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Female and Male Sex Hormones Differentially Regulate Expression of *Ifi202*, an Interferon-Inducible Lupus Susceptibility Gene within the *Nba2* Interval

Ravichandran Panchanathan,* Hui Shen,* Melanie Gubbels Bupp,**† Karen A. Gould,‡ and Divaker Choubey*‡

Increased expression of IFN-inducible *Ifi202* gene in certain strains of female mice is associated with susceptibility to systemic lupus erythematosus (SLE). Although, the development of SLE is known to have a strong sex bias, the molecular mechanisms remain unknown. Here we report that in vivo treatment of orchiectomized (NZB × NZW)F₁ male mice with the female sex hormone 17β-estradiol significantly increased steady-state levels of *Ifi202* mRNA in splenic cells, whereas treatment with the male hormone dihydrotestosterone decreased the levels. Moreover, increased expression of *Ifi202* in B6.Nba2 B cells and reduced expression in T cells were associated with increased levels of estrogen receptor-α (ERα) and androgen receptor, respectively. Furthermore, the steady-state levels of *Ifi202* mRNA were higher in splenic cells from C57BL/6, B6.Nba2, NZB, and (NZB × NZW)F₁ female mice as compared with males. 17β-estradiol treatment of B cells and WT276 cells increased *Ifi202* mRNA levels, whereas treatment with dihydrotestosterone decreased the levels. Interestingly, overexpression of ERα in WT276 cells increased the expression of *Ifi202* and stimulated the activity of the 202-luc-reporter through the c-Jun/AP-1 DNA-binding site. Accordingly, ERα preferentially associated with the regulatory region of the *Ifi202* gene in female B6.Nba2 B cells than in males. Furthermore, *Ifi202* mRNA levels were detectable in splenic cells of wild-type (*Esr1*+/+), but not null (*Esr1−/−), (NZB × NZW)F₁ female mice. Collectively, our observations demonstrate that the female and male sex hormones differentially regulate the expression of *Ifi202*, thus providing support for the role of *Ifi202* in sex bias in SLE.

*Abbreviations used in this paper: SLE, systemic lupus erythematosus; AR, androgen receptor; Chip, chromatin immunoprecipitation; DHT, dihydrotestosterone; E2, 17β-estradiol; ER, estrogen receptor; ERE, estrogen responsive core element.

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7031–7038.
Binding of 17β-estradiol (E2) to ERs results in activation of ERs and transcriptional activation of ER target genes (15–17). Many ER target genes contain a minimal estrogen responsive core element (ERE) sequence (GGTCANNNTGACC) in the 5’ regulatory or promoter region. The ERE sequence functions in an orientation- and distance-independent manner, both of which are characteristics of an enhancer (21). Moreover, ER is also known to bind DNA through half ERE sites (GGTCAN) (15, 21). Because molecular mechanisms of the recruitment of ER to the promoter region of its target genes remain relatively complex, it remains an actively investigated research area. Importantly, proteins that are encoded by the ER target genes mediate many of the biological activities of female sex hormone estrogen (15–17).

Male sex hormone androgen signals via the intracellular androgen receptor (AR), a member of the superfamily of nuclear hormone receptors (22). Androgen-dependent activation and nuclear translocation of the AR is followed by its binding to specific response elements in the promoter regions of target genes to modulate gene expression either positively or negatively (22). Interestingly, expression of AR mRNA has been reported in enriched populations of CD4+ T lymphocytes, CD8+ T lymphocytes, and macrophages (23). However, the enriched populations of B lymphocytes expressed only low levels of AR mRNA (23).

IFN-inducible Ifi200 gene family includes structurally and functionally related murine (E.G., Ifi202a, Ifi202b, Ifi203, Ifi204, Ifi205, and AminH) and human (E.G., IFI16, MNDAA, AIM2, and IFIX) genes (24–30). The Ifi202a and Ifi202b are highly homologous murine genes that encode p202a and p202b proteins, respectively (26, 27). These two proteins differ in only 7 amino acids (out of 445 amino acids) (27). Because Abs, which have been raised against the p202a protein (31), also detect p202b protein (26, 27), in this study, we have referred both p202a and p202b proteins as p202 protein.

Generation of B6.Nba2 congenic (congenic for NZB-derived Nba2 interval on C57BL/6 genetic background) mice and gene expression analyses identified Ifi202 (probably both Ifi202a and Ifi202b genes) gene as a lupus susceptibility gene (26, 32). Importantly, consistent with promoter polymorphisms contributing to differential expression of Ifi202a gene between C57BL/6 and NZB mice (26, 32), increased steady-state levels of Ifi202a and Ifi202b mRNAs (as compared with C57BL/6 mice) are detectable in splenic cells from NZB and B6.Nba2 mice (33). Moreover, levels of Ifi202a mRNA are relatively higher than the Ifi202b mRNA (33). Interestingly, increased expression of p202 protein (probably both p202a and p202b proteins) in B6.Nba2 splenic B and T cells (more in B cells than in T cells) is associated with defects in apoptosis of B cells and increased susceptibility to develop lupus-like disease (26, 32). Furthermore, the B6.Nba2 congenic female mice produce higher levels of antinuclear autoantibodies than did the age-matched male mice, and (B6.Nba2 × NZW/WF1) female mice develop severe proteinuria with much higher frequency (34). These observations prompted us to investigate whether sex hormones could regulate expression of the Ifi202 gene. Here, we report that female and male sex hormones differentially regulate the expression of Ifi202.

Materials and Methods

Mice, orchietomy, and sex hormone treatment

Spleens were isolated from wild-type (Esr1+/+) or null (Esr1−/−) (NZB × NZW/WF1, female mice) (20) (age ~10 wk) that were housed in animal facilities of University of Nebraska Medical Center (Omaha, NE). Age-matched (6–8 wk old) male and female nonautoimmune (C57BL/6J) and preautoimmune (B6.Nba2, NZB, and (NZB × NZW/WF1) mice were purchased from The Jackson Laboratory and housed in the animal facilities of the University of Cincinnati. All animal experimental protocols were approved by the Institutional Animal Care and Use Committee at the institution where animals were housed.

Male (NZB × NZW/WF1) mice were orchietomized at 3 mo of age as described (35). After orchietomy, pellets (Innovative Research of America) releasing E2, dihydrotestosterone (DHT), or placebo for up to 3 wk were inserted s.c. with a 10-gauge needle. Serum was collected on day 7, 8, or 9 and then analyzed by the Endocrine Laboratory at Colorado State University (Fort Collins, CO) for luteinizing hormone, estradiol, and testosterone by radioimmunoassay as described (35). Orchietomized male mice that were treated with E2 pellets exhibited serum E2 levels of 162 pg/ml, while levels in placebo-treated mice were not detectable (data not shown). Intact and orchietomized mice treated with DHT had similarly low luteinizing hormone levels (<1 ng/ml), while orchietomized mice treated with placebo had >20 ng/ml luteinizing hormones (data not shown). Orchietomized male mice treated with placebo demonstrated testosterone levels <2 ng/ml, while intact (NZB × NZW/WF1) male mice exhibited 12 ng/ml testosterone (35).

Splenocyte isolation, purification of B or T cells, cell culture, and treatments

Total single-cell splenocytes were prepared from age-matched male or female mice as described previously (36). After lysis of RBC, splenocytes were resuspended in RPMI 1640 medium supplemented with 10% FBS, 1% penicillin/streptomycin/glutamate, and 1 mM, nonessential amino acids/sodium pyruvate. Unless otherwise indicated, splenic cells from two or more age-matched male or female mice were pooled to purify B or T cells and to prepare total RNA or protein extracts. B or T cells were purificed from splenic cells using magnetic beads (purification kit purchased from Milteny Biotec), allowing the positive selection of either B or T cells. The purified (>90–95% pure) T and B cells were used immediately for additional experiments.

Estrogen-responsive mouse breast cancer cell line WT276 (37) was provided by Dr. J. Welsh (University of Notre Dame, Notre Dame, IN). Cells were maintained in DMEM medium supplemented with 10% FBS and 1× antibiotic-antimycotic solution (Invitrogen). For treatment of WT276 cells with E2 or DHT, cells were cultured in phenol red-free RPMI 1640 medium (Invitrogen) and the medium was supplemented with 10% charcoal-stripped FBS (Invitrogen). Cells were treated in vitro with either DHT (26) or E2 (38) at the concentration used previously.

Plasmids and expression vectors

Dr. P. Chambon (Centre National de la Recherche Scientifique, France) provided ERα expression plasmid that allowed expression of ERα (39). The 202-luc-reporter (36) and the mutant 202AP-1mutCS1-luc reporter (40) plasmids have been described.

Reporter assays

For reporter assays, subconfluent cultures of WT276 cells (in 6-well plates) were transfected with the 202-luc (2.5 μg) or the 202AP-1mutCS1-luc (with mutated AP-1 CS1, Ref. 40) reporter plasmid along with pRL-TK reporter plasmid (0.5 μg), using calcium phosphate transfection kit (Invitrogen), as suggested by the supplier. When indicated, cells were either treated with ethanol (vehicle) or the indicated concentration of E2 or DHT for 16–24 h. Unless otherwise indicated, cells were harvested between 40 and 45 h after transfections. Cells were lysed, and the firefly and Renilla dual luciferase activities were determined as described previously (36). Student’s t test for paired samples was used to determine statistical significance of the reporter activity data. Differences were considered statistically significant at p ≤ 0.05.

Isolation of RNA from splenocytes and RT-PCR

Splenocytes (5–8 × 10^6 cells) were used to isolate total RNA using TRIzol (Invitrogen). Total RNA was digested with DNase I (to remove any contaminating genomic DNA in the preparation), and 0.5–2 μg of RNA was used for RT-PCR reaction using a pair of the Ifi202 primer (forward, 5′-ctgctatcagctacagctag-3′; reverse, 5′-ctcctagagcaagctgggt-3′). For RT-PCR, we used the SuperScript One-Step RT-PCR system from Invitrogen. Primers for Esr1 gene (forward, 5′-aattctgacaatcgacgccag-3′; reverse, 5′-ctcctagagcaagctgggt-3′).

Quantitative real-time TaqMan PCR technology (7300 Real-Time PCR System; Applied Biosystems) and commercially available real-time TaqMan probes were used to compare expression levels of Ifi202 between male and female mice. The PCR cycling program consisted of denaturation at 95°C for 10 min, 40 cycles at 95°C for 15 s, followed by annealing and elongation at 60°C for 1 min. The TaqMan assays for Ifi202
Chromatin immunoprecipitation (ChIP) assays

Splenic B cells were purified using magnetic beads, and the purified splenic B cells (−2–4 × 10⁶) were processed for ChIP assays using Champion-ChIP One-Day Kit (SA Bioscience) as suggested by supplier. In brief, cell lysates containing equal amounts of protein were immunoprecipitated with either an isotype Ab or anti-ERα Abs (sc-542X; Santa Cruz Biotechnol-ogy). The immunoprecipitates were collected using protein A beads. Immune complexes were eluted from beads, proteins were digested, and DNA was collected. The isolated DNA was purified, precipitated, washed, and dissolved in water. Semiquantitative PCR was performed with DNA samples for 28 cycles. PCR products were resolved in an agarose gel and visualized. The PCR primers that were used have been described previously (41). Quantitative PCR was performed using commercially available primer set (GPM0141367; SA Bioscience) corresponding to the genomic 5′ regulatory region of the Ifi202 gene and real-time PCR conditions as de-sccribed above.

Immunoblotting

Total splenocytes, WT276, or NIH 3T3 cells were collected in PBS and resuspended in a modified radioimmunoprecipitation assay (RIPA) lysis buffer (50 mM Tris-HCl (pH 8.0), 250 mM NaCl, 1% Nonidet P-40, 0.5% sodium deoxycholate, 0.1% SDS), supplemented with 1× protease inhibitor (Roche Diagnostics), and incubated at 4°C for 30 min. Cell lysates were sonicated briefly before centrifugation at 14,000 rpm in a microcen-trifuge for 10 min at 4°C. The supernatants were collected, and the protein concentration was measured by Bio-Rad protein assay kit. Equal amounts of protein were processed for immunoblotting. Antiserum to p202 protein has been described previously (31). The p202 antiserum detects both p202a and p202b proteins in immunoblotting (27, 31). Abs to detect mouse ERα (sc-542; MC-20), AR (sc-816), and β2-microglobulin (sc-13565) were purchased from Santa Cruz Biotechnology. Abs to detect β-actin (no. 4967) were purchased from Cell Signaling Technology.

Statistical analyses

Data are presented as mean ± SEM. A p value of <0.05 was considered statistically significant. These methods were performed using GraphPad Prism 5.02 software for Windows (GraphPad Software).

Results

In vivo treatment of orchietomized male mice with estrogen increased steady-state levels of Ifi202a mRNA, whereas treatment with DHT reduced the mRNA levels.

To investigate the role of sex hormones in the regulation of the Ifi202 gene, we chose to compare expression levels of Ifi202a mRNA (because splenic cells express both Ifi202a and Ifi202b genes and steady-state levels Ifi202a mRNA are more than Ifi202b; see Ref. 33) in (NZB × NZW)F1 male mice (age 12 wk) that were orchietomized and reconstituted with slow-releasing pellets releasing E2, DHT, or placebo. After 3 wk of the reconstitution, splenic cells were analyzed for steady-state levels of Ifi202a mRNA by real-time PCR. As shown in Fig. 1, treatment of male mice with the E2-releasing pellet, under our experimental conditions (35), measurably increased levels of Ifi202a mRNA in most of the E2-treated mice as compared with control mice. Interestingly, treatment of mice with DHT decreased the Ifi202a mRNA levels to a measurable extent in most mice as compared with the control mice. These observations are consistent with the possibility that increased in vivo levels of female sex hormone E2 in orchietomized (NZB × NZW)F1 male mice contribute to increased steady-state levels of Ifi202a mRNA. Moreover, our observations also indicated that increased levels of male sex hormone androgen in mice contribute to decreased steady-state levels of Ifi202a mRNA.

Increased expression of Ifi202 in splenic B cells in female mice is associated with increased levels of ERα and reduced levels of AR

Our earlier studies had revealed that splenic B cells from preautoimmune (4-mo-old) B6.Nba2 female mice express higher levels of Ifi202 mRNA as compared with T cells (32). Therefore, our above observations (Fig. 1) that in vivo treatment of orchietomized (NZB × NZW)F1 male mice with estrogen increased Ifi202a mRNA levels, whereas treatment with DHT reduced the mRNA levels, prompted us to compare steady-state levels of Ifi202 mRNA in splenic B and T cells from male and female preautoimmune (age ~10 wk) B6.Nba2 mice (32). We noted that steady-state levels of Ifi202 mRNA were significantly (~7-fold) higher in splenic B cells than in T cells in the female mice (Fig. 2A). Because sex hormones, such as E2 and DHT, regulate gene expression through binding to their respective receptors (15, 22), we also compared expression levels of ERα, AR, and p202 proteins in B and T cells from female mice and the age-matched male mice (Fig. 2A). Because sex hormones, such as E2 and DHT, regulate gene expression through binding to their respective receptors (15, 22), we also compared expression levels of ERα, AR, and p202 proteins in B and T cells from female mice and the age-matched male mice (Fig. 2A). Because sex hormones, such as E2 and DHT, regulate gene expression through binding to their respective receptors (15, 22), we also compared expression levels of ERα, AR, and p202 proteins in B and T cells from female mice and the age-matched male mice (Fig. 2A). Because sex hormones, such as E2 and DHT, regulate gene expression through binding to their respective receptors (15, 22), we also compared expression levels of ERα, AR, and p202 proteins in B and T cells from female mice and the age-matched male mice (Fig. 2A).
Sex-dependent regulation of *Ifi202* expression

Our earlier studies had revealed that increased expression of *Ifi202* in preautoimmune B6.Nba2 female mice (as compared with nonautoimmune C57BL/6 mice) is associated with increased lupus susceptibility (26, 32). Because steady-state levels of *Ifi202* mRNA are very low (as compared with age-matched B6.Nba2 mice), but detectable in C57BL/6 mice (33), we decided to compare the steady-state levels of *Ifi202* mRNA between male and female C57BL/6 mice. We noted that basal levels of *Ifi202* mRNA were detectable in splenic cells from C57BL/6 male and female mice (age ~10 wk), and IFN treatment of cells increased the levels further (Fig. 3A). Interestingly, the basal levels of *Ifi202* mRNA were ~5-fold higher in C57BL/6 females than age-matched males. This observation indicated that steady-state levels of *Ifi202* mRNA are regulated in sex-dependent manner in C57BL/6 splenic cells.

Encouraged by the above observations, we also compared the steady-state levels of *Ifi202* mRNA between young male and female lupus-prone mice (B6.Nba2, NZB, and (NZB×NZW)F₁). As shown in Fig. 3B, basal steady-state levels of the *Ifi202* mRNA were consistently higher in splenic cells from the females than from the age-matched males. Taken together, these observations suggested that the steady-state levels of *Ifi202* mRNA in splenic cells are regulated in a sex-dependent manner.

Treatment of WT276 cells with female or male sex hormone regulates *Ifi202* expression

The mouse mammary tumor cell line WT276 has been reported to be estrogen-responsive (37). Therefore, to identify molecular mechanisms by which E2 treatment up-regulates expression of *Ifi202* gene, we explored whether treatment of WT276 cells with sex hormones could regulate the expression of *Ifi202* gene. For this purpose, we treated cells with increasing concentrations (1, 5, or 10 nM) of female sex hormone E2 (these concentrations were chosen based on an earlier study; see Ref. 38) for 16 h. As shown in Fig. 4A, the treatment with E2 resulted in an increase in steady-state levels of *Ifi202* mRNA as determined by semiquantitative RT-PCR. Moreover, consistent with E2-mediated up-regulation of *Ifi202* expression by E2α, treatment of cells with tamoxifen (100 nM), a selective estrogen receptor modulator (42), which resulted in increases in E2α mRNA and protein levels (data not shown), abrogated the E2-mediated increases in *Ifi202* mRNA levels (data not shown). Consistent with the above observations, we also noted increases in p202 protein levels after treatment of cells with increasing concentrations of E2 (Fig. 4B). Interestingly,
treatment of cells with 5 nM concentration of E2 resulted in increases in p202 protein levels (compare lane 3 with lane 2). However, treatment of cells with 10 nM E2 resulted in moderate decreases in p202 protein levels (compare lane 4 with lane 3). Furthermore, treatment of WT276 cells with the male sex hormone DHT decreased basal levels of p202 protein in a dose-dependent manner. Taken together, the above observations demonstrated that in vitro treatment of WT276 cells with female sex hormone E2 or male sex hormone DHT differentially regulated the levels of the p202 protein.

**Estrogen through ERα up-regulates expression of the Ifi202 gene**

To further investigate whether estrogen treatment of cells activates transcription of the Ifi202 gene through ERα, we first transfected estrogen-responsive WT276 cells with 202-luc-reporter plasmid and treated cells with increasing concentrations of E2. As shown in Fig. 5A, treatment of cells with increasing concentrations of E2 stimulated the activity of the 202-luc-reporter in a concentration-dependent manner. Next, we transfected WT276 cells with 202-luc-reporter along with an empty vector or a plasmid encoding the ERα receptor. After transfections, cells were either treated with ethanol alone (vehicle) or 10 nM E2. As shown in Fig. 5B, transfection of cells with the plasmid encoding ERα protein stimulated the activity of the 202-luc-reporter. Interestingly, transfection of cells with the plasmid encoding the ERα protein and subsequent treatment of the transfected cells with E2 strongly stimulated the activity of the reporter. To further test whether ERα regulates expression of the Ifi202 gene, we transfected NIH 3T3 mouse fibroblasts (we chose these cells because basal levels of p202 protein are detectable and these cells are not known to express ERα) with either an empty vector or the plasmid encoding ERα (cells treated with 5 nM E2 in phenol red-free medium) and analyzed the expression of p202 protein. As shown in Fig. 5C, ectopic expression of ERα protein in NIH 3T3 cells resulted in increases in p202 protein levels. Taken together, these observations suggested that activation of ERα by E2 in WT276 and NIH 3T3 cells up-regulates the expression of the Ifi202 gene.

**Transcriptional activation of Ifi202 by ERα**

The 5′-regulatory region (~800 bp) of Ifi202 gene contains at least two potential ERE half-sites (Fig. 6A). Moreover, one of the potential ERE half-sites is located next to an AP-1 DNA-binding site (AP-1CS1) that can bind to c-Jun/AP-1 in gel-mobility shift assays (40). Molecular mechanisms through which the ERα regulates the transcription of its target genes through the half ERE sites are relatively complex (15, 16) and known to involve collaborations with other transcription factors, such as c-Jun/AP-1 (15, 16). Therefore, to investigate the role of ERα in the regulation of Ifi202 expression, we compared the activity of the wild-type (202-luc) and the mutant (202-AP-1CS1-luc, in which the AP-1CS1 site is mutated) reporters without or after E2 treatment of WT276 cells. As shown in Fig. 6B, the activity of the wild-type reporter was stimulated ~2.5-fold by the treatment of cells with E2. However, the mutation in the AP-1CS1 site in the 5′ regulatory region of the Ifi202 gene abrogated the stimulation of the activity of the reporter after E2 treatment. This suggested that E2 treatment of WT276 cells stimulates transcription of the Ifi202 gene through the AP-1CS1 site. To further examine the role of ERα in the transcriptional activation of the Ifi202 gene by E2, we also compared in vivo association of ERα with the 5′ regulatory region of the Ifi202 gene in splenic B6.Nba2 B cells between female and age-matched males by ChIP assays. As shown in Fig. 6C, some binding of ERα to Ifi202 regulatory region was detected in male B cells (lane 5). Interestingly, relatively more ERα bound to the Ifi202 regulatory region in female B cells (compare lane 6 with lane 5). Moreover, a quantitative real-time pPCR revealed (Fig. 6D) that there was ~4-fold more binding of ERα to the Ifi202 regulatory region in the female B cells than in males. These observations indicated that relatively higher levels of ERα associated with the regulatory region of the Ifi202 gene in female B6.Nba2 B cells than in males.
Female mice were next to an AP-1 DNA-binding site. This allowed us to visualize the potential DNA-binding site in the 5′ regulatory region of the Ifi202 gene. Because our above observations demonstrated that activation of ERα by E2 up-regulates the expression of the Ifi202 gene in WT276 cells, we compared steady-state levels of Ifi202 mRNA in (NZB × NZW)F1 lupus-prone mice (35). The study also identified the Trp53 gene (encoding the p53 protein), whose expression is up-regulated in male mice (as compared with female mice). p53 represses transcription of the Ifi202 gene (46). Therefore, our observation that expression of the Ifi202 gene is down-regulated by male sex hormone (DHT) in orchietomized (NZB × NZW)F1 male mice (Fig. 1) makes it likely that increased levels of male sex hormone negate the regulation of Ifi202 expression, in part by up-regulating the p53 expression. Moreover, the study by Xin et al. (35) also identified other genes that are known to encode proteins with immunomodulatory functions. However, none of the identified estrogen-responsive genes mapped within the NZB-derived Nba2 lupus susceptibility interval, which is syntenic to the human lupus susceptibility locus (26, 32).

Promoter polymorphisms-dependent increased expression of Ifi202 gene in female mice of certain strains is associated with defects in apoptosis of B cells and the development of lupus-like disease (26, 32, 33). Because the development of lupus-like disease in B6.Nba2 mice has sex bias (34), we investigated whether sex hormones could regulate the expression of the Ifi202 gene. Here, we demonstrate that sex hormones regulated the expression of the Ifi202 gene in female mice of certain strains is associated with defects in apoptosis of B cells and the development of lupus-like disease. We also show that increased expression of Ifi202 gene is associated with increased levels of female sex hormone and male sex hormone DHT decreased the mRNA levels (Fig. 1); (2) increased steady-state levels of Ifi202 mRNA and protein in splenic B cells from B6.Nba2 male and female mice were associated with increased levels of ERα and reduced levels of AR (Fig. 2); (3) steady-state levels of Ifi202 mRNA were relatively higher in C57BL/6, B6.Nba2, NZB, and (NZB × NZW)F1 female mice than in age-matched male mice (Fig. 3); (4) treatment of E2-responsive WT276 cells with increasing concentrations of E2 increased the steady-state levels of Ifi202 mRNA and protein, whereas treatment of cells with DHT decreased the p202 protein levels (Fig. 4); (5) treatment of WT276 cells with E2 or overexpression of ERα stimulated the activity of 202-luc-reporter plasmid (Fig. 5, A and B), and overexpression of ERα in NIH 3T3 cells up-regulated the p202 protein levels (Fig. 5C); (6) E2 treatment of WT276 cells activated transcription from 202-luc-reporter gene. Therefore, inactivation of the 5′ regulatory region of the Ifi202 gene by E2 or overexpression of ERα stimulated the activity of 202-luc-reporter plasmid (Fig. 5, A and B), and overexpression of E2 or overexpression of ERα up-regulated the p202 protein levels (Fig. 5C); (6) E2 treatment of WT276 cells activated transcription from 202-luc-reporter gene. Therefore, inactivation of the 5′ regulatory region of the Ifi202 gene by E2 or overexpression of ERα stimulated the activity of 202-luc-reporter plasmid (Fig. 5, A and B), and overexpression of ERα up-regulated the p202 protein levels (Fig. 5C); (6) E2 treatment of WT276 cells activated transcription from 202-luc-reporter gene.
hormone differentially regulate the expression of the Ifi202 gene in immune cells.

Of note, our observations revealed that basal and IFN-induced steady-state levels of Ifi202 mRNA were significantly higher in nonautoimmune C57BL/6 females than in age-matched males (Fig. 3A). Moreover, steady-state levels of Ifi202 mRNA were also significantly higher in preautoimmune B6.Nba2, NZB, and (NZB × NZW)F1 female splenic cells than in age-matched males (Fig. 3B). Similarly, steady-state levels of Ifi202 mRNA and protein were significantly higher in preautoimmune B6.Nba2 female splenic B or T cells than age-matched males (Fig. 2). Taken together, these observations are consistent with sex hormone-dependent (and disease-independent) regulation of Ifi202 expression.

ERα-deficient mice are reported to have elevated levels of estrogen and testosterone (47). Therefore, our observations that in vivo treatment of orchietomized (NZB × NZW)F1 male mice with DHT reduced the mRNA levels (Fig. 1) and the lack of detection of Ifi202 mRNA in Egr1-null mice (Fig. 7) make it likely that increased levels of testosterone in ERα-deficient female mice through AR down-regulate the expression of Ifi202 gene. Further work will be needed to test this possibility.

A study (48) has revealed that treatment of BALB/c mice with ER subtype-selective agonists that results in activation of ERα, but not ERβ, plays a major role in estrogen-induced thymic atrophy and thymic T cell and splenic B cell phenotype alterations. Moreover, the study also revealed that ERα, but not ERβ, mediates the estrogen-induced up-regulation of IFN-γ. Consistent with a role for ERα in the IFN-γ production, our study (20) involving generation of ERα knockout (NZB × NZW)F1 mice and their characterization revealed that estrogen through ERα promotes lupus disease, at least in part, by inducing IFN-γ production. Moreover, estrogen is known to enhance IFN-γ production by CD11c+ cells (49). Taken together, these observations raise the possibility that activation of ERα by E2 in immune cells of certain strains of female mice up-regulates Ifi202 expression in part by increasing IFN-γ production. Therefore, further work will be needed to test this hypothesis.

Previous studies (43, 44, 50) have suggested that estrogen treatment of R4A-γ2b BALB/c mice (transgenic for the H chain of an anti-DNA Ab) with E2 leads to the survival and activation of autoreactive cells in a naïve repertoire. Moreover, studies (43) also revealed that estrogen treatment of B cells also up-regulates the expression of several genes, such as cd22, shp-1, and bcl-2, which are involved in B cell activation and survival. Interestingly, treatment of mice with tamoxifen, a selective ER modulator (42), blocked estrogen-induced B cell maturation but not survival (50). Because increased expression of p202 protein in splenic B cells from B6.Nba2 congenic mice is associated with defects of apoptosis of cells (26, 32, 33) and down-regulation of expression of p53 and E2F-responsive proapoptotic genes (36, 41), we compared basal transcriptional activity of NF-κB between B6.Nba2 female B cells and age-matched males. These experiments indicated that increased levels of p202 in female B cells (as compared with males) are associated with increased transcriptional activity of NF-κB (data not shown). Collectively, our observations support the idea that female sex hormone-dependent increased levels of p202 protein in B cells by increasing cell survival contribute to sex bias in lupus susceptibility (Fig. 8).

The 5′ regulatory region (~800 bp) of Ifi202 gene contains at least two ERE half-sites (Fig. 6A). Interestingly, one of the ERE half-sites is located next to an AP-1 consensus DNA-binding site (the AP-1CS1), which can bind to AP-1 and can stimulate the transcription of the Ifi202 gene (40). Because the ERα can regulate transcription of its target genes through the c-Jun/AP-1 DNA-binding sites (15, 16), we tested whether ERα regulates transcription of the Ifi202 gene through the AP-1CS1 site-dependent manner. Our experiments demonstrated that E2-mediated stimulation of the activity of 202-luc-reporter was abrogated due to a mutation in the AP-1CS1 site. Further work will be needed to investigate how ERα and c-Jun/AP-1 collaborate with each other to up-regulate the expression of the Ifi202 gene in B cells.

A search for an AR-responsive element (ARE) in the 5′-regulatory region of the Ifi202 gene did not result in identification of an ARE. Therefore, further work will be needed to determine whether 5′ regulatory region of the Ifi202 gene, which is upstream to an ~800-bp region, contains an ARE. In this regard, note that bone marrow stromal cells mediate androgenic suppression of B lymphocytes development through up-regulation of TGF-β expression in response to DHT treatment (51). Consistent with these observations, we have noted that TGF-β treatment of splenic cells reduced steady-state levels of Ifi202 mRNA (data not shown). Therefore, further work will be needed to determine how androgens negatively regulate the expression of Ifi202 in T cells.

Interestingly, a recent study (29) has provided evidence that the p202 protein can recognize double-stranded DNA in cytoplasm (29). Moreover, the study proposed that increased levels of p202 protein in immune cells could inhibit the ability of the AIM2 protein, a pyrin domain containing member of the p200-family (28, 30), which can also sense DNA in cytoplasm and can form a caspase-1-activating inflammasome (30). In light of these observations it will be important to investigate whether the expression of the murine AimII is also differentially regulated by the female and male sex hormones in immune cells.

In summary, our observations provide support for our model (Fig. 8). The model predicts that increased levels of male hormone androgen through activation of AR down-regulate the expression of the Ifi202 gene. In contrast, increased levels of female hormone estrogen through activation of ERα up-regulate the expression of Ifi202. Consequently, increased levels of p202 protein in immune cells of certain strains of female mice contribute to increased survival of autoreactive cells, resulting in increased susceptibility to lupus disease. Our observations will serve as a molecular basis to identify signaling pathways and molecules that contribute to sex bias in the development of SLE in human patients.
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References