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An Acquired Defect in IgG-Dependent Phagocytosis Explains the Impairment in Antibody-Mediated Cellular Depletion in Lupus

Anupama Ahuja,* Lino L. Teichmann,* Haowei Wang,* Robert Dunn,† Marilyn R. Kehry,† and Mark J. Shlomchik*,‡

B cells play important roles in autoimmune diseases ranging from multiple sclerosis to rheumatoid arthritis. B cells have also long been considered central players in systemic lupus erythematosus. However, anti-CD20-mediated B cell depletion was not effective in two clinical lupus studies, whereas anti-B lymphocyte stimulator, which inhibits B cell survival, was effective. Others and we previously found that anti-CD20-based depletion was surprisingly ineffective in tissues of lupus-prone mice, but that persistent high doses eventually led to depletion and ameliorated lupus. Lupus patients might also have incomplete depletion, as suggested in several studies, and which could have led to therapeutic failure. In this study, we investigated the mechanism of resistance to Ab-mediated cellular depletion in murine lupus. B cells from lupus-prone mice were easily depleted when transferred into normal environments or in lupus-prone mice that lacked serum Ig. Serum from lupus-prone mice transferred depletion resistance, with the active component being IgG. Because depletion is FcγR-dependent, we assay macrophages and neutrophils exposed to lupus mouse serum, showing that they are impaired in IgG-mediated phagocytosis. We conclude that depletion resistance is an acquired, reversible phagocytic defect depending on exposure to lupus serum IgG. These results have implications for optimizing and monitoring cellular depletion therapy. The Journal of Immunology, 2011, 187: 3888–3894.

Animal models would enable insight into these issues. For this purpose, two groups developed similar strains of mice in which hCD20 was expressed via a transgenic bacterial artificial chromosome (9, 10). B cells can be depleted in these animals using anti-hCD20. Two groups have also developed murine (m)CD20 mAbs that can deplete B cells (10, 11). These models provided insight into the mechanisms by which B cell depletion ameliorates autoimmune disease. They also elucidated the mechanisms of in vivo depletion, which mainly depend on FcγR-mediated phagocytosis of opsonized B cells (9, 11).

A number of groups have used these approaches to demonstrate efficacy of B cell depletion in murine models of lupus (10, 12, 13). Although these studies showed definite effects of B cell depletion, others and we were surprised to find that it was relatively difficult to deplete B cells in lupus-prone mice (10, 13), including MRL/MpJ-Fasstr (MRL.Fasstr), MRL/MpJ.Fastint, and NZB/W. Mild defects in B cell depletion were also seen in NOD mice, another spontaneous model of autoimmunity (14). In the case of MRL.Fasstr mice, even high doses of anti-CD20 did not reverse the defect acutely; strikingly, B cells that were fully coated with Ab were not cleared in these animals. However, persistent administration of high doses of Ab did eventually lead to depletion, which became apparent between 7 and 10 wk sustained treatment. At these time points, a progressive therapeutic effect was also observed, demonstrating that B cells can be a therapeutic target in ongoing disease (10). These observations suggest that there is a kinetic, but not absolute block in the clearance of B cells in lupus mice. The block to B cell depletion is age-dependent, similar to disease itself, although defects are also observed in young lupus-prone mice. Lupus-prone strains that have less severe disease also show a more mild deficiency in B cell depletion. Taken together, these results suggest that the disease process itself could be responsible for ineffective therapeutic B cell depletion (10, 13).

Given the difficulty in depleting B cells in murine lupus models, it seems possible that similar issues could be at play in lupus...
patients. There are anecdotal reports of poor B cell depletion in peripheral blood (PB) of treated patients, and that poor depletion correlated with poor clinical response (15–17). One caution is that very limited information is available about B cell depletion in secondary lymphoid tissues of patients (16). In this regard, mouse studies suggest that assessment of PB overestimates B cell depletion in tissues in the context of lupus (10).

If the mechanism that obstructs B cell depletion were understood, this would be useful for devising strategies to enhance the therapeutic effect of B cell depletion. Elucidation of the mechanism might also provide insight into the disease itself, as inability to deplete B cells likely relates to generalized defects in cellular clearance, a pathway that is involved in lupus pathogenesis (18). In this study, we have used variations on the MRL.Fas<sup>br</sup> model, including in vivo and in vitro studies, to investigate the reasons for defective B cell depletion in lupus. Our findings have implications for cellular depletion therapy as well as for understanding the immune dysregulation underlying lupus.

Materials and Methods

Mice

hCD20 bacterial artificial chromosome transgenic mice (10) were backcrossed >20 generations either on MRL.Fas<sup>br</sup> or BALB/c backgrounds. The Jh<sup>−/−</sup> (19) mice were backcrossed >20 generations with MRL.Fas<sup>br</sup> mice as described (20). The mlgM mice (20) that lack circulating Abs, by virtue of expression of a BCR transgene that lacks the secreted exon of IgM, were also maintained on a Jh<sup>−/−</sup> background and were fully backcrossed to MRL.Fas<sup>br</sup> mice. hCD20.MRL.Fas<sup>br</sup> mice were crossed with mlgM mice to produce hCD20.mlgM mice, which were maintained as Jh<sup>−/−</sup> on the MRL.Fas<sup>br</sup> background. MRL.Fas<sup>br</sup> mice were purchased from The Jackson Laboratory, BALB/c mice were from Charles River Laboratories, and C3.H-12<sup>2</sup> (BALB.K) mice were from Harlan, U.K. F<sub>1</sub> progeny of BALB/c × BALB.K mice were used as recipients. All mouse experiments were approved by the Yale Institutional Care and Use Committee.

Immunodepletion

Mice were injected with either 2H7 (IgG2b) or 18B12 (IgG1) mAbs (10, 21) in sterile PBS as described in the figure legends. Age-matched control mice were treated simultaneously with same amount of total mouse IgG (Rockland) dialedyzed into PBS.

Serum transfer

Serum from BALB/c (>6 wk old) or old MRL.Fas<sup>br</sup> (>14 wk old) mice was collected either by retro-orbital or cardiac puncture and was filter-sterilized. Pooled BALB/c and MRL.Fas<sup>br</sup> sera were analyzed for Ig isotypes, anti-Smith Abs, and anti-nucleosome Abs as described (22). Presence of anti-nuclear and anti-DNA Ab in MRL.Fas<sup>br</sup> serum was confirmed using indirect immunofluorescence of fixed HEp-2 cells (Anti-nuclear Ab;_light), as described (22). BALB/c or MRL.Fas<sup>br</sup> serum (0.5 ml) was injected i.v. into hCD20.BALB/c mice on days 0 to 6 once per day. In a similar experiment, mlgM mice received 1 ml donor serum (BALB/c or MRL.Fas<sup>br</sup>) on day 0, followed by 0.5 ml on days 2 to 6 once per day.

Separation of IgG and non-IgG components of serum followed by transfer and immunodepletion

Sera from 4 mo and older MRL.Fas<sup>br</sup> mice were pooled and passed through a 0.45-μm filter to remove particulates and run over a protein G column (Bio-Rad) equilibrated in binding buffer (20 mM sodium phosphate [pH 7.0]). The flow-through was collected as the non-IgG fraction. After washing, the column was eluted with 0.1 M glycine-HCl buffer at pH 2.7 and eluates were immediately neutralized with 0.5 M Tris (pH 8.0) (IgG fraction). Both fractions were spin dialyzed into PBS, and the volume of each fraction was adjusted to equal the volume of the serum that had been applied to the column. The purity of the fractions was checked by ELISA for IgG and SDS-PAGE gel. ELISA results showed that, compared with intact serum, >90% of the IgG was recovered in IgG fraction and 1% IgG was in the non-IgG fraction (data not shown). Protein gel results demonstrated a very high degree of purity of the IgG fraction and a high level of depletion from the non-IgG fraction (Supplemental Fig. 1).

Six- to 10-wk-old hCD20 transgenic (Tg) BALB/c mice, or Tg-negative controls, were randomly divided into four groups to receive 1) PBS, 2) a pooled, unfraccionated serum, 3) an IgG fraction, or 4) a non-IgG fraction. One-half milliliter was administered i.p. every day from day 0 to day 6. On day 7, 0.5 mg 2H7 was given i.v. to each mouse. On day 11, mice were sacrificed and splenocytes, lymph node (LN) cells, and PB were analyzed by FACS. For comparisons, untreated mice were also analyzed at same time.

Cell transfer

B cells were purified to >90% purity from the spleens of hCD20.BALB/c and hCD20.MRL.Fas<sup>br</sup> mice with the EasySep negative selection mouse B cell enrichment kit (StemCell Technologies), followed by labeling with either CFSE (Invitrogen) or CellTracker Orange (5-((and-6)-((4-chloromethyl)benzoyl)amino) tetramethylrhodamine [CMTMR]; Invitrogen), according to the manufacturers’ instructions. Approximately equal numbers of CFSE- and CMTMR-labeled cells were injected into the recipients on day 0. Mice were treated with a single i.v. dose of 100 μg 2H7 the following morning and sacrificed on day 3.

Phagocytosis assay

Fresh SRBCs (MP Biomedicals) were labeled with PKH26 (Sigma-Aldrich) according to the manufacturers’ instructions. The labeling reaction was stopped with RPMI 1640 containing 1% BSA. SRBCs were then psonitized with rabbit anti-SRBC IgG (Fitzgerald Industries) or used non-sonopsonized. CD11b<sup>+</sup> splenocytes were isolated from mice as indicated using streptavidin microbeads (Miltenyi Biotec) after labeling with biotinylated anti-CD11b (M1/70). CD11b<sup>+</sup> cells were then incubated together with SRBCs at a ratio of 1:2 in serum-free RPMI 1640 for 60 min at 37°C. Phagocytosis was stopped with ice-cold 2 mM EDTA in PBS. SRBCs that had not been internalized were lysed with ammonium chloride buffer.

FACS analysis

Abs to CD4 (GK1.5), CD5 (53.7), CD8 (TIB105), CD21 (7G6), CD22 (2D6), IgM<sup>+</sup> (RS3.1), CD11b (M1/70), and FcγR (2.4G2) were prepared and labeled in our laboratory, as described (23). The mAbs against mCD20 (18B12, mouse IgG1) and hCD20 (2H7, mouse IgG2b) were generated and purified as described (10, 21). The 2H7 hybridoma was a gift from Dr. E. Clark (University of Washington, Seattle, WA). Abs against CD3e (145-2C11), CD19 (1D3.2), CD23 (B384), F4/80, and Gr-1 were purchased from BD Pharmingen. Anti-mouse IgD (11-26c.2a), anti-mouse CD11c (N418), and streptavidin PE/Cy7 were from BioLegend and streptavidin-PE/Cy7 were from eBioscience. Samples were analyzed on a FACS Calibur or LSR II (BD Biosciences) and the data were analyzed using FlowJo software (Tree Star).

Results

Reduced efficacy of Ab-mediated depletion in MRL.Fas<sup>br</sup> mice could stem from defects in elements of the clearance mechanisms, such as macrophages or FcγRs, or from cell-intrinsic resistance by lymphocytes to depletion. We designed a series of experiments to distinguish among these possibilities. Our central hypothesis was that, in lupus, an excess of immune complexes (ICs) disables FcγR function on macrophages (and other phagocytic cells), limiting the ability of macrophages to deplete even fully opsonized cells.

Depletion is restored in MRL.Fas<sup>br</sup> mice that have B cells but lack Ab

If our hypothesis were correct, mlgM.MRL.Fas<sup>br</sup> mice that have B cells and systemic disease but lack secreted Ab (20) would be susceptible to B cell depletion (Fig. 1). To test this we crossed the hCD20 Tg onto this background and used anti-hCD20 (2H7) to deplete B cells. Treatment of 6- to 8-wk-old hCD20.mlgM.MRL.Fas<sup>br</sup> and hCD20.BALB/c mice with 4 mg/wk 2H7 for 2 wk resulted in >90% B cell depletion in spleen and PB for both strains whereas hCD20.MRL.Fas<sup>br</sup> mice were significantly more refractory to depletion (spleen, 30%; PB, 47%; Fig. 1A). B cells in LN of hCD20.mlgM mice had an intermediate susceptibility to depletion (76%) compared with the hCD20.BALB/c mice (97%) and hCD20.MRL.Fas<sup>br</sup> mice (66%). Similar results were obtained in a second set of experiments in which we treated 6- to 8-wk-old...
Efficient depletion of B cells in MRL.Faslpr mice lacking circulating Ab. A, Six- to 8-wk-old hCD20.BALB/c (n = 2), hCD20.MRL.Faslpr (n = 5), and hCD20.mlgM.MRL.Faslpr (n = 6 for control, referred to as hCD20.mlgM on the figure) mice were treated with 4 mg/wk 2H7 Ab i.p., given in divided doses twice each week for 2 wk. Residual CD22+ B cells in spleen, LN, and PB as percentage of the average B cells in IgG-treated, age-matched control mice are shown. B, Eight- to 10-wk-old hCD20.BALB/c and hCD20.MRL.Faslpr mice were derived from figures 2C and 3B, respectively, of Ahuja et al. (10) and are shown for comparison. B, Six- to 8-wk-old mlgM (n > 6) and BALB/c (n > 3) mice were treated with either 1 mg/wk or 3 mg/wk 18B12 mAb i.p. twice each week for 2 wk. Residual CD19+ B cells are plotted as in A for all sites including peritoneal cavity (PerC). The BALB/c data are derived from figure 3C of Ahuja et al. (10) and are shown for comparison. Data are from one experiment for hCD20.mlgM mice and from two experiments for mlgM mice. Error bars represent SEM. *p < 0.05, **p < 0.01 by Mann–Whitney U test.

mlgM.MRL.Faslpr mice with either 1 mg/wk or 3 mg/wk anti-mCD20, 18B12, twice weekly for 2 wk (depletion was: spleen, 70%; LN, 70%; PB, >95%; peritoneal cavity, >50%; Fig. 1B). B cell depletion in BALB/c from a previously reported similar experiment (figure 3C of Ref. 10) is shown for comparison. Again, depletion in the spleen, peritoneal cavity, and PB was comparable in the mlgM.MRL.Faslpr and BALB/c strains. Depletion in the LN was less effective in mlgM.MRL.Faslpr mice, a difference that could relate to the accumulation of double-negative T cells in the enlarged LN that occurs in lpr mice, which might prevent macrophages from coming in contact with B cells (9).

Taken together, these experiments demonstrate that in the absence of circulating Abs B cells are readily depleted even in autoimmune-prone mice. In light of the finding that despite identical genetic backgrounds, depletion efficacy is markedly different in MRL.Faslpr and mlgM.MRL.Faslpr mice, which differ in the presence or absence of serum IgM and not in other disease parameters (20), we conclude that genetic aspects of the MRL.Faslpr strain background alone cannot explain the defects in B cell depletion and that a factor related to secreted Ig must play an important role.

**Serum from lupus-prone mice, but not from normal mice, blocks depletion**

The prior results implicate serum IgG in the inhibition of B cell depletion in a lupus-prone background. To obtain direct evidence for this, we transferred serum from lupus-prone or normal mice to a host that is normally capable of depletion and assessed the ability of this serum to inhibit subsequent depletion. We initially chose to use serum rather than purified IgG, as it seemed possible that isolation of the IgG could change its properties (e.g., by dissociating from ICs), or that serum could contain important non-IgG factors.

Eight- to 10-wk-old hCD20.BALB/c mice and 9- to 11-wk-old mlgM.MRL.Faslpr mice were infused with serum from BALB/c (>6 wk old) or MRL.Faslpr (>14 wk old) mice (Fig. 2). As expected, the MRL.Faslpr serum had elevated concentrations of total IgG, as well as auto-Ab, anti-chromatin Ab, anti-nucleosome Ab, and anti-Smith Ab compared with the BALB/c serum, which was negative for these autoantibodies. On day 7 after serum infusion, mice were treated with a single i.v. dose of 0.5 mg 2H7 (hCD20.BALB/c mice; Fig. 2A, 2B) or 18B12 (mlgM.MRL.Faslpr mice; Fig. 2C, 2D). Enumeration of B cells on day 11 showed that infusion of MRL.Faslpr serum significantly and very substantially inhibited B cell depletion in spleen, LN, and PB (Fig. 2A, 2C). In the spleen, inhibition of depletion was seen both in follicular and marginal zone subpopulations in hCD20.BALB/c mice (Fig. 2B) but only in follicular B cells in mlgM.MRL.Faslpr mice (Fig. 2D). Therefore, serum transfer can render...
a permissive recipient substantially less capable of B cell depletion. MRL.Fas<sup>lpr</sup> serum was potent in these experiments compared with BALB/c serum.

The active component in serum that blocks B cell depletion is IgG

We hypothesized that the active factor in serum mediating inhibition of depletion contained IgG. Alternatively, as MRL.Fas<sup>lpr</sup> mice have intense inflammation, it was possible that the component was cytokine-derived. To test this, we depleted IgG from pooled serum of older MRL.Fas<sup>lpr</sup> mice using protein G (Supplemental Fig. 1). hCD20.BALB/c mice were then infused with either the depleted serum (non-IgG fraction) or the IgG fraction. Control mice received either unfractionated serum or PBS. Recipients were then given anti-hCD20 i.v. as above (Fig. 2) and assessed 4 d later for depletion. As shown in Fig. 3, the unfractionated serum inhibited depletion, confirming results in Fig. 2. Critically, the IgG-containing eluate inhibited B cell depletion to the same degree as did the unfractionated serum. Conversely, serum depleted of IgG had no effect, demonstrating that none of the “non-IgG components” were capable of inhibiting B cell depletion with anti-CD20.

Both MRL.Fas<sup>lpr</sup> and BALB/c B cells are efficiently depleted in a permissive host

The above experiments did not test whether inherent resistance in the B cells of MRL.Fas<sup>lpr</sup> mice could also contribute to defective B cell depletion. Such B cell resistance could have been genetic or acquired over time as a result of B cell exposure to the autoimmune-prone environment. To test whether MRL.Fas<sup>lpr</sup> B cells were more resistant to depletion, we compared the ability of MRL.Fas<sup>lpr</sup> or BALB/c B cells to be depleted upon transfer to a permissive environment. To generate a strain that would accept B cells from both MRL.Fas<sup>lpr</sup> and BALB/c mice, we crossed BALB.K mice (BALB/c congenic for the H<sup>2</sup>a haplotype matching that of MRL.Fas<sup>lpr</sup>) to BALB/c mice to obtain H<sup>2</sup>dxk heterozygous BALB/c mice and used these as the recipient strain. Purified B cells from hCD20.BALB/c and hCD20.MRL.Fas<sup>lpr</sup> mice were labeled with CFSE and CMTMR, respectively, prior to co-transfer. The recipient mice were treated on day 1 with 100 µg 2H7 or control IgG Ab i.v. and analyzed for B cell depletion on day 3 (Fig. 4). Mice treated with 2H7 showed ~90% depletion of both hCD20.BALB/c and hCD20.MRL.Fas<sup>lpr</sup> donor B cells, compared with the control IgG-treated mice (Fig. 4A). Note that the CD19<sup>+</sup>, CFSE<sup>−</sup>, CMTMR<sup>−</sup> cells in these plots are recipient B cells. A summary of B cell depletion from 11 mice is shown in Fig. 4B. The similar degrees of depletion indicate that there is no inherent difference, whether genetic or acquired, in the susceptibility of B cells from either lupus-prone or normal mice. Rather, when transferred to a permissive environment, MRL.Fas<sup>lpr</sup> B cells are readily depleted; our prior studies demonstrated that the same MRL.Fas<sup>lpr</sup> B cells are dramatically resistant to depletion when they remain in situ in their original hosts (10).

Ab-mediated phagocytosis by macrophages and neutrophils is less efficient in MRL.Fas<sup>lpr</sup> versus B cell-deficient MRL.Fas<sup>lpr</sup> mice

Because it has been shown that depletion is chiefly mediated by macrophages in an FcγR-dependent way (9, 11), we hypothesized that efficacy of macrophage-mediated phagocytosis should depend on whether the macrophages have developed in the presence or absence of serum Ab/ICs. To test this, we compared macrophages from B cell-deficient mice that had never been exposed to serum IgG to those from B cell intact mice, both of the MRL.Fas<sup>lpr</sup> genetic background. We isolated splenic CD11b<sup>+</sup> cells and used them in an IgG-dependent phagocytosis assay. The isolation procedure included both macrophages and CD11b<sup>+</sup> neutrophils, allowing us to assess the function of each cell type, as distinguished by F4/80 and Gr-1 staining (Supplemental Fig. 2). As predicted, both populations isolated from Ab-deficient mice phagocytosed IgG-opsonized cells significantly better than did the comparable populations from B cell intact MRL.Fas<sup>lpr</sup> mice (Fig. 5). Thus, genetically identical macrophages and neutrophils that developed in the presence of Ig are inferior at phagocytosis of IgG-opsonized cells compared with those derived from an Ig-deficient environment.

Discussion

Lupus is considered the prototypical Ab-mediated autoimmune disease. Because the first evidence that B cells could be effective therapeutic targets came from lupus models (4, 20), it was particularly surprising that two controlled clinical trials of rituximab in SLE failed to show efficacy. These failed trials even caused the basic tenet that B cells are important in the pathogenesis of human SLE to be questioned (24). The reasons for the failure of these trials are not clear, but they are important to understand given the ongoing clinical and translational efforts in this area. The interest is further heightened because anti-B lymphocyte stimulator, which likely depends on B cell depletion and inhibition, has shown efficacy in two separate trials and has recently been U.S. Food and Drug Administration approved for treatment of some SLE patients, suggesting that B cells are indeed a valid target in SLE (7).

When attempting to model B cell depletion using anti-CD20 in murine SLE, others and we were struck by the difficulty in

![FIGURE 3](http://www.jimmunol.org/) The active component in serum contains IgG. Serum from older MRL.Fas<sup>lpr</sup> mice was collected and separated into IgG- and non-IgG-containing fractions. Unfractionated serum (black bars), IgG fraction (hatched bars), non-IgG fraction (gray bars), or PBS (open bars) was then infused into hCD20.BALB/c mice as described in Materials and Methods. Mice were then treated with 2H7 and the fraction of B cells in the indicated tissues was measured by FACS as previously reported (10) and is plotted as a percentage of the average of the value of untreated control mice analyzed simultaneously. A, PB, B, LN, and C, spleen. Bars show the mean, and error bars show the SEM. Data are combined from two independent experiments with seven to eight mice per group. ***, p ≤ 0.0001 by Student unpaired t test.
achieved B cell depletion in several different lupus-prone mouse strains (10, 13). In the present report, we have advanced our understanding of the mechanism behind such difficulty in depleting B cells. Using transfer experiments, we showed that serum IgG, but not other serum components, can induce a severe and reversible defect in Ab-mediated depletion in vivo. This was traceable to a defect in macrophage (and neutrophil) IgG-dependent phagocytosis. The mechanism is consistent with data showing that in vivo cellular depletion with anti-CD20 is largely FcγR-mediated (9, 11). Genetic background, aside from its role in inducing the presence of hypergammaglobulinemia and serum ICs, was not a factor since B cells from either MRL.Fas<sup>ly</sup> or BALB/c were equally depleted in a permissive host; additionally, B cells in MRL.Fas<sup>ly</sup> mice lacking serum Ig (mlgM) or those in BALB/c mice were similarly sensitive to depletion.

At present it is unclear which forms of serum IgG are most potent in blocking FcγR-mediated clearance of opsonized B cells. Presumably ICs are the key element, as the ligand for the relevant FcγRs is considered to be ICs and not monomeric IgG. In support of this, depletion defects, albeit more mild, are seen in younger MRL.Fas<sup>ly</sup> mice, prior to significant serum hypergammaglobulinemia (10). However, further studies of different types of ICs, including those purified from sera as well as reconstructed using defined sera and/or mAbs, will be required to better define this. It is possible that particular types of autoantibodies/ICs will be more potent, in part because of their ability to also stimulate macrophages and neutrophils via TLRs (25, 26). In the same vein, glycosylation status of IgG and/or its ability to ligate inhibitory FcγR2b could also be important (27). We hope in the future to be in a position to perform studies to test these various ideas.

Although we focused our studies on depletion of B cells in the MRL model, it is reasonable to think that the same mechanism extends to other murine SLE models that also show defects in depletion of both B cells and T cells (10, 13, 28), as well as SLE patients. This is because the proposed mechanism depends on ICs and autoantibodies present in the IgG fraction of serum. The production of autoantibodies and thus ICs is a common feature in virtually all lupus patients and mouse models. Therefore, this mechanism is predicted to be common among lupus mice and patients as long as sufficient levels of IgG ICs are present. Indeed, we propose that since autoantibodies are common in a wide variety of autoimmune syndromes, some degree of resistance to depletion may be present, possibly subclinically, in a range of conditions. Notably this could include rheumatoid arthritis, a disease in which rituximab can be an accepted therapy, where extent of depletion in synovium (29) or in PB (30) can vary and be associated with clinical response. Unfortunately, it is not currently possible to routinely determine the extent of cellular depletion in secondary lymphoid tissue of treated patients. Notably, results in the mouse models underscore the notion that monitoring PB can lead to an overestimation of the full extent of depletion in tissues.

Supporting the idea that FcγR-mediated phagocytosis is defective in lupus in general is older literature describing difficulty in clearing opsonized RBCs in lupus patients (31). Furthermore, there are reports of an in vitro defect of macrophage IgG-dependent phagocytic function in lupus-prone NZB, NZB/W, and MRL.Fas<sup>ly</sup> mice (32). The studies presented in Fig. 5 confirm these earlier in vitro data. In this connection, it is notable that even with respect to PBLs, there are a number of reports of relative difficulty in depleting B cells in SLE patients (16, 33, 34). Lupus patients also often show faster B cell reconstitution, another measure of reduced efficacy of anti-CD20 Ab in depleting B cells, which could in turn reflect both impaired phagocytic clearance function and reduced therapeutic Ab half-life (16, 33, 34).

Prior to embarking on an expensive course of mAb therapy, it may be cost-effective to determine the likelihood that B cell depletion will be effective. Our studies suggest approaches for developing biomarkers to identify patients at risk for incomplete depletion (35); these could include amounts of circulating ICs or measures of macrophage/monocyte function. Such biomarkers...
could be used for selecting patients to receive Ab-mediated depletion, tailoring dosing regimens, and for monitoring efficacy. Our findings also provide a rationale to develop noninvasive tests for directly monitoring the degree of B cell depletion in secondary lymphoid or target tissue, which has only rarely been done via biopsy (16, 29).

In our model, defects were acquired and not the result of genetic differences in macrophages or neutrophils. Macrophage defects in lupus mice have been reported by several investigators (36–39); however, some reports studied uptake of apoptotic (40) and not opsonized cells, or described defective IC degradation (41). Although some defects were thought to be genetically determined (42) in part due to FcγR polymorphisms (43), it is likely that some were due to acquired dysfunction. Notwithstanding our evidence of acquired defects in MRL.Faslpr mice, it is possible that in some patients or murine models, primary macrophage defects based on genetic differences could also contribute to impaired ability to deplete B cells. For example, polymorphisms in FcγRIIIA that adversely affect B cell depletion in patients are well described (33).

The fact that defects in anti-CD20-mediated B cell depletion can be acquired and result from serum IgG or ICs suggests that they could be reversible. In patients, one might postulate that plasmapheresis or column-based IgG depletion could eliminate enough IgG and ICs to potentiate depletion. Similarly, treatments that ameliorate autoantibody production, such as TACI-Ig (44), could be synergistic with anti-CD20. Indeed, there is evidence for synergism in both SLE and rheumatoid arthritis between non-specific immunosuppression and anti-CD20 therapy, although the mechanism has not been defined (7). In this sense our results are encouraging and suggest that further investigation could improve the therapeutic approach by enabling enhanced depletion of B cells.

One alternative approach is based on the idea that anti-CD20–based depletion might be effective if maintained over a long period of time. This is because effective B cell depletion will, in turn, reduce autoantibodies (10, 45), many of which derive from short-lived plasmablasts (46) that are replenished by CD20+ precursors. A small amount of initial depletion should lead to a small reduction in serum autoantibodies and ICs. This could set up a positive feedback loop, as the first small decrement in IC levels would enable the next round of depletion to be somewhat more effective until ultimately depletion goes to completion over time.

This feedback mechanism might explain why long-term treatment of MRL.Faslpr mice was effective in our prior study (10). Interestingly, persistent treatment that did lead to effective B cell depletion was accompanied by only small reductions in total IgG levels. It was, however, accompanied by a substantial reduction in serum anti-chromatin, anti-DNA, and anti-IgG; this suggests that the component in serum that inhibits depletion is autoantibody-containing ICs rather than just nonspecific IgG. Although rituximab is known to have a long terminal half-life in vivo in lymphoma patients (47), the pharmacokinetics of rituximab are little understood. In patients, one might postulate that they could be reversible. In patients, one might postulate that they could be reversible. In patients, one might postulate that they could be reversible.
Supplemental Figure 1. SDS PAGE analysis of MRL/lpr serum and after separation by Protein G chromatography into IgG and IgG-depleted fraction. Serum was collected and fractions purified as described in Methods. Fractions were all adjusted to a volume equal to the original unseparated serum applied to the column and then equal volumes were applied to the column. The gel was stained with Coomassie blue. Lanes are as indicated, with lane 2 being a purified IgG2b standard.
Supplemental Figure 2. Sample FACS data from phagocytosis assays represented in Figure 5. (A) Pseudocolor plot illustrates the gating of macrophages (F4/80hiGr1int) and neutrophils (Gr1hiF4/80lo) after gating on CD11b+ cells (about 95%). (B) Uptake of PKH26 labeled SRBC. Representative flow cytometric data on gated macrophages (left) and neutrophils (right).