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J Immunol 2010; 185:7309-7316; Prepublished online 15 November 2010;
doi: 10.4049/jimmunol.1000095
http://www.jimmunol.org/content/185/12/7309

Supplementary Material http://www.jimmunol.org/content/suppl/2010/11/15/jimmunol.1000095.DC1

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Print ISSN: 0022-1767 Online ISSN: 1550-6606.
Inhibition of TLR4-Induced IκB Kinase Activity by the RON Receptor Tyrosine Kinase and Its Ligand, Macrophage-Stimulating Protein

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The RON receptor tyrosine kinase regulates the balance between classical (M1) and alternative (M2) macrophage activation. In primary macrophages, the ligand for Ron, macrophage-stimulating protein (MSP), inhibits the expression of inducible NO synthase, a marker of classically activated macrophages, whereas promoting the expression of arginase I, a marker of alternative activation. Ron−/− mice express increased levels of IL-12, a product of classically activated macrophages, after endotoxin administration, resulting in increased serum IFN-γ levels and enhanced susceptibility to septic shock. In this study, we demonstrate that MSP inhibits LPS-induced IL-12p40 expression, and this inhibition is dependent on the docking site tyrosines in Ron. To further define this inhibition, we examined the effect of Ron on signaling pathways downstream of Ron. We found that MSP does not inhibit the MyD88-independent activation of IFN regulatory factor 3 and production of IFN-β in response to LPS, nor does it inhibit MyD88-dependent TGF-β–activated kinase phosphorylation or MAPK activation in primary macrophages. However, the induction of IκB kinase activity, IκB degradation, and DNA binding of NF-κB after LPS stimulation is delayed in the presence of MSP. In addition, Ron inhibits serine phosphorylation of p65 and NF-κB transcriptional activity induced by LPS stimulation of TLR4. Finally, MSP inhibits the NF-κB–dependent upregulation of the nuclear IκB family member, IκBζ, a positive regulator of secondary response genes including IL-12p40. LPS also induces expression of Ron and an N-terminally truncated form of Ron, Sf-Ron, in primary macrophages, suggesting that the upregulation of Ron by LPS could provide classical feedback regulation of TLR signaling. The Journal of Immunology, 2010, 185: 7309–7316.

Macrophages are a heterogeneous population of cells that exhibit a wide range of functions depending on their physical location, as well as external cues received from the tissue microenvironment (1, 2). Classical, or M1, macrophages are the most commonly described subset of macrophages. These macrophages are activated by proinflammatory cytokines and pathogen-associated molecular patterns that result in stimulation of pattern recognition receptors including the family of TLRs. Classically activated macrophages, in turn, produce proinflammatory cytokines and an array of cytotoxic molecules that aid in the clearance of invading pathogens and stimulate the acquired immune response. Alternative, or M2, macrophages, in contrast, develop in response to Th2 cytokines and produce anti-inflammatory cytokines, express an array of phagocytic receptors, and release products that promote tissue regeneration and healing. The characteristic hallmarks of these diverse macrophage populations are typified by the expression of inducible NO synthase (iNOS) and arginase I by M1 and M2 macrophages, respectively. These enzymes share a common substrate, L-arginine, but whereas iNOS promotes the production of NO, which is cytotoxic, arginase promotes the production of polyamines and proline, which promote the healing process.

Although classically activated macrophages promote immunity to infection, products from these cells can also be damaging to host tissues and are implicated in the progression of a number of chronic inflammatory diseases. Thus, the study of signals that tip the balance of macrophage activation away from the M1 phenotype and toward the M2 phenotype could have wide-ranging therapeutic implications. A critical role for receptor tyrosine kinases in the regulation of macrophage activation is emerging (3, 4). The RON receptor tyrosine kinase is expressed by tissue-resident macrophages and inhibits classical macrophage activation whereas promoting hallmarks of alternative macrophage activation. Macrophage-stimulating protein (MSP), the ligand for the RON receptor, regulates arginine metabolism in macrophages by inhibiting the expression of iNOS and promoting the expression of arginase in primary resident peritoneal macrophages (5–7). Furthermore, mice with a targeted deletion in the gene encoding Ron exhibit enhanced inflammation and tissue damage in models of septic shock, delayed type hypersensitivity, experimental autoimmune encephalitis, and acute lung injury (8–11). Thus, understanding the mechanism by which the RON receptor promotes these responses is of potential clinical significance.
We have shown previously that the RON receptor is a critical regulator of IL-12 production by macrophages both in vitro and in vivo (12, 13). IL-12, a product of classically activated macrophages, plays a central role in promoting both the innate and acquired immune response to infection (14). In the early stages of infection, IL-12 produced by macrophages induces the expression of IFN-γ by NK cells, γδ T cells, and innate CD8+ T cells. At later stages, IL-12 promotes the differentiation of Th1 cells, which produce increased levels of IFN-γ and drive the acquired immune response to infection. IFN-γ produced by both innate and acquired immune cells feeds back to prime macrophages for further activation of the M1 phenotype. Our previous studies demonstrate that enhanced production of IL-12 by Ron−/− splenocytes after infection with LPS, a ligand for TLR4, results in increased IFN-γ production by NK cells and enhanced susceptibility of Ron−/− mice to endotoxic shock. Furthermore, MSP stimulation of the RON receptor on primary macrophages inhibits the response of these cells to IFN-γ through inhibition of STAT1 tyrosine phosphorylation induced by IFN-γ. MSP stimulation of primary macrophages induces the expression of suppressor of cytokine signaling (Socs) 1 and Socs3 delineating a mechanism by which the inhibition of IFN-γ–dependent responses in these cells could occur (13).

Several studies have demonstrated that the RON receptor inhibits LPS and cytokine-induced NF-κB activation in a variety of macrophage populations (6, 15, 16); however, the molecular targets of Ron downstream of TLR4 activation in primary macrophages have not been systematically addressed. In this report, we show that stimulation of primary peritoneal macrophages with MSP, the ligand for Ron, does not inhibit receptor proximal signaling events initiated by LPS stimulation of TLR4, but rather specifically inhibits the activation of IκB kinase (IKK) and its downstream targets by LPS. Furthermore, we demonstrate the induction of Ron expression by LPS, identifying this as a potential negative feedback response for regulating TLR4 signaling. These studies will contribute to the identification of potential regulatory mechanisms that can be targeted in the treatment of a wide range of inflammatory diseases.

Materials and Methods

Animals

The generation of mice with a targeted mutation in Ron was described previously (8). Primary peritoneal macrophages or bone marrow-derived macrophages were isolated from wild-type and Ron−/− mice on the BALB/c or C57BL/6 backgrounds. The Pennsylvania State University Institutional Animal Care and Use Committee approved all animal experiments.

Reagents

The following reagents were obtained from the indicated sources: FBS and PBS (Invitrogen, Grand Island, NY); recombinant human MSP (R&D Systems, Minneapolis, MN); LPS (055:B5, Sigma, St. Louis, MO); DMEM, 1-glutamine, and nonessential amino acids and sodium pyruvate (cellgro; Mediatech, Manassas, VA). Abs for Western blot against IκBα, phosphoY539/638IFN regulatory factor 3 (IRF3), total IRF3, phosphoThr381/386/VIVO TGF-β–activated kinase (Tak1), total Tak1, IκBα, phosphopT325/325+358/p44/42, total p44/42, phosphoY1061Y1085/Y1049/Y1050/Y1055stress-activated protein kinase/JNK, total stress-activated protein kinase/JNK, phosphoY185stress-activated protein kinase/JNK, total stress-activated protein kinase/JNK, phosphoY185/187,y38, total p38, phosphoY92/Gab1, total Gab1, phosphoY532/Gab2, total Gab2, phosphoS636, phosphoP90PP65, and hemagglutinin (HA)-tag were purchased from Cell Signaling (Beverly, MA). Anti-Bcl3 (C-14) was purchased from Santa Cruz Biotechnology (Santa Cruz, CA). Abs for flow cytometry, F4/80-R-phycocerythrin, CD11b-allophycocyanin-750, and Ron were purchased from AbD Serotec (Raleigh, NC), ebioscience (San Diego, CA), and BD Biosciences (San Jose, CA), respectively.

Cells

Murine resident peritoneal macrophages were harvested and cultured as described previously (17). Thygcollate-elicted macrophages were prepared by injecting 2-mo-old BALB/c mice with 2 ml 3% aged thioglycollate media (Difco, Sparks, MD). On day 4, cells were harvested by lavage, and macrophages were isolated and cultured as described for resident macrophages. 293 cells stably transfected with TLR4 and MD-2 (a kind gift from Dr. Golenbock, University of Massachusetts, Worcester, MA) were maintained in complete cell culture medium (DMEM with 1-glutamine, nonessential amino acids, sodium pyruvate, ciprofloxacin, and 10% FBS) supplemented with 0.5 mg/ml G418. Bone marrow-derived macrophages were differentiated from progenitors in the bone marrow by culturing bone marrow cells in macrophage differentiation media (30% L929 conditioned media, 20% FBS, 50% DMEM) for 5 d.

Luciferase assays

293 cells stably expressing TLR4 and MD-2 were plated overnight in 24-well plates and then transfected using the Mirus transfection reagent (Mirus Bio, Madison, WI), according to the manufacturer’s recommendations. The cells were transfected with 5 ng ELAM or AP-1 luciferase and the indicated concentrations of PCDNA3.1 or PCDNA3.1-Ron-IA. Twenty-four hours after transfection, the cells were stimulated for 4 h with 100 ng/ml LPS. Cell lysates were harvested and the luciferase assay was performed using a Turner Designs TD-20/20 luminometer (Sunnyvale, CA). Luciferase assay reagents were purchased from Promega (Madison, WI).

Retroviral transduction

Bone marrow–derived macrophages were infected twice (once for 6 h, followed by another round of infection overnight) with media containing 75% retroviral supernatants. 25% fresh complete media, and 5 μg/ml polybrene. Viral supernatants were generated by transiently transfecting 293 cells using the Mirus reagent with pEco, pVsvg, and murine stem cell virus-based vectors containing the indicated cDNAs. Twenty-four hours after transfection, the media was changed, and after another 24 h, the culture supernatants containing the packed virus were collected and filtered.

Cytokine analysis

Cells were cultured with or without 100 ng/ml MSP overnight followed by simulation for 24 h with 100 ng/ml LPS. Cell supernatants were collected and assayed for IL-12p40 by ELISA using the manufacturer’s protocol (BD Biosciences, San Jose, CA).

Western blot analysis

Cell lysates were collected using the following lysis buffer (150 mM NaCl, 10 mM Tri-HCl [pH 7.5], 0.5% Triton X-100), supplemented with protease and phosphatase inhibitors (10 μg/ml leupeptin, 2 μg/ml pepstatin A, 10 μg/ml aprotinin, 1 mM sodium orthovanadate, 1 M sodium fluoride, and 1 mM PMSF). SDS-PAGE was performed using 10% bis-acrylamide gels. The protein was transferred onto Immobilon P polyvinylidene difluoride membranes (Millipore, Bedford, MA), and the blots were blocked with 5% nonfat dry milk or 5% BSA (Rockland, Gilbertsville, PA) in TBS containing 0.1% Tween 20 for 1 h at room temperature. The blots were then incubated overnight with primary Ab at 4°C. Anti-rabbit or anti-mouse secondary Abs conjugated with HRP were used at concentrations according to the manufacturer’s instructions for 1 h at room temperature. ECL Plus (Amersham, Piscataway, NJ) was used to develop the blots.

In vitro kinase assay

Cell lysates from thioglycollate-elicted macrophages were prepared using the M-PER buffer (Pierce, Rockford, IL) with protease and phosphatase inhibitors as described earlier. The cell lysates (100 μg protein) were incubated with GST-IFA-α, ATP, and [γ-32P]ATP (1 μCi; Amersham Biosciences) for 1.5 h in the kinase wash buffer (50 mM Tris-Cl, pH 8.0, containing 100 μM NaCl, 10 mM MgCl2, 1 mM DTT, 10 mM β-glycophosphate, 10 mM NaF, and 1 mM sodium orthovanadate) on a shaker at room temperature. Glutathione Sepharose beads were added to the mixture and incubated at room temperature for 1 h with gentle shaking followed by centrifugation. The pellet was washed twice with sterile PBS and phosphosubstituted GST-IFA-α was separated by SDS-PAGE, followed by autoradiography.

Electrophoretic mobility shift assay

Nuclear proteins were isolated from thioglycollate-elicted macrophages as described previously (18). The DNA sequence of the sense strand specific for NF-κB was 5′-AGTGGAGGGGCATCTCAGGC-3′, and the complementary strand was annealed. The double-stranded oligonucleotides were labeled with [γ-32P]ATP (3000 Ci/mmol and 10 μCi/ml by the T4 polynucleotide kinase (NEB, Beverly, MA). Ten micromolars of nuclear proteins was incubated in a binding buffer specific for NF-κB as described previously (19) in the presence of 2 μg poly(dI-dC) for 20 min at room temperature. The nuclear extract was then incubated for 20 min with

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10,000 cpm $^{32}$P-labeled NF-κB probes, and the samples were analyzed as described previously (18).

**NF-κB binding assay**

Thioglycollate-elicited macrophages were cultured with or without 100 ng/ml MSP overnight followed by stimulation with 100 ng/ml LPS for the indicated times. Nuclear proteins were collected and assayed for NF-κB binding activity by ELISA according to the manufacturer’s protocol (Active Motif, Carlsbad, CA).

**Flow cytometry**

Peritoneal lavage cells were plated overnight with or without MSP stimulation. Twelve hours after adherence to the plate, cells were collected. One × 10$^6$ cells were incubated per tube in 100 μl FACS buffer (ice-cold PBS + 2% FBS). Nonspecific binding was blocked by the addition of 1 μl Fc-block for 10 min on ice. F4/80-R-phycocerythrin, CD11b-allophycocyanin-750, and Ron Abs were added to the tubes for 30 min on ice. Cells were washed two times with FACS buffer and resuspended in 100 μl FACS buffer. Cells were incubated with anti-goat Alexa 647 as a secondary Ab to Ron for 30 min on ice. Cells were washed two more times, resuspended in 500 μl FACS buffer, and analyzed on a Beckman Coulter FC500 (Fullerton, CA). Ron expression was measured in macrophages positive for F4/80 and CD11b.

**RNA extraction and real-time PCR**

RNA was extracted from cultured cells using the RNeasy mini column kit (Qiagen, Santa Clara, CA). RNA was treated with DNase 1 (Invitrogen) according to the manufacturer’s recommendations. Real-time PCR was carried out on Applied Biosystems 7500 (San Francisco, CA). Reverse transcription for real-time PCR experiments was carried out using the Applied Biosystems High Capacity RT Kit according to the manufacturer’s instructions. Gene-specific primers and probes were designed by the Nucleic Acid Facility at the Pennsylvania State University using Primer Express V.1.0 (Applied Biosystems). Primer and probe sequences for Sf-Ron and Ron receptor, we treated primary macrophages from wild-type and Ron$^{-/-}$ mice with MSP before stimulation with IFN-γ. Results from these studies demonstrate that the inhibition of IL-12p40 by MSP is dependent on the RON receptor (Fig. 1B). To confirm that MSP induces Ron-dependent signaling in primary macrophages, we examined the phosphorylation of Gab1 and Gab2, two adaptors that are recruited to Ron and mediate signaling downstream of MSP stimulation of the RON receptor inhibits IL-12p40 production by primary macrophages in response to LPS We have shown previously that MSP stimulation of primary peritoneal macrophages inhibits the induction of IL-12p40 expression in response to IFN-γ and LPS. This inhibition is associated with reduced IFN-γ-dependent STAT1 phosphorylation and IFN consensus sequence binding protein upregulation, and the upregulation of Socs1 and Socs3 by MSP (12). However, the effect of the MSP/ Ron signaling pathway specifically on LPS/TLR4 signaling in primary macrophages has not been investigated. In this study, we cultured primary peritoneal macrophages overnight in the presence of MSP, followed by stimulation with LPS in the presence or absence of MSP, and assessed IL-12p40 production by ELISA (Fig. 1A). Results from these studies demonstrate that MSP inhibits IL-12p40 expression in response to LPS stimulation after overnight culture in the absence of MSP. However, this inhibition was significantly enhanced when the cells were first cultured overnight in the presence of MSP. The sustained expression of Ron on primary macrophages after overnight culture was confirmed by flow cytometry (Supplemental Fig. 1). These conditions were used for all subsequent experiments.

To determine whether this inhibition is mediated by the RON receptor, we treated primary macrophages from wild-type and Ron$^{-/-}$ mice with MSP before stimulation with IFN-γ. Results from these studies demonstrate that the inhibition of IL-12p40 by MSP is dependent on the RON receptor (Fig. 1B). To confirm that MSP induces Ron-dependent signaling in primary macrophages, we examined the phosphorylation of Gab1 and Gab2, two adaptors that are recruited to Ron and mediate signaling downstream of

**Statistical analysis**

Statistical analyses were performed using the Student t test.

**Results**

MSP stimulation of the RON receptor inhibits IL-12p40 production by primary macrophages in response to LPS We have shown previously that MSP stimulation of primary peritoneal macrophages inhibits the induction of IL-12p40 expression in response to IFN-γ and LPS. This inhibition is associated with reduced IFN-γ-dependent STAT1 phosphorylation and IFN consensus sequence binding protein upregulation, and the upregulation of Socs1 and Socs3 by MSP (12). However, the effect of the MSP/Ron signaling pathway specifically on LPS/TLR4 signaling in primary macrophages has not been investigated. In this study, we cultured primary peritoneal macrophages overnight in the presence of MSP, followed by stimulation with LPS in the presence or absence of MSP, and assessed IL-12p40 production by ELISA (Fig. 1A). Results from these studies demonstrate that MSP inhibits IL-12p40 expression in response to LPS stimulation after overnight culture in the absence of MSP. However, this inhibition was significantly enhanced when the cells were first cultured overnight in the presence of MSP. The sustained expression of Ron on primary macrophages after overnight culture was confirmed by flow cytometry (Supplemental Fig. 1). These conditions were used for all subsequent experiments.

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**FIGURE 1.** MSP stimulation of the RON receptor inhibits IL-12p40 production in primary macrophages. A, Primary peritoneal macrophages were cultured overnight in the presence or absence of 100 ng/ml MSP followed by stimulation with LPS with or without MSP. IL-12p40 production was assessed by ELISA. *p < 0.01. B, Primary resident peritoneal macrophages were collected from wild-type and Ron$^{-/-}$ BALB/c mice by lavage. Macrophages were isolated by adherence to plastic. The cells were stimulated overnight with 100 ng/ml MSP followed by stimulation with 100 ng/ml LPS for 24 h, and supernatants were collected. IL-12p40 production was assessed by ELISA. C, Primary peritoneal macrophages were isolated from wild-type and Ron$^{-/-}$ mice by lavage, and macrophages were isolated by adherence to plastic. After overnight culture, the cells were stimulated with MSP for the indicated times, and phosphorylation of Gab1 and Gab2 was assessed by Western blot analysis. D, Peritoneal macrophages were stimulated with 100 ng/ml MSP overnight followed by activation with 100 ng/ml LPS for the indicated times. IL-12p40 mRNA expression was assessed by real-time PCR.
Grb2 heterozygous mice (homozygous mutant animals are embryonic lethal). Using these animals, we have shown previously that Grb2 plays an essential role in the progression of erythroleukemia induced by a truncated form of the RON receptor (20). Resident macrophages from Grb2^{−/−} mice or their littermate controls were cultured in the presence or absence of 100 ng/ml MSP for 12 h, followed by stimulation with 100 ng/ml LPS for 24 h. IL-12p40 levels in the supernatants were assessed by ELISA. We observed a statistically significant difference in the level of IL-12p40 suppression by MSP between wild-type and Grb2^{−/−} macrophages indicating that the recruitment of Grb2 to the second docking site tyrosine in Ron may play a role in the inhibition of LPS-induced IL-12p40 after MSP stimulation of Ron (Fig. 2B).

MSP does not inhibit receptor proximal signaling by LPS/TLR4

TLR4 signals through MyD88-dependent and -independent pathways. MyD88-independent signaling by TLR4 requires the adaptor proteins TRIF and TRIF-related adaptor molecule, and results in the phosphorylation of IRF3 and the upregulation of IFN-β. The IFN-β produced in response to TLR4 activation results in delayed activation of STAT1 phosphorylation, which promotes the expression of a secondary wave of LPS-responsive genes, including IL-12p40 (21). To determine whether MSP inhibits the MyD88-independent signaling pathway induced by TLR4, we treated primary peritoneal macrophages with MSP followed by stimulation with LPS, and the phosphorylation of IRF3 and upregulation of IFN-β were assessed. Our results indicate that MSP does not inhibit the serine phosphorylation of IRF3 in response to LPS (Fig. 3A) or the upregulation of IFN-β as assessed by RT-PCR (Fig. 3B) and ELISA (Fig. 3C).

The TLR4-dependent activation of MAPK signaling promotes the expression of many LPS-dependent genes. Alternatively, recent studies demonstrate that hyperactivation of the ERK signaling pathway by LPS can inhibit IL-12p40 expression through the activation of a repressor element in the IL-12p40 promoter (22). LPS

FIGURE 2. Tyrosines in the C-terminal tail of Ron mediate the inhibition of LPS-induced IL-12p40 production in primary macrophages. A, Bone marrow-derived macrophages were isolated from primary murine bone marrow by adherence to plastic. The cells were cultured in L929 cell conditioned media and infected with murine stem cell virus-based retroviral vectors harboring wild-type or mutant forms of Ron. The cells were then treated overnight with 100 ng/ml LPS followed by stimulation with 100 ng/ml LPS for 24 h. Supernatants were collected, and IL-12p40 levels were assessed by ELISA. B, Resident peritoneal macrophages from wild-type or Grb2^{−/−} animals were cultured in the presence or absence of MSP for 12 h followed by stimulation with 100 ng/ml LPS for 24 h. IL-12p40 levels in the supernatants were assessed by ELISA. *p < 0.05; **p < 0.01.

FIGURE 3. MSP does not inhibit MyD88-independent signaling by TLR4. A, Primary resident peritoneal macrophages were isolated by lavage and adherence to plastic. The cells were treated with 100 ng/ml MSP overnight followed by stimulation with 100 ng/ml LPS. Cell lysates were prepared and IRF3 phosphorylation was assessed by Western blot analysis with a phosphospecific IRF3 Ab. The blot was stripped and reprobed with anti-IRF3. B, Primary resident peritoneal macrophages were pretreated with MSP and stimulated with 100 ng/ml LPS for 6 h. RNA was extracted and expression of IFN-β was assessed by RT-PCR. C, Primary resident peritoneal macrophages were pretreated with 100 ng/ml MSP overnight and then stimulated with 250 ng/ml LPS for the indicated times. Cell culture supernatants were collected at the indicated times, and IFN-β production was assessed by ELISA.
stimulation of TLR4 induces MAPK signaling through the recruitment and activation of the adaptors MyD88 and Mal, and the upregulation of Tak1 kinase activity. We have shown previously that MSP induces robust phosphorylation of ERK in primary peritoneal macrophages (13); however, the effect of MSP on the induction of MAPK signaling in response to LPS has not been assessed. To address this question, we stimulated primary peritoneal macrophages with MSP followed by LPS stimulation. Phosphorylation of ERK, p38, and JNK was assessed at various times after LPS stimulation (Fig. 4A). Results from these studies indicate that LPS induces the phosphorylation of ERK, p38, and JNK in primary macrophages, and this induction is not inhibited by MSP. In fact, we consistently observed an increase in the phosphorylation of these MAPK family members in the presence of MSP.

MAPK signaling promotes inflammatory gene expression, at least in part, through the induction of the transcription factor AP-1. To determine whether Ron likewise induces AP-1 transcriptional activity downstream of LPS/TLR4, we used human embryonic kidney 293 (HEK293) cells stably transfected with TLR4 and MD-2. These cells were transiently transfected with CD14, with or without Ron, and an AP-1 luciferase reporter. The cells were stimulated with or without LPS and luciferase activity was assessed (Fig. 4B). As we have observed previously, Ron induces ligand-independent AP-1 reporter activity in these cells (23, 24). LPS stimulation of TLR4 also induced AP-1 reporter activity, and the combination of Ron and LPS induced levels of AP-1 activity greater than those observed with either Ron or LPS alone. These results indicate that Ron does not inhibit, but rather enhances, LPS-induced MAPK signaling and the upregulation of AP-1 transcriptional activity induced by LPS.

MSP inhibits LPS-induced activation of IKK downstream of Tak1

We have shown previously that Ron inhibits NF-κB activity in primary human monocyte-derived macrophages and RAW264.7 cells transfected with exogenous Ron (6, 15). However, the effect of endogenous Ron expressed in primary peritoneal macrophages on NF-κB signaling and the level at which this regulation occurs have not been assessed. Activation of Tak1 kinase in response to TLR4 mediates the activation of both the MAPK and NF-κB signaling pathways. Therefore, we assessed the effect of MSP on the phosphorylation of Tak1 (Fig. 5A). Consistent with the failure of MSP to inhibit MAPK signaling in response to LPS, we failed to detect a reduction in Tak1 phosphorylation by LPS in the presence of MSP. To determine whether Ron regulates IKK activity induced by LPS in primary macrophages, we used thioglycolate-elicited macrophages. We have confirmed that these cells express Ron, and that MSP inhibits LPS-induced IL-12p40 expression in these cells (data not shown). Elicited macrophages were stimulated with LPS in the presence or absence of MSP, and IKK activity was assessed by in vitro kinase assay using IκB as a substrate (Fig. 5B).

We observed a significant upregulation of IKK activity within 15 min of LPS stimulation in these cells. However, this induction was significantly reduced in the presence of MSP. IKK activity returned to basal levels by 60 min in both the presence and absence of MSP.

IKK phosphorylation of cytoplasmic IκB family members results in the proteasome-mediated degradation of IκB. We next examined IκBα protein levels in primary resident macrophages stimulated with LPS in the presence or absence of MSP (Fig. 5C). We observed a reduced rate of IκB degradation in the presence of MSP; however, by 15 min, IκB protein was undetectable in LPS-stimulated cells in either the presence or absence of MSP. To examine the effect of MSP on LPS-induced NF-κB DNA binding, we extracted nuclear lysates from elicited macrophages stimulated with LPS in the presence or absence of MSP. DNA binding of p65 from nuclear lysates was assessed by ELISA (Fig. 5D). Consistent with the IKK and IκB results, we observed a significant reduction in the DNA binding of NF-κB at 15 min after LPS stimulation in the presence of MSP. These results support our previous observations and suggest that MSP delays the activation of NF-κB signaling through the canonical pathway after stimulation of primary macrophages with LPS.

Ron inhibits serine phosphorylation of p65 and NF-κB transcriptional activity

In addition to its role in the phosphorylation of IκB, the IKK complex also promotes phosphorylation of p65 resulting in enhanced transcriptional activity. To determine whether Ron regulates NF-κB transcriptional activity, we used HEK293 cells stably expressing TLR4 and MD-2. These cells were transiently transfected with CD14 and an NF-κB luciferase reporter in the presence of increasing levels of Ron, and luciferase activity was assessed (Fig. 6A). In this system, we observed a significant reduction in NF-κB–dependent reporter activity in response to LPS in the presence of Ron in a dose-dependent manner. Similar results were observed with both murine and human Ron (data not shown).

Phosphorylation of serine 536 on p65 by IKK enhances its transcriptional activity by promoting its association with the transcriptional activators CREB-binding protein/p300 and a member of the basal transcriptional machinery, TAFII31 (25, 26). To determine whether Ron regulates serine phosphorylation of p65 in these cells, HEK293 cells expressing TLR4 and MD-2 were transiently transfected with or without Ron and stimulated with LPS.

**FIGURE 4.** MSP enhances MAPK signaling and AP-1 activity in LPS-stimulated cells. A. Primary resident peritoneal macrophages were stimulated with 100 ng/ml overnight followed by 100 ng/ml LPS. Cell lysates were extracted at the indicated times, and phosphorylation of ERK, p38, and JNK were assessed by Western blot analysis with phosphospecific Abs. Blots were stripped and reprobed with anti-ERK, anti-p38, and anti-JNK. B. 293 cells stably transfected with TLR4 and MD-2 were transfected with pcDNA3.1 empty vector or pcDNA3.1 Ron-HA, a CD14 expression vector, and an AP-1 luciferase reporter construct. After an overnight transfection, the cells were treated with or without 100 ng/ml LPS for 4 h and luciferase activity was measured.
Nuclear extracts were isolated, and phosphorylation of p65 at serine 536 was assessed by Western blot analysis using a phosphospecific Ab (Fig. 6B). In these cells, we observed a decrease in serine phosphorylation of p65 induced by LPS in the presence of MSP. These results indicate that Ron regulates the transcriptional activity of NF-κB at the level of posttranslational modification, by limiting serine phosphorylation of p65. Consistent with these results, inhibition of p65 serine phosphorylation was also seen in primary peritoneal macrophages within 15 min of LPS stimulation (Fig. 7B).

Unlike conventional IκB family members, nuclear IκBs contain a nuclear localization signal and reside primarily in the nucleus, rather than the cytoplasm. These IκB family members exhibit both positive and negative effects on NF-κB-dependent transcriptional activation (27). IκBζ is a nuclear IκB family member that is upregulated at the transcriptional level by LPS in an NF-κB-dependent manner (28). IκBζ-deficient macrophages display impaired expression of a subset of LPS-inducible genes including IL-12p40 (29). To examine the effect of MSP on the induction of early NF-κB response genes, we assessed the expression of IκBζ in response to LPS in the presence or absence of MSP in primary peritoneal macrophages (Fig. 7A). Results from these studies demonstrate a significant reduction in LPS-induced IκBζ expression in the presence of MSP.

**FIGURE 5.** MSP delays IKK activity, IκB degradation, and NF-κB binding after LPS stimulation. A, Resident peritoneal macrophages were stimulated with MSP overnight followed by stimulation with 100 ng/ml LPS for the indicated times. Tak1 phosphorylation was assessed by Western blot analysis using a phospho-specific Tak1 Ab. B, Thioglycolate-elicited macrophages were isolated by adherence and stimulated with 100 ng/ml MSP overnight followed by 100 ng/ml LPS. Cell lysates were harvested at the indicated times, cell lysates were extracted and IKK activity was measured by in vitro kinase assay using IκB as a substrate. C, Primary resident peritoneal macrophages were stimulated overnight with 100 ng/ml MSP followed by 100 ng/ml LPS. Cell lysates were extracted at the indicated times, and IκBα levels were assessed by Western blot analysis. The blots were stripped and reprobed with anti-actin. D, Thioglycolate-elicited macrophages were isolated by adherence to plastic and stimulated with 100 ng/ml MSP overnight followed by 100 ng/ml LPS. Nuclear extracts were isolated at the indicated times, and binding to DNA was assessed by ELISA. *p < 0.01.

**FIGURE 6.** Ron inhibits NF-κB transcriptional activity and p65 serine phosphorylation in response to LPS. A, HEK293 cells expressing TLR4 and MD-2 were cotransfected with the same empty vector or HA-tagged Ron and an NF-κB luciferase reporter. The transfected cells were stimulated with 100 ng/ml LPS and luciferase activity was assessed. Lysates were also analyzed by Western blot using an anti-HA Ab. B, HEK293 cells expressing TLR4 and MD-2 were transfected with or without Ron and stimulated with 100 ng/ml LPS. Nuclear lysates were isolated and phosphorylation of p65 at serine 536 was assessed by Western blot analysis using a phosphospecific Ab. Blots were stripped and reprobed with anti-p65 and anti-Sp1.

**FIGURE 7.** MSP inhibits NF-κB transcriptional activity and delays serine phosphorylation of p65 in primary macrophages. A, Primary resident peritoneal macrophages were harvested by lavage and isolated by adherence to plastic. The cells were stimulated with 100 ng/ml MSP overnight followed by 100 ng/ml LPS. Cell lysates were harvested at the indicated times, and IκBζ expression was assessed by Western blot analysis. The blots were stripped and reprobed with anti-actin. B, Primary resident peritoneal macrophages were stimulated with LPS for the indicated times in the presence or absence of MSP. Serine phosphorylation of p65 was assessed by Western blot analysis.

**FIGURE 8.** LPS induces expression of Ron and Sf-Ron in primary macrophages. Primary resident peritoneal macrophages were stimulated for the indicated times with 100 ng/ml LPS. Expression of Ron and Sf-Ron were assessed by real-time PCR. *p < 0.001 for all values relative to the zero time point for each gene.
regulation of macrophage activation (30). To determine whether LPS regulates the expression of Ron and/or Sf-Ron in macrophages, we stimulated primary peritoneal macrophages with LPS and examined Ron and Sf-Ron expression by real-time PCR (Fig. 8). Results from these studies indicate that LPS induces expression of both Ron and Sf-Ron in primary macrophages. These results are consistent with a recent report describing two NF-kB sites in the promoter of Ron that promote its expression (31), and suggest that the upregulation of Ron by LPS could serve as a negative feedback signal to limit subsequent LPS stimulation of primary macrophages.

Discussion
Previous studies from our laboratory indicate that the MSP/Ron signaling pathway profoundly impacts the expression of IL-12p40 both in vitro and in vivo, rendering Ron−/− mice more susceptible to septic shock (12, 13). MSP stimulation of the RON receptor also inhibits the expression of iNOS and cyclooxygenase-2 in activated macrophages (5, 6, 32). However, our previous studies also demonstrated that, whereas the production of TNF-α is inhibited in activated macrophages by MSP, IL-1β and IL-10 production by these cells is unaffected by MSP, and IL-6 production by these cells is enhanced in the presence of MSP (12). Taken together, these studies indicate that Ron differentially regulates LPS-dependent gene expression. In this study, we demonstrate that MSP stimulation of Ron does not inhibit LPS-induced TLR4 signaling at a receptor proximal level, but rather inhibits TLR signaling at the level of IKK activity. These results are consistent with a role for the RON receptor in shaping the repertoire of LPS-dependent responses in macrophages.

The data presented in this study indicate that the ability of MSP to inhibit LPS signaling in primary macrophages is enhanced significantly when the cells are cultured overnight in MSP before LPS stimulation. These results suggest that the ability of Ron to promote new protein synthesis plays a central role in the inhibition of LPS signaling by MSP. We have shown previously that MSP stimulation of the RON receptor promotes STAT3 tyrosine phosphorylation in resident macrophages (13). Our studies suggest that the activation of STAT3 by Ron occurs via the recruitment of a Grb2/Gab2 signaling complex to Ron, and the subsequent recruitment of STAT3 to the complex through a YxxQ motif present in Gab2 (33). STAT3 promotes a gene expression program, the products of which inhibit proinflammatory mediator induction. For example, Stat3 induces a gene expression program, the products of which inhibit proinflammatory mediator induction. 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Disclosures
The authors have no financial conflicts of interest.

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