Secretion by Dendritic Cells Is Required for CD154-Dependent IL-12 Secretion by Dendritic Cells

Marie Tourret, Sarah Guégan, Karine Chemin, Stéphanie Dogniaux, Francesc Miro, Armelle Bohineust and Claire Hivroz

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References

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T Cell Polarity at the Immunological Synapse Is Required for CD154-Dependent IL-12 Secretion by Dendritic Cells

Marie Tourret,1 Sarah Guégan,1,2 Karine Chemin, Stéphanie Dogniaux, Francesc Miro,3 Armelle Bohineust, and Claire Hivroz

Ag-specific interaction between T lymphocytes and dendritic cells (DCs) leads to both T cell and DC activation. CD154 (CD40 ligand)/CD40 interactions have been shown to play a major, although not exclusive, role in this functional cross-talk. Interactions between T cells and DCs are structured by an immunological synapse (IS), characterized by polarization of the T cell microtubule cytoskeleton toward the interacting DCs. Yet the role T cell polarization may play in T cell-induced DC activation is mostly unknown. In this study, we address the role of T cell polarity in CD154-dependent activation of DCs in a human model, using two different tools to block T cell polarity (i.e., a microtubule depolymerizing drug and an inhibitor of atypical protein kinase C). We show that CD154 is recruited and concentrated at the IS formed between human primary T cells and autologous DCs and that this recruitment requires T cell polarity at the IS. Moreover, we show that T cell polarization at the IS controls T cell-dependent CD154–CD40 signaling in DCs as well as CD154-dependent IL-12 secretion by DCs. This study shows that T cell polarity at the IS plays a key role in CD154/CD40-dependent cross-talk between CD4+ T cells and DCs. The Journal of Immunology, 2010, 185: 6809–6818.

The online version of this article contains supplemental material.

Materials and Methods

Reagents and Ab

The media used are the following: RPMI 1640 Glutamax, 1% pyruvate, 100 U/ml penicillin, 100 μg/ml streptomycin (Invitrogen, Carlsbad, CA), and 10% FCS (Biowest, Miami, FL). Recombinant human IL-4 and GM-CSF were from Brucells (Brussels, Belgium). Recombinant bacterial superantigens were from Toxin Technology ( Sarasota, FL). Mouse mAbs against human CD4, CD69, CD40, CD80, CD83, CD86, CD154, and isotypic controls coupled to fluorochromes were from BD Biosciences (San Jose, CA). Rabbit polyclonal Abs against human Lamp1 and Lamp2 were from Abcam (Cambridge, MA). Mouse mAbs against human CD3ε UCHT1 and CD43 were a gift, respectively, from D. Cantrell (University of Dundee, Dundee, U.K.) and F. Sanchez-Madrid (Autonomous University of Madrid, Madrid, Spain). Rabbit Ab against human Rab 6 was from Santa Cruz Biotechnology (Santa Cruz, CA), rat anti–ε-tubulin was from Serotec (Oxford, U.K.), and Colxx and brefeldin A were from Sigma-Aldrich (St.

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Louis, MO). The myristoylated protein kinase Cζ-pseudosubstrat was from Invitrogen. The agonist anti-human CD40 mAb was a gift from Y. Richard (Service of Immuno-Virology, Commissariat à l’Energie Atomique, Unité Mixte de Recherche-E1, Université Paris-Sud, Orsay, France), and an IgG1 mouse anti-hemaggulatinin (HA) mAb was used as a control. Secondary Abs used for microscopy (anti-rat, rabbit, and mouse Ig) coupled to Alexa Fluor 568 were from Invitrogen.

Constructs and transfection of primary T cells

Human CD154 cDNA followed by a short gly-gly-gly-ser-gly-gly-gly-gly linker was subcloned into the pcEGFP-C1 vector (BD Clontech, Palo Alto, CA) between the Xhol and BamHI restriction site. Human fresh PBMCs were transfected with the Amaxa Nucleofector technology (Amaxa, Köln, Germany), according to the manufacturer’s instructions, and were used 18 h after transfection.

Cells

DCs were generated from human monocytes of healthy donors as described previously (30). CD4+ T cells were negatively selected from PBMCs, after depletion of CD14+ cells, using the T cell isolation kit II from Miltenyi Biotec (Auburn, CA). Sorted CD4+ T cells were 97–99% CD4+/CD3ε. PBMCs were obtained from a patient presenting with CD154 mutation, resulting in a complete defect in CD154 expression (31). This study was conducted according to the Helsinki Declaration, with informed consent obtained from the patient’s family, as requested by our Institutional Review Board.

Colx pretreatment of T cells

Purified CD4+ T cells were treated with 50 μg/ml Colx for 5 min at 37°C, extensively washed, and cultured for 1 h in complete medium to get rid of the excess of Colx. T cells were then washed again and used in cocultures.

Cytokine detection

Cytokine secretion was measured in the supernatants of DC/T cells cocultures performed in 96 round-bottom well plates by ELISA using matched paired Abs specific for IL-12p70 (DuoSet; R&D Systems, Minneapolis, MN). A cytometric bead array (BD Biosciences) was used to measure IL-8 and IL-6, produced in the cocultures.

Flow cytometric analysis of T cell viability formation and T/DC conjugate

For T cell viability, T cells were left untreated or treated with Colx, or the myristoylated protein kinase Cζ-pseudosubstrat DAPI was added at 0.5 μM just before acquisition on a MACSQuant analyzer (Miltenyi Biotec). For the analysis of T/DC conjugates, cells were cocultured in 96 round-bottom well plates at a 2:1 ratio, washed twice in PBS and BSA 0.5% without EDTA, labeled with anti-CD4 and anti-CD1a mAbs at 4°C, and washed twice again in PBS and 0.5% BSA. Stained cells were then acquired on a FACSCalibur (BD Biosciences) without prior vortexing.

Flow cytometric analysis of CD154 expression

For the surface mobilization assay, CD4+ T cells incubated with a 1:10 dilution of PE-labeled diazylated anti-CD154 (clone TRAP1; BD Biosciences) or control IgG1 were cocultured with DCs in the presence or absence of superantigens. After different coculture times, cells were fixed with 3% formaldehyde (T Pharmingen) and permeabilized with glass coverslips mounted onto glass slides using Fluoromount G (Southern Biotechnology Associates, Birmingham, AL). Images were acquired with a wide-field Eclipse 90i Upright Microscope (Nikon, Melville, NY) equipped for image deconvolution. Acquisition was performed using a ×100 Plan Apo VC 1.4 Oil objective and a highly sensitive cooled interline charge-coupled device camera (Roper CoolSnap HQ2). Z-positioning was accomplished by piezoelectric motor (linear variable differential transformer; Physik Instrument, Karlsruhe, Germany), and a Z-series of images was taken every 0.2 μm. After deconvolution, images were segmented with the multidimensional image analysis interface running under MetaMorph (Universal Imaging, Ypsilanti, MI) based on wavelet decomposition 17. Images were analyzed with Metamorph and ImageJ software. Pearson’s coefficient was quantified using the JACOP plugin from ImageJ (32).

Quantification of MTOC polarization and CD154 and CD3ε enrichment at the IS

MTOC polarization at the IS was scored blinded. Cells were analyzed by three-dimensional microscopy, with the same conditions used for control and drug-treated T cells. The proportion of conjugates with MTOC recruited at the immune synapse was calculated by random selection of >50 conjugates selected from transmission images from three independent experiments. Three-dimensional maximal projections of the centrin or α-tubulin labeling were visualized to determine the MTOC recruitment at a z-stack projection of the region of cell–cell contact. The MTOC was scored as polarized when it was juxtaposed to the contact area with the DCs.

CD154- or CD3-associated fluorescent signals were quantified using the Metamorph software. Detectors were set to detect an optimal signal below the saturation limits. Image sets to be compared were acquired during the same session and using the same acquisition settings. Quantification of the recruitment of CD154 at the IS: average fluorescence intensity of CD154 in a fixed region of the synapse was divided by the average total fluorescence intensities inside T cells measured on three-dimensional projections of the CD154 deconvoluted images (Supplemental Fig. 1A, 1B).

CD154 expression was quantified by three-dimensional microscopy, with the ×40 or ×100 1.4 NA oil immersion objective. Coverslips covered with T cells were placed into a chamber on the microscope at 37°C in a 5% CO2 atmosphere. At time 0, superantigen-loaded or untreated DCs were added, and after 20 s, one-phase contrast image and a stack of z-planes (step 0.3 μm) were collected with the green filter set. Movies consisting in the phase contrast image and the best focus plan of GFP-CD40L were accelerated ×90.

Results

CD154 is recruited at the IS formed between CD4+ human T cells and DCs

CD40L (CD154) expression by both murine and human CD4+ has been shown to play a crucial role in T cell-induced DC activation (1–3, 30, 33, 34). Moreover, others and we have shown that the IS formation plays a key role in T/DC cross-talk (30, 35–38). We thus examined the distribution of CD154 at the IS formed between human monocyte-derived DCs and autologous CD4+ T cells after 6 h of contact. No CD154 labeling was detected in T cells cocultured with DCs alone (data not shown), whereas in T cells for-
ming conjugates with superantigen-pulsed T cells, CD154 was observed at the IS “en face” of the DC in 74% of the CD4+ T cells forming conjugates (Fig. 1A–E and data not shown). CD154 did not colocalize with LAMP1 (Fig. 1A; Pearson coefficient [p], quantification on 15 images 1F; p = 0.085) or LAMP2 (Fig. 1B; quantification on 15 images 1F; p = 0.170)-positive lysosomal compartments in T cells forming an IS but mainly colocalized with the Golgi, trans-Golgi marker Rab6 (Fig. 1C; quantification on 15 images 1F; p = 0.440). Moreover, CD154 did not colocalize with the dSMAC marker CD43 (Fig. 1D; quantification on 15 images 1F; p = 0.060), which is excluded from the IS but colocalized with CD3ε at the center of the synapse (Fig. 1E; quantification on 15 images 1F; p = 0.757).

These results show that CD154 is recruited at the IS formed between human CD4+ T cells and DCs and cosegregates with CD3ε.

CD154 recruitment at the IS formed between CD4+ T cells and DCs requires T cell polarity at the synapse

T cell MTOC polarization at the IS induces a reorientation of secretion of lymphokines (19–21), lytic granules (22, 23), and transmembrane proteins (39, 40) at the synapse. We thus asked whether

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**FIGURE 1.** CD154 localization in human CD4+ T cells forming synapse with autologous DCs. DCs and T cells were incubated in the presence of a superantigen mixture on poly (L-lysine)-coated coverslips during 6 h and observed by three-dimensional microscopy. A. Deconvoluted images are shown. White dotted lines indicate the position of T cells. Three-dimensional reconstruction of the interface (“en face view”) revealed that CD154 is clustered at the center of the synapse wherein CD43 is excluded and that CD3ε and CD40L cosegregate in this central zone. The Pearson’s coefficients (p) calculated for the shown merge images are encrusted in the image. Original magnification ×100. Scale bars, 5 μm. F. Pearson’s coefficients (p) calculated for each pair of markers.
T cell polarity was required for CD154 recruitment. To answer this question, we pretreated human primary CD4+ T lymphocytes with Colx, which blocks polymerization of microtubule. Polarization of T cell MTOC toward superantigen-pulsed DCs was blocked by this pretreatment (Supplemental Fig. 2A) but did not significantly affect T cell viability (Supplemental Fig. 2B), T cell activation as measured...
by CD69 expression (Supplemental Fig. 2C), or conjugate formation (Supplemental Fig. 2D). These results showed that interaction between Colx-treated T cells and DCs did occur and were productive in terms of T cell activation. We then controlled CD154 expression by T cells pretreated with Colx. Conventional methods for surface staining of CD154 at the surface are inefficient because of its high endocytosis and degradation rates (41); we thus adapted a method reported earlier (42, 43) to follow CD154 surface expression. Fluorescently conjugated Ab against CD154 or isotypic controls were added for the duration of the cocultures, allowing the accumulation of fluorescence in cells. Specificity of the labeling was controlled with T cells from a patient presenting with a complete defect in CD154 expression (Fig. 2A, lowest right panel). Untreated and Colx-treated CD4⁺ T cells cultured alone (data not shown) or with DCs showed a low, as reported by others (43, 44), but comparable surface expression of CD154.

In this test, however, addition of superantigen in the cocultures did not significantly increase CD154 surface expression. This was probably due to the instability of the CD154 Ab reported by others (42) that precluded accumulation of the complexes inside the cells. We thus measured the intracellular total amount of CD154 produced by T cells in the presence of brefeldin A, which blocks protein secretion thus allowing its accumulation. As shown in Fig. 2B, Colx neither modified the basal production of CD154 by T cells (Fig. 2B), nor the superantigen-induced CD154 expression confirming that T cell activation was not inhibited by Colx treatment (see Supplemental Fig. 2C for CD69 expression).

Knowing that CD154 was normally expressed by Colx-treated T cells, we then studied the effect of Colx on CD154 enrichment at the IS. Colx-treated T cells showed a scattered distribution of the CD154 labeling by fluorescence microscopy. Moreover, whereas the ratio between the synaptic CD154 fluorescence intensity and the total fluorescence CD154 intensity was ≥1 in untreated CD4⁺ T cells, this ratio was ≤1 in Colx-treated T cells revealing that Colx treatment inhibited CD154 recruitment at the IS (Fig. 2C and quantification in Fig. 2D). In contrast, an enrichment of CD3e at the IS was still observed in Colx-treated T cells (Fig. 2C and quantification in Fig. 2E), showing, as demonstrated by us before (15), that CD3e is still recruited at the IS in unpolarized T cells. This result is different from what has been reported by Das et al. (45), who showed a reduced accumulation of TCR in 30 min synapses formed between Colx-treated Jurkat T cells and superantigen-pulsed Raji B cells. Yet, this discrepancy may be due to the cells and kinetic used in this study (i.e., primary T cells and DCs and 6 h of contact).

We then studied the kinetic of recruitment of CD154 at the IS by introducing a GFP–CD154 chimeric molecule in primary CD4⁺ T cells. As shown in Fig. 3A and Supplemental Movie 1, CD154 was present both at the plasma membrane and in intracellular compartments of T cells. Concentration of CD154 at the IS was observed 3 min after contact and lasted for the whole length of the movie (20 min). In contrast, no recruitment of the GFP–CD154 chimeric molecule was observed in Colx-treated CD4⁺ T cells (Fig. 3B, Supplemental Movie 2).

Altogether, these results demonstrate that CD154 is rapidly polarized and recruited at the center of the IS formed between human CD4⁺ T cells and DCs and that this recruitment requires microtubule polymerization in T cells.

T cell-induced IL-12 secretion by DC requires microtubule polymerization in T cell

The role T cell MTOC polarization plays in T cell/APC cross-talk is mostly unknown. We addressed in this study the role it might play in T cell-induced DC activation. We have previously shown that CD4⁺ T cells induce increased expression of CD40, CD80, CD83, and CD86 in monocyte-derived DCs in the presence of superantigen (30). We thus analyzed the phenotype of DCs cocultured with untreated or Colx-treated T cells. Increased expression of CD40, CD80, CD83, and CD86 by DCs was equivalent when induced by polarized and unpolarized T cells (Fig. 4A). Moreover, Colx-treated T cells and untreated T cells also induced equivalent secretion of IL-6 and IL-8 (Fig. 4B, 4C) by DCs demonstrating that unpolarized T cells can induce DC activation. In contrast, Colx-
treated T cells induced significantly less IL-12p70 production by DCs than untreated CD4+ T cells (Fig. 4D), suggesting that polarization of T cells at the IS is required for T cell induced secretion of IL-12 by DCs.

CD154–CD40 signaling in DCs leading to IL-12 production requires microtubule polymerization in T cells. We have shown in this study that microtubule polymerization in T cells controls CD154 recruitment in the contact zone with DCs and IL-12 secretion by DCs. We thus hypothesized that the polarity-dependent CD154 recruitment at the IS is required for CD154/CD40

The T cell-dependent induction of IL-12p70 secretion by DCs requires microtubule polymerization in T cells. DCs (5 × 10^4) were cultured for 20 h with or without untreated or Colx-treated CD4+ T cells (10^5) and the superantigen (sAg) TSST-1 (100 ng/ml). A, Flow cytometric analysis of DC maturation markers. Histograms of a representative experiment. For each marker, expression was plotted as a ratio between the median fluorescence intensity obtained in the various conditions and the median fluorescence intensity measured on DCs alone (fold increase). White bar, DCs alone; black bar, DC+TSST-1+T; and gray bar, DC+TSST-1+Colx–treated T cells. IL-6 and IL-8 secretion (B, C) and IL-12p70 secretion (D) were measured by cytometric bead array and ELISA, respectively, in the supernatants from individual donors, and each circle represents an individual donor (black, untreated; gray, Colx). Significant differences between the groups were assessed by paired t test (IL-6 and IL-8, p = NS; IL-12p70, p = 0.003).

FIGURE 5. CD154–CD40 signaling in DCs leading to IL-12 production requires microtubule polymerization in T cells. A, DCs were incubated with untreated T cells or Colx-treated CD4+ T cells in the presence of a superantigen (sAg) mixture. Anti-CD40 mAb or anti-HA at 3 μg/ml were added during the 2-h cocultures. Expression of phospho-Stat3 in DCs was measured by flow cytometry. B, DCs were cultured for 20 h with or without untreated (black histograms) or Colx-treated CD4+ T cells (gray histograms) and sAgs. A total of 3 μg/ml anti-HA or anti-CD40 mAb were added at the beginning of the coculture. IL-12p70 secretion was measured by ELISA. One representative experiment of three is shown in A and B.
suggest that unpolarized T cells, although expressing CD154 (Fig. 2B, 2C) are unable to induce CD154/CD40 signaling in DCs. We then studied whether the anti-CD40 agonist mAb, which restores CD154/CD40 signaling in DCs, could also restore IL-12 production by DCs activated by unpolarized T cells. Indeed, addition of the anti-CD40 mAb but not of the control anti-HA mAb restored the IL-12 secretion by DCs to levels that were induced by untreated CD4+ T cells (Fig. 5B).

Colx treatment does not only block MTOC polarity but also destroys the microtubule network in T cells. To demonstrate that MTOC polarity was indeed required for CD154-dependent signaling in DCs, we inhibited atypical PKCs, which have been shown to control the polarity of cells (27) including T cells (28, 29), and tested the effect of this inhibition on CD154-dependent IL-12 production by DCs. A cell-permeable myristoylated pseudosubstrate of atypical PKCs was added to the T cell/superantigen-pulsed DCs cocultures. This inhibitor used at 12.5 μM did not affect significantly T cell viability (Supplemental Fig. 3A), T cell activation as measured by CD69 expression (Supplemental Fig. 3B), or conjugate formation measured by flow cytometry (Supplemental Fig. 3C), showing that T/DC contacts were still productive. In contrast, polarization of T cell MTOC toward superantigen-pulsed DCs was inhibited (Fig. 6A and quantification in Fig. 6B), whereas T cell microtubule network (Fig. 6A) was not affected. This absence of T cell MTOC polarity was accompanied by an inhibition of the recruitment of CD154 at the IS (Fig. 6A and quantification in Fig. 6C). The atypical PKC pseudosubstrate, when added to DC/superantigen/CD4+ T cell cocultures, inhibited IL-12 production by DC (Fig. 6D, 6E). Washing away the atypical PKC pseudosubstrate, which unlike Colx is a reversible inhibitor, partially restored IL-12 production by DC (Fig. 6E), showing that T cells after removal of the inhibitor were still able to induce IL-12 production by DCs. Moreover, as shown above for Colx treatment (Fig. 5B), addition of the anti-CD40 agonist mAb to cocultures containing the atypical PKC inhibitor restored the IL-12 secretion by DCs to levels that were induced by untreated CD4+ T cells, showing that atypical PKC pseudosubstrate that inhibits T cell MTOC polarity and CD154 recruitment at the IS inhibits CD154-dependent IL-12 production by DCs.

Altogether, our results demonstrate that nonpolarized T cells, which show no CD154 recruitment at the synapse, do not induce IL-12 secretion by DCs because of defective CD154-dependent signaling in DCs.

Discussion

Polarization of T cells toward the interacting APCs has been described more than 20 y ago (11). Yet, the role this polarization may play in the immune response is not fully characterized. In CD8+ T cells, MTOC polarization controls the directional secretion of cytotoxic granules toward the target cell (reviewed in Ref. 49). In CD4+ T cells, MTOC polarization coincides with accumulation of some cytokines at the IS (19, 21). However, the role T cell polarity may play has not been envisaged on the APC’s point of view. We demonstrate in this paper that T cell polarity is required for CD154-dependent IL-12 secretion by DCs. Indeed, T cell polarity controls correct CD154 recruitment and concentration at the IS, which are themselves required for CD154/CD40 signaling in DCs. We have investigated in this study the role of CD154 clustering at the IS in T/DC cross-talk. However, CD40–CD154 interactions do not only regulate cross-talk between T cells and DCs but also T-B

FIGURE 6. An aPKC inhibitor blocks T cell polarity, CD154 enrichment at the IS, and CD154–CD40-dependent IL-12 production by DCs. A, DCs were incubated with T cells and superantigens (sAgs) in the absence or presence of 12.5 μM of an aPKC inhibitor on coverslips for 6 h. Cells were fixed permeabilized and stained with α-tubulin (in red) and anti-CD154 (in green). Deconvoluted images. Original magnification ×100. Scale bar, 5 μm. B, MTOC polarity at the IS was scored blindly in T cells in contact with DCs. Percentages of the T cells with a polarized MTOC are shown. C, Enrichment of CD154 in the IS was measured as a ratio of the CD154 fluorescence intensity in the synapse versus the total CD154 fluorescence in the cell. D and E, DCs, sAgs, and autologous CD4+ T cells were cocultured for 20 h with a mixture of sAgs in the absence or presence of 12.5 μM aPKC inhibitor (aPKC inh). D, A total of 3 μg/ml anti-CD40 mAb were added at the beginning of the coculture (gray histograms). IL-12 secretion was measured by ELISA. E, IL-12 secretion is expressed as a percentage of the maximum IL-12 secretion (black, no inhibitor; dashed, aPKC inh). In the right part of the histograms, cocultures were washed twice with medium and left in culture for an additional 12 h. One representative experiment of three (A–D) and two for the “washing” experiments.
cooperation. CD154 has major effects on B cell activation by regulating B cell proliferation, differentiation, and Ig production (reviewed in Ref. 50). This pivotal role of CD40–CD154 in the generation of efficient immune responses in humans is highlighted by the fact that patients presenting with mutations in CD154 develop defects of humoral immunity. Indeed, CD154 deficiency results in the X-linked hyper-IgM syndrome, a severe immunodeficiency characterized by low or absent IgG and IgA (reviewed in Ref. 51). CD154 enrichment has already been described at the IS formed in vitro between a mouse CD4+ T cell line and a B cell hybridoma presenting the right MHC-peptide (52) and in vivo in B/T cell IS formed after viral infection of primate brains (53). CD154-expressing T cells have also been shown to be present in germinal centers of human tonsils (54). Thus, polarization-dependent CD154 enrichment at the IS might be involved in CD154-dependent B cell activation.

What could be the mechanisms for the polarization-dependent enrichment of CD154 at the IS? Specialized endocytic compartments such as secretory lysosomes are polarized toward the APC and fused at the synapse (reviewed in Ref. 55). This contributes to the polarized delivery of transmembrane receptors such as CTLA-4 (25, 26) and Fas ligand (24), which are present in secretory lysosomes. Fusion of the secretory lysosomes at the IS has been shown to require both microtubule polymerization to drive these compartments toward the MTOC (55) and MTOC recruitment at the IS to enable fusion of the compartments (49). We have used in this study two different tools to block T cell polarity at the IS: inhibition of microtubule polymerization in T cells by treatment with Colx and inhibition of T cell MTOC polarity by treatment with a pseudosubstrate of atypical PKcs (aPKcs), which has been shown to inhibit T cell polarity (28, 29) but does not perturb microtubule network formation (Fig. 6). In both cases, CD154 was not enriched at the IS, yet when the microtubule network was absent (Colx treatment; Fig. 2C, Supplemental Fig. 1), CD154 endocytic compartments were scattered inside T cells, whereas when the microtubule network was present but not polarized (aPKC inhibitor; Fig. 6A) CD154 compartments were concentrated around the MTOC. These results suggest that recruitment of CD154 endocytic compartments and secretory lysosomes relies on similar mechanisms. Our results show that CD154 is rapidly recruited at the IS. The dynamic analysis of the GFP–CD154 recruitment at the IS suggests that two pools of CD154 are recruited: one that is at the plasma membrane and one that is intracellular. Because CD154 surface expression is low even in activated T cells (Fig. 2A), recruitment of the endocytic pool is probably crucial to provide a “bolus” of CD154 signaling. Yet, although CD154 was recently shown to be present in secretory lysosomes of mouse CD4+ and CD8+ T cells (43), we did not see any colocalization of CD154 with Lamp1 or Lamp2 in human CD4+ T lymphocytes. Alternatively, because CD154 is rapidly endocytosed (41), its enrichment at the IS may involve polarization of the recycling endosomal compartments toward the T cell–APC contact zone. Such a mechanism has been shown to contribute to the enrichment of TCRs, CD71, and the signaling molecule linker for activation of T cells at the IS (16, 45, 56). This polarization secretion of CD154 at the IS is probably crucial for CD154 signaling by nonactivated memory and naive T cells that express low levels of CD154 (43, 44) and in this study (Fig. 2A, compare CD154 labeling of nonactivated T cells and T cells from CD154-deficient patient). In these cells polarized recruitment of the endocytic pools of CD154 may increase CD154/CD40 signaling in the interacting APC. Requirement for CD154 clustering at the IS of T cell polarization, which itself requires antigenic stimulation of T cells (13–15), could thus secure the antigenic specificity of the CD154-dependent signal. Recruitment of CD154 at the IS might also account for its downregulation (41). Indeed, formation of the IS has been shown to facilitate TCR downregulation in the central zone of the synapse (57); the same may be true for CD154 that is also present in this central zone (Fig. 1D, 1E).

Several lines of evidence suggest that CD40 trimerization is required for initiating downstream signaling cascade (58–60). Clustering of CD154 at the IS may allow an efficient multimerization of CD40 leading to signaling in the interacting T cell. We show in this paper that CD154 clustering in the synaptic zone controls Stat3 phosphorylation induced by CD154/CD40 signaling. The signaling pathways induced downstream of CD40 engagement on DCs activate multiple genes important for DC function. The signaling pathways activated downstream of CD40 depend on JAK/Stat signaling (47) as well as the recruitment of TNFR-associated factor family of proteins, which are recruited by the cytoplasmic tail of CD40 (reviewed in Ref. 61). These signaling pathways have been shown to induce IL-12 production as well as upregulation of costimulatory molecules such as CD80 and CD86 (62). It was thus somehow surprising to observe normal expression of activation markers in DCs activated by unpolarized T cells. Yet, as shown previously in our model (30), CD154 expression by T cells is mandatory for IL-12 production by DCs but not for expression of the CD40, CD80, CD83, and CD86 activation markers, which are probably induced by the TNF-α produced in the cocultures (63). Thus, unpolarized T cells, which are activated as shown by their expression of CD69, can still modulate DC function as revealed by expression by DCs of maturation markers (CD80, CD86, and CD40) and inflammatory cytokines (IL-8 and IL-6). These results imply that, in poor polarizing conditions [i.e., low number of TCR ligands or pathological conditions such as tumoral environment wherein defective T cell polarization has been reported (64)], T cells can induce some activation of DCs but no IL-12 production. Thus, DCs that have been activated by unpolarized T cells are probably unable to induce Th1 responses because Th1 polarization has been shown to require IL-12–dependent signals (reviewed in Ref. 65). CD154–CD40 interactions have also been shown to provide antiapoptotic signal to DCs (66–68), to control cross-presentation of Ag by DCs (4), and to overcome peripheral T cell tolerance (69). The role T cell polarity and CD154 recruitment at the IS might play in these CD154-dependent functions should be investigated.

In conclusion, our results demonstrate that T cell polarity at the IS is required for CD154/CD40-dependent signaling in, and IL-12 secretion by, DCs, thus revealing a new role for CD4+ T cell polarity in functional T cell/DC cross-talk.

Note added in proof. While revising our manuscript, a study from Bertrand et al. (70) came out in The Journal of Immunology, which shows that PKCζ activation in T cells is required for the polarization of CD40L at the immunological synapse and the induction of IL-12 production by cognate DCs.

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Disclosures
The authors have no financial conflicts of interest.
References


SUPPLEMENTARY LEGENDS

Supplementary Movie 1: CD154 is rapidly recruited at the immunological synapse in untreated primary CD4+ human T cells. Images were acquired every 20 seconds for 20 min. This video corresponds to the still images shown in Fig. 3A.

Supplementary Movie 2: CD154 is not recruited at the immunological synapse in colchicine treated CD4+ human T cells. Images were acquired every 20 seconds for 20 min. This video corresponds to the still images shown in Fig. 3B.

Supplementary figure 1: Method of quantification of CD154 and CD3ε enrichment at the IS. A and B: Average fluorescence intensity of CD154 in a fixed region (a) of the synapse was divided by the average total fluorescence intensity inside T cells (b) measured on 3D projections of the CD154 deconvoluted images (A: non treated, B: colchicine treated T cells). C: Fluorescence intensity of CD3ε in a fixed region of the synapse (a) was divided by the mean of the average intensities measured in 3 regions of the same size at the plasma membrane outside of the synapse (b, c and d). These values were calculated on two intermediate z planes. CD154 and CD3 associated fluorescences were quantified using the Metamorph software.

Supplementary figure 2: Colchicine treatment blocks T cell polarization at the IS (A) but does not affect T cell viability (B), CD69 expression (C) and conjugate formation (D). (A) CD4+ human T cells were left untreated or pre-treated for 5 min with 50μg/ml of colchicine (Colx). T cells were incubated on coverslips with autologous DC alone or DC pulsed with a cocktail of superantigens (sAg) for 6h, fixed and labeled with an anti-centrin Ab. Percentage of T cells in conjugates presenting with a polarized MTOC are indicated in the histograms.
(B) CD4⁺ human T cells were left untreated or pre-treated with colchicine and left one hour in culture. Cells were then labeled with DAPI. (C, D) Untreated or Colx treated T cells were co-cultured overnight with DC alone or pulsed with 100ng/ml of superantigens and labeled with CD4, CD1a and CD69 mAbs (C) or CD4 and CD1a mAbs (D). Histograms showing the percentages of CD69⁺/CD4⁺ (C) and CD1a⁺/CD4high conjugates (D) in co-cultures containing untreated (black) or Colx-treated (grey) T cells are shown on the right (mean percentage of triplicates, representative experiment out of 5).

Supplementary figure 3: **Inhibitor of atypical PKC does not affect T cell viability (A), CD69 expression (B) and conjugate formation (C).** (A, B, C) CD4⁺ human T cells were left untreated or pre-treated for 1h with 12.5μM of the myristoylated protein kinase C-zeta pseudosubstrate. (A) Cells were then labeled with DAPI. (C, D) T cells were then co-cultured overnight with DC alone or pulsed with 100ng/ml of superantigens in the presence or absence of the inhibitor and labeled with CD4, CD1a and CD69 mAbs (B) or CD4 and CD1a mAbs (C). Histograms showing the percentages of CD69⁺/CD4⁺ (B) and CD1a⁺/CD4high conjugates (C) in co-cultures in the absence (black) or presence of the atypical PKC inhibitor (grey) are shown on the right (mean percentage of triplicates, representative experiment out of 3).