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Externally Triggered Egress Is the Major Fate of Toxoplasma gondii during Acute Infection

Tadakimi Tomita,* Tatsuya Yamada,* Louis M. Weiss,*† and Amos Orlofsky2*

The apicomplexan parasite Toxoplasma gondii expands during acute infection via a cycle of invasion, intracellular replication, and lytic egress. Physiological regulation has not yet been demonstrated for either invasion or egress. We now report that, in contrast to cell culture systems, in which egress occurs only after five or more parasite divisions (2–3 days), intracellular residence is strikingly abbreviated in inflammatory cells in vivo, and early egress (after zero to two divisions) is the dominant parasite fate in acutely infected mice. Adoptive transfer experiments demonstrate rapid, reciprocal, kinetically uniform parasite transfer between donor and recipient compartments, with a t1/2 of ~3 h. Inflammatory macrophages are major participants in this cycle of lytic egress and reinfection, which drives rapid macrophage turnover. Inflammatory triggering cells, principally macrophages, elicit egress in infected target macrophages, a process we term externally triggered egress (ETE). The mechanism of ETE does not require reactive oxygen or nitrogen species, the mitochondrial permeability transition pore, or a variety of signal transduction mediators, but is dependent on intracellular calcium and is highly sensitive to SB203580, an inhibitor of p38 MAPK as well as a related parasite-encoded kinase. SB203580 both inhibited the initiation of ETE and altered the progression of egress. Parasites recently completing a cycle of egress and reinfection were preferentially restricted in vivo, supporting a model in which ETE may favor host defense by a process of haven disruption. ETE represents a novel example of interaction between a parasite infectious cycle and host microenvironment. The Journal of Immunology, 2009, 183: 6667–6680.

Materials and Methods

Materials

The Abs were used anti-F4/80/647 (Serotec), anti-CD11b/647, anti-B220/647, anti-Thyl.2 allophycocyanin, and anti-I-A*PE (BD Biosciences). SB203580, U0126, Jnk inhibitor II, rotterlin, Go 6976, and BAPTA-AM were obtained from EMD Chemicals. N-iminoethyly-1-lysine (L-NIL), N-nitro-t-arginine methyl ester, N-acetyllysine, pyridoxal-phosphate-6-azophenyl-2′,4′-disulfonate (PPADS), cyclosporine, flavidone, and Ac-cytase were from Sigma-Aldrich. Murine IFN-γ was from Chemicon International. Hydroxydichlorodimethylacridinone succinimimidyl ester (DDAO-SE) (Cell Trace Far Red) was from Invitrogen.

Parasites and mice

Parasites were maintained in human foreskin fibroblasts, as described (11). The transgenic strain expressing GFP has been described (12). The yellow fluorescent protein (YFP)-expressing strain (13) was a gift of B. Striepen (University of Georgia, Athens, GA). The in vitro growth characteristics of the fluorescent strains were similar to wild type. Mice (C57BL/6, 6–8 wk old) were inoculated i.p. with 0.2 ml of PBS containing 2000 tachyzoites harvested from lysed cultures. Some experiments used mice expressing enhanced cyan fluorescent protein (ECFP) behind an actin promoter (stock number 4218, The Jackson Laboratory). Some samples for cytology were obtained from wild type mice on a mixed C57BL/6 - 129/Sv background, as previously described (14). All mice were maintained in a specific pathogen-free facility. All mouse studies were reviewed and approved by the Animal Committee at the Albert Einstein College of Medicine.

3 Abbreviations used in this paper: ETE, externally triggered egress; DAP, donor cell-associated parasite; ECFP, enhanced cyan fluorescent protein; L-NIL, N-iminoethyl-1-lysine; moi, multiplicity of infection; NDAP, non-donor cell-associated parasite; PEM, peritoneal exudate macrophage; PPADS, pyridoxal-phosphate-6-azophenyl-2′,4′-disulfonate; WT-RH, wild-type RH strain; YFP, yellow fluorescent protein.

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The Journal of Immunology
Peritoneal exudate macrophages (PEM) were prepared by lavage of mice injected 4 days previously with 1 ml of 3% thyogloccytol broch (Difco). Stocks of frozen aliquots of PEM were generated from pooled lavage of at least three mice. PEM were also prepared from mice deficient in IFN-γ receptor-1 (stock number 3288). Before infection, thawed PEM were cultured for 1 day in DMEM with 10% FBS.

Cytology
At various times postinfection, mice were sacrificed and the peritoneal cavity was washed with ice-cold PBS containing 1% BSA. Cytospin preparations were then prepared and fixed in methanol, dried, stained with a modified Wright’s stain (Leuko-Stat, Fisher Scientific), and examined at ×100 on a Zeiss Axioskop II. Microscope fields were chosen before observation, and all infected nucleocen cells in the field were scored, except that vacuoles containing debris or degraded parasites (<10% of total vacuoles) were excluded.

Adaptive exudate transfer
On day 5 postinfection, when ascites volume is ~1 ml, 0.1 ml of exudate was collected from all mice by paracentesis and immediately diluted with 4 ml of chilled PBS containing 0.1% BSA, 1 mM EDTA, and 10 U/ml heparin (buffer A); centrifuged at 150 × g for 10 min; and suspended in 100 µl of PBS with 0.1% BSA. The samples were immediately analyzed by flow cytometry to determine percentage of cells infected, parasites/infected cell, and free parasites relative to total cells. Mice showing similar progression of toxoplasmosis by these criteria were selected as donor-host combinations. Donor mice were injected i.p. with 0.1 ml of Hoechst 33342 (200 µg/ml, Molecular Probes), and the abdomen was briefly massaged. In comparison with the dye, genetic marking using CD45 or H2 alleles was less compatible with the use of fluorescent parasites and also provided inferior donor-host discrimination in a complex cell population (data not shown). Transgenic ECFP was used as a marker in some experiments, although it was not uniformly expressed in exudate cells (data not shown).

Depletion of exudate cells
Exudate from WT-RH-infected mice was diluted with Hank’s saline containing 0.1% BSA and 1 mM EDTA. Exudate cells were washed and incubated for 30 min on ice with either anti-I-A8-PE, anti-F4/80-647, or anti-CD11b-PE, or the combination of anti-I-A8-PE, anti-Thy-1.2 allophycocyanin, and anti-B220-647. The cells were washed and then incubated with rat-igG Dynabeads (Dynal Biotech) for 30 min. The cells were either exposed or not (mock depleted) to a Dynal MPCS-S magnet for 10 min, spun, and resuspended in assay buffer at 1 × 107/ml.

To deplete total host cells and enrich extracellular parasites, exudates from YFP-RH-infected mice were washed twice in assay buffer and pooled, and 0.8-ml aliquots (3 × 106 cells/ml) were placed in microcentrifuge tubes on ice. Some samples were then spun for 2 min in a Fisher 59A swing-out microcentrifuge in a cold room at a setting of 1.2 (~100 g). Supernatant was collected from spun samples. Aliquots of the samples were fixed for flow cytometry and the remainder briefly warmed before transfer (0.1 ml) to host mice.

Results

Intravascular parasitic content in acute toxoplasmosis is consistent with early egress
Peritoneal inoculation of mice with 107–108 RH tachyzoites generates a progressive infection with a steadily increasing parasite burden that is predominantly localized to the peritoneal cavity, with involvement of adjacent sites (15–19). The inflammatory response in the peritoneum is characterized by the formation of ascites with a massive influx of mononucleocytes and neutrophils ~3–4 days postinfection and is maintained for 3–4 days until the death of the host (17, 20). In the course of studies using this model, we noticed that peritoneal macrophages collected from mice during the peak period of inflammation rarely had parasitophorous vacuoles containing more than eight parasites, and we undertook to investigate whether early egress might account for this observation.

As a first step, we examined the extent of intravascular parasite proliferation in macrophages from mice acutely infected with T. gondii RH. Counts were performed by cytological examination of lavage samples (Fig. 1A) collected from mice that had entered peak inflammation (~4–5 days postinfection), but did not yet exhibit signs of morbidity. Each intracellular parasite was assigned a proliferation value P equal to the number of divisions completed by that parasite in that cell (for example, in a vacuole with three parasites, two are assigned P = 2 and one is assigned P = 1). The
observed distribution of P was compared with two theoretical distributions representing alternative models of egress regulation (Fig. 1B). Both models assume that the distribution is asynchronous (time invariant), that replication cycles are of constant length, that egress is unbiased with respect to parasite cell cycle stage (so that egress occurs, on average, at the midpoint of a cycle), and that the interval between egress and reinvasion is negligible (confirmed below).

Under the first model (conventional or proliferation-dependent egress), egress occurs at a fixed value of P; that is, egress requires a certain number of parasite divisions. In this case, the time-invariant distribution is that in which the fraction of parasites is constant over P, except reduced by half for P = 0 and P = c, because egress is at mid-cycle (dashed line in Fig. 1B, illustrated for egress at P = 5). In the second model (probabilistic egress), egress occurs with equal probability at all P, consistent with an egress signal that is independent of parasite proliferation. Under this model, the distribution can be derived by iteration (beginning with a synchronous infection), and is given by $f_n = 0.5q$ and $f_n = 0.5(q(1 - q)^n + q(1 - q)^{-1})$ for $n > 0$, where $f_n$ is the fraction of parasites for which $P = n$, and q is the probability of egress during one round of parasite replication (see Appendix). This model assumes the absence of proliferation-dependent egress, which is reasonable for sufficiently high values of q. The gray bars in Fig. 1B illustrate the model for values of q between 0.4 and 0.7; for these values, few parasites progress beyond five divisions.

We found that $f_n$ declined rapidly for $n > 1$ (solid colored lines in Fig. 1B), in accordance with the probabilistic model and in contrast to the even distribution expected for conventional proliferation-dependent egress. When vacuoles with greater than eight parasites were observed, they were almost always found in a cell population displaying the extensive basophilic cytoplasm characteristic of mesothelial cells (Fig. 1A, b and c). These presumptive mesothelial cells constitute ~10% of the recovered infected mononuclear cells, the remainder of which have a macrophage-like morphology (Fig. 1A, a and b). In contrast to the distribution of P observed in total mononuclear cells, the distribution within the mesothelial component (blue dotted line in Fig. 1B) does not fit the probabilistic model, but instead is consistent with conventional egress.

Because many of the infected macrophages (~50–60%) bear multiple vacuoles, we also considered the possibility that egress occurs upon accumulation of a certain number of total parasites per cell, rather than a fixed number of parasite divisions. In this case, cells that bear more vacuoles should contain, on average, fewer parasites per vacuole. This, however, is not the case: vacuole progression was observed to be independent of cell vacuole number (supplemental Fig. S1).4

Parasite transfer in vivo indicates the prevalence of early egress

The distribution data are suggestive of a distinct mode of egress in infected inflammatory macrophages. Alternatively, the findings might be explained by the progressive activation of antiparasitic function in infected macrophages. To directly assess the occurrence of early egress in the inflamed peritoneal cavity, we conducted adoptive transfer experiments to determine the rate of parasite egress from exudate cells in situ. Mice were infected i.p. with transgenic parasites expressing either GFP or YFP (GFP-RH or YFP-RH). At day 5 postinfection, when ascites volume is exponentially, implying a first-order decay process with a $t_{1/2}$ of

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**FIGURE 1.** Distribution of intravacuolar parasite content in acute toxoplasmosis. Mice were infected i.p. with *T. gondii* RH strain. *A*. After 4 or 5 days, cytospin preparations of peritoneal exudate were Wright stained, and parasitophorous vacuoles (arrowheads in *a* and *b*) were evaluated for parasite content. *a*, Representative macrophages containing vacuoles in which one to two divisions have taken place (the number of parasites in each vacuole is indicated). *b*, An infected macrophage adjacent to an infected presumptive mesothelial cell (M). *c*, A mesothelial cell (M) with a vacuole bearing 32 parasites. Scale bar: 5 μm. *B*, Parasite enumeration data were used to calculate the distribution of parasite proliferation value (the number of divisions within the vacuole for each parasite) in infected mononuclear cells. Each of the solid colored lines represents the distribution in total mononuclear cells from one mouse. For each sample, 300–400 vacuoles (~600–800 total parasites) were scored. One sample (green line) is from day 5 postinfection; the others are from day 4. The data are representative of four total mice obtained from three independent experiments. The blue dotted line displays the distribution in mesothelial cells (pool of two mice from independent experiments; 87 total vacuoles, 376 total parasites assessed). Indicate the theoretical distribution under probabilistic egress, with each shade representing a different probability, as indicated by the numbers on the left (see text). The dashed line indicates the theoretical distribution for proliferation-dependent egress (see text).
To confirm that this assay measures parasite transfer, rather than donor cells misidentified as host cells, we examined the intracellular parasite burdens of donor and host cells. At early time points, genuine egress of GFP-RH from donor cells, with subsequent reinvasion of host cells, is expected to result in host cells containing only one or two GFP-RH (data not shown). Each line represents sequential sampling of a single animal. The data are representative of five mice. The corresponding recoveries of total donor parasite (intracellular + extracellular, as a fraction of total parasites) are shown as dashed lines (the value at 1 h posttransfer is set to 1). D, GFP-RH histograms of donor (unshaded, solid line) and host cells (shaded) 1 h after GFP→YFP transfer. A histogram of untransferred donor cells is shown for comparison (dashed line).

~2 h. A composite regression performed on data from six experiments (using a total of 13 donor and 38 host mice) yielded a $t_{1/2}$ of $3.1 \pm 0.3$ h ($r^2 = 0.87$). These kinetics suggest a process that is fairly uniform both over time and throughout the population of donor parasite-bearing donor cells. The total amount of donor parasite residing in the host declined only slightly over time (dashed lines in Fig. 2C), indicating that the decline in the fraction of donor cell-associated donor parasite was the consequence of rapid parasite transfer.

To confirm that this assay measures parasite transfer, rather than donor cells misidentified as host cells, we examined the intracellular parasite burdens of donor and host cells. At early time points, genuine egress of GFP-RH from donor cells, with subsequent reinvasion of host cells, is expected to result in host cells containing only one or two GFP-RH, whereas donor cells will have a greater GFP-RH burden, resembling that of parental donor cells. This prediction was verified (Fig. 2D). We similarly verified the converse prediction that the YFP-RH profile of the newly invaded host cells should be consistent with the YFP-RH profile of host cells rather than that of donor cells, which will not have acquired more than one or two YFP-RH (data not shown). The verification of these predictions establishes that the assay detected actual parasite transfer.

To confirm that the observed egress was a physiological process and not an artifact arising from cell transfer, we re-examined the data to compare the rates of egress in the donor→host and host→donor directions. To accomplish this, we derived an estimate of the egress rate as a linear approximation over a brief interval (between 1 and 3–4 h posttransfer), by determining the amount of non-donor cell-associated parasite (NDAP) at these two time points. The NDAP value is the sum of the host cell-associated donor parasite burden and extracellular donor parasites, and is calculated by assuming that total peritoneal cellularity is unchanged over the brief interval. The gain in NDAP over the interval is divided by the donor cell-associated parasite (DAP) burden (mean of 1- and 3-h values) to generate an egress rate value (donor→host) expressed as the percentage of DAP egressing per hour. To estimate host→donor egress, the converse calculation is performed (i.e., the gain in nonhost cell-associated host parasite as a percentage of the total amount of host cell-associated host parasite). However, in this case, to correct for the fact that most reinvasion events occur in host cells and are not scored, the value for the donor cell-associated host parasite burden is multiplied by the host cell/donor cell ratio (mean of 1- and 3-h values). As shown in Table I, good agreement was obtained for egress in the two directions, confirming the validity of the assay. As an additional control, we have obtained similar egress values in experiments in which the use of vital dye was avoided by using host mice expressing transgenic ECFP from an actin promoter (data not shown). Furthermore, these egress rates are consistent with the modeling of egress based on vacuolar parasite content (Fig. 1B). For example, if the length of a parasite replication cycle is 6 h, then probabilistic egress, with probability per cycle set at 0.7 or 0.6, would result in a parasite transfer $t_{1/2}$ of 3.45 or 4.5 h, respectively. In comparison, under the conventional model of proliferation-dependent egress, with $c = 6$ cycles, the $t_{1/2}$ would be 18 h.
Role of exudate macrophages in early egress

We next sought to determine the exudate cell types in which cycles of early egress and reinvasion were taking place. As expected, the cell type with the greatest parasite burden was the F4/80⁺ macrophage, carrying approximately half the total burden in both donor and host cells (Fig. 3). In comparison, B and T lymphocyte populations each contributed about 5% of the burden. The remaining burden was approximately evenly divided between neutrophils and host cells (Fig. 3). This distribution was confirmed by the host cells/donor cells ratio to correct for undetected reinvasion events occurring in host cells.

The demonstration of early egress from inflammatory macrophages suggested that early egress may also be prominent in this cell type. To test this idea, we followed the kinetics of donor cell disappearance after transfer. Consistent with the previous kinetic analysis of donor parasite transfer, donor cells in the host animal were lost with zero-order kinetics over the first 8 h posttransfer, with a $t_{1/2}$ of 3.6 h (Fig. 4A). Remaining cells then decayed at a slightly slower rate. Donor macrophages were lost at a faster rate than total donor cells ($t_{1/2}$ of 2.0 h), and this corresponded to a higher frequency of initial infection in this cell type: 73% of input donor macrophages were infected, compared with 41% for total donor cells. In contrast, when uninfected thioglycollate-elicited peritoneal macrophages (PEM) were labeled and transferred to hosts undergoing thioglycollate peritonitis, little donor cell loss was observed (Fig. 4B), as reported previously (22). Consistent with these findings, we observed that peritoneal recruitment of circulating myeloid leukocytes was greatly elevated in T. gondii-infected mice compared with mice with thioglycollate peritonitis (Fig. S3). When $^{51}$Cr-labeled PEM were injected i.p. in mice undergoing either thioglycollate- or T. gondii-induced peritonitis, no significant difference was observed with respect to appearance of label in the draining thoracic lymph nodes (<1% of input label; data not shown), suggesting that rapid macrophage clearance in infected mice was not due to enhanced drainage.

The correspondence between the high frequency of macrophage infection and rapid macrophage turnover suggests that this turnover may be driven by parasite egress. If this is the case, it would be expected that donor macrophages already infected at the time of transfer will be lost more rapidly than their uninfected counterparts. This prediction was tested by assuming that donor cells bearing donor parasite had acquired the parasite pretransfer, because most posttransfer infection is with host parasite. As shown in Fig. 4C, donor parasite-bearing donor macrophages indeed decayed faster than donor parasite-negative cells ($t_{1/2}$ of 1.8 and 2.4 h, respectively, over the first 8 h of transfer). We next asked whether the decay of the donor parasite-negative macrophages could potentially be accounted for by egress subsequent to invasion by host parasite after transfer. As shown in Fig. 4D, by 3 h posttransfer, the frequency of donor macrophages bearing host parasite was nearly equal to the steady-state frequency of infection observed in host macrophages (dashed line). These data are consistent with an egress-dependent mechanism of cell loss for the preponderance of donor macrophages.

An in vitro model of externally triggered egress

The demonstration of early egress from inflammatory macrophages in vivo suggested that egress might be triggered by components of the inflammatory environment. To address this issue, we sought to develop an in vitro assay in which regulation of the process could be studied. As a first step, we prepared donor and host mice as for the adoptive transfer assay, and then, instead of performing exudate transfer, assessed parasite transfer in ex vivo cultures in which these exudates had been combined. One hour of coculture of a 1:1 mixture of washed exudate cells resulted in a substantial increase in NDAP (Fig. 5A). The gain in NDAP (that is, the combined gain in extracellular plus host cell-associated donor parasites) was equal to 20 ± 1% of initial DAP, a value that becomes 42 ± 1% when corrected for presumed unscored reinvasion events occurring in donor cells. This result suggested that cellular elements of the exudate might be sufficient to generate early egress. To confirm that soluble factors in the exudate were not required for the ex vivo assay, we compared egress in coculture of washed cells with mixtures of whole unseparated exudates (Fig. 5B). In this experiment, mixtures are 1:9 (donor:host). The lower egress values for whole exudates may be related to partial coagulation that occurs in unseparated samples, despite the use of anticoagulants (data not shown).
To ask whether early egress was triggered by exudate cells, we added these cells, prepared from mice infected with nonfluorescent *T. gondii* (WT-RH), to monolayers of YFP-RH-infected PEM. Time-lapse imaging, performed both before and after addition of exudate cells, revealed that the addition of these cells induced egress in 40% of infected PEM within 120 min (Fig. 6, A–C, and supplemental Videos 1 and 2). This egress rate (20%/h) is consistent with the rates observed in vivo (Table I). Few egress events occurred before 20–30 min after addition of exudate cells (Fig. 6D), coinciding with the interval required for complete settling of these cells onto the monolayer (data not shown). Subsequently, the egress rate was relatively uniform over time, suggesting that the egress was not a transient response to the addition of exudate cells. Flow cytometric analysis indicated that the majority of the egressed parasites were associated with exudate cells, which were present in excess over PEM (data not shown). These data indicate that early egress can arise as an acute response to a microenvironmental stimulus. We refer to this phenomenon as externally triggered egress (ETE).

We adopted the PEM-exudate cell coculture system for further studies of ETE, with the modification that, to ensure quantitative cell recovery for flow cytometry, nonadherent conditions were used, although this entailed a reduction in egress rate (data not shown). To minimize loss of cells and parasites to aggregation in the nonadherent system, cocultures were routinely performed in the presence of EDTA, which had a minimal effect on egress (data not shown). Even with this precaution, parasite loss of up to 20% was sometimes observed (as in Fig. 5A), limiting sensitivity for the detection of NDAP gain. We therefore chose to quantify egress in the PEM-exudate coculture experiments by determining the NDAP/total parasite fraction, which proved to be a more robust and sensitive indicator than the measurement of absolute NDAP gain. This NDAP fraction displayed a time-dependent increase during 60 min of PEM-exudate coculture (Fig. S4).

A control experiment was performed to confirm that, in this nonadherent system, egress measured in this way reflected an exudate cell-dependent event, rather than simply an increased ability

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**FIGURE 4.** Kinetics of inflammatory cells in acute toxoplasmosis. A, C, and D, Samples from the adoptive transfer described in Fig. 3 were stained with F4/80 to identify macrophages. Data are representative of three to four experiments. A, The recovery of donor macrophages and total donor cells (in each case as a ratio to total cells, relative to 1 h posttransfer) was determined by flow cytometry. Linear regressions (dashed lines) were calculated based on 1- to 8-h data. B, Donor and host mice were each injected i.p. with thioglycolate. After 5 days, donor exudate cells were harvested, labeled with CellTracker Orange, and injected into host mice. Groups of three mice were sacrificed at the indicated times to determine donor cell recovery. C, Comparison of the recovery of donor macrophages infected or uninfected with donor parasites (reflecting infection status at the time of transfer). D, The percentage of donor macrophages infected with host parasite was monitored at the indicated times and compared with the frequency of host parasite infection of host macrophages (dashed line).

**FIGURE 5.** Ex vivo egress in toxoplasmic exudates. A, Washed exudate cells (host) prepared from mice 5 days postinfection with GFP-RH were labeled in situ with Hoechst 33342 and mixed 1:1 with donor exudate cells prepared from mice infected with YFP-RH. Each preparation represents a pool of two to three mice. Cocultures were conducted for 1 h in nonadherent dishes in the presence of 1 mM EDTA plus 10 U/ml heparin to reduce aggregation. The fate of donor YFP-RH was assessed by flow cytometry. The identity of donor and host cells bearing YFP-RH was verified by monitoring the intensity of GFP and YFP signals, as discussed for Fig. 2 (data not shown). The value for donor cell-associated parasites at 0 h was set to 100. B, An experiment similar to A was conducted, except that cocultures (1.5 h) of washed exudate cells were compared with similar cocultures of whole exudate (fluid + cells). Instead of Hoechst dye, one exudate was labeled with DDAO-SE. The donor:host ratio was 1:9. Each of the three mixes represents a separate pair of donor and host mice. Whole exudate samples were filtered before analysis to remove any coagulated material. The data in the figure display mean ± SE of three replicates and are representative of either seven (A) or two (B) similar experiments.

To ask whether early egress was triggered by exudate cells, we added these cells, prepared from mice infected with nonfluorescent *T. gondii* (WT-RH), to monolayers of YFP-RH-infected PEM. Time-lapse imaging, performed both before and after addition of exudate cells, revealed that the addition of these cells induced egress in ~40% of infected PEM within 120 min (Fig. 6, A–C, and supplemental Videos 1 and 2). This egress rate (~20%/h) is consistent with the rates observed in vivo (Table I). Few egress events occurred before 20–30 min after addition of exudate cells (Fig. 6D), coinciding with the interval required for complete settling of these cells onto the monolayer (data not shown). Subsequently, the egress rate was relatively uniform over time, suggesting that the egress was not a transient response to the addition of exudate cells. Flow cytometric analysis indicated that the majority of the egressed parasites were associated with exudate cells, which were present in excess over PEM (data not shown). These data indicate that early egress can arise as an acute response to a microenvironmental stimulus. We refer to this phenomenon as externally triggered egress (ETE).

We adopted the PEM-exudate cell coculture system for further studies of ETE, with the modification that, to ensure quantitative cell recovery for flow cytometry, nonadherent conditions were used, although this entailed a reduction in egress rate (data not shown). To minimize loss of cells and parasites to aggregation in the nonadherent system, cocultures were routinely performed in the presence of EDTA, which had a minimal effect on egress (data not shown). Even with this precaution, parasite loss of up to 20% was sometimes observed (as in Fig. 5A), limiting sensitivity for the detection of NDAP gain. We therefore chose to quantify egress in the PEM-exudate coculture experiments by determining the NDAP/total parasite fraction, which proved to be a more robust and sensitive indicator than the measurement of absolute NDAP gain. This NDAP fraction displayed a time-dependent increase during 60 min of PEM-exudate coculture (Fig. S4).

A control experiment was performed to confirm that, in this nonadherent system, egress measured in this way reflected an exudate cell-dependent event, rather than simply an increased ability...
to detect reinvasions that, in the absence of exudate, occurred in PEM and were therefore not scored. To allow the detection of reinvasions in PEM as well as exudate cells, dye-labeled, YFP-RH-infected PEM were mixed with a 9-fold excess of unlabeled PEM infected with WT-RH. This base mixture was then incubated with either blank medium (None) or a 10-fold excess of exudate cells (Exudate). Basal egress was then monitored by incubation of this mixture (None) for 1 h in nonadherent dishes, followed by determination of the percentage of YFP-RH positive donor cell-associated NDAP (NDAP). ETE was measured as the additional NDAP generated by the addition of a 5-fold excess of washed exudate cells (Exudate). Control incubations were performed in the presence of a similar excess of uninfected or WT-RH-infected PEM.

The combined depletion resulted in effective removal of macrophages and neutrophils using a combination of F4/80 and anti-CD11b, which reacts with both cell types. Neutrophil depletion with 1A8 served as a control. The combined depletion resulted in effective removal of macrophages and neutrophils using a combination of F4/80 and anti-CD11b, which reacts with both cell types. Neutrophil depletion with 1A8 served as a control.

**Inhibitor studies of ETE**

We next used a panel of inhibitors to investigate the mechanistic basis of ETE. No consistent effect on ETE was observed using antagonists of reactive oxygen or nitrogen species (N-acetylcysteine or l-NIL, respectively), or with cyclosporine A or PPADS, which can protect macrophages from cell death pathways that act, respectively, via opening of the mitochondrial permeability transition pore or purinergic receptor activation (24–26) (Fig. 8A). Inhibition of either classical or novel protein kinase C signaling (using Go-6976 or rottlerin, respectively) was also without effect, as was inhibition of signaling through either ERK or JNK MAPKs (Fig. 8A). We also observed no effect upon inhibition of either PI3K by wortmannin.

**FIGURE 6.** Triggering of egress in vitro. Egress was assessed in cocultures of YFP-RH-infected thioglycolate-elicited macrophages (PEM) with washed exudate cells obtained from mice infected for 5 days with WT-RH. A and B, YFP fluorescence is displayed from time-lapse images (10-min intervals) of adherent PEM exposed to either blank medium (A) or a 10-fold excess of exudate cells (B). Egress events that had occurred by 2 h are indicated by arrows (lower panels). Scale bar: 20 μm. The quantitation of these egress events (C) represents the mean ± SE from either three (exudate) or eight (medium) fields. The data are representative of two similar experiments. D, The time distribution of exudate cell-induced egress events from the same experiment is displayed as a histogram. The settling of most exudate cells onto the PEM monolayer requires 15–20 min. E, Flow cytometric assay of ETE. Donor PEM infected overnight with YFP-RH (moi = 0.2) were labeled with DDAO-SE and mixed (1:9) with PEM similarly infected with WT-RH. Basal egress was then monitored by incubation of this mixture (None) for 1 h in nonadherent dishes, followed by determination of the percentage of YFP-RH positive donor cell-associated NDAP (NDAP). ETE was measured as the additional NDAP generated by the addition of a 5-fold excess of washed exudate cells (Exudate). Control incubations were performed in the presence of a similar excess of uninfected or WT-RH-infected PEM. ***, p < 0.01 (n = 4). The data are representative of two similar experiments.
or G protein signaling by pertussis toxin (data not shown). In contrast, ETE was reduced by 69% in the presence of SB203580, an inhibitor of stress-sensitive kinases of the MAPK family in both Toxoplasma and mammalian cells (27, 28) (Fig. 8A). Furthermore, chelation of intracellular calcium using BAPTA-AM strongly reduced ETE (Fig. 8B), consistent with previous reports describing the inhibition of inducible T. gondii egress with this compound (3, 5, 6). Therefore both stress-sensitive MAPK and calcium signaling are implicated in the generation of ETE. A recent report indicates that calcium-dependent T. gondii egress can be blocked by fluridine, an inhibitor of the synthesis of abscisic acid, a substance that controls intracellular calcium release (29). However, we observed

FIGURE 7. Role of exudate cell types in ETE. Exudate cells were prepared from mice infected for 5 days with WT-RH and depleted of specific cell types. In experiment 1 (A–E), exudates were depleted of either 1A8+ cells (neutrophils), the combination of 1A8+ cells, B220− cells, and Thy1+ cells (neutrophil/B cell/T cell triple depletion), or F4/80− cells (macrophages), or else mock depleted. A and B, Dot plots of cells triply stained with 1A8-PE/B220-allophycocyanin/Thy1-allophycocyanin Abs, before (A) or after (B) depletion. Histograms of cells stained with F4/80 before (C) or after (D) depletion illustrate the partial depletion of macrophages. In experiment 2 (F and G), exudates were depleted of either 1A8+ cells or the combination of CD11b+ cells and F4/80− cells (neutrophil/macrophage double depletion). F, Histogram of exudate cells stained with F4/80 before (gray) or after (black) double depletion of neutrophils and macrophages. E and G, Donor PEM infected overnight with YFP-RH (moi = 1) were labeled with DDAO-SE and cultured for 1 h in nonadherent dishes with either medium (None) or a 5-fold excess of exudate cells depleted as indicated. NDAP (as percentage of total donor parasites) was determined before (□) and after (■) coculture. Bars represent mean ± SE (n = 3). *, p < 0.05. **, p < 0.01 (relative to mock depletion).

FIGURE 8. Effect of signaling inhibitors on ETE. ETE assays were performed as described for Fig. 7E. Bars represent mean ± SE (n = 4). A and B, PEM were cultured with exudate in the absence (Control) or presence of the indicated inhibitors. The inhibitors used were N-acetylcysteine (NAC, 10 mM), t-NIL (1 mM), cyclosporine A (CsA, 5 µM), PPADS (30 µM), Go 6976 (Go, 1 µM), rollelin (5 µM), U0126 (10 µM), JNK inhibitor II (Jnk-II, 10 µM), SB203580 (10 µM), and BAPTA-AM (BAPTA, 10 µM). A and B, Represent separate experiments. *, p < 0.05. C, The ETE assay was performed on donor PEM that were first cultured overnight in either control medium (Control) or 50 µM fluridine, and then incubated in the absence (□) or presence (■) of exudate. The data in the figure are representative of either eight experiments (SB203580) or two to three experiments (other inhibitors).
FIGURE 9. Inhibition of egress by SB203580. A and B, YFP fluorescence in time-lapse images (10-min intervals) of adherent PEM exposed at 0 h to egress in the presence of either DMSO vehicle (A) or 10 μM SB203580 (B). Egress events that had occurred by 2 h are indicated by arrows (lower panels). Scale bar: 20 μm. C, The proportion of YFP-RH⁴ cells egressing over 2 h is displayed as the mean ± SE from either three (DMSO) or eight (SB203580) fields. The data are representative of two similar experiments. D, Inhibition of egress in ex vivo mixed exudate. Washed donor exudate cells from mice infected for 5 days with YFP-RH were labeled in situ with DDAO-SE, mixed (1:9) with unlabelled exudate from WT-RH-infected mice, and incubated for 1 h in the presence of either vehicle or 10 μM SB203580. Each of the three mixes represents a separate pair of donor and host mice. Bars = mean ± SE (n = 3). The data are representative of three similar experiments.

little effect of fluridone on ETE (Fig. 8C), suggesting that alternative mediators of calcium release may govern this form of Toxoplasma egress.

To obtain more definitive evidence for the effect of SB203580 on ETE, time-lapse experiments were performed to directly compare the rates of exudate-dependent egress in the presence or absence of the inhibitor. The results showed that the inhibitor profoundly blocked ETE, reducing the frequency of egress in infected PEM by 83% (Fig. 9, A–C, and supplemental Videos 2 and 3). To confirm this finding in a more physiological setting, we determined the effect of SB203580 on egress in an ex vivo culture of mixed exudates. We again observed a marked reduction of egress by the inhibitor (Fig. 9D). Over the course of three such experiments, the inhibition observed was always at least 80%.

Closer examination of the time-lapse images revealed that residual egress events in SB203580-treated cocultures had distinct features (Fig. 10). In control cultures, vacuole disruption was, as expected, closely followed by parasite dispersal from the host cell, so that the entire process was always completed within one 10-min interval. In contrast, in the presence of the inhibitor, there was typically a substantial delay between vacuole disruption and parasite dispersal, allowing these two events to be separately visualized in images 10 min apart. In ~10% of these events, dispersal failed to occur during the interval of observation, suggesting arrest at the disruption stage (SB Cell-3 in Fig. 10). These data suggest that the inhibitor may affect one or more parasite functions required for the efficient execution of egress.

In vivo fate of egressed T. gondii

We next sought evidence for a potential impact of the egress-reinvasion cycle on T. gondii pathogenesis. Use of SB203580 for this purpose was problematic, because this inhibitor can also suppress Toxoplasma growth (30). We therefore performed an experiment to compare the fate of exudate parasites residing in either the intracellular (nonegressed) or extracellular (recently egressed) compartments. We first identified conditions of brief centrifugation of exudate cells that substantially pelleted intracellular, but not extracellular parasites. A pool of exudates from YFP-RH-infected mice was then separated into aliquots that were either depleted by centrifugation or not (control). These aliquots, in which the intracellular proportion of total parasite burden was 10 or 93%, respectively, were transferred to WT-RH-infected hosts, and the recovery of total YFP-RH was then monitored by sequential peritoneal sampling and normalized to the recovery obtained at 1 h posttransfer for each host animal. By 1 h, parasites from the centrifuged aliquot had efficiently reinfection host cells: the cell-associated proportion of donor-derived parasites at this time point was 96 ± 0.4% for both aliquots (n = 5), indicating that parasites from the two aliquots were similarly viable and infectious. As in previous experiments, by 3 h posttransfer, the majority of intracellular donor parasite had egressed from donor cells (data not shown). Therefore, the 1- to 3-h posttransfer interval represents the period in which parasites from the two aliquots exist in substantially different states: either primarily residing in cells of origin (control) or having just completed a cycle of egress and reinfection (centrifuged). In the subsequent intervals, in contrast, the state of the two parasite populations is similar.

In accordance with this kinetic of parasite distribution, we observed a fate difference between the two aliquots that was specific to the 1- to 3-h interval (Fig. 11). The recovery of control parasites following this interval was 50% greater than that of the centrifuged aliquot, but no further difference occurred subsequently. The overall recovery of control parasite was reduced in comparison with
earlier adoptive transfer experiments (Fig. 2D). This is attributable to the need in this case to store exudate before transfer; we have observed that storage for 1 h significantly reduces subsequent recovery (data not shown). The differential recovery of egressed and nonegressed parasites suggests that nonegressing parasites may survive and expand preferentially in comparison with an egressing parasite compartment, consistent with a potential role for ETE in host defense.

The reduced recovery of recently invaded parasites suggests that many exudate macrophages possess activated antiparasitic function. To verify this expectation, we examined the ability of cell-free exudate fluid to induce antiparasitic function in macrophages. The results demonstrated that pretreatment of PEM with exudate fluid was comparable in this respect to pretreatment with IFN-γ, and that this activity was dependent on IFN-γ signaling and NO generation (Fig. 12). Induction of NO production by exudate fluid was comparable with the level induced by IFN-γ (data not shown). These data suggest that ETE could potentially act to limit parasite expansion by redistributing parasites to activated host macrophages (see Discussion and Fig. 13).

**Discussion**

The infective cycle of the tachyzoite form of *T. gondii* has been intensively investigated; however, studies to date have largely focused on elucidating the mechanisms of a fixed sequence of events, from invasion through replication and egress, rather than on the physiological regulation of these processes. It is indeed unclear how much regulation occurs for any of these events, either at the level of the parasite or initiated by the host. The current study

![FIGURE 10. Altered progression of egress by SB203580. The time-lapse series are from the experiment described in Fig. 9, A and B. A random number generator was used to select three events each from the cultures treated with either DMSO (Con; 101 total events) or SB203580 (SB; 27 total events). Arrowheads indicate vacuoles that have undergone disruption, but not dispersal. Scale bar: 10 μm.](http://www.jimmunol.org/)

![FIGURE 11. Fate of egressed parasites. Donor and host mice were infected for 5 days with YFP-RH or WT-RH, respectively. Exudates from four donors (verified to have similarly progressed toxoplasmosis) were pooled and divided into aliquots that were either centrifuged for 2 min (discarding the pellet) to enrich recently egressed parasites (Spun), or held at 4°C (Control). Each aliquot was transferred to a group of five host mice, of which three expressed transgenic cyan fluorescent protein to mark host cells to monitor egress (data not shown). Recovery of donor parasite was monitored by sequential paracentesis and normalized to the 1-h values for each host animal. *, p < 0.05. The data are representative of two similar experiments.](http://www.jimmunol.org/)

![FIGURE 12. Activation of antiparasitic function by exudate fluid. WT or IFN-γR1−/− PEM (1 x 10⁵) were cultured for 1 day in adherent 96-well dishes and then treated overnight with medium containing either 100 U/ml IFN-γ, 20% cell-free exudate fluid prepared from infected mice (Exudate fluid), or no addition (Medium). Cells were then infected for 1 day with GFP-RH at moi of either 0.1 or 2, in the presence or absence of the NO synthase inhibitor N-nitro-L-arginine methyl ester (2.5 mg/ml). Data from the two infection levels were similar and were pooled for analysis. Cells were harvested by treatment with Accutase for 10 min and assessed for parasite content by flow cytometry, using fluorescent beads to normalize sample volume (n = 4). *, p < 0.05. The data are representative of two experiments.](http://www.jimmunol.org/)
A key finding of this study is that, in contrast to in vitro models of tachyzoite growth, in which the interval between parasite invasion and egress is typically 36–48 h, in the macrophages that host a large fraction of the parasite burden during the acute inflammatory response this interval is dramatically shortened. Our data imply that a majority of the parasite residing in peritoneal exudate cells egresses approximately every 3–4 h, and the kinetics of macropage turnover suggest an even shorter interval (2–3 h) for this cell type. Our conclusion that egress is a dominant event in this setting is based on four lines of evidence. First, the distribution of vacuolar parasite content in macrophages is consistent with a probabilistic, rather than proliferation-dependent model of egress. Second, rapid parasite transfer between donor and host cells is observed in adoptive transfer experiments. Third, the turnover of donor cells in these experiments is consistent with lytic egress as the mechanism of parasite transfer. Fourth, a similar rate of egress was induced in infected adherent macrophages cocultured with exudate cells. The finding that donor-to-host parasite transfer continues to occur at a steady rate many hours after transfer, and that this transfer is mirrored by transfer at a similar rate from host to donor cells, strongly argues against an artifactual explanation for these observations.

Because T. gondii has been shown to egress efficiently in response to two cell death-inducing signals (Fas and perforin) (9), it is plausible that cytotoxic signals derived from activated leukocytes in the inflammatory exudate could provide the basis for high rates of egress in vivo. However, we found that the presence of lymphocytes was not necessary for ETE, arguing against a role for perforin. It remains possible that NK cell-derived perforin contributes to ETE. We have observed no consistent effect on ETE following magnetic depletion using the NK Ags NK1.1 and DX5; however, these small cell populations were not sufficiently well distinguished for us to be certain of their efficient removal (data not shown). Because depletion of macrophages reduced ETE (whereas neutrophil depletion had no effect), our data are most consistent with the view that the inflammatory macrophage is the major inducing cell for ETE, as well as the major responding cell from which egress is initiated. Our data suggest that macrophages acquire ETE-inducing capability as a result of the activating environment in infected mice, because macrophages elicited by a sterile irritant (thioglycolate) were less effective ETE inducers. However, the nature of the upstream activating signals that elicit ETE-inducing function requires further investigation.

The mechanism by which macrophages trigger egress remains to be determined. Activated macrophages release Fas ligand, and are also capable of undergoing Fas-mediated death in certain circumstances (31, 32). Immature monocytes are also susceptible to Fas-mediated death (33), and this susceptibility might be retained in many of the macrophages in the toxoplasmic exudate, because our data imply that these cells are predominantly recent emigrants. In addition, macrophages can release a variety of other mediators capable of triggering apoptosis, including reactive oxygen and nitrogen species. We have previously documented the occurrence of reactive oxygen-producing macrophages in the toxoplasmic inflammatory exudate (21), and we have also detected the progressive accumulation of NO in these exudates (data not shown). We have observed that 1–2% of the uninfected macrophages in the inflammatory exudate are TUNEL positive, whereas infected cells are uniformly TUNEL negative (20) (data not shown), so it is plausible that ETE represents a response to the induction of early stages of apoptosis (preceding endonuclease activation). However, our data do not support a role for reactive species-induced apoptosis in ETE, because we observed no inhibition of ETE with N-acetylcysteine or i-NIL. In addition, no inhibition was seen with cyclosporine, an inhibitor of the mitochondrial permeability transition pore that prevents apoptosis in many settings, including NO-mediated apoptosis in macrophages (24). We also failed to observe ETE inhibition by blockade of purinergic receptor signaling. We have attempted similar experiments using either soluble Fas ligand to assess the role of the Fas pathway or the pan-caspase inhibitor zVAD to more broadly address the role of apoptosis; however, each of these agents triggered significant egress in the absence of exudate, and the results were consequently inconclusive (data not shown). Further studies will be needed to more fully elucidate the role of apoptotic signaling in ETE.

Although the mechanism that governs ETE remains unknown, it is clear from our data that this mechanism includes a step that is highly and specifically sensitive to the MAPK inhibitor, SB203580. Importantly, the effect of SB203580 was validated in an ex vivo culture of mixed inflammatory exudates, eliminating the possibility that the result was an artifact of the ETE assay system. There are at least three sites at which this inhibitor might potentially exert its effect in the ETE assay: the triggering exudate cell (presumably a macrophage), which generates an ETE-inducing signal that acts on infected PEM; the host compartment of the target cell (PEM) within which parasites initiate egress; and the parasite itself. We are as yet unable to distinguish these possibilities, because SB203580 is a reversible inhibitor and can act on both mammalian p38 MAPK27 and a related T. gondii-encoded kinase, TgMAPK1 (28). Both of these kinases are capable of responding to external stress signals.

If the inhibitor exerts its effect by acting on p38 MAPK in the triggering cell, a potential mechanism is the inhibition of cytokine production, which can be blocked in macrophages by SB203580 at both the transcriptional and translational levels (34, 35). An alternative mechanism, which might operate on both triggering and target cells, is the inhibition of cell locomotion and/or the
formation of appropriate cell-cell contacts, because many studies have implicated p38 MAPK in the regulation of the cytoskeleton and the promotion of cell migration (27). We have not obtained direct evidence for a role of cell-cell interaction in ETE, because the standard two-compartment assay used for this purpose is complicated by the fact that the triggering exudate cells also serve as a major destination for egressing parasites. However, it is noteworthy that we observed the efficiency of egress induction to be increased by approximately an order of magnitude when the target PEM were adherent (as in the time-lapse experiments) rather than nonadherent, as in our standard ETE assay (Fig. 6, compare C and E). Also, in the adherent system, egress induction continued for at least 2 h of coculture, whereas in the nonadherent system most of the observable events were limited to the first hour (data not shown). These observations are consistent with a role for substrate-facilitated migration and cell-cell interaction in ETE.

Alternatively, if the relevant inhibitor target is TgMAPK1, then the inhibition of ETE could be due to either an effect on regulatory signals that govern parasite commitment to egress or alternatively to a blockade in the execution of egress. It may be pertinent in this respect that we have noted some inhibition by SB203580 of PEM invasion by T. gondii (data not shown), which has also been reported by others in the monocytic cell line THP-1 (36). Although these invasion studies do not distinguish between SB203580 effects on host and parasite kinases, the findings are consistent with the notion that ETE inhibition occurs via a blockade in egress execution, because parasite egress and invasion share mechanistic features (3, 7). The finding that residual egress in SB203580-treated cultures shows an abnormal delay between vacuole disruption and parasite dispersal is also supportive of an inhibitor effect in the execution phase. However, because recent evidence suggests that host plasma membrane disruption is not a simple consequence of parasite exit per se (7), delayed dispersal may also reflect alteration of a host, rather than parasite function. It will be of interest to determine more precisely the events that are delayed or abrogated by SB203580. If a fundamental execution step of egress is affected, it might be expected that conventional proliferation-dependent egress during cell culture would also be sensitive to SB203580; however, this is difficult to test because parasite replication is inhibited by the action of the drug on TgMAPK1 (30).

The mechanistic relationship between ETE and proliferation-dependent egress will be an important subject for future enquiry. Most studies of T. gondii egress have been restricted to egress events artificially induced by agents such as calcium ionophores, DTT, or bacterial toxins. However, the recent work of Lavine and Arrizabalaga indicates that proliferation-dependent egress is most likely driven by stress of the host cell membrane due to the growing parasitophorous vacuole, and is mechanistically distinct in several respects from artificially induced egress. In particular, a key feature common to all instances of artificially induced egress is dependence upon intracellular calcium, as evidenced by the prevention of egress with BAPTA-AM. Similar calcium dependence has been recently observed for egress induced by the ligation of death receptors in infected B lymphoma cells (9). In contrast, Lavine and Arrizabalaga report that proliferation-dependent egress is completely resistant to BAPTA-AM. These findings throw into question the physiological relevance of the earlier mechanistic findings. The current study, however, shows that inducible egress occurs physiologically and that it is dependent on intracellular calcium, in this respect resembling egress induced artificially or via death receptors, rather than proliferation-dependent egress. It is possible, then, that ETE represents a mechanistically distinct egress program that can be regulated by the parasite in response to environmental cues. It is possible, for example, that macrophages as host cells provide a favorable environment for ETE, because infection of macrophages with RH strain T. gondii up-regulates host cell p38 MAPK function in a calcium-dependent manner (37). It will be of interest to determine whether the induction of ETE is specific to certain host cell types, and also whether features of artificially induced egress, such as host cell permeabilization and potassium entry, contribute to the mechanism of physiological ETE. A feature of artificially induced egress that may not be shared with ETE is dependence on the calcium-dependent protein phosphatase calcineurin, since the calcineurin inhibitor cyclosporine A, which blocks calcium ionophore-induced egress (6), had no effect on ETE.

Alternatively, it remains possible that all T. gondii egress is calcium dependent. A recent study, using the same parasite strain and host cell as Lavine and Arrizabalaga, indicates that proliferation-dependent T. gondii egress is controlled through the production of abscisic acid (29). This finding suggests that the egress pathway is a calcium-dependent process, because abscisic acid stimulates calcium release in plants and other species (38–40), and activates calcium-dependent secretion in T. gondii (29). It is possible that the BAPTA-AM resistance of egress observed by Lavine and Arrizabalaga (10) reflects a difference in the calcium concentration required for rapid responses to external triggers, in comparison with a process that occurs over 1–2 days. However, it is also possible that proliferation-dependent egress can occur by more than one mechanism. We did not detect abscisic acid dependence of ETE, implying, at least, that the initiating signals upstream of calcium release are distinct for ETE and proliferation-dependent egress. Whether this is the case for downstream events remains a question for further investigation.

The notion that ETE represents an environmentally cued parasite response suggests that this form of egress has developed as an adaptation of the parasite, for example, as a means to escape inhospitable or dying cells. It has been suggested that induced egress may favor dissemination in the face of an immune response (9). However, our results suggest a second scenario, not exclusive of the first, in which the benefit to the parasite, if any, comes at a price in the setting of acute inflammation. Our data indicate that parasite sites that have recently reinfected new cells after egress fare poorly compared with those that have maintained longer residence in their host cells. The simplest explanation for this finding is based on the likelihood of heterogeneity among host cells (Fig. 13). We have shown that the environment of the peritoneal exudate is capable of inducing antiparasitic activity through the IFN-γ-inducible NO synthase axis. Because we have also shown that most macrophages in the exudate are likely to be very recent emigrants from the circulation, with varying degrees of exposure to these activating stimuli, and because the parasite can suppress IFN-γ-mediated activating signals that only arrive after infection (41), it is likely that exudate macrophages are highly heterogeneous with respect to activation state and consequent permissiveness. Consistent with this idea, we have demonstrated the existence of a subpopulation of reactive oxygen-expressing macrophages that appear resistant to infection in vivo (21). Heterogeneity of permissiveness is in fact characteristic even of resident peritoneal macrophages in normal mice (42, 43). Because parasites will expand preferentially in more hospitable cells, such cells will perform contain a disproportionately high share of total parasite. Therefore, any process, such as ETE, that tends to randomize parasite distribution will result in a net shift of parasite from more to less hospitable host cells, with benefit to the host. Such a host strategy of haven disruption could
in principle provide benefit in any setting in which an intracellular parasite was subject to an inducible release-reinfection cycle. The ultimate balance between parasite and host benefit might then depend on the signaling context of the setting in which egress was induced.

These considerations suggest that ETE may potentially contribute both to parasite expansion and to host defense. The ability to manipulate and exploit this modality of parasite transmission may therefore yield multiple avenues of investigation and intervention in Toxoplasma pathogenesis. In particular, the view of T. gondii egress as a microenvironmentally regulated event promises to provide further insight into the emerging picture of host-parasite interaction as a coevolved and highly adapted network.

Appendix

Probabilistic egress model

A theoretical distribution for parasite proliferation value (P) under the model of probabilistic egress (Fig. 1B) is derived as follows:

It is assumed that initially all parasite cycles are synchronous, and that all egress occurs at the parasite cycle midpoint (because egress is assumed to be unbiased with respect to cycle stage). We then consider two distributions: \( D_m(g) \) is the distribution of \( P \) immediately following the completion of the mid-cycle egress plus reinvasion process in two distributions: \( D_m(g) \) is the distribution of \( P \) immediately following parasite division after the completion of generation \( G \). A value for \( D_m \) or \( D_c \) has the form \((f_0, f_1, \ldots)\), where \( f_n \) is the fraction of total parasite for which \( P = n \).

Suppose that for \( g = a \), \( D_m(a) = (m_0, m_1, \ldots) \). Then \( D_c(a) = (0, m_0, m_1, \ldots) \) (because all vacuoles now have at least two parasites). It follows that:

\[
D_m(a + 1) = \{q(1 - q)m_0, q(1 - q)m_1, \ldots\},
\]

where \( q \) is the probability of egress.

Iteration then yields the series:

\[
D_m(a + 1) = \{q(1 - q)m_0, q(1 - q)m_1, \ldots\},
\]

\[
D_m(a + 2) = \{q(1 - q)q, q(1 - q)m_1, \ldots\},
\]

\[
D_m(a + 3) = \{q(1 - q)q^2, q(1 - q)q, \ldots\}.
\]

It is then evident by induction that:

\[
D_m(g) = \{q(1 - q)n, 1 - q^n, \ldots\}.
\]

Because \( D_m \) and \( D_c \) represent distributions that occur over equal time intervals (one half cycle each), the asynchronous distribution \( D \) is their mean:

\[
D = (0.5q, 0.5(q(1 - q^n) + q(1 - q^{n-1}))\ldots).
\]

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Disclosures

The authors have no financial conflict of interest.

References


Supporting information.

Supplementary figures.

Figure S1. Vacuolar parasite content is independent of the number of vacuoles per cell. The datasets employed for Fig. 1B were examined to determine the number of parasites per vacuole as a function of the number of vacuoles in each inflammatory macrophage. Each curve represents one mouse, corresponding to one of the three mice used for the data displayed in Fig. 1B.

Figure S2. YFP-RH fluorescence measures intracellular parasite content. Exudate cells were collected by lavage from three mice infected 4d previously with YFP-RH. Cells were pooled, fixed with and equal volume of 4% paraformaldehyde, and three populations of different mean fluorescence (MFI) were obtained by sorting on a MoFlo cell sorter. The sorted populations were analyzed for parasite content / cell by fluorescence microscopy of cytospin preparations (total of 597 vacuoles counted). The plot displays the correlation between MFI and parasite content for the three populations (r² = 1.00). Dotted line = linear regression of the data.

Figure S3. Enhanced recruitment of myeloid cells to peritonitis induced by T. gondii compared to thioglycolate. Groups of 5 recipient mice (C57BL/6; CD45.1) were either infected with wild-type RH or else injected with thioglycolate. After 5d, pooled donor blood was obtained from 3 SJL mice (CD45.2) by cardiac puncture into EDTA-saline and transfused into recipient mice by tail vein injection (0.2 ml). After 3h, blood and peritoneal lavage samples were obtained and analyzed by flow cytometry after hypotonic lysis of red blood cells, followed by incubation for 1 h with Fc block, FITC-anti-CD45.1, PE-anti-CD454.2 and either 647-anti-Thy1 to detect T cells or 647-anti-CD11b to detect myeloid cells. F4/80 was not used since it is not sufficiently expressed on monocytes (1)(data not shown). Samples were fixed with an equal volume of 4% paraformaldehyde and fluorescent beads were added to normalize the analyzed volume. No Thy1+ donors were detected in lavage. The results show that while chimerism (as indicated by donor Thy1+ levels) was comparable in the two groups, the distribution of donor myeloid cells was strongly shifted from blood to the peritoneal cavity in infected, compared to thioglycolate-treated hosts.

Figure S4. Time-dependence of ETE. ETE assays were performed as described in the legend to Fig. 7E, except that the time of co-culture was varied as indicated. Data represent the mean ± S.E. of four replicates. *, p < 0.05 compared to time 0.

Figure S5. Uniformity and stability of Hoechst 33342 labeling of donor cells. Data are derived from the experiment displayed in Fig. 2. (A) Dot plot (gated on total viable cells, excluding free parasites) displaying the Hoechst 33342 labeling in untransferred donor cells from a GFP-RH-infected donor mouse. The donor and host gates used for egress analysis are shown, as well as the percent of donor cells in each gate. (B) Uniformity of labeling in infected donor cells. The sample displayed in panel A was analyzed for Hoechst staining as a function of GFP-RH content. Infected donor cells (GFP+) are 99
percent labeled. (C, D) Stability of Hoechst 33342 labeling. Panel C (gated as for panel A) illustrates the retention of label in donor cells in a sample removed at 8 h post-transfer from a host mouse injected with the donor cells displayed in panel A. Panel D displays the mean Hoechst 33342 intensity (MFI) of cells in the donor gate at 0 - 8 h post-transfer. Three curves are displayed, each corresponding to a host animal injected with the donor cells displayed in panel A.


**Videos**

**Videos 1 - 3. Regulation of egress by exudate cells and SB203580.** PEM were infected with YFP-RH overnight and then exposed for 2h to either blank medium (Video 1), exudate cells + DMSO (Video 2), or exudate cells + SB203580 (Video 3). YFP fluorescence was collected at 10 min intervals. Cells that undergo egress are indicated by arrows.
Parasites per vacuole vs Number of vacuoles in the cell.