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Eosinophils Preserve Parasitic Nematode Larvae by Regulating Local Immunity

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Eosinophils play important roles in regulation of cellular responses under conditions of homeostasis or infection. Intestinal infection with the parasitic nematode, *Trichinella spiralis*, induces a pronounced eosinophilia that coincides with establishment of larval stages in skeletal muscle. We have shown previously that in mouse strains in which the eosinophil lineage is ablated, large numbers of *T. spiralis* larvae are killed by NO, implicating the eosinophil as an immune regulator. In this report, we show that parasite death in eosinophil-ablated mice correlates with reduced recruitment of IL-4+ T cells and enhanced recruitment of inducible NO synthase (iNOS)-producing neutrophils to infected muscle, as well as increased iNOS in local F4/80+Ly6C+ macrophages. Actively growing *T. spiralis* larvae were susceptible to killing by NO in vitro, whereas mature larvae were highly resistant. Growth of larvae was impaired in eosinophil-ablated mice, potentially extending the period of susceptibility to the effects of NO and enhancing parasite clearance. Transfer of eosinophils into eosinophil-ablated ΔdblGATA mice restored larval growth and survival. Regulation of immunity was not dependent upon eosinophil peroxidase or major basic protein 1 and did not correlate with activity of the IDO pathway. Our results suggest that eosinophils support parasite growth and survival by promoting accumulation of TH2 cells and preventing induction of iNOS in macrophages and neutrophils. These findings begin to define the cellular interactions that occur at an extraintestinal site of nematode infection in which the eosinophil functions as a pivotal regulator of immunity. The Journal of Immunology, 2012, 188: 417–425.

Investigations of infections caused by helminths that are natural parasites of rodents have revealed a number of mechanisms of protective immunity. Studies of the intestine-dwelling nematodes *Heligmosomoides polygyrus*, *Nippostrongylus brasiliensis*, *Trichuris muris*, and *Trichinella spiralis* have documented TH2-driven immune responses that incorporate production of IL-4, IL-5, IL-9, IL-10, and IL-13, as well as basophilia, eosinophilia, and alternative activation of macrophages (1). Parasite clearance from the intestine is abrogated in the absence of Stat6, IL-4, and/or IL-13, confirming the importance of these mediators; however, because of differences in habitats and life cycles, the specific effector mechanism that clears worms from the intestine varies among infections. For example, mast cells are crucial to expulsion of intraepithelial *T. spiralis* (2) but dispensable for clearance of *T. muris* and *N. brasiliensis* (3, 4) during primary infection. Among the cells that are prominent in immune responses to intestinal helminths, perhaps the most enigmatic is the eosinophil. Eosinophilia is a hallmark of nematode infection, yet infection of eosinophil-ablated mice with *T. muris*, *Schistosoma mansoni*, or *T. spiralis* has failed to reveal a key role for eosinophils in clearance of intestinal worms (5–7).

Immune responses and mechanisms of helminth clearance from extraintestinal sites have been less thoroughly studied in natural rodent hosts. It has been shown that clearance of *Litomosoides sigmodontis* is promoted by the presence of eosinophil granular proteins, major basic protein 1 (MBP) and eosinophil peroxidase (EPO) (8). Furthermore, eosinophils are necessary for development of immunity that limits the early tissue migratory larval stage during secondary infections by *N. brasiliensis* (9). These findings support the paradigm of eosinophils as defenders against worm infection.

* T. spiralis* occupies both intestinal and extraintestinal sites during the course of its life cycle. Adult worms in the intestine release newborn larvae (NBL) that migrate to skeletal muscle and initiate chronic infection. Arrival of NBL in muscle is coincident with an intestinal TH2 immune response that expels adult worms and induces profound blood and tissue eosinophilia (7). Despite the magnitude of the local inflammatory response, intracellular muscle larvae mature to become infectious. We have shown previously that although eosinophil-ablated mice clear intestinal *T. spiralis* normally, immunity to the muscle stage of infection is impacted dramatically (7, 10). Muscle larvae die in large numbers (50–75%) coincident with enhanced IFN-γ and decreased IL-4 production in draining lymph nodes. In the absence of eosinophils, leukocytes at sites of infection produce inducible NO synthase (iNOS) and parasite survival improves when mice are treated with specific iNOS inhibitors. Introducing IL-10 deficiency into the PHIL background dramatically enhanced NO production and increased...
parasite killing to ≥90%. These observations suggest that eosinophils protect developing larvae against NO-mediated killing (7).

In this study, we extend our earlier findings by showing that accumulation of IL-4+ T cells to sites of infection is reduced in eosinophil-ablated mice, and this correlates with infiltration of iNOS+ neutrophils and inflammatory macrophages during a time at which the growing larva is vulnerable to the effects of NO. Re-storing eosinophils to infected mice improved Th2 cell recruitment, parasite growth, and survival, clearly implicating eosinophils as crucial to immune regulation that supports parasite survival.

Materials and Methods

Rats and mice

Adult Albino Oxford strain rats were produced and maintained in the Baker Institute Vivarium. IL-10−/−, PHIL, and ΔdblGATA (C57BL/6 background) (11) mice were bred at Cornell Transgenic Mouse Core Facility, and progeny were transferred to the Baker Institute. PHIL and IL-10−/− mice were genotyped as described previously (7, 12). IL-5 transgenic, EPO−/−, and MBP−/− mice were described previously (12–14). C57BL/6 mice (henceforth referred to as wild type [WT]) were purchased from Taconic. Animal care was in accordence with the guidelines of the Association for Assessment and Accreditation of Laboratory Animal Care, and experiments were performed with the approval of the Institutional Animal Care and Use Committee of Cornell University.

Parasite and Ags

*T. spiralis* first-stage larvae (L1) and NBL were recovered from rats as described previously (15, 16). For synchronous infection, 20,000 NBL were administered by gavage. Mice were euthanized by CO2 inhalation at the time indicated in each experiment. Muscle larvae burdens were assessed 24 days postinfection (dpi) or later in whole carcasses as described previously (15). In some experiments, larvae were recovered from diaphragms 12–18 dpi by digesting minced tissue for 15 min at 37°C in 5 mg/ml collagenase I (Sigma). Somatic Ags from L1 were prepared as described previously (17).

Eosinophil transfer experiments

Eosinophils were obtained from the peritoneum (by lavage) and spleens (SPLs) of infected IL-5 transgenic mice 12–20 dpi. Cells were pooled and purified by either positive or negative MACS bead selection. For positive selection, eosinophils were enriched using PE-conjugated anti–Siglec-F Ab (BD) and anti-PE microbeads (Miltenyi Biotec), a procedure that yielded eosinophil preparations of ≥95% purity. For negative selection, contaminating cells were labeled with PE-conjugated rat anti-mouse CD90.2, Leukocytes were recovered from cell surfaces as described above using PE-conjugated anti-CD11b, PE-Cy7-conjugated anti-F4/80, and FITC-conjugated anti-Ly-6C (eBioscience). For basophil counts, cells isolated from CLN, mesenteric lymph node (MLN), and SPL were stained with FITC-conjugated anti-CD49b (BioLegend), PE-conjugated anti–FcR1-α (BioLegend), and allophycocyanin-conjugated anti-c–Kit (BioLegend).

Parasite measurements and susceptibility to NO

Developing L1 were recovered by digesting minced diaphragms for 15 min at 37°C in 5 mg/ml collagenase I (Sigma). To prevent curling, we treated larvae with 70% ethanol at 56°C and left them overnight at room temperature. Straightened larvae were centrifuged and resuspended in 5% glycerol/70% ethanol to soften and clear them before preparation for cytopsin. The cytopsin slides were stained with HEMA-3 (Fishner Health-care), and measurements were performed using 10× and 20× objectives on a Leica microscope (Olympus) by fitting a polygon around the boundary of the larva and computing the area (Microsuite Basic Olympus software). At least 20 larvae were measured per mouse, and values are expressed in micrometers squared. To evaluate susceptibility of larvae to NO-mediated killing, developing L1 recovered from synchronously infected WT mice at 8, 11, 14, and 21 d postinfection or NBL were cultured with indicated concentrations of (Z)-1-[N-(2-aminoethyl)-N-(2-ammonioethyl)amino]diazen-1-ium-1,2-diolate (DETA) for 48 h (37°C, 8% CO2) in DMEM (Mediatech) supplemented with 30% fetal bovine serum (Gibco). Parasite death (larvae that were immobile and granular) is reported as a percentage of the total.

Quantitative RT-PCR

Total RNA was isolated from diaphragm tissue using TRIzol reagent (Invitrogen), and cDNA was prepared using SuperScript III First-strand cDNA Synthesis System (Invitrogen). A sample lacking reverse transcriptase served as negative control. Quantitative RT-PCR (qRT-PCR) was performed for Nos2, Arg1, Fizz1, Ym1, and Gapdh using the following TaqMan Gene Expression primers and probes: GAPDH: forward, 5′-GTTCAAGCTCTTCCGTGATA-G-3′; reverse, 5′-CCTACTCTTGOG-AGGCCATGTG-3′; probe, 5′-TACCTTAAACCAC-3′. IL-4, IL-5, IL-10, IL-13, and IFN-γ were assayed from CLN, mesenteric lymph node (MLN), and splenic (SPL) cell populations. IL-4 (eBioscience). For intracellular iNOS detection, permeabilized diaphragm leukocytes were incubated with a rabbit polyclonal anti-iNOS (NeoMarkers) followed by allophycocyanin-conjugated goat anti-rabbit (Jackson). Cells were first stained for cell surface Ags as described previously (15). Cells were resuspended in 200 μl sterile PBS and injected i.v. into 8-week-old C57BL/6J mice through the lateral tail vein. Animal Care and Use Committee of Cornell University.

Histology and immunohistochemistry

Histochemical staining and immunohistochemistry were performed as described previously (15). To recover focally damaged organs and cells were prepared for cytologic staining as previously described (15). Slides were stained with rabbit polyclonal anti-iNOS (NeoMarkers) and hematoxylin (Fisher), and differential counts were performed under 40× magnification using a BX51 microscope (Olympus).

Cytokine ELISA

Cells from cerebral lymph nodes (CLNs) were obtained and cultured as described previously (10). IL-4, IL-5, IL-10, IL-13, and IFN-γ were assayed in culture supernatants by ELISA as described previously (7).

Flow cytometry

Cells were recovered from individual diaphragms as described previously (10). For intracellular IL-4 detection, cells were cultured ex vivo for 5 h with 250 ng/ml ionomycin (Sigma-Aldrich), 50 ng/ml PMA (Sigma-Aldrich), and 1 μg/ml brefeldin A (BD Pharmingen). After a 15-min incubation with Fc block (eBioscience) and 10% normal mouse serum, cells were incubated for 15 min with FITC-conjugated anti-CD8 and PE-Cy7-conjugated anti-CD4 (eBioscience). Samples were treated with fixation/permeabilization buffer (eBioscience), and permeabilized cells were stained using PE-conjugated anti–IFN-γ and allophycocyanin-conjugated anti–IL-4 (eBioscience). For intracellular iNOS detection, permeabilized diaphragm leukocytes were incubated with a rabbit polyclonal anti-iNOS (NeoMarkers) followed by allophycocyanin-conjugated goat anti-rabbit (Jackson). Cells were first stained for cell surface Ags as described above using PE-conjugated anti-CD11b, PE-Cy7-conjugated anti-F4/80, and FITC-conjugated anti-Ly-6C (eBioscience). For basophil counts, cells isolated from CLN, mesenteric lymph node (MLN), and SPL were stained with FITC-conjugated anti-CD49b (BioLegend), PE-conjugated anti–Fcr1-α (BioLegend), and allophycocyanin-conjugated anti–c–Kit (BioLegend).

Results

Cellular sources of iNOS in eosinophil-ablated mice

We have reported previously that iNOS contributes to clearance of muscle larvae in eosinophil-ablated mice (7). To better characterize the destructive immune response, we used qRT-PCR to...
document phenotypic changes in local macrophages. We analyzed the expression of genes associated with alternatively activated (M2) macrophages (ARG1, YM1, and FIZZ1) and classically activated (M1) macrophages (NOS2) in diaphragms of infected WT, ΔdblGATA, and PHIL mice. Although all three strains dramatically upregulated M2 markers, eosinophil ablation was associated with a marked increase in NOS2 transcription and decreases in M2 marker expression at 17 dpi (Fig. 1A). Immunohistological analysis of leukocytes recovered from diaphragms confirmed the presence of iNOS+ macrophages but also revealed large numbers of iNOS+ neutrophils in PHIL mice (Fig. 1B). Although the representation of iNOS+ cells among neutrophils was markedly greater than that of macrophages, the number of each cell type in infected muscle (7) is such that iNOS+ macrophages outnumbered neutrophils in diaphragms of eosinophil ablated mice by a ratio of 2:1. Overall, macrophage accumulation was not affected by eosinophil ablation (Fig. 1C), but among phenotypically distinct macrophage subsets, Ly-6C+ CD11b+ F4/80+ inflammatory macrophages produced significantly more iNOS in PHIL mice compared with WT (Fig. 1D).

**Impact of eosinophil deficiency on leukocyte recruitment**

Histologic examination of tongues revealed that the cellular infiltrates around nurse cells in PHIL mice were reduced compared with WT (Fig. 2A). Enumeration of leukocytes recovered from diaphragms confirmed a reduction in absolute cell numbers at sites of infection in PHIL mice (Fig. 2B). Flow cytometric evaluation of T cell subsets documented fewer CD4+ and CD8+ T cells (Fig. 2C), as well as significantly reduced frequencies of IL-4+ CD4+ and IL-4+ CD8+ T cells in diaphragms of PHIL mice (Fig. 2D). The results suggest that eosinophils promote local recruitment of IL-4-producing T cells.

**Potential for IDO to regulate T cell responses**

IDO mediates oxidation of tryptophan to KYN and has been implicated in regulating Th1 responses. Human eosinophils produce IDO in response to IFN-γ, thereby promoting apoptosis or inhibiting cellular proliferation of Th1 cells (18). We investigated whether IDO might be the link between eosinophils and reduced Th1 responses in WT mice by investigating gene expression and enzyme activity during infection. IDO gene expression was similar in infected muscles of PHIL, ΔdblGATA, and WT mice (Fig. 3A). IDO enzymatic activity, measured by KYN production in cultures of Ag-stimulated CLN cells or diaphragm leukocytes, was similar across strains (Fig. 3B). Immunohistochemical staining of diaphragm leukocytes showed that the percentages of IDO+ cells were similar between WT and PHIL mice (Fig. 3C). Large mononuclear cells, but not eosinophils, were IDO+ in tissues of WT mice. The results do not support a role for IDO in eosinophil-dependent regulation of local T cell responses.

**Impact of eosinophil deficiency on basophilia**

We examined the impact of eosinophil ablation on basophilia by counting basophils in MLN, CLN, and SPL at 0, 2, 5, and 12 dpi (Fig. 3D). Uninfected ΔdblGATA mice had significantly fewer basophils in the SPL, and this trend was evident in the MLN, CLN, and SPL on 2 and 5 dpi, although differences were statistically significant only in the SPL at 2 dpi. Numbers of basophils in all tissues were similar in the two mouse strains on 12 dpi, before the time at which parasite compromise and altered immunity are evident in eosinophil-ablated mice.

**Eosinophil granular proteins do not influence survival of muscle larvae**

We investigated the impact of eosinophil granular proteins on the progression of infection and modulation of immune responses by infecting mice deficient in either EPO or MBP. Muscle larvae burdens in both strains were similar to WT (Fig. 4A). Cytokine production by CLN cells in response to Ag restimulation was not altered (Fig. 4B), nor was there an effect on the numbers of eosinophils, CD4+ cells, or CD8+ cells at sites of infection (Fig. 4C). Thus, MBP or EPO were dispensable for regulation of T cell responses and parasite survival.

![Figure 1](http://www.jimmunol.org/)
Larval growth is impaired in the absence of eosinophils

Microscopic examination of H&E-stained tongue sections revealed that both nurse cells and larvae appeared to be smaller in PHIL versus WT mice (Fig. 2A). To determine whether larval growth was compromised in the absence of eosinophils, we measured the dimensions of larvae recovered from diaphragms between 12 and 18 dpi. The mean area of larvae was similar in PHIL versus WT mice on 12 dpi but was reduced in PHIL mice on 15 and 18 dpi (Fig. 5A). Similarly, a significant difference in larval area was detected in DdblGATA versus WT mice on 17 dpi (Fig. 5B). Larvae grew normally in MBP2/2 and EPO2/2 mice (Fig. 5C). To test whether iNOS activity, in the presence of eosinophils, would cause impaired larval growth, we infected IL-102/2 mice. This strain has dramatic, local iNOS+ cellular infiltrates and some reduction in larval burden (7, 10). Parasite growth was not altered by IL-10 deficiency (Fig. 5D), indicating that nitrosative stress does not compromise parasite growth.

Developing T. spiralis larvae are susceptible to direct killing by NO

To determine whether death of larvae is the direct result of exposure to NO, we tested the larvicidal effect of NO on T. spiralis larvae in vitro. Larvae that recovered from WT mice at different times postinfection were cultured for 48 h with the artificial NO donor (DETA-NONOate or with the vehicle control DETA. To generate larvae that were at the same stage of development, we infected mice by i.v. injection of NBL to achieve synchronous muscle infections. The results show that NBL and muscle larvae up to 11 d of age were killed in significant numbers (Fig. 6A, 6B), whereas relatively mature muscle larva (≥14 d old) were resistant (Fig. 6C). Note that because of the asynchronous nature of NBL production by adult worms in the intestine, NBL colonize the muscle between 4 and 14 d after oral infection (19). Thus, susceptible larvae would be present in the muscle between 4 and 28 d after oral infection, with the proportion of susceptible larvae declining between 14 and 28 d. In support of the conclusion that only growing muscle larvae are susceptible to the effect of NO, we found that parasite clearance in PHIL mice is completed by 28 dpi, with no additional reduction in burden evident when estimated 40 dpi (Fig. 6D).

Adoptive transfer of eosinophils improved larvae growth and survival

To confirm the role of the eosinophil in parasite growth and retention, we transferred eosinophils isolated from infected IL-5 transgenic mice to infected DdblGATA mice (Fig. 7A). Based on results of replicate experiments showing that transferred eosinophils extravasated in skeletal muscle and persisted there for 24 h...
FIGURE 4. Effects of MBP and EPO on parasite survival and immunity. A, Larval burdens in muscles of WT, EPO−/−, and MBP−/− mice 28 dpi. B, Cytokines measured in Ag-stimulated cultures of CLN cells collected from WT, MBP−/−, and EPO−/− mice 17 dpi. C, CD4+, CD8+, and Siglec-F+ cells recovered from diaphragms of infected WT, MBP−/−, and EPO−/− mice 17 dpi. Values were determined twice with similar results. Values represent means ± SD; n = 3–4 mice. No significant differences were found.

but not 48 h (not shown), we designed the experimental protocol to incorporate cell transfers on alternate days between 5 and 15 dpi. Four experiments, conducted using positively selected (n = 2) and negatively selected (n = 2) eosinophils, yielded similar results, and statistical analysis documented that the improvement in larval burdens in this group of experiments averaged 36% and was highly significant (p = 0.005; Fig. 7B). One experiment performed with PHIL mice showed that transfer of positively selected eosinophils improved larval burdens by 42% (p = 0.04). Results from a single experiment performed with negatively selected eosinophils in ΔdblGATA mice (Fig. 7C–E) show that transfer of eosi

Discussion

Host adaptation is highly evolved among parasitic worms, and there is ample evidence that helminths manipulate the immune response in ways that prolong their survival in the host or promote their dispersal in the susceptible host population. The importance of obtaining a detailed and thorough understanding of the means by which nematodes interfere with immunity lies in the potential for such knowledge to inform the design of approaches to controlling infection and preventing disease. Our previously published findings revealed the potential for eosinophils to promote survival of T. spiralis in muscle. Specifically, we reported that when PHIL or ΔdblGATA mice are infected with T. spiralis, larvae are cleared from skeletal muscle by an iNOS-dependent mechanism (7). We show in this study that neutrophils and macrophages produce iNOS at sites of infection. Although markers of M2 macrophages were dramatically upregulated in diaphragm tissue of both eosinoph

FIGURE 5. Larval growth in eosinophil-ablated mice. Estimated area of larvae recovered from (A) PHIL and WT mice between 12 and 18 dpi. (B) ΔdblGATA and WT mice 17 dpi. (C) EPO−/−, MBP−/−, and WT mice 18 dpi, and (D) IL-10−/− and WT mice 18 dpi. A total of 25–30 larvae were evaluated per mouse. Values represent means ± SD; n = 3–4 mice. Bars indicate the mean values from 75–90 larvae pooled from three mice (B–D). Experiments were performed twice to four times with similar results. Significant differences were determined by Student t test (A, B, D) or by ANOVA and Tukey’s test (C). **p < 0.001, ***p < 0.0001.

Taken together, the results support a model in which eosinophils act locally, either directly or indirectly, to prevent the development of iNOS-producing neutrophils and macrophages capable of parasite killing. A direct effect of eosinophils on macrophage phenotype has been described recently. By virtue of their production of IL-4, eosinophils have been shown to promote alternative activation of macrophages in mouse adipose tissue, thereby promoting glucose tolerance and protecting against diet-induced obesity (21). Furthermore, a recent report has shown that M2 macrophages proliferate in the pleural cavities of mice infected with L. sigmodonitis, and that proliferation in situ is under the influence of IL-4 (22). Our findings are not incompatible with local proliferation of M2 macrophages in muscles of T. spiralis-infected mice, making the potential influence of eosinophils on local macrophage populations of considerable interest.

Previous work did not determine whether NO-dependent clearance of T. spiralis resulted from direct toxicity to larvae or from nitrosative damage to nurse cells (7). Our results demonstrate that T. spiralis larvae are susceptible to direct killing by NO. Suspe-
tibility is evident in newborns but increases during the period of rapid growth between 4 and 14 d after invasion of muscle cells (23). These results suggest that NO can mediate killing of parasitic worms in response to infection in a natural host. Similarly, NO has been implicated in helminth killing in mice vaccinated against *S. mansoni* or colonized with *Brugia malayi* (24, 25). NO produced by endothelial cells and macrophages kills *S. mansoni* larvae in vitro (26), and susceptibility is age dependent; however, in contrast with *T. spiralis*, older *S. mansoni* larvae show greater susceptibility (27). In the *B. malayi* model, treatment of mice with an inhibitor of NO synthase abrogates resistance (24, 25). Therefore, it is apparent that in contrast with the well-established role for Th2 immune responses in expulsion of intestinal worms, reactive nitrogen species produced during Th1 immune responses can be effective in host defense against tissue-dwelling parasitic worms.

We found that *T. spiralis* larvae became remarkably resistant to NO as they approached maturity. Antioxidant enzymes likely afford protection to the parasite (28), and expressed sequence tag analysis predicted that mature *T. spiralis* muscle larvae transcribe three types of antioxidant genes, specifically, thioredoxin peroxidase, peroxiredoxin, and glutathione peroxidase (29, 30). These transcripts are less frequent in NBL, compatible with immature larvae being more vulnerable to oxidative or nitrosative damage (29–31) and consistent with results of our in vitro and in vivo experiments. Eosinophils appear to protect larvae during a window of susceptibility to oxidative and nitrosative stress.

Before destruction of larvae in eosinophil-deficient mice, nurse cell development was impaired and larval growth was inhibited. We have not yet determined whether this inhibition is the result of an immune response that develops in the absence of eosinophils or it reflects the parasite’s dependence upon factors produced or induced by eosinophils. Parasite growth modulation by the immune system has been shown in other helminth infections. For example, T cells facilitate growth of *S. mansoni* by exerting noncognate influence on MHC class II+ APCs (32). In addition, development of the filarial nematode *L. sigmodontis* is transiently delayed in the absence of IL-5 or eosinophils, and *B. malayi* development improves in the presence of T cells and NK cells (33–35). Although the role of lymphocytes may vary across infections, the accumulating evidence supports a model in which innate immune cells influence the rate of growth and development of parasitic helminths.

We speculate that inhibition of larval growth would lengthen the period of susceptibility to NO-mediated killing and promote clearance of larvae. Consistent with this notion was the finding that larvae grew normally in IL-10–deficient mice, which demonstrated less dramatic parasite clearance but strong iNOS production (10). The mechanism(s) behind compromise of larval growth in eosinophil-ablated mice remains to be elucidated. Angiogenesis is a prominent feature of nurse cell development, during which infected myotubes develop a surrounding vascular network that is presumed to support the parasite (36). Eosinophils and M2 macrophages can promote angiogenesis (37–40). If vascularization is compromised in the absence of eosinophils, both nurse cell differentiation and parasite growth may be inhibited, a hypothesis that we are currently testing.

*T. spiralis* can live for years in skeletal muscles of its host (41). Prolonged survival requires that the worm suppress the host immune response or block its effects. Our data indicate that eosinophils directly or indirectly inhibit a Th1 immune response that induces the production of larvicidal NO; survival of the parasite is correlated with Th2 immunity. Eosinophils can promote Th2 responses by different mechanisms. It has been reported that human eosinophils expressing IDO catabolize tryptophan to KYN.

**FIGURE 6.** Susceptibility of muscle larvae to NO-mediated killing. Susceptibilities of different larval stages of *T. spiralis* to NO-mediated killing were determined by culturing (A) 8- to 14-d-old larvae isolated from C57BL/6 mice, (B) NBL, or (C) mature L1 (24 d old) with DETA/NO or DETA for 48 h in vitro. Viability was estimated for 60 larvae/well, three wells per mouse, mean ± SD for three mice per time point. D. Muscle burdens in PHIL and WT mice, 28 and 40 dpi. Experiments were performed twice with similar results. Values represent means ± SD; n = 3–4 mice. Significant differences were determined by Student t test. *p < 0.05, **p < 0.001.
that subsequently causes apoptosis of Th1 cells (18). This phenomenon has not been documented in murine eosinophils; however, we hypothesized that mouse eosinophils might have a similar influence, possibly by promoting IDO production in other cells. We evaluated the numbers of IDO-producing cells and IDO gene expression in diaphragms of WT, PHIL, and C57BL/6 mice at 2 dpi. It is not obvious that these differences would affect immunity to *T. spiralis* (43). Eosinophil ablation in *dblGATA* mice does not clear muscle larvae, whereas IL-10 deficient mice upregulate clear cells rather than granulocytes were the dominant sources of these measures was affected by eosinophil deficiency. Mononuclear cells rather than granulocytes were the dominant sources of IDO in all strains. These results do not support an eosinophil-dependent role for IDO in inhibiting a Th1 response in WT mice.

We found that *T. spiralis* infection progressed normally in the absence of the MBP or EPO, indicating that these granular proteins do not contribute to parasite growth and survival, or to immune modulation. Another granular protein, human eosinophil-derived neurotoxin (an ortholog of the mouse eosinophil-associated RNase-2), has been shown to induce dendritic cell maturation and expansion of Th2 responses by virtue of its ability to activate TLR2 (42). We did not detect a difference in dendritic cell numbers or maturation (as evidenced by MHC class II and CD86 expression) between WT and eosinophil-ablated mice in the CD8+ infiltrates and Th2 cytokine production in the lung (44, 45), and supplementing *dblGATA* mice with CCL11 enhances the Th2 response (44). Adoptive transfer of eosinophils or eosinophils and CD4+ T cells reconstitutes disease in *dblGATA* and PHIL mice, respectively (44, 45). Transfer of eosinophils deficient in IL-13 failed to restore disease, documenting IL-13 as a critical mediator of the regulatory effect of eosinophils in allergic airway disease (44, 47). Our previous studies showed that STAT6-deficient mice do not clear muscle larvae, whereas IL-10 deficient mice upregulate NOS2 and clear parasites, suggesting that IL-10 may be more
The authors have no financial conflicts of interest.

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