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Micrometer-Sized Titanium Particles Can Induce Potent Th2-Type Responses through TLR4-Independent Pathways

Pankaj K. Mishra,* Wenhui Wu,* Cristina Rozo,* Nadim J. Hallab,† Joseph Benevenia,‡ and William C. Gause*

Wear debris in joint replacements has been suggested as a cause of associated tissue-damaging inflammation. In this study, we examined whether solid titanium microparticles (mTi) of sufficient size to accumulate as wear debris could stimulate innate or adaptive immunity in vivo. mTi, administered in conjunction with OVA, promoted total and Ag-specific elevations in serum IgE and IgG1. Analysis of transferred transgenic OVA-specific naive T cells further showed that mTi acted as an adjuvant to drive Ag-specific Th2 cell differentiation in vivo. Assessment of the innate response indicated that mTi induced rapid recruitment and differentiation of alternatively activated macrophages in vivo, through IL-4– and TLR4-independent pathways. These studies suggest that solid microparticles alone can act as adjuvants to induce potent innate and adaptive Th2-type immune responses and further suggest that wear debris in joint replacements may have Th2-type inflammatory properties.

and Dentistry of New Jersey Research Animal Facility. All of the mice were maintained in a specific pathogen-free, virus-free facility during the experiments. The studies have been reviewed and approved by Institutional Animal Care and Use Committee at New Jersey Medical School. The studies conformed to the principle for laboratory animal research outlined by the Animal Welfare Act and the Department of Health, Education and Welfare (National Institutes of Health) guidelines for the experimental use of animals.

**Administration of Ti particles and Alum/OVA and quantitation of total and OVA-specific IgE, IgG1, and IgG2a**

Mice were inoculated i.p. with either PBS alone, 50 μg endotoxin-free OVA protein (grade V; Sigma-Aldrich, St. Louis, MO) alone, 12.5 mg Ti particles plus 50 μg OVA, or 4 mg Alum plus 50 μg OVA. All groups were subsequently challenged i.p. with OVA alone on day 7. Twenty-one days after the last immunization, sera were collected and total and OVA-specific IgG1, IgG2a, and IgE Abs were determined by ELISA, as previously described (16).

**OVA, Alum, and mTi preparations**

EndoGrade OVA (Hyglos, Regensburg, Germany), Injekt Alum (Thermo Scientific, Rockford, IL), and mTi (<20 μm; Alfa Aesar, Ward Hill, MA) were used in all experiments described. Ti particles were washed three times with absolute alcohol followed by three washes with sterile water and dried at 65°C under vacuum with a vacuum concentrator (Eppendorf Vacufuge) and redissolved in sterile PBS. In additional experiments, the particles were further treated to remove residual endotoxin either by 1) treating with 1% acetic acid and boiling for 3 h, or 2) autoclaving for 1 h and incubating in a heated oven at 175°C for 3 h, as previously described (17). Using an endoplasmic chromomeric Limulus amebocyte lysate assay (QCL-1000; Lonza, Walkersville, MD), the LPS levels of Ti particles treated with either method were found to be less than or equal to the negative control (≤0.01 endotoxin units/ml), whereas Ti particles treated with alcohol alone had detectable levels of LPS at ∼0.38 endotoxin units/ml. Ti particles were further characterized by low-angle laser light scattering using previously described methods (18–20). The average size of the Ti particles plus 50 μg OVA were further characterized by low-angle laser light scattering using previously described methods (16).

**IgG1, IgG2a, and IgE Abs were determined by ELISA, as previously described**

After the last immunization, sera were collected and total and OVA-specific IgG1, IgG2a, and IgE Abs were determined by ELISA, as previously described (16).

**Adoptive transfers of D011.10 TCR CD4+ T cells**

Peripheral lymph nodes and spleen were harvested from wild-type (WT) D011.10 TCR-transgenic mice, which express a TCR specific for chicken OVA peptide 323–339-α-ad complexes and which are uniquely recognized by the K(1–26) anti-IgM mAb (21). Single-cell suspensions were prepared, and D011.10 TCR transgenic CD4+ T cells were isolated, CFSE labeled, and adoptively transferred to age- and sex-matched BALB/c mice i.v., as previously described (22). OVA peptide alone (30 μg) or with either 12.5 mg Ti particles or with 4 mg Alum was injected i.p. into recipient mice 2 d after transfer. HPLC-purified OVA323–339 (OVA peptide) with the sequence ISQA VHAAHAEINEAGR-COOH was synthesized at the University of Medicine and Dentistry of New Jersey Core Facility.

**Flow cytometry**

Peritoneal exudate cell suspensions were blocked with Fe Block (BD Pharmingen, San Jose, CA) and then stained with various Abs, including anti-LY6G-FITC, anti-CD11c-PE, anti-CD11b-PerCP-C55.5, Siglec-F-PE, anti–c-Kit-allophycocyanin, F4/80-allophycocyanin (BD Pharmingen), and CD206-FITC (AbD Serotec, Raleigh, NC). For CFSE-labeled cells, anti-CD4-PerCP (BD Pharmingen, San Jose, CA) and KJ1-26 mAb (BD Pharmingen, CA) were used to phenotype DO11.10 T cells. Cell cycle progression was monitored by measuring sequential reductions in CFSE fluorescence of KJ1-26+ CD4+ cells, and the proliferation index was calculated using FlowJo software (Tree Star, Ashland, OR).

**ELISPOT**

Single-cell lymph node suspensions were prepared in RPMI 1640 containing 10% heat-inactivated FCS, 100 U/ml penicillin, 100 μg/ml streptomycin, and 2 mM l-glutamine (all from Invitrogen). Lymph node cells (0.5 × 10^6) were cultured with 10 μg/ml OVA peptide for 3 d on anti–IL-4 (clone BV4D-1D11.2; BD Pharmingen, San Jose, CA)-coated plates/anti–IFN-γ (BD Biosciences)-coated plates. ELISPOT was developed as previously described (16). After 72 h the plates were washed several times with PBS followed by PBS with 0.5% Tween 20 (PBST). Secondary biotinylated anti–IL-4 Ab/anti–IFN-γ Ab (BD Biosciences) was diluted in PBST plus 5% FCS, added at 100 μl/well, and incubated overnight at 4°C. Plates were then washed, and a 1:2000 dilution of streptavidin/alkaline phosphatase (Jackson ImmunoResearch Laboratories) was added. Plates were developed and results quantitated by counting the number of positive colonies under a microscope.

**Sorting of CD4+ T cells or DO11.10 T cells after immunization**

For CD4+ T cell sorting, lymph node cell suspensions were prepared from D011.10-inoculated mice, incubated with anti-CD4 microbeads (Miltenyi Biotec), and CD4+ T cells were purified by positive selection. The purities were >98%. For cell sorting of OVA-specific DO11.10 T cells, mesenteric lymph node (MLN) cells were stained with anti-mouse D011.10 TCR biotin conjugate Ab (Caltag Laboratories) and then labeled with anti-biotin Ab beads (Miltenyi Biotec). Labeled cells were passed through MS columns (Miltenyi Biotec) according to the protocol provided by the manufacturer. The KJ1-26+ population was positively selected and assessed for purity using FACS analysis. The purities were 85–90% in all sorts. RNAs were extracted from sorted cells using RNazol (Intronigene).

**Cytokine gene expression by RT-PCR**

For RT-PCR, total RNA was extracted from peritoneal exudate cells by using RNazol (Intronigene) and then reverse transcribed with SuperScript II reverse transcriptase (Intronigene) as per the manufacturer's protocol. Gene-specific TaqMan assays along with the TaqMan Gene Expression Master mix (Applied Biosystems) were used for amplification and detection of different genes in an Applied Biosystems 7300 sequence detector. Gene expression fold changes of different mRNA of treated samples were quantified relative to the untreated samples by employing the 2^(-∆∆CT) relative quantification method using 18S RNA as the housekeeping gene, as previously described (23).

**Statistical analysis**

Data are presented as means ± SEM, indicated by error bars. Statistical significance was determined by ANOVA followed by individual comparisons with a Tukey test using SigmaPlot version 11.0 (Systat Software, San Jose, CA). Statistical difference at a level of p < 0.05 between groups was considered significant.

**Results**

Ti particles can enhance elevations of serum Igs in response to OVA protein immunization

We first investigated the effect of mTi, ranging from 0.2 to 88 μm, on the development of the adaptive in vivo humoral immune response. BALB/c female mice were initially inoculated i.p. with 12.5 mg OVA and 50 μg OVA protein alone. Seven days after primary inoculation, all treatment groups were inoculated with OVA protein (50 μg) alone, and 21 d after secondary inoculation serum was collected and assayed for Igs. Alum, a well-studied adjuvant that can promote a potent Ag-specific Th2-type immune response (24, 25), was included for comparison. As shown in Fig. 1, administration of Alum or Ti in combination with OVA markedly enhanced serum elevations of total IgE and IgG1 compared with OVA alone, with negligible effects on IgG2a. Ag-specific serum Ig levels were also examined using an OVA-specific ELISA assay. As also shown in Fig. 1, Ag-specific serum Ig levels were also elevated in mice inoculated with OVA plus Ti compared with mice inoculated with OVA alone. Both total and Ag-specific serum Ig elevations were comparable in groups given either mTi or Alum in combination with OVA, suggesting that mTi can function under these conditions as a potent adjuvant.

Ti can function as an effective adjuvant to promote naive Ag-specific Th2 cell differentiation

To examine whether mTi can function as an adjuvant to promote naive T cell expansion and differentiation into effector T helper cells, CFSE-labeled transgenic OVA-specific DO11.10 CD4+ cells were adoptively transferred to age- and sex-matched recipient BALB/c mice. Two days later, recipient mice were inoculated i.p. as determined using QIAVHAHAAHAEINEAGR-COOH was synthesized at the University of Medicine and Dentistry of New Jersey Core Facility.
experiments were repeated twice with similar results. **SEM of eight individual mice within each treatment group, and all

C

used to determine the number of IL-4– and IFN-

B

administered OV A plus mTi compared with mice administered

nounced cell cycle progression of OV A-specific T cells in mice

dicating that mTi as well as Alum could promote Ag-specific

IgG1, and IgG2a (Fig. 1). In additional experiments mice were immunized with mTi plus OV A, and 21 d after final inoculation serum was collected from each mouse and individually assayed for total and Ag-specific Igs. Both

mTi administered with OVA Ag promotes serum IgE and

IgG1 elevations. Mice (eight per treatment group) were administered OVA protein alone, OVA plus mTi, or OVA plus Alum and 7 d later challenged with OVA alone, and 21 d after final inoculation serum was collected from each mouse and individually assayed for total and Ag-specific Igs. Both total serum IgE and IgG1 were markedly elevated, whereas slight increases in total serum IgG2a (A) were observed. Similarly, OVA-specific IgE, IgG1, and IgG2a (B) were elevated. Data are expressed as the means ± SEM of eight individual mice within each treatment group, and all experiments were repeated twice with similar results. **p < 0.01.

with OVA peptide (30 μg), OVA peptide (30 μg) plus mTi (12.5 mg), or OVA peptide (30 μg) plus Alum (4 μg), and 7 d after inoculation MLNs were collected and cell suspensions analyzed by flow cytometric analysis or OVA-specific ELISPOT. As shown in Fig. 2A, the percentage of OVA-specific T cells (KJ-126+) was increased in mice inoculated with either OVA plus mTi or OVA plus Alum compared with mice inoculated with OVA alone, indicating that mTi as well as Alum could promote Ag-specific T cell expansion in vivo. CFSE analysis further showed pronounced cell cycle progression of OVA-specific T cells in mice administered OVA plus mTi compared with mice administered OVA alone (Fig. 2B). An OVA-specific ELISPOT assay (23) was used to determine the number of IL-4– and IFN-γ–secreting cells following in vitro restimulation with OVA. As depicted in Fig. 2C, immunization with OVA plus mTi or OVA plus Alum resulted in pronounced increases in IL-4, but not IFN-γ. Sorted KJ-126+ T cells showed pronounced IL-4, IL-10, and IL-13 gene expression in mice inoculated with OVA plus Ti or OVA plus Alum. Taken together, these studies indicate that mTi is an effective adjuvant that can promote naïve T cell differentiation to proliferating and cytokine-producing Ag-specific Th2 cells.

**FIGURE 1.** mTi administered with OVA Ag promotes serum IgE and IgG1 elevations. Mice (eight per treatment group) were administered OVA protein alone, OVA plus mTi, or OVA plus Alum and 7 d later challenged with OVA alone, and 21 d after final inoculation serum was collected from each mouse and individually assayed for total and Ag-specific Igs. Both total serum IgE and IgG1 were markedly elevated, whereas slight increases in total serum IgG2a (A) were observed. Similarly, OVA-specific IgE, IgG1, and IgG2a (B) were elevated. Data are expressed as the means ± SEM of eight individual mice within each treatment group, and all experiments were repeated twice with similar results. **p < 0.01.

**Ti induces a robust innate response characterized by M2 cell differentiation**

mTi may act as an adjuvant to stimulate Th2 cell differentiation by triggering an appropriate innate immune response or alternatively by enhancing antigenicity either through a depot effect, thereby increasing the time of Ag release, or by facilitating Ag uptake (6). Previous studies have suggested that mTi preparations similar to those used in this investigation are inert in biological systems (26). To investigate whether Ti may be triggering activation of innate immune cell populations characteristic of a Th2-type innate immune response, mice were inoculated i.p. with Ti alone, and 48 h later immune cell populations in the peritoneum were phenotyped using flow cytometric analysis. Additionally, IL-4−/− BALB/c mice were inoculated with Ti to examine whether early production of IL-4 might be contributing to the development of a Ti-induced innate response. As shown in Fig. 3A, pronounced increases in neutrophils (Ly6G+, CD11b+) were observed following Ti inoculation in both WT and IL-4−/− mice, whereas eosinophils (Siglec-F+, c-Kit+) were increased >10-fold following Ti inoculation, consistent with a Th2-type innate response phenotype (Fig. 3B). Eosinophil increases were blocked in IL-4−/− mice, indicating that at this early time point after Ti inoculation IL-4 functions to promote eosinophil recruitment and expansion in the peritoneal cavity.

During a Th2-type innate immune response to helminths, macrophages characteristically differentiate to alternatively activated (M2) macrophages (27). To determine whether M2 macrophages were increased in Ti-inoculated mice, cell suspensions from the peritoneal cavity were dual stained for F480 and CD206, a characteristic M2 macrophage phenotype. As shown in Fig. 3C, the frequency of M2 macrophages was markedly increased in mTi-inoculated WT mice compared with untreated controls. Intriguingly, increased M2 macrophages were also observed in Ti-inoculated IL-4−/− mice, indicating that mTi induced M2 macrophage differentiation without IL-4 signaling. Gene expression analysis of characteristic M2 macrophage markers showed pronounced increases in RELM-α, Ym-1, and Arg-1 in peritoneal cells of mTi-inoculated mice compared with untreated controls; however, inducible NO synthase, an M1 macrophage marker, was elevated to a lesser degree (Fig. 4). Furthermore, these M2 markers were similarly increased in Ti-inoculated IL-4−/− mice.

Cytokines and notch ligands typically elevated during Th1-type or Th2-type responses were also examined at 48 h after mTi inoculation. As shown in Fig. 5, IL-4, IL-5, and IL-13 were elevated in mTi-inoculated WT mice but blocked in Ti-inoculated IL-4−/− mice, whereas Jagged-1, IL-25, IL-6, and IL-33 were increased in both Ti-inoculated WT and IL-4−/− mice. In contrast, Delta-4, IL-12, and IFN-γ showed at most modest elevations. Alum, a well-known Th2 adjuvant, was used for comparison; in most cases mTi showed a comparable ability to activate the Th2-type innate immune response.

**Adjuvant functions of Ti are TLR4 independent**

The presence of LPS, which signals through TLR4, is ubiquitous and can sometimes confound results during adjuvant studies. Although we used commercially available endotoxin-free mTi, previous findings have shown that under some circumstances even low levels of LPS can trigger a Th2-type immune response through TLR4 signaling (10), raising the possibility that some of the adjuvant functions observed with mTi may be due to associated LPS. As described in Materials and Methods, two previously published methods were used to effectively remove residual LPS from the mTi. Treatment with either method triggered a similar activation of innate cell populations, including neutrophils, eosinophils, and M2 macrophages, as compared with untreated mTi (Supplemental Fig. 1). In additional experiments mice were immunized with treated mTi plus OVA, and 21 d later marked increases in serum IgE and IgG1 were detected (Supplemental Fig. 2).

To further exclude the possibility that LPS-induced TLR4 signaling was contributing to the adjuvant function of mTi, TLR4-sufficient (C3H/OuJ) and TLR4-deficient (C3H/HeJ) mice were inoculated with mTi plus OVA or OVA alone and 7 d later given a secondary inoculation with OVA alone. In both mTi-inoculated TLR4-deficient and TLR4-sufficient mice, total serum IgE and IgG1, but not IgG2a, were markedly elevated 21 d after inoculation (Fig. 6).

The innate immune response was also examined in TLR4-deficient mice at 48 h after inoculation. As shown in Fig. 7, after

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Ti inoculation, pronounced and similar increases in neutrophils, eosinophils, and M2 macrophages were observed in TLR4-deficient and TLR4-sufficient mice. In both strains, the phenotype was similar to that observed in Ti-inoculated BALB/c mice, indicating that this response is sufficiently robust to be reproducible in different background murine strains. We also examined the

![Figure 3](http://www.jimmunol.org/)
innate immune response in Ti-inoculated BL/6 MYD-88−/− mice, which lack this important TLR signaling pathway, and found similar increases in neutrophils, eosinophils, and M2 macrophages (data not shown). Taken together, these results indicate that potential low-dose LPS contamination is not playing an important role in affecting the innate or adaptive Th2-type immune response following mTi inoculation.

**Discussion**

Our findings indicate that a micrometer-sized Ti particle can act as an adjuvant to promote naive Ag-specific CD4 T cells to expand and differentiate into cytokine-producing Th2 effector cells in vivo. Our findings further show that rather than simply acting as a depot or through general enhancement of Ag presentation, mTi elicit a potent innate Th2-type immune response associated with IL-4− independent M2 macrophage activation. This effect has implications for Ti implants, suggesting that associated wear debris may contribute to harmful Th2-type inflammation.

Previous studies have suggested that wear debris, present in periprosthetic tissues and ranging in size from 0.5 to 100 μm, may contribute to osteolysis and aseptic loosening of total joint prostheses (2, 28, 29). We demonstrated that the commercial mTi preparations used had a very similar size range, from 0.2 to 88 μm. In vitro cultures with similar preparations of mTi can stimulate macrophages to secrete inflammatory cytokines, raising the possibility that mTi may also trigger an inflammatory response in vivo. However, such inflammatory responses may in many cases result from endotoxin adhering to the mTi, as mTi with adherent LPS preferentially induce inflammatory cytokines, including IL-1, IL-6, and TNF-α (3, 5). Our studies indicate that the very different Th2-type immune response induced by mTi in vivo is LPS-independent, as Ti-inoculated TLR4-deficient mice supported potent innate and adaptive Th2-type responses, and as effective removal of endotoxin did not impair induction of the innate or adaptive Th2-type responses.

Our observation that total and Ag-specific serum IgG1 and IgE were elevated following administration of mTi with specific Ag
FIGURE 6. TLR4-deficient mice administered OVA plus Ti show elevations in serum IgG1 and IgE. TLR4-deficient C3H/HeJ and control C3H/HeOuJ mice were administered OVA protein alone or OVA plus Ti and 7 d later challenged with OVA alone, and 21 d after final inoculation serum was collected from each mouse and individually assayed for total and Ag-specific Igs. Both total serum IgE (A) and IgG1 (B) were markedly elevated, whereas slight increases in total serum Ig2a (C) were observed. Similarly, OVA-specific IgE (D), IgG1 (E), and IgG2a (F) were elevated. Data are expressed as the means ± SEM of five individual mice within each treatment group treated compared with the untreated (OVA) group. This experiment was repeated twice with similar results. **p < 0.01.

suggested that mTi could act as an adjuvant to drive a potent humoral response and raised the possibility that mTi could promote Th2 cell differentiation. Previous studies have shown that Ti dioxide (TiO2) nano-particles can enhance allergy-associated inflammation in the lung; however, the small nanoparticle size along with associated charge have been thought to be important characteristics in driving this response (6). Our studies indicate that micron sized particles can also be potent adjuvants. This is particularly relevant to inflammation of joint replacements as wear debris that deposits and accumulates in surrounding tissue is primarily in the micron size range (28, 29). Through adoptive transfer of CFSE-labeled OVA-specific DO11.10 T cells, we were able to demonstrate that mTi increased cell cycle progression and expansion of Ag-specific T cells and promoted their differentiation into IL-4 secreting Th2 cells in vivo. In the same experiment the adjuvant effect of mTi on Th2 cell differentiation was similar to that observed with Alum. Alum is a well-documented potent Th2 adjuvant (12, 30) and the only vaccine adjuvant currently approved by FDA (31). Our inclusion of Alum in the described experiments allowed us to examine the relative strength of mTi as an adjuvant. Our findings indicate that mTi is as potent as Alum in driving both Ag-specific Th2 cell expansion and differentiation in vivo and also in promoting serum IgG1 and IgE elevations.

The mechanisms through which adjuvants drive Th2-type responses remain unclear. Traditionally, Alum was thought to work through a “depot” effect stabilizing adherent Ag and thereby prolonging exposure of the Ag to the immune system (32, 33). Alternatively, Alum has also been shown to trigger a strong innate immune response resulting in activation of the nod-like receptor protein 3 (NLRP3) inflammasome, resulting in caspase-1-mediated conversion of members of the IL-1 family to active cytokines (34, 35). More recent studies, however, suggest that blockade of this innate inflammasome response does not affect Alum-mediated stimulation of the Th2-type immune response (30, 36–39). Most recently PGE2 has been implicated in promoting IgE production (39) while other studies suggest Alum may affect Ag presentation by directly perturbing dendritic cell membranes (40).

Our studies indicate that Alum and mTi both trigger a potent Th2-type innate response previously associated primarily with helminth infection. Intriguingly, components of this innate response varied in their IL-4 dependence, as eosinophil recruitment was blocked in response to both Alum and Ti particles in IL-4–deficient mice, whereas M2 cell activation remained intact. The IL-4–independent increases in IL-33, IL-25, and Jagged-1, all of which are associated with initiation of Th2-type responses (41, 42), indicate a robust Th2-type innate response even in the absence of IL-4 signaling. The overall innate Th2-type responses induced by both Alum and Ti particles were remarkably similar, suggesting the involvement of similar signaling pathways. M2 cells have been shown to develop in response to a number of stimuli, including IL-4, adenosine, CSF, IL-10, and helminths (27, 43–46). Previous studies have shown macrophage infiltration (47, 48) at the periprosthetic site of osteolysis. Intriguingly, recent studies have also shown increased gene expression of M2 markers in synovial-like tissue in late-stage osteolysis patients and also elevations in chemokines potentially involved in recruitment of osteoclast precursor cells, whereas elevations in TNF-α and other Th1-type inflammatory markers were not detected (49). Note that these studies with patients examined sites of chronic inflammation, whereas our studies examined the acute response that may include infiltration by cell populations, such as neutrophils, that may only transiently infiltrate the site of osteolysis. Also, our study likely resulted in a more systemic response, including increases in serum Igs, compared with the more local response associated with peri-implant inflammation.

Helminths can induce Th2 cell differentiation in the absence of IL-4, although subsequently IL-4 is required for Th2 cell expansion (23, 50, 51). Our findings with Ti and Alum suggest that at least M2 cell differentiation is IL-4–independent.
products that trigger Th2-type responses include both glycans and phosphorylcholine-containing proteins (e.g., ES62), both of which can induce Th2-type responses through TLR4-dependent pathways (7, 52, 53). Our results show that mTi, as well as Alum, are TLR4-independent. Interestingly, the Th2-type response induced by many live helminths is also TLR4-independent (7, 14, 15), suggesting that both mTi, Alum, and helminths can all induce polarized Th2 cell differentiation in vivo in the absence of TLR4 signaling. Cell interactions associated with relatively large solid structures in tissue, such as frustrated phagocytosis or cell and tissue damage (8, 40, 54), may be an important mechanism and common pathway through which an innate response is stimulated that ultimately contributes to Th2 cell differentiation. It will thus be of interest to examine whether a Th2-type response is associated with other implant materials, particularly since inflammatory infiltrates are observed with non-metal-on-metal as well as metal-on-metal implants (55).

These studies thus indicate that mTi can act as potent adjuvants to stimulate an innate Th2-type in vivo immune response, characterized by M2 cell activation, and subsequently the development of Ag-specific Th2 cells and associated serum IgE and IgG1 elevations. Their similarity to wear debris in prosthetic implants of Ag-specific Th2 cells and associated serum IgE and IgG1 characterized by M2 cell activation, and subsequently the development of chronic intestinal nematode infection. Eur. J. Immunol. 33: 2797–2807.


