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Many pathogenic enveloped viruses, including HIV-1, escape complement-mediated virolysis by incorporating host cell regulators of complement activation into their own viral envelope. The presence of complement regulators including CD59 on the external surface of the viral envelope confers resistance to complement-mediated virolysis, which may explain why human pathogenic viruses such as HIV-1 are not neutralized by complement in human fluids, even in the presence of high Ab titers against the viral surface proteins. In this study, we report the development of a recombinant form of the fourth domain of the bacterial toxin intermedilysin (the recombinant domain 4 of intermedilysin [rILYd4]), a 114 aa protein that inhibits human CD59 function with high affinity and specificity. In the presence of rILYd4, HIV-1 virions derived from either cell lines or peripheral blood mononuclear cells of HIV-1-infected patients became highly sensitive to complement-mediated lysis activated by either anti–HIV-1 gp120 Abs or by viral infection-induced Abs present in the plasma of HIV-1-infected individuals. We also demonstrated that rILYd4 together with serum or plasma from HIV-1-infected patients as a source of anti–HIV-1 Abs and complement did not mediate complement-mediated lysis of either erythrocytes or peripheral blood mononuclear cells. These results indicate that rILYd4 may represent a novel therapeutic agent against HIV-1/AIDS. The Journal of Immunology, 2010, 184: 359–368.
of CD59 that would sensitize HIV-1 virions or HIV-infected cells to the lytic effect of complement has been actively sought by us and others (11).

In this study, we report the development of rILYd4, a high-affinity specific inhibitor of human CD59 (hCD59). rILYd4 is the recombinant form of the 114 aa domain 4 (D4) of intermediolysin (ILY), a cell lytic toxin secreted by Streptococcus intermedium. ILY is a pore-forming toxin that exclusively lyases human cells, because it binds with high affinity and specificity to hCD59 but not to CD59 from other species (15, 16). Binding of ILY to hCD59 occurs through D4, whereas the three other domains (domains 1, 2, and 3 of ILY) form the lytic transmembrane pore (15). Because D4 of ILY binds to a region of hCD59 to encompass its active site (aa 42–58) (15, 17), we reasoned that rILYd4 would inhibit hCD59 function (2) and thereby enhance Ab-dependent complement-mediated virolysis of HIV-1. Our results show that rILYd4 potently enhances complement-mediated HIV-1 virolysis activated by anti–HIV-1-specific Abs with no or minimal bystander effects. We conclude that rILYd4 has strong potential as an anti–HIV-1 therapeutic agent, a notion that warrants further testing in animal studies and in human clinical trials.

Materials and Methods
Preparation of rILY

For the generation of a truncated rILYd4 fragment, sequences encoding the fragments were cloned into an expression vector pTrcHis A with a His6 tag. The 114 aa rILYd4 fragment contains only D4 of ILY: GALTINLHDAFGAVFPPYTVWEELGHDDAGYTTSRBSWGNYGNAHSTYLLTLEFGVNRVKVLGALGWEPLLWRLYSNLWPLVPQRNI- STGWHTILIFQEDFKVKDN. This-tagged rILYd4 was expressed in Escherichia coli and purified with the His•Bind purification kit (Novagen, San Diego, CA) as described previously (15).

FACS analysis

Erythrocytes from human and hCD59transgenic mouse expressing hCD59 only on the mouse erythrocytes published previously (16) were preincubated with rILYd4 (1 μg/ml) or PBS for 10 min at room temperature, incubated with mouse anti-hCD59 monoclonal Ab (0.2 μg/ml; BRIC 229; Bioreclamation, Office, Bristol, U.K.) at room temperature for 30 min, washed, and incubated with a FITC-conjugated corresponding secondary Ab. The cells were washed with PBS three times before analyzing the fluorescence intensity using a FACScan (Becton Dickinson, Franklin Lakes, NJ).

Hemolytic assay

Human or mouse blood was obtained by venipuncture into a syringe containing sodium citrate (105 mM) as an anticoagulant (blood:buffer = 9:1 v/v). The erythrocytes were washed four times by PBS, stored in Alsever’s solution at 4˚C, and used for hemolytic assays, as described previously (16). The amount of hemoglobin released from lysed erythrocytes was determined by the absorbance of the supernatant at 414 nm, and percent lysis was calculated as follows: [([experimental OD414] – blank OD414)]/(total lysis OD414 – blank OD414)]×100. The total lysis sample was obtained by adding pure water to the erythrocyte pellet.

Complement-mediated lysis on human erythrocytes

The sensitivity of human erythrocytes to human complement-mediated lysis in the presence or absence of rILYd4 was assessed by two different methods: cobra venom factor (5 mg/L) lysis assay and anti-human erythrocyte Ab-sensitized erythrocyte method, as described previously (16). As the source of complement, we used human serum (HS; 50% v/v diluted in GVB++) as described previously (15, 17), we reasoned that rILYd4 would inhibit hCD59 function (2) and thereby enhance Ab-dependent complement-mediated virolysis of HIV-1. Our results show that rILYd4 potently enhances complement-mediated HIV-1 virolysis activated by anti–HIV-1-specific Abs with no or minimal bystander effects. We conclude that rILYd4 has strong potential as an anti–HIV-1 therapeutic agent, a notion that warrants further testing in animal studies and in human clinical trials.

Preparation of rILY–11, 12

Briefly, 5 × 10⁵ human non-Hodgkin lymphoma RL cells purchased from the American Type Culture Collection (Manassas, VA) were suspended in 100 μl RPMI 1640 medium supplemented with 10% heat-inactivated FBS and plated in 96-well plates. After an additional 100 μl of medium containing different concentration of rILYd4, rituximab (2 μg/ml), and 10% HS as the source of complement was added to the wells, the cells were incubated for 4 h at 37˚C. Next, 70 μl culture medium plus 30 μl Alamar blue solution (Sorotec) was added to the wells and incubated at 37˚C overnight. Cell lysis was assessed by reading the plates in an F-2000 fluorescence spectrophotometer (Hitachi, Tokyo, Japan; excitation, 560 nm; emission, 590 nm). Percent cell lysis in each well was calculated as: (reading from the well without any treatment – reading from testing well)/(reading from the well without any treatment)×100. All pooled HS used as a source of complement in this study was purchased from Complement Technology (Tyler, TX).

Preparation of HIV-1 from HIV-1-infected patients

HIV-1 primary isolates were generated by coculture of PBMCs from HIV-1–infected and healthy donors, following methods described previously (19). PBMCs were prepared from heparinized peripheral blood donated by six HIV-1–seropositive patients naive for antiretroviral therapy (patients 1–6 in Table I) and by HIV-1–seronegative donors. PBMCs from seronegative and seropositive individuals were stimulated separately for 2 d with PHA (5 μg/ml) and cocultured at a 1:3 ratio in the presence of IL-2 (10 ng/ml) in complete RPMI 1640 medium (200 μl per well) in 96-well round-bottom plates. After 7 d of coculture, supernatants were harvested, aliquoted, and stored at −80˚C as HIV-1 primary isolate stocks for virolysis assay.

Anti-gp120/gp160 Ab activated complement-mediated virolysis

HIV-1 virions (20 μl containing 5 ng HIV-1 p24/ml) were preincubated for 30 min at 37˚C with or without rILYd4 (20 μg/ml) before exposure to anti–HIV-1-specific Abs (anti–HIV-1 gp120 monoclonal Ab, IgG1B12, National Institutes of Health AIDS Research and Reference Reagent Program [Germantown, MD]) and to the pooled HS as a source of complement (1:10 dilution in GVB++) buffer. Heat-inactivated HS was used as a negative control. Virolysis of HIV-1 was quantified using HIV-1 ELISA Ag assay (PerkinElmer or XpressBio). The lower limit of sensitivity of the assay for HIV-1 p24 was 26 pg/ml.

Preparation of six primary isolates of HIV-1 from HIV-1-infected patients

HIV-1 primary isolates were generated by coculture of PBMCs from HIV-1–infected and healthy donors, following methods described previously (19). PBMCs were prepared from heparinized peripheral blood donated by six HIV-1–seropositive patients naive for antiretroviral therapy (patients 1–6 in Table I) and by HIV-1–seronegative donors. PBMCs from seronegative and seropositive individuals were stimulated separately for 2 d with PHA (5 μg/ml) and cocultured at a 1:3 ratio in the presence of IL-2 (10 ng/ml) in complete RPMI 1640 medium (200 μl per well) in 96-well round-bottom plates. After 7 d of coculture, supernatants were harvested, aliquoted, and stored at −80˚C as HIV-1 primary isolate stocks for virolysis assay.

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Virolysis was calculated as follows: (p24 released in the presence of complement-competent serum − p24 released in the presence of heat inactivated serum) / (p24 released from Triton X-100 treated virions − p24 released by medium only) × 100%. Means ± SD of three experiments were compared using the paired two-tailed Student t test.

Complement-mediated virolysis activated by anti–HIV-1 Abs in plasmas of HIV-1-infected patients

Viral preparations (20 μl; 5 ng HIV-1 p24/ml) derived from the chronically-infected cell line OM10 or from primary HIV-1 isolates were preincubated for 30 min at 37˚C with either rILYd4 (20 μg/ml) or neutralizing anti-hCD59 monoclonal Ab (30 μg/ml; BRIC229). After preincubation, heat-inactivated plasma from either HIV-1–infected or healthy individuals (1:5 at final dilution) were individually added as a source of endogenous Abs, followed by the exposure to either complement-competent or heat-inactivated HS diluted in GVB++ buffer. Triton X-100 was used for determining the total virolysis. Experiments were conducted in duplicates and the paired two-tailed Student’s t test was used to compare the means ± SD.

Infectivity assay

Five microliters of reaction mixture from each condition in the virolysis experiment described above (patients 1–4 in Fig. 3A,3B) were added to fresh H9 cells (0.2 × 10^5 cells per well in 200 μl complete RPMI 1640 medium), and cultured for 7–10 d. The infectivity was then assessed by measuring HIV-1 p24 in the culture supernatant using HIV-1 p24 ELISA Antigen Assay (PerkinElmer). The lysis buffer included in the ELISA kit was used to lyse the viral particles for measuring HIV-1 core protein p24.

Plasma collection from HIV-1–infected patients

Plasma specimens were obtained from the repository at the Hawaii Center for AIDS, University of Hawaii. All samples were from patients who signed informed consent forms in accordance with the guidelines for conduction of clinical research by the University of Hawaii Institutional Review Board. Plasma HIV-1 levels (AmpliCord HIV-1 Monitor Ultra Sensitive Test, Roche Diagnostics, Basel, Switzerland) and CD4 cell counts were measured in a certified clinical laboratory, as previously reported (20).

Measurement of HIV-1 p24 in plasma samples from HIV-1–infected patients

Plasma specimens were tested for HIV-1 p24 Ag using the Perkin Elmer HIV-1 ELISA kit as described above. Each plasma sample was treated with the lysis buffer included in the ELISA kit to lyse the viral particles for releasing HIV-1 core protein p24, which was then measured.

Measurement of anti–HIV-1 envelope Abs from HIV-1–infected patients

Anti–HIV-1 envelope Ab was measured using an ELISA Kit for Antibody to Human Immunodeficiency Virus 1&2 (BioChain, Hayward, CA) according to the manufacturer’s protocol. The microplates included in this kit were coated with the rHIV Ags (gp120, gp36, and gp47), which specifically capture anti–HIV envelope Abs. Test results for HIV-2 specific Ab, as measured by the FDA-licensed HIV-2 enzyme immunoassay, were negative in all subjects. Therefore, Abs measured by this ELISA kit were specifically against HIV-1 envelope.

**FIGURE 1.** Generation of rILYd4 and characterization of its functional activity as an inhibitor of hCD59. A, SDS-PAGE separation and Coomassie Blue staining of rILYd4 after purification. B, Confirmation of rILYd4 binding to hCD59 by FACS analysis. Preincubation of rILYd4 inhibits binding of anti-hCD59 Abs to mouse erythrocytes from hCD59^hRBC mice (14) (top panel) and to human erythrocytes (bottom panel). The red curve represents isotype-matched Ab + FITC secondary Abs staining (negative control). The black curve represents ± rILYd4 (0 or 1 μg/ml) + anti-hCD59 Abs (0.2 μg/ml) + FITC secondary Abs. C, rILYd4 blocked the lysis of human erythrocytes induced by 1.2 nM ILY, which mediates ~90% hemolysis in vitro. D and E, rILYd4 abrogates hCD59 function in complement-mediated hemolytic assays. HS = 50% HS; HIS = 50% heat-inactivated HS. D, Alternative pathway assay activated by 5 mg/l cobra venom factor. E, Classical pathway assay with human RBC sensitized with anti-human RBC Ab. Results in C, D, and E are mean ± SD from four independent experiments.
Assessment of the nonspecific cytolytic effect of rILYd4 on erythrocytes and PBMCs from HIV-1–infected patients

Fresh whole blood from HIV-1–infected patients (patients 14 and 17 in Table I) were collected into two tubes, one containing no anticoagulant reagent for serum isolation and one containing potassium EDTA as an anticoagulant for erythrocyte preparation. The tube without anticoagulation was kept at room temperature for 30 min and subjected to centrifugation at 10,000 rpm at 4˚C for 15 min to separate serum from erythrocytes. The serum was aliquoted and stored at 280˚C until used as a source of complement. Some serum aliquots were heated at 56˚C for 30 min to inactivate complement. Erythrocytes and PBMC from the same HIV-1-infected patients were prepared as described above and their sensitivity to complement-mediated lysis in the presence of rILYd4 was assessed as follows. Erythrocytes were washed with PBS, suspended in GVB++ (hematocrit: 2%), treated with rILYd4 (20 μg/ml) at 37˚C for 15 min, and then incubated with or without rabbit anti-human erythrocyte polyclonal Abs at 37˚C for 30 min. Both nonsensitized and sensitized erythrocytes were exposed to 50% serum or plasma (as a source of complement) from the same HIV-1–infected patient. The effect of rILYd4 on PBMCs from the same HIV-1–infected patient was also assessed as follows. PBMCs were prepared as described above, pretreated with rILYd4 (20 μg/ml) at 37˚C for 15 min, suspended in GVB++ and exposed to the same patient’s serum or plasma (50%) as a source of complement. PBMC lysis was assessed by the release of lactate dehydrogenase measured with the CytoTox-ONE kit according to the manufacturers’ instructions (Promega, Madison, WI) (21). The fluorescent signal was recorded at 560/590 nm using Fluostar Optima (BMG Labtech, Cary, NC). The background fluorescence of the PBS buffer was subtracted. Total lysis of PBMCs was induced with Triton X-100. Percent lysis was calculated as: (experimental fluorescent signal – blank fluorescent signal)/(total lysis fluorescent signal)×100.

Results

Generation of human CD59 specific inhibitor rILYd4

rILYd4 (MW, 18.6 kDa) was expressed in E. coli. (Fig. 1A) and purified as described in Materials and Methods. FACS analysis of both human and mouse erythrocytes that transgenically express hCD59 (16) demonstrated that rILYd4 blocked anti-hCD59 monoclonal Ab binding to membrane hCD59 (Fig. 1B). At the functional level, rILYd4 induced a significant and dose-dependent inhibition of human erythrocytes lysis mediated by the intact ILY (1.2 nM; Fig. 1C) and a significant increase of complement-mediated lysis of human erythrocytes triggered by either the alternative (Fig. 1D) or the classical activation pathways (Fig. 1E). No lysis was observed when human or hCD59 transgenic mouse erythrocytes were exposed to rILYd4 alone. Together, these experiments demonstrate that rILYd4 binds and inhibits the function of hCD59. These results confirm and extend our previous communication (22) and are comparable to those recently published by Hughes et al. (23).
rILYd4 sensitizes the HIV-1 virions derived from CD59-positive cell lines to complement-mediated virolysis activated by anti–HIV-1 envelope Abs

As mentioned above, hCD59 protects HIV-1 from complement-mediated virolysis (3, 24). To test whether inhibition of hCD59 by rILYd4 sensitizes HIV-1 to complement-mediated virolysis, we derived HIV-1 particles from two different HIV-1–infected host human monocyte cell lines: THP-1 expresses and U1 is devoid of hCD59 in the cell membranes, as documented by FACS analysis with an anti-hCD59 specific Ab (Ab 2F1). Virolysis was quantitated by an ELISA that measured the release of HIV-1 core protein p24 from the lysed viral particles. HIV-1 virions derived from hCD59-negative U1 cells were highly sensitive to virolysis induced by anti–HIV-1 gp120 monoclonal Ab plus HS as a source of complement (Fig. 2B). In contrast, HIV-1 virions derived from hCD59-positive THP-1 cells were resistant to complement-mediated virolysis (Fig. 2B). Remarkably, preincubation of the HIV-1 virions derived from the infected THP-1 cells with rILYd4 rendered the virus sensitive to complement-mediated lysis. This virolysis enhancement did not occur in HIV-1 virions derived from hCD59-negative U1 cells (Fig. 2C). In the presence of 20 μg/ml rILYd4, THP-1 cell-derived virions were almost as sensitive to complement-mediated lysis as the virions derived from CD59 negative U1 cells (Fig. 2C). Similar dose-dependent results were obtained with 1) HIV-1 virions derived from two additional human T-lymphocytic cell lines (OM10 and H9), which express a high density of hCD59 on their cell membranes (Fig. 2D), and 2) with a different anti–HIV-1 Ab (anti-gp120/160 polyclonal Abs; Fig. 2E). Blocking hCD59 function with a neutralizing anti-hCD59 Ab increased complement-mediated virolysis, albeit with less potency than with rILYd4 (Fig. 2F). In all experiments depicted in Fig. 2B, 2C, 2E, and 2F, virolysis in the presence of HS is complement-mediated because it is totally abrogated by preincubation of serum at 56°C for 1 h. These results demonstrate that inhibition of hCD59 function by rILYd4 sensitizes HIV-1 virions to complement-mediated virolysis.

Abs in the plasma from the HIV-1–infected subjects exhibit their anti–HIV-1 activity in the presence of rILYd4

The presence of anti–HIV-1 envelope Abs in the blood of HIV-1–infected subjects fostered the notion that a hCD59 inhibitor would not protect and thereby sensitize circulating HIV-1 virions to complement-mediated virolysis. We tested this predication ex vivo by exposing HIV-1 virions derived from a hCD59 positive cell line to heat-inactivated plasma samples from either HIV-1–infected (HIV-1plasma) or control plasma from HIV-1 seronegative donors (Controlplasma), followed by incubation with a pool of normal HS as a source of complement. Table I shows that the plasma levels of Abs against HIV-1 envelope measured by ELISA. Although the Ab titer varied among HIV-1 patients, every plasma sample from this cohort contained anti–HIV-1 envelope Abs. Preincubation with rILYd4 dramatically increased complement-mediated virolysis of CD59-positive virions exposed to HIV-1plasma, but not to the Controlplasma (Fig. 3A–C). This effect of rILYd4 was comparable with, albeit much stronger than, the effect mediated by the anti-hCD59 monoclonal Ab BRIC229 (Fig. 3B).

To understand the functional consequence of complement-mediated virolysis we used an HIV-1 infectivity assay. HIV-1-permissive H9 cells were exposed for 10 d to each conditioned medium from the virolysis experiments depicted in Fig. 3A and 3B (samples 1–4). The culture supernatants were then collected to determine HIV-1 infection intensity by measuring HIV-1 p24 (ELISA). The higher the level of p24 in the culture supernatant, the higher the number of infectious viral particles that remained in the conditioned medium of the virolysis experiments. Fig. 3D shows that p24 was undetectable in the supernatant from H9 cells exposed to conditioned medium from Triton X-100 treatment (total lysis), indicating that potentially infective particles were totally lysed and no infectious viral particles remained. The supernatants from cells exposed for 10 d to control conditioned medium (i.e., conditioned by the virions not treated with rILYd4 before exposure to endogenous anti–HIV-1 Ab and complement) had high titers of p24, indicating that the viral particles were not lysed and their infectivity was preserved. In contrast, supernatants from cells exposed for 10 d to conditioned medium from rILYd4 pretreated virions (treated with rILYd4 before exposure to endogenous anti–HIV-1 Ab and complement) showed low levels of p24, an indication that rILYd4 allowed the anti–HIV-1 Abs present in the plasma of HIV-1–infected patients to regain their activity in triggering complement-mediated virolysis and thereby reduce the infective potential of HIV-1 virions.

Table I. Profiles of HIV-1–infected patients

<table>
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<tr>
<th>Patient No.</th>
<th>Clinical Stage</th>
<th>CD4+ T Cell (count/μl)</th>
<th>Plasma HIV-1 RNA (copies/ml)</th>
<th>Treatment at Times of Study</th>
<th>Duration HIV-1 Diagnosis (y)</th>
<th>Duration of Antiviral Therapy (y)</th>
<th>p24 (pg/ml)</th>
<th>Anti-HIV-1 Envelope Ab Titer</th>
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The sera from patients 1–16 were used for the experimental results shown in Fig. 3. The serum from patient 17 and plasma from patient 14 were used for the experimental result shown in Fig. 6.

NA, not available.
rILYd4 sensitizes primary HIV-1 isolates to complement-mediated virolysis activated by anti–HIV-1 Abs present in the plasmas of HIV-1–infected patients

Results obtained from primary HIV-1 isolates are considered more representatives of the physiologic conditions of HIV-1–infected patients than those obtained from cultured cell lines. In the specific case of hCD59, primary HIV-1 isolates derived from PBMC of HIV-1–infected patients are likely to carry a load of membrane-derived human CD59, which confers resistance to the Ab-dependent complement-mediated virolysis in the presence of rILYd4. For these reasons, we generated primary HIV-1 isolates from six HIV-1–seropositive individuals who were naive for antiretroviral therapy, and we tested whether rILYd4 sensitizes these virions to complement-mediated virolysis. In the same experiment, we assessed the relative potency of endogenous anti–HIV-1 Abs developed by HIV-1–infected patients to promote complement-mediated virolysis of PBMC-derived HIV-1 primary isolates in the presence and absence of rILYd4. To this end, we pretreated the primary HIV-1 isolates with or without rILYd4 and exposed them to heat-inactivated HIV-1plasma (patients 1 to 5 in Table I), followed by incubation with pooled normal HS as a source of complement. The results showed that rILYd4 sensitized each of the six primary HIV-1 isolates to complement-mediated virolysis activated by HIV-1plasma (Fig. 4). In the presence of rILYd4, each of the five different HIV-1plasma samples tested significantly increased complement-mediated lysis of each of the six primary HIV-1 isolates (Fig. 5). These effects of rILYd4 were comparable with, albeit much stronger than, those mediated by the anti-hCD59 monoclonal Ab BRIC229 (Figs. 4 and 5). These results confirm that rILYd4 sensitizes HIV-1 to complement-mediated virolysis not only under experimental conditions using cell lines and commercially available Abs, but also of primary HIV-1 isolates sensitized by the endogenous anti–HIV-1 Abs naturally present in the blood of HIV individuals. These results also indicate that inhibition of hCD59 with rILYd4 unprotects HIV-1, unleashing the ability of complement to lyse the virions sensitized by anti–HIV-1 Abs present in the circulation of patients with HIV-1.

rILYd4 does not mediate cytolytic effect on erythrocytes and PBMC in the blood from HIV-1–infected patients

Patients infected with HIV-1 are well known to mount a vigorous and sustained Ab response to the virus (25–30). We consistently detected anti–HIV-1 envelope Abs in all 16 plasma samples from HIV-1–infected patients.
patients (Table I). When hCD59 activity is inhibited by rILYd4, these Abs could conceivably trigger unwanted complement-mediated effects, such as hemolysis. To address this issue, we investigated the potential lytic effect of rILYd4 on erythrocytes and PBMC from the HIV-1–infected patients with high levels of plasma HIV-1 RNA (patients 14 and 17 in Table I). We found that incubation of either erythrocytes or PBMC from HIV-1–seropositive individuals with 50% of their own serum (patient 17) or plasma (patient 14) in the presence of rILYd4 at the concentration that completely blocked hCD59 function did not result in any lytic effect of either cell type (Fig. 6A, 6B). This experiment indicates that rILYd4 does not induce complement-mediated lysis of cells not infected by HIV-1, such as erythrocytes. Although the infected CD4+ T cells in PBMC are susceptible to HIV-1 infection and might be lysed in the presence of rILYd4, they represent a small proportion of the overall PBMC population (0.001%–10%) (31), and their potential complement-mediated lysis in the presence of anti–HIV-1 Abs and rILYd4 is probably undetectable with the methods used for these experiments. These results suggest that bystander lytic effects induced by rILYd4 in the circulation of HIV-1–infected individuals are unlikely.

Discussion
In this study, we report the development of rILYd4, a potent and specific inhibitor of hCD59. We show that rILYd4, in conjunction with anti–HIV-1 Abs, either exogenous like the anti-gp120/160 polyclonal Abs or endogenous such as those in plasma from HIV-1–infected patients, efficiently abrogates hCD59 function and renders complement-resistant laboratory strains of HIV-1 sensitive to Ab-dependent, complement-mediated lysis. We also show that, in the presence of rILYd4, anti–HIV-1 Abs in the circulation of HIV-1–infected individuals are capable of triggering complement-mediated virolysis of primary HIV-1 isolates. Inhibition of hCD59 activity by rILYd4 in erythrocytes or PBMC from HIV-1–infected individuals does not induce unwanted lytic effects.

The complement system, a key member of innate immunity, is a first-line defender against foreign pathogens such as HIV-1. However, HIV-1 in the circulation escapes complement-mediated attack and remains highly infective, even though there is strong experimental evidence that both the virus itself and anti–HIV-1 Abs in the blood of HIV-1–infected individuals are capable of activating the complement cascades (32). Indeed, HIV-1 virions from infected individuals accumulate C3 on their surface, an indication of complement activation (33). Normally, component C3 activation generates C3a and C3b, which then trigger a cascade of activation events that eventually result in formation of the MAC, an end-product of all the three complement activation pathways (2, 34). The MAC forms a lytic pore in the lipid bilayer membrane that destroys membrane integrity, allows the free passage of solutes and water, and eventually kills pathogens including viruses and/or infected cells. In HIV-1 infection, however, activation of complement, as evidenced by accumulation of C3 on the surface of the HIV-1 virions, fails to induce the HIV-1 lysis and enhances the viral infectivity by facilitating the interaction of the HIV-1 particles with complement-receptor–positive cells, including B and dendritic cells (35). Incomplete activation of complement enabling HIV-1 to escape its immune detection and increasing its pathogenic potential represents a major challenge for vaccine developers.

FIGURE 4. HIV-1 primary isolates become sensitive to complement-mediated virolysis in the presence of rILYd4. HIV-1 primary isolates derived from six HIV-1–infected patients. PBMCs preincubated with rILYd4 (20 μg/ml), medium only, or anti-hCD59 monoclonal Ab (BRIC 229) were treated with heat-inactivated plasma from 5 HIV-1–positive individuals containing anti–HIV-1 envelope Abs (patients 1–5 shown in Table I) followed by exposure to 10% normal HS as a source of complement (heat-inactivated normal serum was used as a negative control). Each panel represents the sensitivity of HIV-1 virions derived from one patient to complement-mediated virolysis activated by the endogenous anti–HIV-1 Abs developed in five HIV-1–infected patients who were naive for antiretroviral therapy. Horizontal lines represent the mean. Statistical significance (p < 0.01 versus medium treatment group) is indicated by an asterisk.
Ab-dependent, complement-mediated lysis is due in part to the presence of hCD59 in the viral envelope, which the virus recruits from the host cell in the budding process (3, 10, 11). In addition, binding of the fluid phase complement regulator factor H to HIV-1 confers onto the virus further protection from complement attack (36). In summary, the HIV-1 virus, like other pathogens such as *Schistosoma mansoni* (37), manipulates the delicate balance between complement activation and restriction in a manner that is favorable to the virus.

For these reasons, it has long been suggested that an inhibitor of hCD59 or an agent that would abrogate factor H binding to the HIV-1 envelope could have a beneficial therapeutic effect against HIV-1 infection and AIDS (11, 33). The experiments reported in this paper indicate that rILYd4, a high affinity inhibitor of hCD59, could represent such a sought after therapeutic tool. Whether rILYd4 also abrogates the protective effect of factor H binding to HIV-1 is not known at present. More importantly, our experimental results show that rILYd4 abrogates hCD59 function and fosters complement-mediated cytolysis in human nucleated cells such as lymphocytes, which are the natural reservoir of HIV-1 in infected individuals (Supplemental Fig. 1). It has been documented that patients with HIV-1 infection also have diminished expression of the GPI-anchored cell surface proteins CD55 and CD59 on the erythrocytes and granulocytes (38). Because the density of hCD59 on the surface of HIV-1–infected cells appears to be reduced, one would expect that primary virions would carry less hCD59. Different density of CD59 in the membrane of laboratory or primary HIV-1 virions could explain different sensitivities to induction of complement-mediated lysis by rILYd4 (Fig. 3). In patients, a lower density of hCD59 in infected than in noninfected cells would enhance the efficacy of rILYd4 in specifically eliminating HIV-1 virions and HIV-1–infected cells. In summary, the results presented in this paper indicate that rILYd4 represents a preclinical candidate that deserves further investigation as a potential therapeutic agent against HIV-1 infection and AIDS.

However, in the discussion of a potential therapeutic use of rILYd4, it is important to highlight that abrogation of hCD59 function in humans has the potential of inducing complement-mediated side effects. A pertinent example of the potentially harmful effects of abrogating homologous restriction on the surface of “self” cells is illustrated by the human disease paroxysmal nocturnal hemoglobinuria (PNH), in which bone marrow–derived circulating cells are deficient in CD59 and other GPI-anchored proteins (39–43). Patients with PNH usually exhibit a mild hemolytic anemia, attributed to complement activation at its basal “tick over” rate, and develop paroxysmal hemolytic crisis when infections of other stressors trigger more massive complement activation (39, 40). Of note, patients with a variety of autoimmune disorders also have diminished expression of the GPI-anchored cell surface proteins CD55 and CD59 on erythrocytes and granulocytes (44). Whether administration of rILYd4 to humans with PNH or autoimmune disorders will trigger hemolysis and/or other unwanted complement-mediated phenomena is not known at present and raises a question that can be answered only by human studies. Our preliminary attempts to address the potential complement-mediated adverse effects of inhibiting hCD59 function ex vivo have shown that incubation of erythrocytes and PBMCs from normal or HIV-1–infected individuals in 50% HS or human plasma (HP) in the presence of rILYd4 at a concentration that maximally inhibits hCD59 function did not result in any spontaneous cytolysis. This finding indicates that the basal activity of complement in plasma/serum (as assessed ex vivo) is not sufficient to lyse human erythrocytes and PBMCs in the presence of...
An interesting advantage of rILYd4 over current anti–HIV-1 pharmacologic therapies derives from the fact that hCD59 present in the viral surface is not encoded in the viral genome, but rather a human cell protein recruited from the membrane of the host cell in the budding process. For this reason, it is less likely that the HIV-1 virus in infected individuals would acquire resistance to rILYd4 through a high rate of mutation and recombination of viral proteins, as it does to all currently available anti–HIV-1 drugs and vaccines (48–50).

In conclusion, it is tempting to postulate that administration of rILYd4 to HIV-1–infected individuals would allow anti–HIV-1 Abs to unleash their capacities to induce complement-mediated virolysis of both free viral particles and infected cells with no or minimal effect on other bystander cells. Further investigation of the potential therapeutic applications of rILYd4 for HIV-1 treatment certainly deserves consideration toward preclinical development and eventually clinical trials.

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We dedicate this article to the memory of Dr. Daniel. C. Tosteson, Dean Emeritus of Harvard Medical School, who was an uniring mentor and role model to us.

**Disclosures**

The authors have no financial conflicts of interest.

**References**


A rILYd4 concentration that dramatically increased lysis when complement was activated by either the classical or alternative pathways (Figs. 1D, 1E, 6).

Plasma from HIV-1–infected patients, but not from healthy individuals, contains anti–HIV-1 Abs that promote complement-mediated virolysis of HIV-1 primary isolates in the presence of rILYd4 (Fig. 3). These endogenous anti–HIV-1 Abs developed by HIV-1–infected individuals in both the acute and chronic phases of infection activate complement through the classical pathway, as indicated by published reports that C1q-deficient serum as a source of complement fails to induce anti–HIV-1 Ab-mediated virolysis (45). Fig. 3B shows that plasmas from HIV-1–infected individuals segregate into two distinct populations: one with higher and one with lower capacity to induce complement-mediated virolysis in the presence of rILYd4. This finding may be attributed to the different Ab titers and/or different complement-activating capacity of the endogenous anti–HIV-1 Abs present in those plasmas. Additional specificity for complement activation on the viral surface would be provided by direct activation of complement triggered by gp120, two complement-activating proteins present in the HIV-1 envelope (46, 47). Direct complement activation by viral proteins explains the complement-mediated virolysis seen in the presence of rILYd4, but in the absence of sensitizing anti–HIV-1 Abs (Fig. 3C).

![FIGURE 6. Incubation of HIV-1–infected patient’s erythrocytes and PBMCs with 50% HIV-1–infected patient’s serum or plasma in the presence of rILYd4 did not result in a nonspecific lytic effect on the erythrocytes and PBMCs. A, The HS and from HIV-1–infected patient (patient 17 in Table 1) was used as a source of complement and anti–HIV-1 Abs to investigate the bystander effect of rILYd4 on the erythrocytes. B, The HP from an HIV-1–infected patient (patient 14 in Table 1) was used as a source of complement and anti–HIV-1 Abs to investigate the bystander effect of rILYd4 on the RBCs or PBMCs. The patient’s erythrocytes were sensitized by anti-human erythrocyte Abs and then exposed to 50% HS or HP as well as heat-inactivated HS or HP; rILYd4 + Abs, in the presence of rILYd4 (20 μg/ml), HIV-1–infected patients’ erythrocytes were sensitized by anti-human erythrocyte Abs and then exposed to 50% HS or HP and heat-inactivated HS or HP; sp < 0.01 versus heat-inactivated HS or HP treatment in rILYd4 + Abs.](http://www.jimmunol.org/content/jimmunol/67/12/3674/F6.expansion.png)
INHIBITION OF HUMAN CD59 FOR THE TREATMENT OF AIDS INFECTION


