXA1 was cleaved in activated mast cells, and this cleavage was required for the phosphorylation of cPLA2 and subsequent inflammatory reactions, such as the production of eicosanoids. In resting RBL-2H3 cells, full-length ANXA1 interacted with cPLA2 in the cytosol. However, when cells were activated by DNP-BSA or A23187, both ANXA1 cleavage and cPLA2 phosphorylation were observed. Furthermore, cleaved ANXA1 and phosphorylated cPLA2 interacted and colocalized mainly to the membrane fractions of activated mast cell. We also found that mutant ANXA1, which is resistant to cleavage in activated mast cells, exhibited inhibitory effects on both the phosphorylation of cPLA2 and production of eicosanoids.

Different degrees of ANXA1 expression and susceptibility to modulation by allergic stimuli have been observed in connective tissue and mucosal mast cells (33). ANXA1-null mice have shown resistance to the anti-inflammatory actions of glucocorticoids in several experimental models of inflammation (34–36). Mast cells from ANXA1-null mice are more prone to release of histamine and PGD2 than those of wild-type mice in response to compound 48/80 and zymosan, respectively (37). Moreover, ANXA1-derived peptide Ac2-26 and antiflammin, nonapeptide fragments of lipocortin I, also inhibited mast-cell degranulation (38, 39). These inhibitory effects were thought to be mediated by formyl peptide receptor in mast cells. In this study, we found that ANXA1-knockdown RBL-2H3 cells released more eicosanoids and exhibited increased phosphorylation of cPLA2. These effects were mediated through endogenous ANXA1, not formyl peptide receptor downstream signaling. The release of β-hexosaminidase from RBL-2H3 cells did not change with DNP-BSA treatment, as has been shown with release of histamine by compound 48/80 treatment (Fig. 2). ERK phosphorylation is known to be necessary for the generation of inflammatory mediators in mast-cell activation. Activated ERK has been shown to phosphorylate cPLA2 for AA release (40). To determine whether the increased phosphorylation of cPLA2 in ANXA1-knockdown cells was due to changes in ERK phosphorylation, we measured ERK phosphorylation in RBL-2H3 cells transfected with ANXA1 siRNA. In this experiment, we found that the level of ERK phosphorylation in ANXA1-knockdown RBL-2H3 cells was the same as that in control cells. However, cPLA2 phosphorylation was markedly increased in the knockdown cells (Fig. 2E). These data suggest that ANXA1 may regulate ERK-mediated cPLA2 phosphorylation during mast-cell activation.

Eicosanoids are produced predominantly by inflammatory cells, such as PMN cells, macrophages, and mast cells. The FcεRI-mediated mast-cell activation signal pathway for eicosanoid production is regulated by a complex series of signaling molecules. In particular, cPLA2 activation by ERK and intracellular Ca2+ is
increased intracellular Ca\(^{2+}\) synergistically promote the full acti-
phosphorylation of cPLA\(_2\) acts to regulate catalytic activity, but
leased less AA (46). In addition, a recent study revealed that
membrane in response to a calcium ionophore, although it re-
translocation to the nuclear envelope and endoplasmic reticulum
located mainly in the cytosol in resting mast cells and undergoes
data are expressed as means ± SE of three independent experiments. (B) Total RNA isolated from activated RBL-2H3 cells was subjected to RT-PCR analysis. The expression of TNF-α, IL-4, and IL-6 mRNA was determined for measurement of cytokine production. GAPDH was used as a loading control. Results shown are representative of at least three independent experiments. (C) The levels of secreted cysLTs from activated RBL-2H3 cells and BMMCs were analyzed by an enzyme immunoassay system. Data are expressed as means ± SE of three independent experiments. Significant differences were determined by comparison of wtANXA1 or mtANXA1 constructs with GFP (*p < 0.05).

![Graphs](image)

**FIGURE 7.** A cleavage-resistant ANXA1 mutant inhibited the production of eicosanoids in DNP-BSA–stimulated mast cells. GFP-, wtANXA1-, or mtANXA1-overexpressing RBL-2H3 cells and BMMCs were activated with DNP-BSA for 30 min. (A) Incubation buffer and cell lysates were recovered for quantitative analysis of β-hexosaminidase release as a measure of RBL-2H3 cell degranulation activity. Data are expressed as means ± SE of three independent experiments. (B) Total RNA isolated from activated RBL-2H3 cells was subjected to RT-PCR analysis. The expression of TNF-α, IL-4, and IL-6 mRNA was determined for measurement of cytokine production. GAPDH was used as a loading control. Results shown are representative of at least three independent experiments. (C) The levels of secreted cysLTs from activated RBL-2H3 cells and BMMCs were analyzed by an enzyme immunoassay system. Data are expressed as means ± SE of three independent experiments. Significant differences were determined by comparison of wtANXA1 or mtANXA1 constructs with GFP (*p < 0.05).

important for the production of eicosanoids (27, 41). cPLA\(_2\) is
located mainly in the cytosol in resting mast cells and undergoes
translocation to the nuclear envelope and endoplasmic reticulum in
response to various stimuli (23). Phosphorylation of cPLA\(_2\) and
increased intracellular Ca\(^{2+}\) synergistically promote the full activ-
ation of cPLA\(_2\) for AA release (42, 43). AAs are converted to
eicosanoids. In contrast, when intracellular Ca\(^{2+}\) levels were de-
increased by pretreatment with BAPTA-AM and 2-APB, ANXA1
cleavage, cPLA\(_2\) phosphorylation, and eicosanoid production were
all inhibited (Fig. 3). We also observed that cPLA\(_2\) translocated
from the cytosol to the membrane in activated RBL-2H3 cells, but
downregulation of ANXA1, did not affect the translocation of
cPLA\(_2\). However, cPLA\(_2\) phosphorylation was increased in the
membrane fraction of ANXA1-knockdown cells (Fig. 4D). These
data indicate that ANXA1 regulates cPLA\(_2\) phosphorylation, but
not cPLA\(_2\) translocation.

ANXA1 is present in both the nucleus and cytoplasm of rat mast
cells (48). In this study, we also investigated the intracellular lo-
kalization of ANXA1 in activated RBL-2H3 cells. The level of full-
length ANXA1 decreased in the cytosolic fraction, whereas
ANXA1 cleavage increased in the membrane fraction on activa-
tion of RBL-2H3 cells with DNP-BSA or A23187 (Fig. 4A, 4B,
5B). However, the level of full-length ANXA1 in the membrane
fraction did not change during activation. Together with the data
on cPLA\(_2\) translocation described earlier, these results indicate
that both cleaved ANXA1 and cPLA\(_2\) translocate from the cytosol
to the membrane in activated mast cells.

ANXA1 is a cPLA\(_2\)-binding protein that negatively regulates
cPLA\(_2\) activity both in vitro (28, 49) and in vivo (14, 50). Inter-
actions of cPLA\(_2\) with either full-length or cleaved ANXA1 were
observed in whole-cell lysates (Fig. 5A). Using subcellular frac-
tions, we found that cPLA\(_2\) mainly interacted with full-length
ANXA1 in the cytosol and cleaved ANXA1 in the membrane (Fig.
5B). Furthermore, phosphorylated cPLA\(_2\) was mainly ob-
served in the membrane fraction of activated RBL-2H3 cells.

The N-terminal peptide of ANXA1 is cleaved by several pro-
teases, including elastase, calpain, plasmin, cathepsin D, and pro-
teinase 3 (11–14). ANXA1 cleavage promotes neutrophil trans-
endothelial migration (17) and regulates EGF-triggered signaling
and AA production in normal and malignant squamous epithelial
cells (14). Recently, an ANXA1 mutant (A11R/V22K/V36K)
was reported to be resistant to cleavage by all proteases present
in PMN cell extracts (18). In this study, we tested the effects of
N-terminal cleavage on inflammatory reactions using RBL-2H3 cells and BMCCs expressing mANXA1. Expression of this mANXA1 protein blocked the phosphorylation of cPLA2 in activated RBL-2H3 cells and BMCCs, but ERK phosphorylation was not affected (Fig. 6B, 6C, Supplemental Fig. 3B, 3C). The release of cysLTs and PGE2 was inhibited in mtANXA1-expressing RBL-2H3 cells and BMCCs; however, there were no changes in degranulation activity or cytokine production (Fig. 7). This could be attributed to the inhibition of cPLA2 phosphorylation by the mutant full-length ANXA1.

In summary, we suggest that ANXA1 cleavage is a necessary process for inflammatory reactions in mast cells, cPLA2, which constitutively interacts with ANXA1, is phosphorylated by cleavage of ANXA1 during mast-cell activation. Phosphorylated cPLA2 then produces AA, which is converted into various eicosanoids. Cleavage of ANXA1 may result in the loss of ANXA1's inhibitory effect on cPLA2 phosphorylation and may promote eicosanoid production during mast-cell activation. Although the mechanism of ANXA1 cleavage has not been fully elucidated, we found that intracellular Ca2+ concentration is one of the most important regulatory factors for ANXA1 cleavage in mast-cell activation.

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Disclosures
The authors have no financial conflicts of interest.

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