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Mast Cells and Neutrophils Release IL-17 through Extracellular Trap Formation in Psoriasis

Andrew M. Lin,* Cory J. Rubin,* Ritika Khandpur,* Jennifer Y. Wang,* MaryBeth Riblett,* Srilakshmi Yalavarthi,† Eneida C. Villanueva,† Parth Shah,* Mariana J. Kaplan,† and Allen T. Bruce*

IL-17 and IL-23 are known to be absolutely central to psoriasis pathogenesis because drugs targeting either cytokine are highly effective treatments for this disease. The efficacy of these drugs has been attributed to blocking the function of IL-17–producing T cells and their IL-23–induced expansion. However, we demonstrate that mast cells and neutrophils, not T cells, are the predominant cell types that contain IL-17 in human skin. IL-17+ mast cells and neutrophils are found at higher densities than IL-17+ T cells in psoriasis lesions and frequently release IL-17 in the process of forming specialized structures called extracellular traps. Furthermore, we find that IL-23 and IL-1β can induce mast cell extracellular trap formation and degranulation of human mast cells. Release of IL-17 from innate immune cells may be central to the pathogenesis of psoriasis, representing a fundamental mechanism by which the IL-23–IL-17 axis mediates host defense and autoimmunity. The Journal of Immunology, 2011, 187:490–500.

Although IL-17 production by T cells is widely studied, it is increasingly appreciated that diverse types of innate immune cells also can produce IL-17 (17). Recent studies demonstrate that IL-17+ mast cells and neutrophils can be found in complicated atherosclerotic plaques (18). Similarly, IL-17+ mast cells are also present in inflamed synovium of rheumatoid arthritis (19). Mast cells, particularly a subset containing tryptase and chymase (MC1C5), are enriched in the papillary dermis of psoriasis lesions (20, 21). Mast cells frequently degranulate in early eruptive and recurring psoriasis lesions and have been described as “ghost cells” (22–24). Interestingly, mast cell numbers are decreased in psoriasis lesions after successful treatment with anthralin, psoralen plus UVA light therapy, or cyclosporine (25–27). Neutrophils also are enriched in psoriasis lesions, especially in the epidermis where they aggregate in Munro’s microabscesses (MMs) in the stratum corneum and spongiform pustules of Kogoj (28). Although the precise function of neutrophils is unknown in psoriasis, a critical pathogenic role is supported by case reports of psoriasis remission during drug-induced agranulocytosis and its reappearance after the normalization of neutrophil numbers (29). Furthermore, ruxolizumab, a drug effective against all forms of psoriasis and psoriatic arthritis, causes a dose-dependent depression of neutrophil counts (30).

Very recently, a study indicated that mast cells and neutrophils contain IL-17 in psoriasis plaques of three patients (31). However, the significance of these numbers relative to normal-appearing skin and the mechanism of IL-17 release by innate immune cells in psoriasis remain unknown.

IL-17 orchestrates innate immune responses against extracellular pathogens by inducing expression of antimicrobial peptides (AMPs) and neutrophil-tropic chemokines CXCL1, CXCL2, and IL-8 (32–35). The same AMPs and chemokines are found at extremely high levels in psoriatic epidermis (36–38). Not surprisingly, mice and humans with deficiencies in IL-17 production or signaling are highly susceptible to infection with extracellular bacteria and fungi (39–42). Similarly, effective antimicrobial activity by neutrophils and mast cells depends on the formation of structures called extracellular traps (ETs), termed NETs and...
MCETs, respectively (43–45). ETs are formed through a specialized process of cell death termed ETosis (46), where chromatin extends into fine, weblek threads to which proteins are bound. In particular, NETs can contain myeloperoxidase (MPO), proteinase 3, and AMPs such as cathelicidin (LL-37) (47). The process of NET formation (NETosis) can be triggered by extracellular bacteria and fungi or their components (48, 49). MCETs contain tryptase, IL-37, and chromatin (45), forming in response to bacteria, H2O2, or PMA (45, 48). In humans, NETs have been visualized in physiologic host responses to infections (44, 50) and have been implicated in the pathology of anti-neutrophil cytoplasmic Ab-induced vasculitis (51). Additionally, a recent study showed that lupus nephritis is associated with an inability to degrade NETs in blood (52). Furthermore, although MCETs have been studied in a human mast cell line and mouse bone marrow-derived mast cells (45), it is unclear whether the process of MCET formation (MCETosis) occurs in human tissue under normal or pathologic conditions.

To understand the complex pathophysiology of psoriasis, it is imperative to define the precise cellular sources of IL-17 and the mechanisms mediating IL-17 release. Therefore, we investigate the production of IL-17 by innate immune cells in psoriasis and explore a potential role for ETs formed by these cells in human skin. Surprisingly, we observe that most IL-17+ cells in normal and psoriatic skin are mast cells, not T cells. Additionally, neutrophils in well-developed psoriasis lesions also express IL-17. We observe that mast cells and neutrophils release IL-17 into the skin though ETosis as well as conventional degranulation. Furthermore, a subtype of human neutrophils, low-density granulocytes (LDGs), isolated from psoriatic blood are increased in psoriatic compared to control human skin explants treated with IL-23 and IL-1b, and have been implicated in the pathology of anti-neutrophil cytoplasmic Ab-induced vasculitis (51). Additionally, a recent study showed that lupus nephritis is associated with an inability to degrade NETs in blood (52). Furthermore, although MCETs have been studied in a human mast cell line and mouse bone marrow-derived mast cells (45), it is unclear whether the process of MCET formation (MCETosis) occurs in human tissue under normal or pathologic conditions.

Materials and Methods

Human subjects

All of the subjects provided written, informed consent. Skin punch biopsies (6 μm) were obtained from patients with normal healthy skin (NN), normal-looking symptomless psoriatic skin (PN), and psoriasis plaques (PP). Before biopsy, subjects were required to be off all systemic therapies for at least 2 wk and off topical antipsoriatic medications for at least 1 wk. Control donors were identified from respondents to advertisements in the area (Ann Arbor, MI), had no personal or family history of psoriasis, and were free of inflammatory skin disease at the time of biopsy. This study was approved by the Institutional Review Board of the University of Michigan Medical School.

Immunofluorescence

Five-micrometer sections of skin were deparaffinized on a hot plate at 65°C for 1 h. They were rehydrated by incubation and heat retrieval in CCI1 cell conditioning solution (Ventana Medical Systems, Tucson, AZ). The sections were blocked with horse serum (0.2%; Invitrogen, Carlsbad, CA) for 30 min at room temperature. Then simultaneous staining first was performed for 30 min at room temperature with goat anti-IL-17 (100 μg/ml; R&D Systems, Minneapolis, MN) and another primary Ab, either rabbit anti-CD3 (0.4 μg/ml; Ventana Medical Systems), rabbit anti-CD4 (1:25; Cell Marque, Rocklin, CA), mouse anti-CD8 (1:25; Leica Biosystems, Newcastle, U.K.), mouse anti-mast cell tryptase (1:150; Dako, Carpinteria, CA), mouse anti-mast cell chymase (1:1000; ABD Serotec, Oxford, U.K.), or rabbit anti-MPO (1:1500; Dako). Triple staining was conducted with IL-17, MPO, and mouse anti-human IL-37 Abs (1:200; Abcam, Cambridge, MA). This was followed by 30 min of incubation at room temperature with matched secondary Abs: chicken anti-goat IgG Alexa Fluor 488 (1:300; Invitrogen), chicken anti-mouse IgG Alexa Fluor 594 (1:300; Invitrogen), or chicken anti-rabbit IgG Alexa Fluor 594 (1:300; Invitrogen). For triple staining, donkey anti-rabbit Alexa Fluor 350 (1:300; Invitrogen) also was used. ProLong Gold antifade reagent with DAPI was added to the slides prior to mounting with coverslips (Invitrogen). Images were captured with a fluorescence microscope (BX50; Olympus, Essex, U.K.) using DP Controller and DP Manager (Olympus) software and a confocal microscopic (FV-500; Olympus).

Cell quantification from immunofluorescence

The number of cells expressing one or both markers of interest in each ×200 field was counted manually by two independent blinded observers on Adobe Photoshop. From these numbers, we calculated the proportions of cells of different populations [e.g., the proportion of IL-17+ of all tryptase+ cells is calculated as (number of IL-17+tryptase+cells)/(number of IL-17+ tryptase+ cells + IL-17+ tryptase+ cells)]. Each data point in Figs. 2 and 5 represents the average of one to six independent fields for each patient sample.

Skin explant cultures

Punch biopsies were obtained from healthy donors without psoriasis as above and were quartered longitudinally. Each divided section was incubated in R10 media (RPMM 1640 [Life Technologies] with 0.4 mM Ca2+ and 10% FCS) with soy bean trypsin inhibitor (100 μg/ml; Sigma-Aldrich) or R10 media with soy bean trypsin inhibitor containing either Substance P (10 μM; Sigma-Aldrich), Compound 48/80 (10 μg/ml; Sigma-Aldrich), IFN-γ (200 ng/ml; Miltenyi Biotec), IL-1β (2.5 ng/ml; R&D Systems), IL-23 (10 ng/ml; R&D Systems), or both IL-1β and IL-23 together for 2 or 3 d. The tissues were fixed in 10% neutral buffered formalin and subjected to immunofluorescence staining as above. The proportions of intact mast cells and degranulated mast cells were counted by direct visualization through the microscope by two independent observers.

Isolation of neutrophils and LDGs

Normal-density neutrophils and LDGs were as described (53). The purity of the neutrophil fraction was at least 95% and was determined by CD15, CD14, CD10, and negative CD3 expression by flow cytometry. In some experiments, neutrophils were sorted after dextran gradient isolation to >95.5% purity using FACSARia (BD Biosciences, San Jose, CA) gating on CD15+CD3− cells with neutrophil size and granularity characteristics as in Fig. 4A. For LDGs, PBMCs were isolated using Ficoll-Hypaque. RBCs were lysed using hypotonic/hypertonic saline, and then T and B lymphocytes, NK cells, monocytes, and residual RBCs were removed by negative selection. LDGs were identified as CD15+CD14+ or CD10+CD14+ (CD10+CD14+).

Western blot analysis

Total cell lysates were obtained by Laemmli extraction. A total of 10 μg of each sample and 40 ng of the recombinant cytokines were loaded. Samples were separated on 15% SDSPolyacrylamide gels and transferred to Immobilon-P membranes (Millipore, Billerica, MA). Western blots were blocked with PBS/0.2% Tween 20 for 1 h at RT, washed twice, and incubated with goat anti-human IL-17A Ab (1:250; R&D Systems) or mouse anti-human IL-17A Ab (1:1000; eBioscience, San Diego, CA) overnight at 4°C. The blots were washed three times with PBS/0.02% Tween 20 and then incubated with horseradish peroxidase-conjugated mouse secondary Ab (1:2500; GE Healthcare, Piscataway, NJ) for 1 h at room temperature. Blots were developed using ECL (Thermo Scientific, Rockford, IL) and X-Omat film (Kodak, Rochester, NY). Anti-β-actin (1:2500; Sigma-Aldrich) was used as a loading control.

NET immunofluorescence staining and quantification

A total of 1× 10⁶ neutrophils or LDGs per milliliter were seeded in poly-l-lysine–coated coverslips and incubated at 37°C, 5% CO2 for 15 min. Cells were washed with ice-cold PBS, fixed with cold paraformaldehyde for 15 min, and blocked with 10% FBS, 1% BSA, 0.05% Tween 20, and 2 mM EDTA in 1× PBS at 4°C overnight. Stimulated cells were incubated for 2 or 3 h in 1× RPMI 1640 with glutamine and 2% BSA with 20 nM PMA (Sigma-Aldrich). For detection of NETs, fixed cells were washed with ice-cold 10% FBS in 1× PBS for 5 min, incubated with primary Abs for neutrophil elastase (Abcam) or rabbit isotype control (Abcam) for 45 min at 4°C, washed twice, and incubated with secondary Abs for 45 min at 4°C. Nuclear material was detected by incubating cells with Hoechst 33342 (Invitrogen) for 10 min at room temperature. Coverslips were placed in slides with ProLong Gold antifade
extracellular traps and IL-17 in mast cells and neutrophils

portions of IL-17 + T cells of all CD3 + T cells were calculated to and NN skin (25 ± 7 versus 16 ± 4 versus 7 ± 4 cpf; \( p_{PP:PN} < 0.01, p_{PP:NN} < 0.001, p_{PN:NN} < 0.01 \); Fig. 2J). Importantly, the majority of IL-17 + cells were mast cells, regardless of disease status. Chymase + mast cells accounted for nearly all of the IL-17 + cells, in contrast to CD3 + T cells in PP (89.0 ± 10.6 versus 7.3 ± 7.2%; \( p = 5.9 \times 10^{-15} \)), PN (96.3 ± 3.6 versus 7.3 ± 7.1%; \( p = 1.5 \times 10^{-14} \)), and NN skin (92.2 ± 12.7 versus 5.0 ± 5.0%; \( p = 1.4 \times 10^{-11} \); Fig. 2C, 2K). Additionally, the proportions of chymase + IL-17 + cells of all chymase + cells were also significantly greater in PP and PN skin compared with that in NN skin (94.7 ± 5.5 versus 95.5 ± 5.0 versus 66.6 ± 24.5%; \( p_{PP:NN} < 0.001, p_{PN:NN} < 0.001 \); Fig. 2L). Experiments with tryptase staining revealed similar findings (Fig. 2E-H). These data demonstrate that mast cells, primarily MC_TcS, represent the majority of IL-17-containing cells in human skin. Furthermore, in psoriasis lesions, the density of MC_TcS is increased, and a greater proportion of these cells contain IL-17.

Neutrophils containing IL-17 also are enriched in psoriasis lesions

We noticed that many CD3 + IL-17 + cells in the epidermis of psoriasis plaques had trilobed nuclear morphology and were enriched in areas resembling SPKs and MMs. Dual-color immunofluorescence studies of IL-17 and MPO on PP skin revealed that most of these IL-17 + cells in the stratum spinosum and stratum corneum of well-developed psoriasis lesions contained with MPO, verifying their identity as neutrophils (Fig. 1D, Supplemen tal Fig. 4). Quantification of PP, PN, and NN sections stained with MPO and IL-17 confirmed the increased number of total neutrophils in PP skin compared with scant numbers in PN or NN skin (56 ± 47 versus 4 ± 4 versus 2 ± 3 cpf; \( p_{PP:PN} < 0.001, p_{PP:NN} < 0.001 \); Fig. 2A). However, the number of CD3 + T cells containing detectable IL-17 in PP lesions compared with those in PN and NN skin was not significantly different (1.1 versus 0.8 versus 0.5 cpf, Fig. 2B). To our surprise, we found that T cells represent only a minority of all IL-17 + cells in PP, PN, and NN skin (7.3 ± 7.2, 7.3 ± 7.1, and 5.0 ± 5.0%; Fig. 2C). The proportions of IL-17 + T cells of all CD3 + cells T cells were calculated to be 3.4 ± 5.8 (PP), 10.1 ± 11.9 (PN), and 4.4 ± 4.9% (NN) (Fig. 2D). This finding is concordant with studies by our group and others defining the proportion of T cells capable of producing IL-17. Majority of IL-17 + cells, especially those cells most intensely staining for IL-17, did not express CD3, suggesting that most cells containing IL-17 in psoriasis lesions and normal skin are not T cells.

Mast cells are the majority of IL-17–containing cells in human control and psoriatic skin

Because most IL-17 + cells were not T cells, we performed dual-color immunofluorescence for IL-17 with markers of innate immune cells. To identify subsets of skin mast cells, we stained for the mast cell-specific enzymes tryp tase and chymase. Most mast cells in human skin express both enzymes and are designated MC_TcS. Containing revealed that most tryptase + cells and chymase + cells contained brightly for IL-17 (Fig. 1B, 1C, Supplementary Figs. 2, 3). We observed that the number of chymase + mast cells significantly increased in PP skin compared with those in PN and NN skin (25 ± 7 versus 16 ± 4 versus 11 ± 3 cpf; \( n_{PP} = 11, n_{PN} = 10, n_{NN} = 10; p_{PP:PN} < 0.01, p_{PP:NN} < 0.001, p_{PN:NN} < 0.05 \); Fig. 2F). The densities of chymase + IL-17 + cells were significantly increased in PP and PN skin compared with that in NN skin (23 ± 7 versus 16 ± 4 versus 7 ± 4 cpf; \( p_{PP:PN} < 0.01, p_{PP:NN} < 0.001, p_{PN:NN} < 0.01 \); Fig. 2J). Neutrophils were enriched in superficial MMs and SPKs, analysis of the cells was by necessity more epidermally focused but also included superficial dermal regions. A significant number of IL-17 + neutrophils was seen in PP lesions but rarely present in PP skin and essentially absent in NN skin (18 ± 18 versus 1 ± 1 versus 0 ± 0 cpf; \( p_{PP:PN} < 0.01, p_{PP:NN} < 0.001 \); Fig. 2N). Neutrophils were present in PP lesions in epidermal clusters, the proportion of neutrophils of all IL-17 + cells in PP skin shows substantial variability but is significantly higher than those in PN or NN skin (32.4 ± 20.6 versus 8.1 ± 5.7 versus 1.3 ± 4.3%; \( p_{PP:PN} < 0.001, p_{PP:NN} < 0.001 \); Fig. 2O). Neutrophils in PP or PN tended to contain more IL-17 compared with those in NN skin (33.8 ± 19.4 versus 50.7 ± 45.9 versus 0.8 ± 2.6%; \( p_{PP:NN} < 0.01 \); Fig. 2P). Together, these data reveal that IL-17 + neutrophils are enriched in psoriasis lesions, primarily in focal epidermal collections.

IL-17 is released during the formation of NETs in psoriasis lesions

At low and medium powers, we observed that the nuclei of IL-17 + neutrophils in MMs frequently displayed the morphology of NETs. Indeed, high-power microscopic examination of dual-color immunofluorescence revealed that IL-17 and MPO were associated frequently with NET-like formations of DNA, visualized with DAPI staining. These NETs were observed frequently in psoriatic lesional epidermis (Fig. 3A), particularly in MMs and SPKs, as demonstrated by the thin extensions of nuclear material coexisting with MPO (Fig. 3B). To confirm the functional relevance of NETs, tricolor immunofluorescence staining showed that dermal NETs frequently contained both IL-17 and LL-37 (Fig. 3C). Further visualization through confocal microscopy confirmed that the morphology of NETs varied from cell to cell even within aggre-
gates (Fig. 3D), suggesting that newly arriving neutrophils may be undergoing NETosis. H&E-stained slides confirmed that not all of the neutrophils in a MM had altered nuclear morphology (Fig. 3E). The NETs with the longest chromatin projections tended to costain with MPO but often not with IL-17. In contrast, NETs with shorter extrusions often stained with IL-17, presumably representing less mature NETs. Because IL-17 is not seen in all of the NETs in PP, we hypothesize that IL-17 does not bind to extruded chromatin nearly as strongly or abundantly as MPO.

To explore the sites of IL-17 production, human neutrophils isolated using the standard dextran protocol were enriched further by flow cytometric sorting to >99.5% purity (Fig. 4A). Lysates of these flow-sorted cells contained IL-17A but not IL-17F protein, as detected using Western blot analysis (Fig. 4B).

**Increased LDGs in psoriatic blood frequently undergo NETosis**

Seminal studies almost three decades ago described a subpopulation of neutrophils in psoriatic blood that are polar in shape and have ruffled cell membranes (54). Similarly, a recent study
revealed a population of LDGs in the blood of patients with systemic lupus erythematosus that share some properties with activated neutrophils (53). We found increased numbers of LDGs in the blood of patients with psoriasis compared with those in control blood (34.6 ± 5.1 versus 11.0 ± 2.0%, *p*, 0.01; Fig. 4C).

We measured NET formation in blood neutrophils from psoriasis or controls and visualized NETosis using stains for neutrophil elastase and DAPI. In four sets from patients and controls, we observed that psoriasis LDGs tended to form NETs without any stimulation, in contrast to control or psoriasis neutrophils. After 2 h of incubation in PMA, control neutrophils, psoriasis neutrophils, or LDGs all formed NETs. However, compared with control neutrophils, psoriasis neutrophils tended to form more NETs after culture without PMA treatment although this was not statistically significant (Fig. 4D). These findings indicate that populations of circulating neutrophils in psoriasis readily undergo NETosis.

**MCets in psoriasis lesions are associated with IL-17 release**

In immunofluorescence studies, we noticed that MC TCs in the papillary dermis of psoriasis plaques were often not intact. On initial inspection, they appeared to be degranulated, which is consistent with scanning electron microscopy studies showing the presence of extracellular material. However, upon closer examination, we observed the presence of IL-17+ mast cells and neutrophils, which are known to be involved in the pathogenesis of psoriasis (31). We quantified the number of single CD3+, tryptase+, chymase+, and MPO+ cells (A, E, I, M) and their means (bars) in each ×200 field were quantified as described in Materials and Methods (each, *n* ≥ 10). The double CD3+IL-17+, tryptase”IL-17”, chymase”IL-17”, and MPO“IL-17” cells (B, F, J, N) and their means (bars) in each ×200 field also were quantified. The proportions of these double-positive cells out of all IL-17+ cells and their means (bars) were calculated (C, G, K, O). In addition, the proportions of double-positive cells of all CD3+, tryptase+, chymase+, and MPO+ cells and their means (bars) were calculated (D, H, L, P). *p < 0.05, **p < 0.01, ***p < 0.001 (one-way ANOVA, Tukey’s post hoc test).
FIGURE 3. Neutrophils frequently undergo ETosis in psoriasis. A, Punch biopsies of skin from subjects with lesional psoriasis plaques (PP, n = 12) were subjected to dual-color immunofluorescence staining for IL-17 (green) and MPO (red), with DAPI counterstain (blue). An epidermally centered field of a mature psoriasis plaque at ×200 magnification reveals IL-17–containing neutrophils in a characteristic MM and SPK. Scale bar, 100 μm. B, NETs magnified past ×1000 were observed to extrude nuclear material (open arrows) and contained both MPO and IL-17. Scale bar, 10 μm. C, PP skin also was subjected to tricolor immunofluorescence staining (n = 3) for IL-17 (green), LL-37 (red), and MPO (blue), revealing many triple-positive NETs in the dermis (white cells, solid arrow) as well as other nonneutrophil IL-17+ cells (green, open arrow). Scale bar, 100 μm. D, NETs in a MM were examined further by confocal microscopy at ×1000 (solid arrows) and approximately ×5000 magnification, showing elaborate distortion of nuclear material. Scale bar, 10 μm. E, H&E-stained sections of a MM at ×200 and past ×1000 magnification also revealed altered nuclear morphology resembling NETs. Scale bars, 100 μm (upper panel), 10 μm (lower panel).
FIGURE 4. Neutrophils and LDGs from the blood of patients with psoriasis exhibit frequent NET formation. A. Neutrophils isolated from human blood were sorted further by FACS Aria with forward light scatter, side scatter of light, CD15, and CD3 gates (red) to >99.5% purity. Flow cytometry plots from one representative sample are shown. B. Western blot analysis of cell lysates from postsort neutrophils with anti-IL-17A and anti-IL-17F Abs was performed to confirm the type of IL-17 contained in control neutrophils (n = 3). Staining for β-actin served as the loading control. C. A higher number of LDGs was present in blood from psoriasis patients (n = 15) compared with that in control blood (n = 6). **p < 0.01 (unpaired two-tailed Student t test). D. Neutrophils and LDGs isolated from psoriatic blood and neutrophils from control blood were stained at time 0, 2 h without stimulation, and 2 h after incubation in PMA. In vitro NETs were examined at ×400 magnification using stains for neutrophil elastase (green) and Hoechst 33342 (blue). Scale bar, 20 μm. A, rIL-17A; F, rIL-17F; γ, rIFN-γ; N, neutrophils; P, PBMCs.
FIGURE 5. IL-1β and IL-23 promote mast cell degranulation or MCET formation. 

A, MCETs observed in PP lesions at ×1000 magnification showed extruded nuclear material (open arrows) and contained both chymase (red) and IL-17 (green). Scale bar, 10 μm. 

B, MCETs were observed further in the dermis of psoriatic skin by zooming in on a confocal image past ×1000 magnification. Scale bar, 25 μm. 

C, Punch biopsies of skin from normal controls (NN) were incubated for 3 d in various conditions: media alone, IFN-γ, Substance P, Compound 48/80, IL-1β, IL-23, and IL-1β + IL-23 as described in Materials and Methods. Tissue sections were subjected to dual-color immunofluorescence staining for IL-17 (green) with chymase (red) or tryptase (data not shown), with DAPI counterstain (n = 3). Representative images from one NN series stained with chymase and IL-17 are shown at ×400 magnification and compared with a typical PP section. Intact cells (solid arrows) and MCETs or degranulated cells (open arrows) are highlighted. Scale bar, 100 μm. 

D, The proportion of MCETs or degranulated tryptase+ cells was observed to increase when exposed to IL-23 alone or IL-1β and IL-23. *p < 0.05 for media versus IL-23 or IL-1β + IL-23 (one-way ANOVA, Dunnett’s test). 

E, Similarly, chymase+ cells also displayed more frequent MCETs and degranulation when exposed to IL-1β and IL-23. **p < 0.01 for media versus IL-23 or IL-1β + IL-23.
degranulated MCs in developing or recurrent psoriasis plaques (22, 23). However, high-power microscopic visualization of the superficial papillary dermis revealed that although many of these MCETs had undergone conventional degranulation, a significant number of cells revealed mast cell tryptase and chymase colocalized with linear DNA (Fig. 5A, 5B). We identified these structures as MCETs, based on their similarity to MCETs formed in vitro (45). Although essentially all mast cells in PP contain IL-17, a minority of MCETs contained only for IL-17, and many were IL-17-. This suggests that IL-17 is released at low levels upon MCETosis, diffusing to a level below our threshold of detection faster than tryptase or chymase. The shapes of MCETs in skin are more compact than those formed in vitro, likely due to the physical constraints of surrounding tissue. MCETs or degranulated mast cells were not seen frequently in NN or PN samples (Supplemental Figs. 2, 3). These data demonstrate that MCETs can form in human psoriasis lesions and represent a distinct mechanism of IL-17 release by mast cells in vivo.

Treatment of normal human skin explants with IL-1β and IL-23 results in mast cell degranulation and MCETosis

Because IL-1β and IL-23 promote increased production of IL-17 from T cells, we evaluated whether these cytokines similarly might promote production or release of IL-17 from mast cells in situ. Tissue sections from normal skin biopsies (n = 3) incubated for 3 d in control media contained mostly intact mast cells at a density comparable to that of tissue sections from the same patient fixed immediately after biopsy. After 3 d of treatment with IL-1β and IL-23, there was no increased expression of IL-17. However, there were significant increases in degranulated and ETosed mast cells in treated tissue, as measured by staining for tryptase, chymase, and IL-17 (Fig. 5C). Treatment of normal skin with either IL-1β or IL-23 alone led to fewer degranulated/ETosed mast cells than treatment with both compounds together as measured with tryptase (Pmedia:IL-23 < 0.05, Pmedia:IL-1β/IL-23 < 0.05; Fig. 5D) or chymase (Pmedia:IL-23 < 0.01, Pmedia:IL-1β/IL-23 < 0.01; Fig. 5E). Treatment with Substance P or Compound 48/80, classic mast cell degranulating agents, elicited degranulation of mast cells but did not yield significant ETs. Consistent findings were observed in separate experiments from four independent healthy skin donors in the 3-d cultures. These findings indicate that IL-1β and IL-23 can rapidly induce mast cell degranulation and MCETosis in human skin, resulting in the release of IL-17 and other mast cell products (Fig. 6).

Discussion

Our findings suggest prominent roles for IL-17 released by mast cells and neutrophils in psoriasis as outlined in our model (Fig. 6). In normal skin that encounters trauma or microbial infection, mast cells detect stimuli such as necrotic tissue or microbial invasion through TLRs and other pattern recognition receptors. This detection can lead to direct release of preformed mediators including TNF-α, IL-17, CXCL2 (55), chymase, and tryptase through degranulation or MCETosis. These mediators promote endothelial activation, vascular permeability, and rapid influx of neutrophils. Encounter with extracellular bacteria or products of necrotic cells will promote NETosis with the release of mediators including IL-17, which can amplify further neutrophil accumulation by increasing CXCL1, CXCL2, and IL-8 expression. IL-17 produced by neutrophils also has been shown to be critical for neutrophil accumulation (56). Interestingly, these CXCR2 agonists are known to participate in NETosis in vivo because treatment with CXCR2 antagonists block NETosis and improve lung function in cystic fibrosis (57). Activation of CXCR2 by increased release of CXCL1, CXCL2, and IL-8 similarly may mediate NETosis in psoriasis. Although the relevant targets of IL-17 in psoriatic skin are unclear, IL-17 can increase the expression of AMPs human β-defensin-2, S100A7, S100A8, S100A9, and LL-37 in keratinocytes (33–35). These AMPs can stimulate immune cell infiltration, and NET-derived LL-37–DNA complexes may promote IFN-α release from plasmacytoid dendritic cells (58). Indeed, MCET- or NET-derived nucleic acids may be responsible for the plasmacytoid dendritic cell activation seen after tape-stripping of human skin, a model for sterile mechanical skin injury (59, 60), accounting in part for the proposed role of mast cells in the Koebner phenomenon (24). IFN-α and TNF-α can stimulate influx of inflammatory dendritic cells and macrophages, which in the presence of IFN-γ and other danger signals produce cytokines including IL-23 and IL-1β. These cytokines also can stimulate IL-17 release by T cells, neutrophils, and mast cells to further promote the cycle of inflammation.

In a typical host response to infection, TLR stimulation diminishes as pathogens and necrotic tissue are cleared, thereby interrupting the cycle of inflammation with decreased release of IL-17, TNF-α, IFN-α, IFN-γ, and IL-23. However, in people carrying polymorphisms in the psoriasis susceptibility genes IL12B, IL23A, or IL23R, IL-23 signaling may be dysregulated, resulting in inappropriately sustained release of IL-17 and other mediators from mast cells, neutrophils, and T cells. Likewise, polymorphisms in genes modifying signaling downstream of IL-17 (TRAF3IP2, TNF-α, and TLRs (TNFAP3 and TNIP1) may fail to appropriately dampen or extinguish these pathways. Either scenario would result in prolonged inflammation in the absence of infection.

Although our Western blot and immunofluorescence studies indicate the presence of IL-17A protein, mRNA from human neutrophils from several subjects showed either low or undetectable levels of IL-17 using real-time RT-PCR and Affymetrix gene array experiments (M.J. Kaplan, unpublished observations). This may be explained by early production and storage of IL-17A by developing neutrophils prior to their release from bone marrow. Given their short life spans, upon entering the circulation neutrophils may downregulate transcription of many mRNAs. An
alternative explanation is that neutrophils and mast cells may bind to bacteria, fungi, bacterial products, activated platelets, or IL-8 through activation of TLRs and cytokine and Fc receptors (47, 64). The ETosis-promoting activities of IL-23 and IL-1β were not suspected previously, and the precise signaling mechanisms regulating this process remain to be defined. On the basis of our results, we suggest that modulation of mast cell and neutrophil ETosis and release of IL-17 is a novel therapeutic mechanism of action for drugs targeting IL-23.

In conclusion, this study demonstrates that increased numbers of mast cells and neutrophils in psoriasis lesions contribute to the release of the pathogenic cytokine IL-17 through the formation of ETs and that IL-23 and IL-1β provide a novel mechanistic stimulus for this phenomenon. These observations support a model in which mast cells and neutrophils play significant roles in the pathophysiolog of psoriasis and potentially other autoimmune diseases driven by the IL-23–IL-17 axis. This model presents opportunities to better understand current treatments and develop novel therapies that inhibit ETosis and release of IL-17 in a novel therapeutic mechanism of action for drugs targeting IL-23.

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Disclosures
The authors have no financial conflicts of interest.

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