This information is current as of November 13, 2017.

IgG Autoantibodies against Deposited C3 Inhibit Macrophage-Mediated Apoptotic Cell Engulfment in Systemic Autoimmunity

Karla D. Kenyon, Caroline Cole, Fran Crawford, John W. Kappler, Joshua M. Thurman, Donna L. Bratton, Susan A. Boackle and Peter M. Henson

*J Immunol* 2011; 187:2101-2111; Prepublished online 3 August 2011;
doi: 10.4049/jimmunol.1003468
http://www.jimmunol.org/content/187/5/2101

**Why The JI?**

- **Rapid Reviews!** 30 days* from submission to initial decision
- **No Triage!** Every submission reviewed by practicing scientists
- **Speedy Publication!** 4 weeks from acceptance to publication

*average

**References**

This article cites 22 articles, 5 of which you can access for free at:
http://www.jimmunol.org/content/187/5/2101.full#ref-list-1

**Subscription**

Information about subscribing to *The Journal of Immunology* is online at:
http://jimmunol.org/subscription

**Permissions**

Submit copyright permission requests at:
http://www.aai.org/About/Publications/JI/copyright.html

**Email Alerts**

Receive free email-alerts when new articles cite this article. Sign up at:
http://jimmunol.org/alerts
IgG Autoantibodies against Deposited C3 Inhibit Macrophage-Mediated Apoptotic Cell Engulfment in Systemic Autoimmunity

Karla D. Kenyon,* Caroline Cole,* Fran Crawford,† John W. Kappler,‡ Joshua M. Thurman,† Donna L. Bratton,* Susan A. Boackle,§,∥ and Peter M. Henson*

Defective clearance of apoptotic cells has been shown in systemic lupus erythematosus (SLE) and is postulated to enhance auto-immune responses by increasing access to intracellular autoantigens. Until now, research has emphasized inherited rather than acquired impairment of apoptotic cell engulfment in the pathogenesis of SLE. In this study, we confirm previous results that efficient removal of apoptotic cells (efferocytosis) is bolstered in the presence of wild-type mouse serum, through the C3 deposition on the apoptotic cell surface. In contrast, sera from three mouse models of SLE, MerKΔ, MRLpr, and New Zealand Black/WF1 did not support and in fact actively inhibited apoptotic cell uptake. IgG autoantibodies were responsible for the inhibition, through the blockade of C3 recognition by macrophages. Consistent with this, IgG removal reversed the inhibitory activity within autoimmune serum, and purified autoimmune IgG blocked both the detection of C3 on apoptotic cells and C3-dependent efferocytosis. Sera from SLE patients demonstrated elevated anti-C3b IgG that blocked detection of C3 on apoptotic cells, activity that was not found in healthy controls or patients with rheumatoid arthritis, nor in mice prior to the onset of autoimmunity. We propose that the suppression of apoptotic cell disposal by Abs against deposited C3 may contribute to increasing severity and/or exacerbations in SLE. The Journal of Immunology, 2011, 187: 2101–2111.

Systemic lupus erythematosus (SLE) is a chronic, multiorgan autoimmune disease that is characterized by the production of high titers of Abs against nuclear Ags, including dsDNA, histones, and small ribonuclear proteins (1, 2). Dying cells are thought to be the primary source of intracellular autoantigens in lupus, through the release of nucleosomes (3–5) and the display of nuclear Ags on membrane blebs (6, 7) in situations in which apoptotic cell clearance is defective.

The death and disposal of aged and damaged cells are essential for the maintenance of healthy tissues. In healthy individuals, apoptotic cells are cleared with remarkable efficiency. This is in part due to alterations of the dying cell surface, which are recognized as “eat me” signals by phagocytes, which ingest apoptotic cells through a process that has been termed efferocytosis (3). Numerous serum proteins, including C1q, IgM, C-reactive protein, and mannan-binding lectin, deposit on apoptotic cells, initiating and amplifying the deposition of C3 and its degradation products C3b and iC3b (5) and resulting in enhanced removal via recognition by complement receptors CR3 and CR4 (4, 6, 7).

The processes that govern dying cell clearance are defective in SLE. As a result, apoptotic cells accumulate in affected tissues (8–12). The persistence of apoptotic cells is thought to result in secondary necrosis and the release of proinflammatory and proimmunogenic intracellular constituents that contribute to the pathogenic autoantibody production. Therefore, insight into the pathways that govern apoptotic cell ingestion may be critical to uncovering the mechanisms of disease progression in SLE.

Reports correlating inefficient clearance with systemic autoimmunity have evoked inherited defects in either components of the apoptotic cell recognition mechanisms, such as C1q (13), or defects in the macrophages’ ability to phagocytose apoptotic cells (14–17). However, the contribution of acquired defects in efferocytosis (those that arise as a consequence rather than the cause of disease progression) in SLE has not been fully explored.

In the present work, we investigate the hypothesis that breaks in lymphocyte tolerance may precede and be the cause of apoptotic cell clearance defects in systemic autoimmunity. We show that IgG Abs that inhibit apoptotic cell uptake develop in three different strains of autoimmune mice. The inhibitory IgG Abs were directed against C3b components on the apoptotic cells, yet did not alter the total amounts or composition of the bound C3. Instead, they are suggested to block the interaction between C3b bound to the apoptotic cell surface and phagocyte C3 receptors. In mice, these Abs became apparent only as the animals developed their autoimmune state. Further analysis revealed higher titers of these Abs in patients with SLE compared with healthy control (HC) subjects or patients with rheumatoid arthritis (RA). Therefore, anti-C3 Abs that develop as a consequence of deficient lymphocyte tolerance may serve to initiate and/or exacerbate defects in apoptotic cell clearance and thereby accentuate the autoimmune and/or inflammatory processes in SLE.

*Department of Pediatrics, National Jewish Health, Denver, CO 80206; †Department of Immunology, National Jewish Health, Denver, CO 80206; ‡Division of Nephrology and Hypertension, University of Colorado Denver School of Medicine, Aurora, CO 80045; §Department of Medicine, University of Colorado Denver School of Medicine, Aurora, CO 80045; and ∥Department of Immunology, University of Colorado Denver School of Medicine, Aurora, CO 80045

Received for publication October 19, 2010. Accepted for publication June 18, 2011.

This work was supported by National Institutes of Health Grants GM61031 and HL81151. Additional support was provided by National Institutes of Health Grants DK076690 (to J.M.T.), AI070304 and 5K24AI078004 (to S.A.B.), and A1058228 (to D.L.B.).

Address correspondence and reprint requests to Dr. Peter M. Henson, Department of Pediatrics, National Jewish Health, A539, 1400 Jackson Street, Denver, CO 80206. E-mail address: hensonp@njhealth.org

Abbreviations used in this article: HC, healthy control; HMDM, human monocyte-derived macrophage; NZB, New Zealand Black; RA, rheumatoid arthritis; SLE, systemic lupus erythematosus.

Copyright © 2011 by The American Association of Immunologists, Inc. 0022-1767/11/$16.00 www.jimmunol.org/cgi/doi/10.4049/jimmunol.1003468
Materials and Methods

Mice
C57BL/6, BALB/c, ICR, MRL, MRLfp, and New Zealand Black (NZB)/WF1 mice were purchased from The Jackson Laboratory. MertK mice were provided by Dr. Douglas Graham at the University of Colorado Denver Anschutz Medical Campus (Aurora, CO). C3−/− mice were from Michael Carroll at Harvard University (Boston, MA). MRL mice were used as a control for MRLfp to develop autoimmune disease until later in life. All animal studies were performed in compliance with the United States Department of Health and Human Services Guide for the Care and Use of Laboratory Animals. The Institutional Animal Care and Use Committee at National Jewish Health approved all experimental procedures performed on the animals.

Human cells and serum
Whole blood was collected from healthy donors for the isolation of mononuclear cells using CPT tubes (BD Biosciences) or sera in accordance with the guidelines of the Institutional Review Board at National Jewish Health (Denver, CO). Deidentified serum samples from 43 SLE patients, 18 patients with RA, and 19 nonautoimmune controls (HC) were obtained from a research protocol approved by the Institutional Review Board at University of Colorado, in which subjects had provided informed consent to allow their leftover samples to be used for additional research in association with linked clinical data. Deidentified serum samples from 10 additional SLE patients were obtained from the University of Colorado Clinical Rheumatology Laboratory and represent leftover samples after performance of clinical tests. Samples from lupus patients obtained from the Clinical Rheumatology Laboratory were identified based on ICD9 codes, but information was not recorded that could link the samples back to the patients; thus, the research involving these samples is not considered human subjects research and is exempt from the requirements of the United States Department of Health and Human Services under 45 CFR 46.101(b) (4). Deidentified serum samples from 16 additional HC subjects, also obtained from the University of Colorado Clinical Rheumatology Laboratory, were from anonymous blood bank donors used by the laboratory for quality control purposes, and thus, the research involving these samples is not considered human subjects research. SLE and HC sera were used in experiments involving macrophage uptake so that differences in how sera were handled or changes in sera complement levels for each subject would not affect the total amount of complement in each sample. SLE sera with the greatest ability to block C3 detection and HC sera that did not block C3 detection in initial experiments were selected for further studies involving protein G treatment.

Abs and other reagents
The FITC- or HRP-conjugated goat anti-mouse C3 and goat anti-human C3 Abs were purchased from MP Biomedicals and used at 1:25 for flow cytometry and 1:1000 for Western blots along with anti-actin (Chemicon) (1:2000), peroxidase-conjugated rabbit anti-mouse IgG, and rabbit anti-human IgG Abs (Jackson ImmunoResearch Laboratories) (1:10,000 for Western blot and 1:1,000 for ELISA). Control (isotype) goat IgG was from Jackson ImmunoResearch Laboratories and used at the same concentration as the anti-C3. Mouse serum was prepared from blood collected via cardiac puncture or tail vein. Purified human C3 protein was purchased from Quidel (San Diego, CA). Human TruStain Fc Receptor Blocking Solution was purchased from BioLegend (San Diego, CA) and used to treat human monocyte-derived macrophages (HMDM) at 1:20 in X-Vivo for 30 min prior to initiation of uptake assays. Mouse anti-human CD3 (BD Biosciences) was used at 1 μg/106 Jurkat cells for 30 min prior to washing and initiation of Fe-mediated uptake assay.

Cells and cell culture
Murine J774 macrophages (American Type Culture Collection) were cultured in DMEM (Mediatech) containing 10% heat-inactivated FBS supplemented with 2 mM l-glutamine in humidified 5% CO2 at 37°C. A primary CD4+ T cell clone isolated from NOD mice was a gift of D. Tonkin at National Jewish Health (Denver, CO). Murine splenocytes were prepared from C57BL/6 with a 100 μM cell strainer and were cultured similarly to Jurkats.

Induction of apoptosis
Apoptosis of Jurkat T cells, CD4+ T cells, or splenocytes (106 cells/ml in complete media) was induced by exposure to UV irradiation at 254 nm for 10 min, followed by 2–3 h of culture at 37°C. Percentage of apoptosis was confirmed by evaluation of nuclear morphology by light microscopy after PROTOCOL Hema 3 staining.

Phagocytosis assay
J774 macrophages were plated at 5 × 105 cells per 24-well 2 d before initiation of the assay and primary mouse macrophages at 2.3 × 105 cells/well for 1 d. HMDM were cultured at 1 × 105 cells per well for 7 d. Macrophages were washed thoroughly with PBS and cultured for 2–3 h in serum-free media before the phagocytosis assays were performed. A total of 0.5–1 × 105 targets, either apoptotic cells, viable cells, or anti-CD3- coated Jurkats (Fc-mediated uptake), was cocultured with effector macrophages for 1.5 h. Macrophages were washed four times with ice-cold PBS, fixed, and stained with PROTOCOL Hema 3 staining. Phagocytosis (minimum 200 cells, 2–3 replicate wells per condition) was scored by visual inspection of blinded samples using light microscopy. Results are displayed as phagocytic index, which is defined as the total number of apoptotic targets ingested divided by the total number of phagocytes counted × 100.

For serum pretreatment, apoptotic Jurkat T cells were pelleted and resuspended in DMEM alone or with 10% C57BL/6 mouse serum for 30 min at 37°C, washed with DMEM, and resuspended in DMEM alone or with 10% C57BL/6 mouse serum for phagocytosis or flow cytometry. For C3 blockade, apoptotic Jurkat T cells were resuspended in DMEM plus 10% freshly prepared C57BL/6 mouse serum with anti-mouse C3 or isotype IgG before coinubcation with macrophages. For mixed sera samples, 2.5% autimmune serum was added to 5% control sera just before the initiation of the phagocytosis assay. For HMDM, uptake of apoptotic Jurkats was done with 10% fresh human sera alone (control) or mixed with 5% heat-inactivated SLE or HC sera in X-Vivo.

Flow cytometry
Apopotic Jurkat T cells were resuspended in DMEM containing 10% mouse serum, incubated for 30 min at 37°C, washed, and stained at 0.75–1 × 105 cells in 0.1 ml vol on ice with the FITC-conjugated goat anti-human C3 or isotype Ab. Ten percent nonautoimmune serum plus 5, 10, or 20% autoimmune serum was used for mixed samples. For experiments with human sera, 15% fresh human sera in X-Vivo was used to opsonize Jurkats with C3. Cells were washed and incubated with X-Vivo (control) and 5% heat-inactivated SLE or HC sera, and then washed and stained with FITC-conjugated goat anti-human C3 or isotype Ab. Cells were analyzed by flow cytometry using either the FACScan or FACSCalibur systems running on CellQuest software (BD Biosciences), and plots were rendered using FlowJo software (Tree Star).

Mouse serum fractionation
DMEM with 6% sucrose and either MertK or C57BL/6 sera was loaded onto a Superdex 200 gel filtration column, and 31 0.5–ml–vol samples were collected. For each sample, the protein concentration was determined by absorbance at 280 nM, and phagocytosis assays were performed after adding C57BL/6 serum (5% final concentration) to duplicate wells for each fraction.

Depleting and purifying IgG from sera
Protein G-Sepharose (Zymed) or Sepharose 4B (Amersham) was washed with PBS, blocked with 1% BSA, washed again, and resuspended in a 50:50 bead:media (either DMEM + l-glutamine or X-Vivo) slurry. A total of 50 μl sera and 300 μl beads was incubated for 2 h while rotating at 4°C and pelleted, and supernatant was collected. Beads were washed with media and mixed with the original supernatant to make 1 ml total volume. All samples were filtered through a 0.45-μm filter before phagocytosis or flow cytometry. Fc-mediated uptake assays were performed, depletion of IgG from sera was confirmed by Western blot. IgG was eluted with 0.1 M glycine HCl (pH 2.8), neutralized with Tris (pH 9.0), and concentrated with an Amicon Ultra-4 (100,000 MWCO) filter.
Western blots

Lysates of apoptotic cells (0.5–1 × 10⁶) served in SDS-PAGE and transferred to polyvinylidene fluoride membranes. Membranes were blocked with 5% milk, probed with Abs at concentrations listed above, washed, incubated with West Femto Maximum Sensitivity Substrate (Pierce), and visualized on film. Degradation of the C3 α-chain produces proteins of 115, 68, and 43 kDa.

ELISA for C3b-specific, dsDNA-specific, or chromatin-specific IgG

A total of 200 µg purified human C3b protein (Comptech) was coated onto ELISA plates (Falcon PVC) in PBS overnight at 4°C. The plates were blocked with 1% BSA in PBS for 2 h at room temperature and washed four times with PBS plus 0.05% Tween-20 before adding 100 µl 1:800 dilution of human serum (SLE, RA, or HC) or 1:100 dilution of mouse sera (C57BL/6 or Mer) to each well. After 2 h at room temperature, plates were washed and 100 µl peroxidase-conjugated anti-human IgG (Jackson Immunoresearch Laboratories) or 100 µl peroxidase-conjugated anti-mouse IgG (ICN Pharmaceuticals) was added for 1 h at room temperature. Plates were washed and developed with 100 µl ABTS (Thermo Scientific). OD was read at 405 nm on a microplate reader.

Anti-chromatin and anti-dsDNA Abs were detected by a sandwich ELISA, as described (18), except SNF1 sera was used as a positive control and BALB/c sera as a negative control. Statistical analysis was performed by one-way ANOVA.

Results

Nonautoimmune mouse serum enhances murine macrophage-mediated apoptotic cell clearance in a C3-dependent fashion

To examine the effect of autoimmune serum on the uptake of apoptotic cells by macrophages, we first examined the effect of nonautoimmune serum on this process. As shown in Fig. 1, ingestion of apoptotic cells by murine macrophages was markedly enhanced in the presence of mouse serum compared with standard tissue culture conditions using heat-inactivated FBS. The enhancement was equally effective with serum from three different mouse strains (Fig. 1D) and did not occur with viable cells (Fig. 1C). In keeping with studies using human serum reported earlier by Matsui et al. (4) and Mevorach et al. (6), the enhancement in this murine system was also shown to depend on complement activity and the presence of C3, that is, it was abrogated by heat inactivation of the serum, blockade with an anti-C3 Ab, or by use of serum from C3−/− mice (Fig. 1E, 1F). Pretreatment of apoptotic cells with fresh mouse serum resulted in deposition of C3 on the apoptotic cell surface, as demonstrated by FACS analysis (Fig. 1D, 1G). The latter approach indicated that C3b and iC3b could be detected on the apoptotic cells. By itself, the deposition of C3 on the apoptotic cells was insufficient to mediate uptake into macrophages because the subsequent removal of the serum prevented the ingestion of C3-opsonized apoptotic cells (Fig. 1H). These experiments support the notion that C3 is necessary, but not sufficient, for the effect of serum in supporting uptake of apoptotic cells, and that additional serum factors are required.

Autoimmune serum inhibits apoptotic cell engulfment by blocking C3 detection

Systemic autoimmunity has previously been associated with defects in apoptotic cell clearance that were attributed to either impaired phagocyte activity or lack of serum factors to support uptake (19). Accordingly, serum from the MerKO murine model of systemic autoimmunity did not support the efficient uptake of human or murine apoptotic targets (Fig. 2A), regardless of the macrophages analyzed (primary or immortalized murine macrophages) (Fig. 2B). Moreover, the defect in efferocytosis correlated with the chronological development of autoimmunity, as serum from preautoimmune, 6-wk-old MerKO mice (20) promoted similar levels of apoptotic cell uptake as that from age-matched C57BL/6 controls (Fig. 2C). When sera from two other models of systemic autoimmunity were tested, neither MRL/Lpr nor NZB/WF1 serum proved capable of supporting efferocytosis to the levels seen in control sera (Fig. 2D, 2E). The inability for autoimmune serum to support engulfment may be explained by either the absence of an enhancing factor or the presence of an inhibitor. To distinguish between these possibilities, efferocytosis was assessed in a mixture of nonautoimmune (C57BL/6) or preautoimmune (MRL) and autoimmune sera. The results demonstrated that sera from all three autoimmune strains inhibited apoptotic cell engulfment (Fig. 2F).

To determine whether autoimmune mouse sera blocked effector functions of macrophages, we examined apoptotic cell clearance by FACS analysis and Western blotting. These studies showed that C3 was nearly undetectable when cells were exposed to autoimmune serum from either MRL/Lpr (Fig. 3A) or MerKO (Fig. 3C) mice. Furthermore, C3 levels were sharply reduced when autoimmune serum was mixed with pre- or nonautoimmune serum. By contrast, C3 protein levels (and the distribution of subfragments) were unchanged when analyzed by Western blot (Fig. 3B, 3D). Taken together, these results suggest that autoimmunity leads to development of an inhibitor of efferocytosis in these mice that blocks the macrophage recognition of complement C3 on the surface of apoptotic cells.

Identification of anti-C3 Ab as the inhibitor present in autoimmune serum

Superdex 200 gel filtration of serum was used to identify candidate molecules that inhibited the uptake of apoptotic cells by macrophages. In the absence of an enhancing factor or the presence of an inhibitory factor or Mer KD sera removed the inhibitor of apoptotic cell clearance (Fig. 5A, 5D), and in the case of the MRL/Lpr serum, restored the ability to detect C3 on the apoptotic cells by FACS analysis (Fig. 5B). Despite the observation that protein G treatment removed the inhibitory activity present in MerKD serum, this treatment did not restore our ability to detect C3 on the surface of apoptotic cells by flow cytometry (Fig. 5E). Actual C3 deposition was unaltered by the presence of either MRL/Lpr or MerKD autoimmune serum (Fig. 5C, 5F). However, protein G-mediated removal of the inhibitory activity did not render autoimmune sera capable of supporting high levels of apoptotic cell uptake, like that which occurred in the presence of pre- or nonautoimmune serum (Fig. 5A, 5D). Protein G treatment may remove enhancing factors that are required to promote optimal clearance or autoimmune serum may inherently lack these factors.

Protein G-purified IgG from MRL/Lpr, MerKO, and their control serum counterparts was examined to determine whether IgG could directly block efferocytosis and C3 detection. IgG from both autoimmune strains reduced phagocytosis by 90% (Fig. 6A, 6C) and extinguished C3 protein detection on the surface of apoptotic cells.

Downloaded from http://www.jimmunol.org/ by guest on November 13, 2017
Western blot analysis confirmed that IgG purified from all sera did not alter the levels of C3 protein deposited on apoptotic cells (Fig. 6D). IgG from control sera had no effect. 

**FIGURE 1.** C3 deposition was necessary, but not sufficient for serum enhancement of apoptotic cell engulfment. A, Comparison of apoptotic Jurkat T cell uptake by J774 macrophages in 10% heat-inactivated FBS or 10% fresh C57BL/6 mouse serum (BL/6). Arrows indicate engulfed apoptotic cells or cell fragments. Original magnification ×40. B, Quantification of engulfment assay described in A showed enhanced uptake using serum from BL/6, ICR, or BALB/c mice compared with FBS (n = 3). C, Engulfment of apoptotic, but not viable, Jurkat T cells was seen using BL/6 serum (n = 3). D, C3 on the surface of apoptotic Jurkats was detected by FACS after treatment with BL/6 serum, but not after no serum or heat-inactivated BL/6 serum (ΔH BL/6). E, Compared with BL/6 serum, engulfment of apoptotic Jurkats was reduced in media containing ΔH BL/6 and C3 knockout mouse serum (C3−/−) (n = 3). F, Treatment of BL/6 serum with neutralizing goat anti-mouse C3 also reduced efferocytosis compared with nontreated (NT) or control Ab (IgG) (n = 3). G, C3 could be detected on apoptotic Jurkats by FACS after treatment with BL/6 serum, even after a second incubation in serum-free media (BL/6 Serum → No Serum). H, Phagocytosis of apoptotic Jurkats was observed when the assay was performed in BL/6 serum, but not without serum despite the presence of C3 on apoptotic Jurkats preincubated in BL/6 serum (gray bar) compared with no serum (black bar) (n = 3). The results in this figure are expressed as phagocytic index (PI), which is defined as the total number of apoptotic targets ingested divided by the total number of phagocytes counted multiplied by 100. PI is expressed as a percentage of nonautoimmune serum control. Unfilled histograms are isotype labeling; filled histograms are anti-C3 labeling. *p < 0.05.
significant due to the much lower number of mice that lacked antichromatin autoantibodies in this analysis (Fig. 6). There was no correlation of anti-C3b levels with age of the mice or titers of autoantibodies (data not shown). Taken as a whole, these data suggest that IgG autoantibodies are elicited in the course of autoimmune disease that can inhibit apoptotic cell engulfment by binding to C3 on the surface of apoptotic cells and blocking its interaction with macrophage complement receptors.

C3 detection is blocked by Ig from the sera of SLE patients

Human sera contains natural Abs to C3 (9) as well as induced autoantibodies (immunoconglutins) that are thought to be generated in response to acute or chronic inflammation involving complement activation (10), but these Abs have not been shown to directly interfere with macrophage uptake of C3-opsonized apoptotic cells. To test whether Abs to C3 contribute to the clearance defects in human SLE as in murine models, the capacity of sera from SLE patients to block C3 accessibility and phagocytic uptake of serum-exposed apoptotic cells was evaluated. Sera from SLE patients contained significantly higher titers of anti-C3 Abs when compared with sera from nonautoimmune controls or patients with RA (Fig. 7A). Consistent with this, C3 detection on apoptotic cells incubated in fresh sera mixed with heat-inactivated HC sera (Fig. 7B). Protein G treatment of sera from the SLE patients with the highest level of inhibition of C3 detection restored the ability to detect C3 on apoptotic cells (Fig. 7C), and IgG purified from these sera blocked C3 detection (Fig. 7C). Deposition of C3 was not affected by SLE sera. Detection and deposition of C3 by sera from three HC were not changed by these treatments (Fig. 7C,7G). As previously shown by Mevorach et al. (6), human sera enhanced the uptake of apoptotic Jurkat cells by HMDM, and this was blocked by heat inactivation (Fig. 7D). However, the efferocytosis mediated by fresh sera was not blocked in these experiments by mixing either SLE or HC sera (Fig. 7E). Additional experiments were performed after blockade of macrophage Fcγ receptors to isolate complement-mediated uptake. In these experiments, protein G treatment of the majority of serum samples in both groups improved macrophage uptake of C3-opsonized cells, and IgG purified from these sera had no effect (Fig. 7F).

Discussion

The data presented in this work suggest that deficiencies in apoptotic cell engulfment need not precede the onset of systemic autoimmunity, but may instead arise as a result of C3-targeted
immune responses. This study describes a mechanism whereby C3-specific IgG Abs delay efferocytosis by blocking macrophage detection of activated C3 on apoptotic cells. Consistent with previous reports, C3 opsonization enhanced efferocytosis (4, 6) and autoimmune sera did not support apoptotic cell clearance (19). The current study expanded on these observations by demonstrating that autoimmune serum can contain Abs against opsonized C3 that inhibit apoptotic cell uptake. We propose a model in which acquired defects in efferocytosis may exacerbate intrinsic, inappropriate autoimmune responses.

The exposure of phosphatidylserine on the surface of dying cells has been suggested to initiate activation of the complement system with subsequent cell surface C3 deposition that can then contribute to removal by CR3 and CR4 receptors (4, 6, 7, 12). However, as noted in the current studies, opsonization of the apoptotic cells with C3 still required additional serum factors for uptake. This raises the possibility that C3 receptors in this system serve primarily as essential tethering ligands that then need additional stimuli to optimally induce the actual ingestion (8). Because removal of the inhibitory anti-C3 IgG from autoimmune serum did not restore the ability of that serum to support efficient apoptotic cell engulfment, even when complement components (C1q or C3) were added back (data not shown), we concluded that C3 was necessary, but not sufficient for clearance, and that additional phagocytic components (stimuli) were also lacking in the autoimmune serum.

Consistent with the hypothesis that serum from autoimmune animals inhibited efferocytosis by blocking the C3-dependent component of the uptake, depletion of IgG from MRL\textsuperscript{1pr} serum prevented its ability to inhibit the efferocytosis as well as the detection of C3 on the apoptotic cells. The situation with the Mer\textsuperscript{KD} system, however, appeared more complex. IgG depletion of Mer\textsuperscript{KD} serum removed its ability to inhibit control serum-mediated uptake, but did not restore the ability to detect C3 on the apoptotic cells with exogenous Ab. One explanation for this result is that the macrophage CR3 and CR4 may recognize different domains of C3, or with greater affinity, than those bound by the C3 detection Ab. Alternatively, additional factors may be present in the Mer serum that mask C3 detection, but do not block macrophage tethering and removal. Attempts to remove IgM autoantibodies as one potential candidate for this factor did not restore the detection (data not shown).

Nonetheless, the IgG isolated from either autoimmune mouse strain was shown to inhibit C3 reactivity and was able to inhibit efferocytosis. Thus, in contrast to the IgG purified from control sera, the IgG purified from either MRL\textsuperscript{1pr} or Mer\textsuperscript{KD} serum inhibited
It has been shown that the paucity of Mer expression in MerKD mice leads to a primary defect in macrophages that prevents apoptotic cell ingestion (11) that contributes to the subsequent development of autoimmunity (20). However, sera from all three murine models of systemic autoimmunity examined exhibited the inhibitory activity, independent of primary defects in macrophage-mediated efferocytosis. This suggests that autoimmunity in the MerKD mice might result from the in vivo clearance deficiency of the macrophages and be exacerbated by the development of the Ab inhibitor to efferocytosis. Our preliminary fractionation data suggested that there may also be other inhibitors present in Mer sera that we did not investigate. In the other autoimmune strains, as well as human patients, additional and/or alternative uptake defects may also contribute to the decreased apoptotic cell clearance that has been hypothesized as the source of autoantigens for the autoimmune response.

It is intriguing to note that autoantibodies termed immunoglobulins, and later shown to react against fixed C3, have been noted since the 1930s (21), and in fact represented a subject of investigation for one of us in his PhD project (22). Elevated levels of immunoglobulins, with varying Ig isotypes, have been reported in the sera from patients with chronic infection or autoimmune diseases, including SLE and RA, in which they increased during exacerbations (10). Whether in humans these exhibit variable effects in apoptotic cell clearance depending on isotype (i.e., ability to react with Fc receptors) or fine specificity against the C3 Ag is Figure 4. The inhibitory activity in autoimmune serum coeluted with Ig. Serum from MerKD mice (A) or control C57BL/6 (B) was fractionated on a Superdex 200 size gel filtration column. For each fraction, the relative serum protein levels (gray histogram) and apoptotic Jurkat T cell engulfment by J774 macrophages after mixing with C57BL/6 serum (black bars) are depicted. For phagocytosis assay, whole serum controls (gray bars) were 5% C57BL/6 serum, 5% C57BL/6 serum plus 2.5% MerKD serum (mixed serum), and 5% MerKD serum. Analysis of the inhibitory activity in serum fractions from each strain was performed separately, and data represent the mean of samples run in duplicate from a single experiment. *Fractions in which both duplicates demonstrated inhibitory activity.

FIGURE 4. The inhibitory activity in autoimmune serum coeluted with Ig. Serum from MerKD mice (A) or control C57BL/6 (B) was fractionated on a Superdex 200 size gel filtration column. For each fraction, the relative serum protein levels (gray histogram) and apoptotic Jurkat T cell engulfment by J774 macrophages after mixing with C57BL/6 serum (black bars) are depicted. For phagocytosis assay, whole serum controls (gray bars) were 5% C57BL/6 serum, 5% C57BL/6 serum plus 2.5% MerKD serum (mixed serum), and 5% MerKD serum. Analysis of the inhibitory activity in serum fractions from each strain was performed separately, and data represent the mean of samples run in duplicate from a single experiment. *Fractions in which both duplicates demonstrated inhibitory activity.

It has been shown that the paucity of Mer expression in MerKD mice leads to a primary defect in macrophages that prevents apoptotic cell ingestion (11) that contributes to the subsequent development of autoimmunity (20). However, sera from all three murine models of systemic autoimmunity examined exhibited the inhibitory activity, independent of primary defects in macrophage-mediated efferocytosis. This suggests that autoimmunity in the MerKD mice might result from the in vivo clearance deficiency of the macrophages and be exacerbated by the development of the Ab inhibitor to efferocytosis. Our preliminary fractionation data suggested that there may also be other inhibitors present in Mer sera that we did not investigate. In the other autoimmune strains, as well as human patients, additional and/or alternative uptake defects may also contribute to the decreased apoptotic cell clearance that has been hypothesized as the source of autoantigens for the autoimmune response.

It is intriguing to note that autoantibodies termed immunoglobulins, and later shown to react against fixed C3, have been noted since the 1930s (21), and in fact represented a subject of investigation for one of us in his PhD project (22). Elevated levels of immunoglobulins, with varying Ig isotypes, have been reported in the sera from patients with chronic infection or autoimmune diseases, including SLE and RA, in which they increased during exacerbations (10). Whether in humans these exhibit variable effects in apoptotic cell clearance depending on isotype (i.e., ability to react with Fc receptors) or fine specificity against the C3 Ag is...
FIGURE 5. IgG depletion with protein G-Sepharose removed the inhibitory activity present in autoimmune serum. Analyses of MRL^lpr^ (A–C) and Mer^KD^ (D, E) sera treated with protein G-Sepharose beads to remove serum IgG. A and D, Engulfment assays using J774 macrophages and apoptotic Jurkat T cells. Single serum samples contained 5% serum each, whereas “Mixed Serum” samples contained 5% untreated pre- or nonautoimmune serum plus 2.5% autoimmune serum. Autoimmune serum was either untreated (black bars), control Sepharose bead treated (dark gray bars), or protein G-Sepharose bead treated (light gray bars). Removing IgG from MRL^lpr^ serum (A) and Mer^KD^ serum (C) using protein G-Sepharose did not restore the ability of these sera to promote uptake of apoptotic cells, but did prevent these autoimmune sera from inhibiting the uptake of apoptotic cells when mixed with nonautoimmune sera (n = 3). B and E, C3 was detected by FACS on apoptotic Jurkats treated with either pre- or nonautoimmune serum (black), autoimmune serum (dark gray), pre- or nonautoimmune serum mixed with autoimmune serum (medium gray), or pre- or nonautoimmune serum mixed with protein G-treated, IgG-depleted autoimmune serum (light gray). Protein G removal of IgG from MRL^lpr^ serum (B), but not Mer^KD^ serum (E), allowed C3 protein to be detected on the surface of apoptotic cells treated with mixed serum (n = 3). Western blot demonstrated that C3 protein and distribution of its fragments bound to the apoptotic cells were unchanged when exposed to MRL, MRL^lpr^, or a mixture of both sera (C), or C57BL/6, Mer^KD^, or a mixture of both sera (F) (results shown here are representative of those obtained in three separate experiments). PI is expressed as a percentage of nonautoimmune serum control. Unfilled histograms are isotype labeling; filled histograms are anti-C3 labeling. The geometric mean fluorescence for C3 (C3 Protein MFI) is expressed as a percentage of nonautoimmune serum control. *p < 0.05.
Apoptotic cell engulfment and C3 protein detection were inhibited by IgG purified from autoimmune sera. A and C, Engulfment of apoptotic Jurkats by J774 macrophages in pre- or nonautoimmune sera alone or mixed with IgG purified from either pre- or nonautoimmune or autoimmune sera. IgG purified from MRL^{lpr} (A) and Mer^{KD} sera (C) inhibited the uptake of apoptotic Jurkats seen in the presence of pre- or nonautoimmune sera (n = 4). B and D, C3 was detected by FACS on apoptotic Jurkat T cells incubated with pre- or nonautoimmune serum alone (black), or pre- or nonautoimmune serum mixed with IgG purified from pre- or nonautoimmune (dark gray) or autoimmune (light gray) mouse serum. The detection of C3 was inhibited by IgG purified from MRL^{lpr} (B) and Mer^{KD} (D) (n = 3). IgG purified from nonautoimmune sera had no effect on uptake or FACS detection of C3 (results shown here are representative of those obtained in three separate experiments). Phagocytic index (PI) is expressed as a percentage of nonautoimmune serum control. Unfilled histograms are isotype labeling; filled histograms are anti-C3 labeling. The geometric mean fluorescence for C3 (C3 Protein MFI) is expressed as a percentage of nonautoimmune serum control. E, C3 deposition was unaltered as detected by Western blot analysis of total C3 protein and distribution of its fragments bound to apoptotic cells exposed to pre- or nonautoimmune serum mixed with IgG purified from either autoimmune serum or pre- or

FIGURE 6.
not clear, but the possible effect described in this work on blocking C3 recognition could be a contributing factor in the progression and/or exacerbations seen in systemic autoimmune diseases.

Disclosures

J.M.T. is a stockholder in and consultant for Taligen Therapeutics, Inc. The other authors have no financial conflicts of interest.

FIGURE 7.  SLE sera contained IgG against C3b that blocked opsonized C3 detection.  A, ELISA showed that the mean anti-C3b titers were higher in sera from SLE patients (n = 53) compared with sera from HC (n = 35) or patients with RA (n = 18).  B, FACS detection of C3 on apoptotic Jurkats preincubated with fresh human sera (Control) was reduced by mixing heat-inactivated SLE (n = 16), but not heat-inactivated HC sera (n = 16).  C, Removal of IgG with protein G treatment restored the FACS detection of C3 from selected SLE sera (n = 3), whereas the IgG purified from these SLE sera blocked C3 detection (n = 3).  Removal of IgG and purified IgG from HC sera had no effect on the detection of C3 (n = 3).  D, HMDM uptake of apoptotic Jurkats was enhanced by fresh, but not heat-inactivated (ΔH) human sera (n = 4).  E, Mixing sera from SLE patients (n = 14) or HC (n = 16) with fresh human sera did not inhibit uptake of apoptotic Jurkats compared with fresh sera alone (Control).  F, Uptake of apoptotic cells with fresh human sera mixed with either SLE (n = 6) or HC (n = 7) was increased by protein G treatment of these sera, whereas the purified IgG had no effect.  G, Western blots of lysates revealed no change in total C3 bound to apoptotic cells incubated in fresh sera and mixed with SLE (n = 6) or HC (n = 7).  Dots represent each individual sera, horizontal lines are the mean for the group, and whiskers are the SEM.  The geometric mean of C3 staining (MFI) on apoptotic Jurkats preincubated in 15% human sera alone was used as the control.  Phagocytic index of apoptotic Jurkats by HMDM in 10% fresh human sera alone was used as the control.  *p < 0.05.

nonautoimmune controls.  F, IgG from autoimmune MRL

IgM and MeC57 sera bound purified C3 and its degradation products by immunoblotting (blot shown here is representative of results obtained in multiple experiments).  G, J774 macrophage phagocytosis of apoptotic Jurkats (Fc-mediated), was enhanced by C57BL/6 sera (n = 3).  H and I, Sera from 23 MeC57 mice were tested by ELISA for anti-C3b and markers of autoimmunity (anti-dsDNA or anti-chromatin).  Each dot represents an individual animal, with the line representing the mean for the group and the whiskers representing the SEM.  Elevated levels of anti-C3b were detected in the sera of MeC57 mice with anti-dsDNA (n = 17) (H) or anti-chromatin (n = 20) (I) autoantibodies compared with the MeC57 mice lacking these autoantibodies (n = 6 and 3, respectively).  *p < 0.05.
References

22. Henson, P. M. 1968. Immunoconglutinins of different immunoglobulin classes demonstrated by the antiglobulin reaction. Immunology 14: 697–705.