Generation and Differentiation of IL-17–Producing CD4+ T Cells in Malignant Pleural Effusion

Zhi-Jian Ye, Qiong Zhou, Yong-Yao Gu, Shou-Ming Qin, Wan-Li Ma, Jian-Bao Xin, Xiao-Nan Tao and Huan-Zhong Shi

J Immunol 2010; 185:6348-6354; Prepublished online 15 October 2010; doi: 10.4049/jimmunol.1001728
http://www.jimmunol.org/content/185/10/6348

Why The JI?

- Rapid Reviews! 30 days* from submission to initial decision
- No Triage! Every submission reviewed by practicing scientists
- Speedy Publication! 4 weeks from acceptance to publication

*average

References
This article cites 38 articles, 15 of which you can access for free at:
http://www.jimmunol.org/content/185/10/6348.full#ref-list-1

Subscription
Information about subscribing to The Journal of Immunology is online at:
http://jimmunol.org/subscription

Permissions
Submit copyright permission requests at:
http://www.aai.org/About/Publications/JI/copyright.html

Email Alerts
Receive free email-alerts when new articles cite this article. Sign up at:
http://jimmunol.org/alerts

Errata
An erratum has been published regarding this article. Please see next page or:
/content/193/9/4748.full.pdf
Generation and Differentiation of IL-17–Producing CD4+ T Cells in Malignant Pleural Effusion

Zhi-Jian Ye,*1 Qiong Zhou,*1 Yong-Yao Gu,*1† Shou-Ming Qin,† Wan-Li Ma,* Jian-Bao Xin,* Xiao-Nan Tao,* and Huan-Zhong Shi*

IL-17–producing CD4+ T (Th17) cells have been found to be increased in some human cancers; however, the possible implication of Th17 cells in regulating antitumor responses in malignant pleural effusion (MPE) remains to be elucidated. In the current study, distribution and phenotypic features of Th17 cells in both MPE and peripheral blood from patients with lung cancer were determined by flow cytometry or double immunofluorescence staining. The impacts of cytokines on Th17 cell generation and differentiation were explored. The chemotactant activity of chemokines CCL20 and CCL22 for Th17 cells in vitro was also observed. It was found that the increased Th17 cells could be found in MPE compared with blood. The in vitro experiments showed that IL-1β, IL-6, IL-23, or their various combinations could promote Th17 cell generation and differentiation from naive CD4+ T cells. MPE was chemotactic for Th17 cells, and this activity was partly blocked by anti-CCL20 and/or CCL22 Abs. Our data also showed that the accumulation of Th17 cells in MPE predicted improved patient survival. It could be concluded that the overrepresentation of Th17 cells in MPE might be due to Th17 cell differentiation and expansion stimulated by pleural proinflammatory cytokines and to recruitment of Th17 cells from peripheral blood induced by pleural chemokines CCL20 and CCL22. Furthermore, the accumulation of Th17 cells in MPE predicted improved patient survival. These data provide the basis for developing immune-boosting strategies based on ridding the cancer patient of this cell population. The Journal of Immunology, 2010, 185: 6348–6354.

Malignant pleural effusion (MPE) is frequently observed in some cancers, especially lung cancer, and appearance of an MPE is an ominous prognostic sign for cancer patients, because the presence of MPE indicates that the tumor is incurable by surgery and life expectancy is short (1). Although there have been no epidemiologic studies, the annual incidence of MPE in the United States is estimated to be >150,000 cases (2). Although MPE is more and more common, very little information is available on the immune mechanisms that are involved in its development. An accumulation of lymphocytes, especially CD4+ helper T cells, frequently occurs in MPE secondary to direct pleural involvement and/or metastases from malignancies (3, 4).

IL-17 (also known as IL-17A) was originally cloned as CTLA8 from a rodent T cell hybridoma by Rouvier et al. (5) in 1993 and was identified by Yao et al. (6) in 1995 as a cytokine produced by activated human CD45RO+ memory T cells. IL-17F, a closely related member with 50% amino acid sequence homology to IL-17A, was later discovered and is also expressed in activated CD4+ T cells (7). The subset of CD4+ T cells that produce both IL-17A and IL-17F is now defined as a separate subset IL-17–producing CD4+ T (Th17) cells. Distinct from Th1 and Th2 cells, Th17 cells are reported to be generated from naive T cells by IL-6 and TGF-β (8, 9) and are expanded and stabilized further by IL-23 (10) and by virtue of expressing the orphan nuclear receptor RORyt as a critical transcription factor (11). Th17 cells have been found in some human cancers (12–15), however, the possible implication of Th17 cells in regulating antitumor responses in MPE remains to be demonstrated. The present study investigated the idea that Th17 cells could be involved in the control of the local immune response in MPE. In this study, we provided the insightful mechanism by which Th17 cells are generated and regulated by cytokines secreted from pleural cells and their immune infiltrates in MPE.

Materials and Methods

Subjects

The study protocol was approved by our institutional review board for human studies, and informed consent was obtained from all subjects. Pleural fluid samples were collected from 30 patients (age range, 32–81 y) with newly diagnosed lung cancer with MPE (Table I). Histologically, 19 cases were adenocarcinoma and 11 were squamous cell carcinoma. A diagnosis of MPE was established by demonstration of malignant cells in pleural fluid and/or on closed pleural biopsy specimen. The patients were excluded if they had received any invasive procedures directed into the pleural cavity or if they had suffered chest trauma within 3 mo prior to hospitalization. At the time of sample collection, none of the patients had received any anticancer therapy, corticosteroids, or other nonsteroid anti-inflammatory drugs.

Sample collection and processing

The pleural fluid samples were collected in heparin-treated tubes from each subject, using a standard thoracocentesis technique within 24 h after hospitalization. Thirty milliliters of peripheral blood was drawn simul-
taneously, MPE specimens were immersed in ice immediately and were then centrifuged at 1200 × g for 5 min. The cell-free supernatants of MPE and serum were frozen at −80°C immediately after centrifugation for later determination of concentrations of cytokines and chemokines. The cell pellets of MPE were resuspended in HBSS, and mononuclear cells were isolated by Ficoll-Hypaque gradient centrifugation (Pharmacia, Uppsala, Sweden) to determine the T cell subsets within 1 h. A pleural biopsy was performed when the results of pleural fluid analysis were suggestive of malignancy.

Cell isolation

Bulk CD4+ T cells from pleural fluid and blood were isolated by negative selection (by depletion of CD8+, CD11b+, CD16+, CD19+, CD36+, and CD56+ cells) with the Untouched CD4+ cell isolation kit (Miltenyi Biotec, Auburn, CA), according to the manufacturer’s instructions. After isolation of bulk CD4+ T cells, the naive CD4+ T cells (CD45RA+CD45RO−) were further purified by EasySep enrichment kits (StemCell Technologies, Vancouver, British Columbia, Canada), according to the manufacturer’s instructions. The purity of naive CD4+ T cells was >97%, as measured by flow cytometry.

Double immunofluorescence staining

Double immunofluorescence staining was performed on cell pellets of MPE to visualize Th17 and Th1 cells. The cell pellets of purified CD4+ T cell and Th17 from MPE were fixed in freshly made 4% paraformaldehyde/PBS for 2 h and washed three times in PBS. The cell pellets were first embedded in 8% agarose gel and then in paraffin according to standard pathology protocols. The paraffin-embedded cell pellets were cut into 4- to 5-μm thick sections. The primary Abs were rabbit polyclonal Ab targeted against human IL-17 and mouse anti-human IFN-γ mAb (both were purchased from Santa Cruz Biotechnology, Santa Cruz, CA). Appropriate species-matched Abs were used as isotype controls. As secondary Abs, rhodamine-labeled affinity-purified goat anti-rabbit IgG was used for labeling the

rabbit anti-IL-17 Ab, and fluorescein-labeled affinity purified goat anti-mouse IgG was used for labeling the mouse anti-IFN-γ mAb. After permeabilizing with 0.1% Triton X-100 in PBS for 15 min at room temperature and washing with PBS, slides were incubated with 10% goat serum in PBS at 4°C overnight, and then incubated at 4°C overnight with 1/50 concentrations of primary Abs as recommended by the manufacturer. After washing, slides were incubated with selected secondary Abs for 40 min at room temperature in the dark, correctly matched to the appropriate species. DAPI mounting medium (Vector Laboratories, Burlingame, CA) was used for cell nuclei staining. Finally, slides were viewed under imaging fluorescence microscope (Olympus BX51; Olympus, Tokyo, Japan).

A similar double immunofluorescence staining technique was used to identify which cell types in MPE were expressing chemokine CCL20. The primary Abs were as follows: rabbit polyclonal Ab targeted against human CCL20 (Abcam, Cambridge, U.K.), mouse anti-human CD3 mAb (BioLegend, San Diego, CA), specific for T cells; and mouse anti-CD68 mAb (Thermo Fisher Scientific Anatomical Pathology, Fremont, CA), specific for microphages, mouse anti-human epithelial membrane Ag (EMA) mAb (Thermo Fisher Scientific Anatomical Pathology) to identify malignant cells. Appropriate species-matched Abs were used as isotype controls. As secondary Abs, rhodamine-labeled affinity purified goat anti-rabbit IgG was used for labeling the rabbit anti-CCL20 Ab, and fluorescein-labeled affinity-purified goat anti-mouse IgG was used for labeling the mouse anti-CD3, -CD68, and -EMA mAbs.

Flow cytometry

The expression markers on T cells from MPE and blood were determined by flow cytometry after surface staining or intracellular staining with anti-human-specific Abs conjugated with either PE or FITC. These human Abs included anti-CD3, anti-CD4, anti-CD45RA, anti-CD58RO, anti-CD62L, anti-CCR2, anti-CCR4, anti-CCR5, anti-CCR6, anti-CCR7, anti-IL-17, and anti-IFN-γ mAbs, which were purchased from BD Biosciences or eBioscience (San Diego, CA). Intracellular staining for IL-17– or IFN-γ–producing cells was performed on MPE cells stimulated with PMA (50 ng/ml; Sigma-Aldrich, St. Louis, MO) and ionomycin (1 μM; Sigma-Aldrich) in the presence of GolgiStop (BD Biosciences) for 5 h, and the intracellular IL-17 or IFN-γ was then stained with anti–IL-17 or IFN-γ conjugated with

![FIGURE 1. Presence of Th17 and Th1 cells in MPE.](image)

A. Expression of IFN-γ was detected by mouse anti–IFN-γ mAb and then fluorescein-labeled goat anti-mouse IgG (green). IL-17 expression was detected by rabbit polyclonal Ab targeted against IL-17, and then rhodamine-labeled goat anti-rabbit IgG (red). DAPI mounting medium was used for cell nuclei staining. Top panels, Purified Th17 cells served as positive controls. Bottom panels, Immunofluorescence double staining of pleural CD4+ T cells indicated that some CD4+ T cells were Th17 cells, some are Th1 cells, and some are Th17/Th1 cells (arrows in the merged panel). Original magnification ×1000. The representative flow cytometric dot plots of expression of Th17/Th1 cells MPE (B) and in peripheral blood (C). Comparisons of percentages of Th17 (D) and Th1 cells (E) in MPE and blood from patients with lung cancer (n = 30). The percentage of Th17 and Th1 cells was determined by flow cytometry. Horizontal bars indicate medians. Comparison was made using a Wilcoxon signed-rank test. F. The percentages of Th17 cells correlated with Th1 cells in MPE. Correlations were determined by Spearman’s rank correlation coefficients.
PE (eBioscience). Flow cytometry was performed on a BD FACSCalibur flowcytometer using FCS Express3 software.

**Measurement of cytokines and chemokine**

The concentrations of cytokines, including IL-1α, IL-1β, IL-2, IL-4, IL-5, IL-6, IL-12p70, IL-23, IFN-γ, and TGF-β1, as well as chemokine CCL20 in both pleural fluids, and sera were measured by ELISA kits according to the manufacturer’s protocols (all kits were purchased from R&D Systems, Minneapolis, MN). All samples were assayed in duplicate.

**Generation and differentiation in MPE of Th17 cells**

Purified naive CD4+ T cells (5 × 10^5) were cultured in 1 ml complete medium containing human IL-2 (2 ng/ml) in 48-well plates and stimulated with plate-bound anti-CD3 (OKT3; 1 µg/ml) and soluble anti-CD28 mAbs (1 µg/ml) for 7 d. The exogenous cytokines used were TGF-β1 (5 ng/ml), IL-6 (100 ng/ml), IL-1β (10 ng/ml), and IL-23 (10 ng/ml). Recombinant human IL-1β, IL-2, IL-6, IL-23, and TGF-β1 were purchased from R&D Systems.

**Th17 cell chemotaxis assay**

Chemotaxis assays were performed using 8-µm pore polycarbonate filters in 24-well Transwell chambers (Corning Costar, Corning, NY). Briefly, Transwell membranes were coated with fibronectin (5 µg/ml; Chemicon International, Schwabach, Germany) for 30 min at 37 °C. Th17 cells isolated from peripheral blood were added to the top chamber resuspended in RPMI 1640 medium plus 0.5% BSA at 1 × 10^5 cells/ml in a final volume of 0.5 ml. MPE from five lung cancer patients were placed in the bottom chamber of the Transwell in a volume of 600 µl, and the chamber was incubated at 37 °C in 5% CO2 atmosphere for 3 h. At the end of incubation, the filter was washed by HBSS lightly, fixed, stained, and mounted on a glass microscope slide. The chemotaxis index was calculated by dividing the numbers of cells migrated in response to test MPE or recombinant human chemokines by the numbers of cells migrated in response to medium alone.

**Statistics**

Data are expressed as mean ± SEM or median (range). Comparisons of the data between different groups were performed using a Kruskal-Wallis one-way ANOVA on ranks. For data in MPE and in the corresponding blood, paired data comparisons were made using a Wilcoxon signed-rank test. Correlations between variables were determined by Spearman’s rank correlation coefficients. Survival was estimated by the Kaplan-Meier method and compared by the log-rank test. Multivariate analysis of prognostic factors for overall survival was performed using the Cox proportional hazards model. Analysis was completed with SPSS version 16.0 Statistical Software (Chicago, IL), and p < 0.05 was considered to indicate statistical significance.

**Results**

**Th17 and Th1 cells were significantly increased in MPE**

To visualize the distribution of Th17 and Th1 cells in MPE, double immunofluorescence staining was performed on the pleural cell pellets with both anti-CD4 and anti-IL-17 or IFN-γ Abs. Because rabbit polyclonal Ab targeted against human IL-17 was used in the current study, and such a polyclonal Ab is notorious for lacking specificity, we performed positive controls using purified Th17 cells, which showed that the immunofluorescent technique used in our study was reliable (Fig. 1A, top panels). We noted that both Th17 and Th1 cells were found in MPE, and some of these pleural CD4+ T cells were Th17/Th1 cells (Fig. 1A, bottom panels).

Consistent with the immunofluorescent findings, Th17 cells, Th1 cells, and Th17/Th1 cells in MPE could also be identified by flow cytometry (Fig. 1B). Percentages of Th17 cells represented the higher values in MPE (3.74 ± 0.34%), showing a significant increase in comparison with those in the corresponding blood (0.59 ± 0.06%) (n = 30; Wilcoxon signed-rank test; p < 0.001) (Fig. 1B–D). Similarly, a significant increase in Th1 cells was observed in MPE (29.81 ± 2.49%) compared with blood (2.92 ± 0.35%) (Wilcoxon signed-rank test; p < 0.001) (Fig. 1B–E). It was noted that pleural Th17 cell frequency was correlated positively with Th1 cell frequency (r = 0.865; p < 0.001) (Fig. 1F).

To identify the phenotypic characteristics of Th17 in MPE, we analyzed the expression profiles of CD45RO, CD62L, and CCR7 and observed that most pleural Th17 cells expressed high levels of CD45RO (89.6 ± 1.3%) but not of CD62L (3.7 ± 0.6%) or CCR7 (5.1 ± 0.7%), indicating that they were effector memory cells (Fig. 2).

**Generation and differentiation of Th17 cells in MPE**

We determined some cytokines that are thought to be related to the generation and differentiation of human Th17 cells in MPE. As shown in Fig. 3A, high levels of IL-1β, IL-6, IFN-γ, and TGF-β1 could be found in MPE, and they were significantly higher than their compartments in the corresponding serum (n = 14; all p < 0.01). The above results suggested that these proinflammatory cytokines might promote the generation and differentiation of Th17 cells in MPE.

To evaluate the contribution of cytokines to the generation and differentiation of pleural Th17 cells, we purified naive T cells and cultured them in the presence of one or more of IL-1β, IL-6, IL-23, and TGF-β. With IL-2–containing medium provided a baseline for comparison, IL-1β, IL-6, or IL-23, but not TGF-β, could promote the differentiation of Th17 cells from naive CD4+ T cells (Fig. 3B). The combination of IL-1β plus IL-6, IL-1β plus IL-23, IL-6 plus IL-23, or IL-1β plus IL-6 plus IL-23 significantly
increased the percentage of Th17 cells at higher extents compared with any single one of above cytokines. Surprisingly, although high concentration of IFN-γ was found in MPE, which was much higher than that in serum, IFN-γ did not affect Th17 cell numbers (Fig. 3B). Likewise, a significant high concentration of TGF-β was also found in MPE, but it did not promote the differentiation of naive CD4+ T cells into Th17 cells. The purified naive CD4+ T cells were stimulated with plate-bound anti-CD3 and soluble anti-CD28 mAbs in the presence of the indicated cytokines, either alone or in various combinations. Seven days after activation, the cells were restimulated with PMA and ionomycin and analyzed for IL-17 expansion after intracellular staining. The percentages of Th17 cells generated in culture medium alone served as a baseline. The results are reported as mean ± SEM from five independent experiments. *p < 0.01 compared with the corresponding blood.

**FIGURE 3.** Generation and expansion of human Th17 cells from MPE and peripheral blood regulated by different cytokines. A, Proinflammatory cytokines in MPE and serum were determined by ELISA and are reported as mean ± SEM (n = 14). *p < 0.01 compared with the corresponding sera. B, Th17 cells detected in naive CD4+ T cells from MPE and blood after culturing with indicated cytokines. The purified naive CD4+ T cells were stimulated with plate-bound anti-CD3 and soluble anti-CD28 mAbs in the presence of the indicated cytokines, either alone or in various combinations. Seven days after activation, the cells were restimulated with PMA and ionomycin and analyzed for IL-17 expansion after intracellular staining. The percentages of Th17 cells generated in culture medium alone served as a baseline. The results are reported as mean ± SEM from five independent experiments. *p < 0.01 compared with the corresponding blood.

**FIGURE 4.** Chemokine receptors expressed on Th17 cells. A, Flow cytometric dot plots of expressions of CCR2, CCR4, CCR5, and CCR6 on Th17 cells from blood and MPE. B, Comparisons of expressions of CCR2, CCR4, CCR5, and CCR6 on Th17 cells in both blood and MPE from patients with lung cancer (n = 14). Comparisons of CCR expressions were made using a Wilcoxon signed-rank test, and there were no significant statistical differences in all CCRs. p > 0.05.
of Th17 cells; in contrast, TGF-β could reduce the increased percentage of Th17 cells stimulated by the above cytokines (Fig. 3B).

We noted that abovementioned proinflammatory cytokines or their combinations could cause generation and differentiation of Th17 cells from blood naive CD4+ T cells in a similar manner (Fig. 3B). We also noted that IL-1β plus IL-6 could induce more Th17 cell generation and differentiation from MPE naive CD4+ T cells than from blood naive CD4+ T cells.

Chemokines CCL20 and CCL22 might be responsible for recruitment of Th17 cells into MPE

Besides local generation and differentiation, recruitment from peripheral blood might also contribute to the increase in pleural Th17 cells. Because lymphocyte migration is tightly regulated by chemokine–chemokine receptor interaction (16), we therefore sought to elucidate the effects of chemokine–chemokine receptor interaction on recruitment of Th17 cells into MPE. In the current study, our results showed that the expressions of CCR2, CCR4, CCR5, and CCR6 could be observed in pleural Th17 cells; among these chemokine receptors, CCR4 and CCR6 exhibited significant high levels of expressions (Fig. 4). The percentages of CCR2+ Th17, CCR4+Th17, CCR5+Th17, and CCR6+Th17 cells were 20.6 ± 2.7, 65.9 ± 2.4, 26.8 ± 2.2, and 48.3 ± 4.8%, respectively. We also noted that the expressions of these CCRs in Th17 cells was similar between MPE and blood (n = 14; all p > 0.05).

In a previous study, we have reported that all cancer cells, most of macrophages and T cells in MPE, expressed high level of CCL22 and that concentration of soluble CCL22 in MPE was much higher than that in serum (17). In the current study, we extended these observations and further demonstrated that significant expression of another chemokine CCL20 was found in all cancer cells identified by anti-EMA mAb; some macrophages identified by anti-CD68 mAb, and some T cells identified anti-CD3 mAb (Fig. 5A). These data suggested that malignant cells, macrophages, and T cells might be the cell sources of pleural CCL20. In addition, our data also showed that CCL20 concentration in MPE (79.6 ± 19.6 pg/ml; n = 14; p < 0.001) (Fig. 5B).

On the basis of our observation that Th17 cells expressed high levels of CCR4 and CCR6, and high levels of CCL20 and CCL22 were found in MPE, we hypothesized that Th17 cells could migrate into the pleural space in response to CCL20 and/or CCL22. Our results showed that MPE exerted a potent chemotactic activity for circulating Th17 cells and that either anti-CCL20 or CCL22 mAb significantly suppressed Th17 cell chemotaxis; the combination of both mAbs exhibited an even more strong suppressive effect (Fig. 5C). Taken together, these data suggested that chemokines CCL20 and CCL22 might be responsible for recruitment of Th17 cells into MPE.

Increase in pleural Th17 cells predicted improved survival of patients with MPE

To test the prediction that pleural Th17 cells would affect survival of patients with MPE, we analyzed survival time with the number of Th17 cells in 30 patient with MPE. The individuals were divided into two equal groups on the basis of Th17 cell numbers. The low Th17 cell group included all those with Th17 cell counts of ≤4% (n = 15) and the high Th17 cells group included those with Th17 cell counts > 4% (n = 15). As shown in Fig. 6, there was a significant correlation between pleural Th17 cell numbers and patient survival (p = 0.001). Patients with higher Th17 cells had significantly longer overall survival (median, 5 mo) than patients with lower Th17 cells (median, 3 mo). Individuals in the low Th17 cell group experienced a 4.5-fold higher death hazard as compared with those in the high Th17 cell group (95% confidence interval, 1.8–11.3). The multivariate Cox proportional hazards analysis was performed, and variables that were associated with survival by univariate analysis were adopted as covariates (Table I). The multivariate analysis revealed that the number of pleural Th17 cells was an independent prognostic factor for survival (hazard ratio = 0.347; p = 0.002) (Table II). However, Th1 number, glucose concentration, and pH value were not independent prognostic factors for survival (all p > 0.05). Therefore, an increased in pleural Th17 cells was a significant predictor of decreased risk for death and for improved survival in patients with MPE.

Discussion

In MPE, CD4+ T cells are dominant, and the proportion of CD8+ T cells is significantly lower than that of CD4+ T cells (18). In contrast, the proportion of CD4+ T cells in the pleural cavity of lung cancer patients without MPE is significantly lower than that of CD8+ T cells (19). Furthermore, the proportion of pleural CD4+...
T cells may help to select patients who are likely to have a poorer prognosis after surgery and therefore may be suitable for consideration of adjuvant treatments (20). Classically, naive CD4+ T cells have been thought to differentiate into two main lineages, Th1 and Th2 cells (21, 22). Regulatory T cells represent only a small subset of CD4+ T cells in the peripheral circulation and are responsible for the balance of immune responses, which is essential for health (23). The identification of Th17 cells not only changes the classical Th1/Th2 paradigm of Th cell differentiation but also markedly facilitates our understanding of human immunity under both physiological and pathological conditions (24, 25). Early studies have suggested that Th1/Th2 cell balance in MPE may influence pathophysiologic process of pleural disease (26, 27). Our previous studies showed that increased regulatory T cells were found in MPE, and these regulatory T cells were recruited into pleural space induced by CCL22 (17, 28). In the current study, we have demonstrated that Th17 cells, Th1 cells, and Th17/Th1 cells were increased in MPE compared with corresponding peripheral blood. The mechanisms of accumulation of Th17 cells in the tumor microenvironment remain largely unknown. We speculated that an optimal cytokine milieu for human Th17 generation (10). On the basis of our observation that the increased Th17 cells could be seen in MPE, we reasoned that a proinflammatory cytokine milieu that facilitates the differentiation of Th17 cells should also be present in MPE microenvironments. Indeed, our data showed that the concentrations of some cytokines, including IL-1β, IL-6, and TGF-β1, in MPE were significantly higher than those in the corresponding serum. Furthermore, our data demonstrated that IL-1β, IL-6, and IL-23 each expanded the Th17 cells in naive CD4+ T cells. The combination of IL-1β plus IL-6, IL-1β plus IL-23, IL-6 plus IL-23, or IL-1β plus IL-6 plus IL-23 significantly increased the percentage of Th17 cells at even higher extents. Some studies demonstrated that TGF-β is required for human Th17 cell differentiation (29–31); however, our data are consistent with the findings reported in the other studies in which TGF-β suppresses the differentiation of Th17 cells (14, 32).

The mechanisms of accumulation of Th17 cells in the tumor microenvironment remain largely unknown. We speculated that an increased percentage of Th17 cells in MPE might be due to local differentiation and/or active recruitment. In the previous study, we provided direct evidence that IL-16 is capable of inducing CD4+ T cell infiltration into the pleural space (33). Therefore, as a subpopulation of CD4+ T cells, Th17 cells might also be recruited into MPE by local production of IL-16, because IL-16 level is significantly higher in MPE than in serum (33).

Although chemokine receptors are important for T cell migration, it has been unclear how they are regulated in Th17 cells. Recently, Su et al. (34) have demonstrated that tumor microenvironmental RANTES and MCP-1 secreted by tumor cells and tumor-derived fibroblasts mediate the recruitment of Th17 cells into the tumor sites. In the current study, we were prompted to evaluate whether pleural CCL20 and CCL22 might be responsible for the influx of Th17 cells into the pleural space. In the previous study (17) and the current study, our results showed that concentrations of both CCL20 and CCL22 in MPE were significantly higher than those in corresponding serum and that malignant cells, macrophages, and T cells might be the cell sources of pleural CCL20 and CCL22. Moreover, Th17 cells in peripheral blood and MPE strongly express CCR4 and CCR6, two chemokine receptors for CCL22 and CCL20, respectively, on their surface. The above data suggested that CCL20 and CCL22 in MPE might be related to the accumulation of Th17 cells in MPE. Indeed, an in vitro migration assay further confirmed that MPE could induce the migration of Th17 cells and that either anti-CCL20 or CCL22 mAb significantly inhibited the ability of the MPE to stimulate Th17 cell chemotaxis, and the combination of both mAbs exhibited an even more strong suppressive effect. Therefore, CCL20 and CCL22 might be able to chemotract Th17 cell recruitment into pleural space.

It should be mentioned that besides Th17 cells, we also noted Th1 and Th17/Th1 cells were increased in MPE compared with

---

**Table I. Characteristics of the study population**

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>n</th>
<th>Mean ± SEM</th>
<th>Median (Range)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sex</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Male</td>
<td>12</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Female</td>
<td>18</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Histological type</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Adenocarcinoma</td>
<td>19</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Squamous cell carcinoma</td>
<td>11</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Age, y</td>
<td>30</td>
<td>60.6 ± 2.7</td>
<td>63.5 (32–81)</td>
</tr>
<tr>
<td>pH</td>
<td>30</td>
<td>7.29 ± 0.02</td>
<td>7.30 (7.17–7.43)</td>
</tr>
<tr>
<td>Protein, g/ml</td>
<td>30</td>
<td>3.9 ± 0.3</td>
<td>4.3 (0.7–6.1)</td>
</tr>
<tr>
<td>Glucose, mg/dl</td>
<td>30</td>
<td>82.1 ± 8.9</td>
<td>72.2 (19.3–198.3)</td>
</tr>
<tr>
<td>LDH, U/l</td>
<td>30</td>
<td>797 ± 135</td>
<td>615 (110–3200)</td>
</tr>
<tr>
<td>Total cell counts, × 10⁷/μl</td>
<td>30</td>
<td>1.54 ± 0.09</td>
<td>1.53 (0.76–2.68)</td>
</tr>
<tr>
<td>Th1 cells, %</td>
<td>30</td>
<td>29.81 ± 2.49</td>
<td>25.64 (11.63–55.20)</td>
</tr>
<tr>
<td>Th17 cells, %</td>
<td>30</td>
<td>3.74 ± 0.34</td>
<td>4.04 (1.16–7.53)</td>
</tr>
</tbody>
</table>

LDH, lactate dehydrogenase.

---

**Table II. Univariate and multivariate analyses of variables associated with survival**

<table>
<thead>
<tr>
<th>Variables</th>
<th>Univariate Value</th>
<th>Hazard Ratio</th>
<th>95% CI</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sex</td>
<td>0.595</td>
<td>NA</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Histological type</td>
<td>0.823</td>
<td>NA</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Age</td>
<td>0.840</td>
<td>NA</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Protein</td>
<td>0.449</td>
<td>NA</td>
<td></td>
<td></td>
</tr>
<tr>
<td>LDH</td>
<td>0.151</td>
<td>NA</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Total cell counts</td>
<td>0.623</td>
<td>NA</td>
<td></td>
<td></td>
</tr>
<tr>
<td>pH</td>
<td>0.042</td>
<td>0.418</td>
<td>0.006–30.489</td>
<td>0.690</td>
</tr>
<tr>
<td>Glucose</td>
<td>0.021</td>
<td>1.001</td>
<td>0.992–1.010</td>
<td>0.842</td>
</tr>
<tr>
<td>Th1 cells</td>
<td>0.001</td>
<td>1.044</td>
<td>0.962–1.134</td>
<td>0.304</td>
</tr>
<tr>
<td>Th17 cells</td>
<td>&lt;0.001</td>
<td>0.347</td>
<td>0.177–0.682</td>
<td>0.002</td>
</tr>
</tbody>
</table>

CI, confidence interval; LDH, lactate dehydrogenase; NA, not adopted.

FIGURE 6. Accumulation of Th17 cells predicts improved survival in patients with malignant pleural effusion. Individuals with higher Th17 cell frequency group (n = 15) had a significant improved survival than those with lower Th17 cell frequency group (n = 15). Overall survival was estimated by the Kaplan-Meier method and compared by the log-rank test.
blood, and Th17 cell numbers were correlated positively with Th1 cell numbers. In addition, IFN-γ released into MPE was also significantly increased. Recently, Yang et al. (35) have demonstrated that inhibition of the transcription factor T-bet expressed in Th1 cells suppresses the differentiation of both Th1 and Th17 cells. Furthermore, T-bet directly regulates transcription of the IL-23R and influenced the fate of Th17 cells, which depend on optimal IL-23 production for survival (36). The insights into the interaction between Th1 and Th17 cells in human were largely unknown so far, and the investigation of impacts and regulation of Th1 cells on Th17 cell generation and functions in MPE is on the way in our laboratory.

Although Th17 cells have been found in tumor microenvironment, their physiological functions in cancer immunity have been poorly defined. It has been suggested that Th17 cells have an indirect role in antitumor immunity by promoting dendritic cell, cytotoxic T cell, and NK cell trafficking to, and retention within, the tumor microenvironment (37). Recently, Martin-Orozco et al. (38) have provided direct evidence for a protective role of Th17 cells in antitumor responses by using IL-17–deficient mice in a model of lung melanoma. Moreover, the adoptive transfer of tumor-specific Th17 cells protect against various tumors and, in the lung melanoma model, promote tumor-specific cytotoxic T cell responses (38). We noted in the current study that the accumulation of Th17 cells in MPE predicted improved patient survival, implying a new role of proinflammatory response in regulating tumor progression in human cancer. Taken together, these data provide the basis for developing novel immune-boosting strategies based on ridding the cancer patient of this cell population.

In conclusion, our data showed that Th17 cell numbers in MPE were significantly increased when compared with the corresponding peripheral blood. Although it cannot be excluded that over-representation of Th17 cells in MPE may be due to increased local Ag stimulation, our findings have demonstrated that pleural proinflammatory cytokines could promote the differentiation of Th17 cells and that chemokines CCL20 and CCL22 might be capable of inducing Th17 cells to MPE.

Acknowledgments
We thank Dr. Jiao Lan, Medical Research Center, People’s Hospital of Guangxi Zhuang Autonomous Region, for her excellent flow cytometric assistance.

Disclosures
The authors have no financial conflicts of interest.
Corrections


The images published in Fig. 5A were mistakenly selected from micrographs showing expression of CCL22 in macrophages, T cells, and malignant cells, respectively, from other experiments in which similar procedures were performed. The correct figure showing expression of CCL20 is published below; the entire figure is shown, but the only change is to panel A. The legend is correct as published and is also shown below for reference.