Identification of Semaphorin 4B as a Negative Regulator of Basophil-Mediated Immune Responses

Yukinobu Nakagawa, Hyota Takamatsu, Tatsusada Okuno, Sujin Kang, Satoshi Nojima, Tetsuya Kimura, Tatsuki R. Kataoka, Masahito Ikawa, Toshihiko Toyofuku, Ichiro Katahama, and Atsushi Kumanogoh

J Immunol 2011; 186:2881-2888; Prepublished online 26 January 2011; doi: 10.4049/jimmunol.1003485
http://www.jimmunol.org/content/186/5/2881

Supplementary Material: http://www.jimmunol.org/content/suppl/2011/01/26/jimmunol.1003485.DC1

References: This article cites 47 articles, 12 of which you can access for free at: http://www.jimmunol.org/content/186/5/2881.full#ref-list-1

Subscription: Information about subscribing to The Journal of Immunology is online at: http://jimmunol.org/subscription

Permissions: Submit copyright permission requests at: http://www.aai.org/About/Publications/JI/copyright.html

Email Alerts: Receive free email-alerts when new articles cite this article. Sign up at: http://jimmunol.org/alerts
Identification of Semaphorin 4B as a Negative Regulator of Basophil-Mediated Immune Responses

Yukinobu Nakagawa,*‡,1 Hyota Takamatsu,*† Tatsusada Okuno,*,‡ Sujin Kang,* Satoshi Nojima,*‡ Tetsuya Kimura,* Tatsuki R. Kataoka,† Masahito Ikawa,‡ Toshikazu Tooyofuku,* Ichiro Katayama,† and Atsushi Kumanogoh*,#

Basophils are strong mediators of Th2 responses during helminthic infections. Recently, basophils were shown to function as APCs and promote both Th2 skewing and humoral memory responses. However, the mechanisms that regulate basophils are still unclear. In this article, we show that a class IV semaphorin, Sema4B, negatively regulates basophil functions through T cell–basophil contacts. In a screen to identify semaphorins that function in the immune system, we determined that Sema4B is expressed in T and B cells. Interestingly, Sema4B−/− mice had considerably increased serum IgE levels despite normal lymphocyte and dendritic cell functions. Recombinant Sema4B significantly inhibited IL-4 and IL-6 production from basophils in response to various stimuli, including IL-3, papain, and FceRI cross-linking. In addition, T cell–derived Sema4B, which accumulated at contact sites between basophils and CD4+ T cells, suppressed basophil-mediated Th2 skewing, suggesting that Sema4B regulates basophil responses through cognate cell–cell contacts. Furthermore, Sema4B−/− mice had enhanced basophil-mediated memory IgE production, which was abolished by treating with an anti-FceRI Ab. Collectively, these results indicate that Sema4B negatively regulates basophil-mediated Th2 and humoral memory responses.


Basophils are rare granulocytes that are found in the circulation and are effector cells of the innate immune system that are associated with allergic inflammation and infections with helminth parasites (1–4). Recent studies indicated that basophils regulate Th1/Th2 homeostasis (5, 6) and humoral immunity (7). Basophils produce large amounts of IL-4, a key cytokine in Th2 skewing, in response to various stimuli, including IL-3, the protease allergen papain, and cross-linking of surface-bound IgE molecules (8, 9). In addition, basophils have been shown to express MHC class II and costimulatory molecules and to function as APCs (10–12), although the role of basophils as APCs in vivo is still controversial (13–16). Previous studies have shown that basophil-mediated Th2 skewing can be promoted when these cells are activated by papain (11) or internalize Ags through Ag-specific IgE on their cell surface (12). In addition, basophils have been implicated in humoral memory responses; on re-exposure to Ags, Ag and Ag-specific IgE complexes activate basophils to release IL-4 and IL-6, resulting in enhanced humoral immune responses (7). However, the mechanisms that regulate basophil-mediated Th2 responses are still unknown.

Semaphorins were originally identified as axon guidance molecules during neuronal development (17). However, cumulative findings indicate that semaphorins have diverse functions in many physiological processes (18–22). Semaphorins have been shown to be involved in various phases of immune responses, including the activation of B cells (23), T cells (24), and dendritic cells (DCs) (25); the regulation of Th differentiation (26); and the navigation of immune cell trafficking (27). In particular, membrane-bound class IV semaphorins, Sema4A and Sema4D/CD100, have been extensively investigated. Sema4D has been shown to be important for B cell and DC activation (23). In addition, Sema4A has been demonstrated to be critical for Th differentiation (26). However, the impact of other class IV semaphorins on other immune functions has not been determined.

In this study, we searched for semaphorins that function in the immune system and identified a class IV semaphorin, Sema4B, as a novel immune semaphorin. To determine the physiological roles of Sema4B, we generated Sema4B−/− mice and determined that Sema4B negatively regulates IL-4 and IL-6 production by basophils. We also determined that T cell–derived Sema4B suppresses basophil-mediated Th2 skewing. In addition, Sema4B−/− mice not only had increased serum IgE levels under steady-state conditions but also enhanced memory IgE responses caused by defects in Sema4B−/− mice, which was abolished by treating with an anti-FceRI Ab. Collectively, these results indicate that Sema4B negatively regulates basophil-mediated Th2 and humoral memory responses.

Received for publication October 25, 2010. Accepted for publication December 19, 2010.

This work was supported by research grants from the Ministry of Education, Culture, Sports, Science and Technology of Japan; grants-in-aid from the Ministry of Health, Labour and Welfare; the program for Promotion of Fundamental Studies in Health Sciences of the National Institute of Biomedical Innovation (to A.K.); and the Target Sciences of the National Institute of Biomedical Innovation (to A.K.).

Address correspondence and reprint requests to Prof. Atsushi Kumanogoh, Department of Immunopathology, World Premier International Immunology Frontier Research Center, Research Institute for Microbial Diseases, Osaka University, Suita, Osaka 565-0871, Japan; Department of Dermatology, Osaka University Graduate School of Medicine, Suita, Osaka 565-0871, Japan; Department of Neurology, Osaka University Graduate School of Medicine, Suita, Osaka 565-0871, Japan; Department of Pathology, Osaka University Graduate School of Medicine, Osaka, Osaka 565-0871, Japan; Department of Diagnostic Pathology, Kyoto University Hospital, Kyoto 606-8507, Japan; Animal Resource Center for Infectious Diseases, Research Institute for Microbial Diseases, Osaka University, Suita, Osaka 565-0871, Japan; and Department of Respiratory Medicine, Allergy and Rheumatic Diseases, Osaka University Graduate School of Medicine, Suita, Osaka 565-0871, Japan.

1Y.N. and H.T. contributed equally to this study.

The online version of this article contains supplemental material.

Abbreviations used in this article: BM, bone marrow; BDMD, bone marrow–derived dendritic cell; DC, dendritic cell; hlgG, human IgG; HSA, human serum albumin; KLH, keyhole limpet hemocyanin; NP, 4-hydroxy-3-nitrophenyl acetyl; NP-CGG, 4-hydroxy-3-nitrophenylacyl-chicken-globulin conjugate; PSDL, postsynaptic density; rSema4B, recombinant Sema4B; Tg, transgenic; WT, wild-type.

Copyright © 2011 by The American Association of Immunologists, Inc. 0022-1767/11/$16.00

www.jimmunol.org/cgi/doi/10.4049/jimmunol.1003485

The online version of this article contains supplemental material.

Abbreviations used in this article: BM, bone marrow; BDMD, bone marrow–derived dendritic cell; DC, dendritic cell; hlgG, human IgG; HSA, human serum albumin; KLH, keyhole limpet hemocyanin; NP, 4-hydroxy-3-nitrophenyl acetyl; NP-CGG, 4-hydroxy-3-nitrophenylacyl-chicken-globulin conjugate; PSDL, postsynaptic density; rSema4B, recombinant Sema4B; Tg, transgenic; WT, wild-type.

Copyright © 2011 by The American Association of Immunologists, Inc. 0022-1767/11/$16.00

www.jimmunol.org/cgi/doi/10.4049/jimmunol.1003485
Materials and Methods

**Mice**

C57Bl/6 and BALB/c mice were purchased from CLEA Japan. Sema4B/ε−/− mice on C57Bl/6 and BALB/c backgrounds were generated as described later. Sema4B/ε−/− mice were backcrossed >6x onto the C57Bl/6 or BALB/c background. Transgenic (Tg) mice expressing the TCR specific for OVA (323–339; BALB/c background) were kindly provided by S. Habu (25). OVA-TCR Tg mice on a Sema4B/ε−/− background were established by crossing the OVA-TCR Tg mice with Sema4B/ε−/− BALB/c mice. All mice were bred in specific pathogen-free conditions. All animal experimental protocols were reviewed and approved by our institutional animal care committees.

**Generation of Sema4B/ε−/− mice**

A targeting vector was designed to replace the fifth to eighth exons of the Sema4B gene with a Neo resistance cassette. In addition, the HSV thymidine kinase gene was inserted to select against random integration of this targeting vector. The linearized targeting plasmid DNA was transfected into embryonic stem cells by electroporation. After double selection with G418 and ganciclovir, we screened for homologous recombination of the Sema4B allele by PCR and FACS analysis. PCR was performed using 35 cycles at 94°C for 30 s, 65°C for 30 s, and 72°C for 60 s. The following oligonucleotide primers were used to identify the rearranged Sema4B locus: forward primer (P1): 5′-TAGTGCCCATATGGGACCTG-3′; reverse primer (P2): 5′-TCTCGGACGACTGACTGTT-3′; and reverse primer (P3) that includes the Neo cassette: 5′-TGCTGACGTTGGCTAAGA-3′.

**RT-PCR**

Sema4B mRNA expression was examined by RT-PCR using a panel of multiple mouse tissue cDNAs (Clontech). RT-PCR was performed using 35 cycles at 94°C for 30 s, 60°C for 30 s, and 72°C for 30 s with the forward primer (5′-AACAGCAACCTCTGGTTCGC-3′) and reverse primer (5′-GGCTCCTATCTGGGCTAAGA-3′).

**Generation of the anti-Sema4B Ab**

Rats were repeatedly immunized with purified mouse recombinant Sema4B (rSema4B) and alum, and their spleens were harvested to generate hybridomas using standard methodologies. The hybridomas were screened by ELISA for the production of Sema4B-specific Abs. To confirm the specificity of the Ab for Sema4B, we transfected COS7 cells with Sema4B, Sema4A, Sema4D, or the control vector using Lipofectamine (Invitrogen). After 48 h, the transfecants were stained with the biotinylated anti-Sema4B Ab (TK-2) followed by allophycocyanin-conjugated streptavidin.

**Reagents and Abs**

Anti-CD4 (OKI.5), anti-CD8 (53-6.7), anti-B220 (RA3-6B2), anti-CD49b (DX5), anti-FcεRⅠx (MAR-1), anti-CD62L (MEL-14), anti-ε-Kit (2B8), MHC class II (I-A/I-E, M5/I-11.15.2), and allophycocyanin-, FITC-, and Cy5-conjugated streptavidin were obtained from eBioscience. Anti-CD16/CD32 (2.4G2), anti-CD11c (HL3), anti-CD40 (HM40-3), and anti-CD3ε (145-2C11) were from BD Pharmingen. Anti-T1/2ST2 was from MD Biosciences. Anti-p44/p42 MAPK (SC-154) and anti-STAT5 (SC-835) were from Santa Cruz Biotechnology. Anti-phospho-p44/p42 MAPK (9101) was from Cell Signaling Technology. Anti–phospho-STAT5 (47) was from BD Biosciences. Papain was purchased from Calbiochem. The monoclonal IgE anti-DNP Ab (SPE-7), OVA, DNP-human serum albumin (HSA), and LPS were from Sigma, 4-Hydroxy-3-nitropheny lacetyl-chicken-y-globulin conjugate (NP-CGG), NP-BSA, and DNP-OVA were from Biosearch Technologies.

**Cell purification**

To prepare bone marrow (BM)-derived basophils, BM cells were cultured for 1 d with IL-3 (30 ng/ml, condition medium from a mouse IL-3-producing cell line) in RPMI 1640 medium supplemented with 10% (v/v) FBS, 50 μM 2-ME, 100 U/ml penicillin, and 100 μg/ml streptomycin (29). Samples were enriched for basophils by single-cell sorting on CD11c ε-Kit FcεRⅠx cells or by positively selecting CD11c ε-Kit DX5 cells with MACS (Miltenyi Biotech). Otherwise, BM-derived basophils were enriched by depleting mast cells using anti-ε-Kit–biotin and streptavidin–coupled Dynabeads (Invitrogen) (30). T cells, B cells, and DCs were isolated from the spleen by MACS. The resulting purity was >95% for each experiment. BM-derived DCs (BMDCs) were generated by stimulating with GM-CSF as previously described (31).

**Flow cytometry**

For the rSema4B binding assay, BM-derived basophils were incubated with either biotinylated rSema4B or biotinylated human IgG (hIgG), followed by streptavidin-allophycocyanin. For in vivo Th2 skewing, single-cell suspensions from the spleens of adoptively transferred mice were stained with allophycocyanin-anti-CD4 and FITC-anti-T1/2ST2. To evaluate Sema4B expression in specific immune cell populations, we identified CD4+ T cells (CD4+CD8+), CD8+ T cells (CD4+CD8+), B cells (B220+CD11c+), DCs (CD11c+B220+), and basophils (FcεRⅠx+DX5+) using the indicated markers.

**In vitro stimulation assays**

c-Kit-depleted BM-derived basophils were stimulated with IL-3 (30 ng/ml) after being starved for 8 h, or stimulated with papain (20 μg/ml) or IgE cross-linking without starvation. Basophils were activated with IgE cross-linking by incubating with IgE anti-DNP (1 μg/ml), followed by various concentrations of DNP-HSA for 16 h. Basophils were stimulated coated with rSema4B (10 μg/ml) or hIgG (10 μg/ml). The concentrations of IL-4 and IL-6 in the culture supernatants were measured by ELISA (R&D Systems). B cells were cultured with anti-CD40 (0.2 μg/ml) and various concentrations of IL-4 for 7 d (32). The IgM, IgG1, and IgE levels in the culture supernatants were measured by ELISA (Bethyl Laboratories).

**In vitro and in vivo basophil-mediated Th2 skewing assays**

For the in vitro basophil-mediated Th2 skewing assay, BM-derived basophils (CD11c ε-Kit FcεRⅠx+) were enriched by flow cytometry sorting. Naive CD4+CD262L+ T cells from OVA-TCR Tg mice (4 × 106 cells/ml) and irradiated basophils (5 × 105 cells/ml) were cultured with the OVA peptide (1 μM), OVA (100 μg/ml) and papain (20 μg/ml), or IgE anti-DNP (1 μg/ml) and DNP-OVA (100 μg/ml) (11, 12) in plates coated with Sema4B (10 μg/ml) or hIgG (10 μg/ml). In some experiments, naive CD4+ CD262L+ T cells from wild-type (WT) or Sema4B/ε−/− mice were used instead of rSema4B. After 5 d of coculture, CD4+ T cells isolated by anti-CD4–conjugated magnetic beads (autoMACS) were restimulated with immobilized anti-CD3 for 24 h. For the in vivo basophil-mediated Th2 skewing assay, naive CD4+CD262L+ T cells (5 × 105 cells/mouse) from WT or Sema4B/ε−/− OVA-TCR Tg mice were i.v. transferred into nude mice. The next day, basophils (CD11c ε-Kit FcεRⅠx+, 5 × 105 cells/mouse) pulsed with IgE anti-DNP (2 μg/ml) and DNP-OVA (100 μg/ml) were transferred through the tail vein into these recipient nude mice. After 4 d, CD4+ T cells (1 × 106 cells/well) isolated from the spleen by flow cytometry sorting were cultured with BMDMC (1 × 105 cells/well) and DNP-OVA (100 μg/ml) in 96-well plates for 36 h. Cytokine concentrations in the supernatants were measured by ELISA (Bethyl Laboratories).

**In vivo T cell priming and Ab production assays**

Mice were immunized in the hind footpads with keyhole limpet hemocyanin (KLH; 100 μg) in CFA or alum. Five days after priming, cells were purified from the draining lymph nodes. Lymph node cells (1 × 105 cells/well) were stimulated with various concentrations of KLH. For priming assays, the cells were cultured with 2 μg/ml [3H]thymidine for the last 16 h (26). To induce Ab responses to T cell-dependent Ags, we immunized mice i.p. with 50 μg NP-CCG as an alum-precipitated complex on day 0 and then boosted on day 14. Serum was collected on days 0, 14, and 21. NP-specific Abs were detected with NP-BSA–coated ELISA plates and quantified using isotype-specific Abs as previously described (33).

**Measurement of serum IgS**

The concentrations of serum IgS were measured using a mouse IgE ELISA quantitation kit (Bethyl Laboratories).

**In vivo memory response assays**

WT and Sema4B/ε−/− mice were immunized i.p. with 100 μg OVA without adjuvant. Four weeks after the primary immunization, the mice were administered 10 μg OVA i.v. (7). Serum was collected on days 0, 5, 7, and 9 after rechallenging with the Ag. To deplete basophils, we injected primary immunized mice twice daily for 3 d with 5 μg anti-FcεRⅠx. The mice were allowed to rest for 2 d and then were i.v. injected with 10 μg OVA. OVA-specific serum IgE was measured with a mouse OVA-IgE ELISA kit (Dainippon Sumitomo Pharma). OVA-specific serum IgG1 was measured with a mouse OVA-IgG1 ELISA kit (Shibayagi).

**Immunohistochemistry**

Sema4B/ε−/− BM-derived basophils (CD11c ε-Kit DX5+) and OVA-TCR Tg-derived CD4+ T cells were cultured with OVA peptide for 1 h. Then the
cells were fixed with 4% paraformaldehyde in phosphate buffer for 15 min, attached to coverslips by cytospin, stained with FITC–anti-CD3ε, PE–anti–I-A/I-E, and anti-Sema4B–biotin plus streptavidin-Cy5, and then examined by confocal microscopy (Zeiss Exciter).

**Western blot analysis**
BM-derived basophils were starved for 16 h and then stimulated with IL-3 (30 ng/ml) in the presence of Sema4B- or hIgG-coated Dynabeads (Invitrogen). The cells were lysed at the indicated times with lysis buffer containing 1% Nonidet P40, 10 mM Tris-HCl, 150 mM NaCl, 1 mM EDTA, 10 mM Na3Vo4, 0.5 mM PMSF, 5 µg/ml leupeptin, 5 µg/ml aprotinin, 1 mM sodium orthovanadate, and a protease inhibitor mixture (Roche). Whole-cell lysates were separated by SDS-PAGE and then electrophoretically transferred to nitrocellulose membranes. The membranes were immunoblotted with various Abs.

**Statistical analysis**
Data are presented as mean ± SD. The p values were calculated with the two-tailed Student t test after the data were confirmed to fulfill the criteria. Otherwise, Mann–Whitney U test was performed.

**Results**
**Sema4B is expressed in T and B cells**
In a screen to identify semaphorins in the immune system, we isolated a cDNA fragment encoding a class IV semaphorin, Sema4B, through PCR-based cloning using degenerate oligonucleotide primers derived from motifs that are conserved across the semaphorin family. RT-PCR analyses showed that Sema4B was abundantly expressed in the spleen (Fig. 1A). Sema4B has been reported to play a role in assembling the postsynaptic specialization at glutamatergic and GABAergic synapses in the nervous system (34). However, the involvement of Sema4B in immune cell regulation has not been determined, and this led us to investigate the immunological function of Sema4B. To examine the expression of Sema4B in more detail, we generated an anti-Sema4B mAb that specifically recognizes Sema4B and does not cross-react with Sema4A or Sema4D (Fig. 1B). As shown in Fig. 1C, Sema4B is constitutively expressed in B and T cells, but not in DCs or basophils. To investigate the physiological role of Sema4B, we generated Sema4B−/− mice by deleting the fifth to eighth exons in the Sema4B gene (Fig. 1D–F). Sema4B−/− mice were born at the expected Mendelian ratio and were fertile.

**Sema4B−/− mice have increased serum IgE levels**
Immune cell populations, such as T cells, B cells, DCs, NK cells, and subpopulations of T cells, were normal in Sema4B−/− mice.
Sema4B negatively regulates basophil functions

We then analyzed the basophil populations in the spleen and BM because basophil numbers have been shown to be critical for Th2 polarization and IgE production (5, 6). The proportion of basophils in WT and Sema4B−/− mice was comparable (Fig. 3A). We next examined whether rSema4B affects basophil responses. As shown in Fig. 3B, rSema4B bound to BM-derived basophils and significantly inhibited the ability of these cells to produce IL-4 and IL-6 in response to IL-3 stimulation (Fig. 3C), and these inhibitory effects were proportional to the rSema4B concentrations (Supplemental Fig. 4). It was previously reported that the FcγRIIA–mediated ITAM-spleen tyrosine kinase and its downstream mediator, ERK, are involved in IL-3–induced IL-4 production (30). In addition, IL-3Rβc–mediated JAK–STAT5 pathways are crucial for IL-3–induced proliferation (36, 37). As shown in Fig. 3D, ERK phosphorylation was inhibited by rSema4B, and similarly, STAT5 phosphorylation was inhibited by rSema4B (Fig. 3E). Basophils produce large amounts of IL-4 and IL-6 after being stimulated with cysteine proteases such as papain (9) or cross-linking of their surface IgE (12). rSema4B also inhibited IL-4 and IL-6 production from BM-derived basophils that were stimulated with papain or IgE anti-DNP and DNP-HSA (Fig. 3F, 3G). These results indicate that Sema4B negatively regulates cytokine production from basophils.

Sema4B suppresses basophil-mediated Th2 skewing

We examined whether rSema4B suppressed basophil-mediated Ag-specific Th2 skewing because basophils have been reported to promote Th2 polarization by functioning as APCs (38). To clarify this point, we cocultured OVA-TCR Tg-derived naive CD4+ T cells with OVA peptide-pulsed BM-derived basophils for 5 d in the presence of rSema4B or hIgG. Interestingly, rSema4B significantly suppressed IL-4 but not IFN-γ production from T cells (Fig. 4A). Furthermore, when the basophil–T cell cocultures were...
treated with papain and OVA or IgE anti-DNP and DNP-OVA. Sema4B also significantly inhibited basophil-mediated Th2 skewing, resulting in suppression of basophil-mediated Th2 skewing.

Sema4B is highly expressed in CD4+ T cells (Fig. 1C). However, intrinsic Sema4B did not affect Th1 and Th2 differentiation in vitro (Supplemental Fig. 2C). To analyze whether T cell-derived Sema4B is critical for regulating basophil-mediated Th2 skewing, we cultured WT or Sema4B−/− OVA-TCR Tg-derived naive CD4+ T cells with BM-derived basophils and OVA peptide or DNP-OVA and IgE anti-DNP for 5 d. Sema4B−/− T cells showed considerably enhanced Th2 skewing (Fig. 4D, 4E). Next, to examine basophil-mediated Th2 skewing in vivo, we adoptively transferred WT or Sema4B−/− OVA-TCR Tg-derived naive CD4+ T cells into nude mice; then BM-derived basophils pulsed with OVA proteins. Sema4B−/− basophils produced comparable levels of IL-4 and enhanced IL-4 production after being stimulated with DNP-OVA and BMDC (Fig. 4G). These results suggest that T cell-derived Sema4B is important for basophil-mediated Th2 skewing.

Basophils have been shown to form an immunological synapse with cognate T cells (11). The cytoplasmic tail of Sema4B contains a PDZ-binding motif and binds to postsynaptic density (PSD)-95 (39) that accumulates at contact sites between thymocytes and DCs (40). Thus, we next examined the localization of Sema4B during basophil and T cell interactions. Consistent with previous reports, papain-activated BM-derived basophils formed an immunological synapse with OVA-TCR Tg-derived CD4+ T cells in the presence of OVA peptide. Of note, Sema4B in T cells colocalized with CD3 and clustered at T cell–basophil contact sites (Fig. 4H). These results suggest that Sema4B in T cells accumulates at the immunological synapse and suppresses basophil functions in a cell–cell contact-dependent manner.

**Sema4B−/− mice have enhanced IgE memory responses**

During secondary Ag exposure, Ag-specific, IgE-bearing basophils capture Ag, become activated, and secrete cytokines, which subsequently facilitates memory B cell responses (7, 41). Although WT and Sema4B−/− basophils produced comparable levels of IL-4 and IL-6 on FcεRI cross-linking (Supplemental Fig. 5), the serum IgE concentrations increased with age in Sema4B−/− mice (Fig. 5A). We hypothesized that this phenomenon can be caused not only by basophil-mediated priming responses but by basophil-mediated memory responses. To determine whether Sema4B is involved in immunological memory responses, we immunized WT and Sema4B−/− mice with OVA proteins and then boosted them with OVA proteins. Sema4B−/− mice had significantly greater levels of serum OVA-specific IgG1 and IgE than WT mice (Fig. 5A). Furthermore, the increased IgG1 and IgE responses in Sema4B−/− mice were suppressed when basophils were depleted before the secondary immunization (Fig. 5B), which suggests that basophils are responsible for the enhanced memory responses in Sema4B−/− mice. Taken together, these results indicate that Sema4B also negatively regulates basophil-mediated humoral memory responses.

**Discussion**

Semaphorins have been shown to play crucial roles in the immune system (21). In this study, we performed a screen to identify novel
FIGURE 4. Sema4B negatively regulates basophil-mediated Th2 skewing. A–C, OVA–TCR Tg-derived naive CD4+CD62L+ T cells and irradiated BM-derived basophils (CD11c−c-Kit− FcεRIα−) were cultured with OVA peptide (A), papain and OVA (B), or IgE anti-DNP and DNP-OVA (C) in the presence of rSema4B (closed circles) or hlgG (open circles) for 5 d. Then the CD4+ T cells isolated by anti-CD4–conjugated magnetic beads were restimulated with various concentrations of immobilized anti-CD3 for 24 h. The cytokine concentrations in the culture supernatants were measured by ELISA. D and E, WT (open circles) or Sema4B−/− (closed circles) OVA–TCR Tg-derived naive CD4+CD62L− T cells and irradiated BM-derived basophils (CD11c−c-Kit− FcεRIα−) were cultured with OVA peptide (D) or IgE anti-DNP and DNP-OVA (E) for 5 d. Then the CD4+ T cells isolated by anti-CD4–conjugated magnetic beads were restimulated with various concentrations of immobilized anti-CD3 for 24 h. The cytokine concentrations in the culture supernatants were measured by ELISA. F and G, WT (+/+ or open bars) or Sema4B−/− (−/− or closed bars) OVA–TCR Tg-derived naive CD4+CD62L− T cells were i.v. transferred into nude mice. The next day, BM-derived basophils (CD11c−c-Kit− FcεRIα−) pulsed with IgE anti-DNP and DNP-OVA were i.v. transferred into these recipients. Four days later, CD4+ T cells isolated from these mice were stained with anti-CD4 and anti-T1/ST2 (AII1/ST2) (red) and anti-Sema4B–biotin plus streptavidin-Cy5 (blue). The localization of Sema4B was evaluated by confocal microscopy. Scale bar, 10 μm.

FIGURE 5. Sema4B−/− mice exhibit enhanced humoral memory responses. A, WT (open circles) and Sema4B−/− (closed circles) mice were immunized i.p. with OVA without adjuvant and then rechallenged i.v. with OVA after 4 wk. OVA-specific IgG1 and IgE in the serum were measured at the indicated times by ELISA. B, WT (open circles) and Sema4B−/− (closed circles) mice were primarily immunized as described earlier and then depleted of basophils. Two days after the final injection of anti-FceRIα, the mice were rechallenged with OVA. The OVA-specific IgG1 and IgE levels were measured by ELISA. *p < 0.05, **p < 0.01 (Mann–Whitney U test).
IgE production in Sema4B induced phosphorylation of ERK and STAT5, suggesting that sofophil functions, we showed that Sema4B suppressed the IL-3–mediated signals in basophils, it is possible that Sema4B recruited to ITIM-containing receptors (45). Although it remains unclear whether ITIM-containing receptor is directly involved in IL-3–mediated signals in basophils, it is possible that Sema4B may regulate basophil functions through ITIM-containing molecules. Regarding the receptors for class IV semaphorins, several molecules such as plexin-Bs (B1, B2, B3) (46), plexin-D1 (47), T cell Ig and mucin domain-containing molecule-2 (Tim2) (24), and CD72 (23) have been shown to bind to class IV semaphorins. In fact, many of these molecules are expressed by basophils as determined by RT-PCR (data not shown). Additional investigations, including the identification of the Sema4B receptor, are required to further examine these mechanisms.

In conclusion, we demonstrated that Sema4B suppressed IL-4 production from basophils, in which T cell-derived Sema4B inhibited basophil-mediated Th2 skewing. In addition, a Sema4B deficiency significantly affected Ag-specific IgE memory responses, possibly in a cell–cell contact-dependent manner. Thus, Sema4B negatively regulates basophil functions during both primary and memory responses. These findings not only provide new insight into mechanisms that regulate basophils but identify a novel therapeutic target for allergic diseases.

Acknowledgments
We thank T. Yazawa for technical support.

Disclosures
The authors have no financial conflicts of interest.

References


