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CD1d Activation and Blockade: A New Antitumor Strategy

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CD1d is expressed on APCs and presents glycolipids to CD1d-restricted NKT cells. For the first time, we demonstrate the ability of anti-CD1d mAbs to inhibit the growth of different CD1d-negative experimental carcinomas in mice. Anti-CD1d mAbs systemically activated CD1d+ APC, as measured by production of IFN-γ and IL-12. Tumor growth inhibition was found to be completely dependent on IFN-γ and IL-12 and variably dependent on CD8+ T cells and NK cells, depending upon the tumor model examined. Anti-CD1d mAb induced greater CD8+ T cell-dependent tumor suppression where regulatory CD1d-restricted type II NKT cells have been implicated, and were less effective in a NK cell-dependent manner against tumors where T regulatory cells were immunosuppressive. The ability of anti-CD1d mAbs to coincidently activate CD1d+ APCs to release IL-12 and inhibit CD1d-restricted type II NKT cells makes CD1d an exciting new target for immunotherapy of cancer based on tumor immunoregulation. The Journal of Immunology, 2009, 182: 3366–3371.

Dendritic cells (DCs) play an important role in the induction of an antitumor immune response (1). Depending on the activation signals received from either Toll-like receptors or interacting lymphocytes, DCs can differentiate and mature as characterized by up-regulation of costimulatory molecules and production of IL-12 and IFN-α (2, 3). Production of these immune activating cytokines often occurs in the context of cross-talk with lymphocytes such as conventional T cells, NKT cells (4), and NK cells (5), depending upon the initiating stimulus. However, in cancer, the interaction of DCs with tumors, tumor-derived factors, or other cells in the tumor microenvironment can result in DC differentiation into tolerogenic types (6). Given the importance of activated/matured DCs in the initiation and activation of tumor-specific T cells, many preclinical studies and clinical trials have examined the use of different agents to activate/mature DC (1).

To date, DCs have been activated/matured directly through the use of agonistic anti-CD40 mAbs (7) or Flt3L (8), or indirectly using CD1d-reactive glycolipid ligands such as α-galactosylceramide (α-GC), that engage cross-talk with CD1d-restricted type I NKT cells (9). However, toxicity issues have been associated with agonistic CD40 mAbs in recent clinical trials (10, 11), and a recent study has demonstrated that signaling of CD40 through endothelial cells can promote tumor growth (12). In addition, qualitative and quantitative defects of type I NKT cells in cancer patients can potentially hamper the use of glycolipid ligands to indirectly activate DCs via type I NKT cell TCR (13–15).

An overlooked alternative strategy to activate DCs involves the use of anti-CD1d mAbs. CD1d is expressed prominently and constitutively on professional APCs such as DCs, macrophages, and B cells (16). We recently demonstrated that anti-human CD1d mAbs matured DCs, resulting in the production of bioactive IL-12p70 (17). In addition to its agonistic activity, anti-CD1d mAbs may potentially block the activation of CD1d-restricted type II NKT cells (18). Two types of NKT cells are restricted by CD1d: type I and type II. Type I NKT cells have a host protective role in natural or α-GC-induced tumor immunity (9, 19, 20). In contrast, type II NKT cells have different tissue distribution in humans and mice and have been implicated in immunosuppression of effector T cell responses to cancer (18, 21, 22).

In this study, we assessed the antitumor efficacy of anti-CD1d mAbs in three different experimental tumor models: R331 renal carcinoma, 4T1 mammary carcinoma, and CT26L5 colon adenocarcinoma. These tumors do not express CD1d, and evidence suggests or shows type II NKT cells play a role in immune suppression in the 4T1 and CT26L5 tumor models (18), whereas regulatory T cells control immune effector response to the R331 tumor (M.J.S., unpublished data). We demonstrate, for the first time, the ability of anti-CD1d mAb monotherapy to inhibit the s.c. growth of three different aggressive experimental tumor models. Interestingly, anti-CD1d mAb therapy resulted in greater suppression of 4T1 and CT26 tumors than R331, and the mechanism of growth suppression was distinct. Given that our results implicate the ability of anti-CD1d mAb to coincidently activate APCs and block type II NKT cell activation, we believe there is a rationale to use similar agonistic mAb reactive with human CD1d in the treatment of human tumors immunoregulated by type II NKT cells.

Materials and Methods

Mice

Inbred BALB/c mice were either bred at the Peter MacCallum Cancer Centre (Peter Mac) or purchased from The Jackson Laboratory or the Walter and Eliza Hall Institute of Medical Research. BALB/c SCID, BALB/c CD1d gene-targeted (CD1d−/−), and BALB/c Jα18 gene-targeted mice (Jα18−/−) (18) were bred and maintained at the Peter Mac or purchased from The Jackson Laboratory. Female mice were used for all experiments with the 4T1 tumor and all experiments were performed in
cytokines assayed for IL-12p70 or IFN-γ production from CD1d+ splenocytes. A and B, Splenocytes from BALB/c WT or BALB/c CD1d−/− mice were stimulated with either soluble or plate-bound anti-CD1d mAbs (10 μg/ml), isotype control, α-GC (100 ng/ml), or LPS (10 ng/ml). Supernatants were collected one (A, C, E) and 3 days (B and D) after stimulation and cytokines assayed for IL-12p70 or IFN-γ by ELISA. F, Purified splenic F4/80+ class II+, CD3+, or CD19+ populations were cultured as shown with plate-bound anti-CD1d or LPS for 24 h and active IL-12p70 measured by ELISA. G and H, Groups of BALB/c WT or BALB/c CD1d−/− mice (n = 2–4) were injected i.p. with anti-CD1d mAb (50 μg) or cIg and serum harvested 24 h later to assay for IL-12 (G) and IFN-γ (H). All cytokines were determined by at least triplicate by ELISA. Results represents mean ± SDs. Representative of two experiments and time points. Statistical differences in cytokine production between isotype- and anti-CD1d mAb-treated WT mice were determined by Mann-Whitney Rank Sum test (*, p < 0.05).

According to [25], St. Louis, MO) were prepared and used as previously described (25). Anti-asialo GM1 (anti-ASGM1) for depletion of NK cells was obtained from Wako Pure Chemical and used as previously described (26). Recombinant mouse IL-12 was provided by the Genetics Institute.

**Tumor cell lines**

BALB/c-derived CD1d-negative TRAIL-sensitive 4T1 mammary carcinoma (23, 24), the TRAIL-sensitive Renca variant R331 (23), and the carcino- gen-induced colon carcinoma line Colon-26 (CT26L5) (18) were maintained as described previously.

**Antibodies**

We prepared and purified agonistic mAbs reactive with mouse CD1d (1B1) or purchased from BD Pharmingen, along with isotype control rat IgG2b. Depleting anti-mouse CD4 (GK1.5), anti-CD8 (53–6.7) mAb, neutralizing mAb to mouse IL-12 (C17.8) and IFN-γ (H22) (provided by Dr. Robert Schreiber, Washington University School of Medicine, St. Louis, MO) were prepared and used as previously described (25). Anti-asialo GM1 (anti-ASGM1) for depletion of NK cells was obtained from Wako Pure Chemical and used as previously described (26). Recombinant mouse IL-12 was provided by the Genetics Institute.

**Measurement of cytokines derived from splenocytes or serum**

Splenocytes (1 × 10^6/well) from BALB/c wild type (WT) or BALB/c CD1d−/− mice were stimulated with soluble or plate-bound anti-CD1d mAbs (10 μg/ml), isotype control (10 μg/ml), α-GC (100 ng/ml) (Kirin Pharma), or O111:B4 *E. coli* lipid A (LPS) (10 ng/ml) (Sigma-Aldrich) in 96-well plates as previously described (17). Supernatants were collected at day 1 and 3 after stimulation and assayed for IFN-γ and IL-12. In some experiments, PE-labeled beads (Miltenyi Biotec) (1 μl per 200 μl cells) were used to prepurify F4/80+, class II+, CD3+, or CD19+ populations from mouse spleen as per the manufacturer’s instructions. These populations (>90% pure) were then cultured (10^5 per well) in a similar manner with plate bound anti-CD1d or LPS (and 10 ng/ml IFN-γ) for 24 h to measure active IL-12p70. For serum cytokines, groups of BALB/c WT (n = 4) or BALB/c CD1d−/− mice (n = 2) were i.p. injected with...
Mice were treated i.p. with anti-CD1d mAbs (200 μg) or isotype control (50 μg). Serum was harvested 24 h later and assayed for IFN-γ and IL-12. All cytokines were determined by at least triplicate samples by ELISA (PBL Biomedical Laboratory; R&D Systems). Limits of detection were 1 pg/ml.

**Therapy of transplanted tumors**

Groups of five BALB/c WT or BALB/c CD1d<sup>−/−</sup> mice (n = 5) were inoculated s.c. with the renal carcinoma cell line, R331 (5 × 10<sup>5</sup>) (A), the mammary carcinoma cell line 4T1 (2 × 10<sup>5</sup>) (B), and the colon carcinoma cell line CT26L5 (1 × 10<sup>5</sup>) (C). On days 11, 15, and 19 after tumor inoculation, WT mice were treated i.p. with anti-CD1d mAbs (200 μg) or cIg as indicated. Tumor sizes are represented as the mean ± SEM and tumor rejection rates are indicated in parentheses. Statistical differences in tumor sizes between cIg- and anti-CD1d-mAb-treated WT mice or CD1d<sup>−/−</sup> mice were determined by Mann-Whitney Rank Sum test (*, p < 0.05).

**Statistics**

Statistical differences in between control Ig (cIg)-treated mice or mAb-treated mice injected with anti-CD1d mAbs were determined by Mann-Whitney Rank Sum test (*, p < 0.05).

**Results**

Anti-CD1d mAb induce APC production of IFN-γ and IL-12

We first tested the ability of anti-CD1d mAb to activate CD1d<sup>+</sup> APCs by testing for the production of IFN-γ and IL-12 (Fig. 1). Mouse splenocytes from BALB/c WT or BALB/c CD1d<sup>−/−</sup> mice were stimulated with plate-bound anti-CD1d mAb, isotype control, α-GC, or LPS for 1 (Fig. 1, A and C) or 3 days (Fig. 1, B and D). Supernatants derived from BALB/c WT splenocytes cultured on anti-CD1d mAbs resulted in more dramatic increases in both IL-12 and anti-CD1d mAb (50 μg) or isotype control (50 μg). Serum was harvested 24 h later and assayed for IFN-γ and IL-12. All cytokines were determined by at least triplicate samples by ELISA (PBL Biomedical Laboratory; R&D Systems). Limits of detection were 1 pg/ml.

**Therapy of transplanted tumors**

Groups of five BALB/c WT, BALB/c CD1d<sup>−/−</sup>, BALB/c Jα18<sup>−/−</sup>, or BALB/c SCID mice were inoculated s.c. with either 1 × 10<sup>5</sup> CT26L5, 2 × 10<sup>5</sup> 4T1, or 5 × 10<sup>5</sup> R331 tumor cells on day 0. On the days indicated after tumor inoculation, groups of mice were treated i.p. with anti-CD1d mAb (200 μg) or cIg (200 μg). In some experiments, mice were depleted of CD<sup>+</sup> (αGC) or LPS for 1 (Fig. 1, A and C) or 3 days (Fig. 1, B and D). Supernatants derived from BALB/c WT splenocytes cultured on anti-CD1d mAbs resulted in more dramatic increases in both IL-12

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and IFN-γ above those cultured with isotype control (Fig. 1, A–D). The increases in cytokine production were CD1d-dependent as only baseline levels of IL-12 and IFN-γ were detected from BALB/c CD1d−/− spleenocytes stimulated with anti-CD1d mAbs (Fig. 1, A–D). The levels of cytokines produced by WT splenocytes activated by anti-CD1d mAb were similar to that induced by CD1d-dependent α-γC, but lower than those stimulated with the CD1d-independent TLR4 agonist LPS (Fig. 1, A–D). Soluble anti-CD1d also produced active IL-12p70, although cross-linking was optimal (Fig. 1E). Consistent with previous reports (27–30), when isolated into purified populations it was clear that F4/80+ macrophages and class II+ cells were stimulated to produce active IL-12p70 by plate bound anti-CD1d, whereas neither T nor B cells released IL-12p70 (Fig. 1F). We next tested the agonistic function of anti-CD1d mAbs in vivo. BALB/c WT or BALB/c CD1d−/− mice were treated i.p. with anti-CD1d mAbs (200 μg) or clg. Tumor sizes are represented as the mean ± SEM, representative of two independent experiments. Statistical differences in tumor sizes between mice injected with clg and anti-CD1d mAbs and mAb-treated mice injected with anti-CD1d mAbs were determined by Mann-Whitney Rank Sum test (*, p < 0.05).

Anti-CD1d induces optimal tumor suppression when CD1d-restricted type II NKT cells regulate growth

We next assessed the antitumor efficacy of anti-CD1d mAbs in three different s.c. tumor models, the renal carcinoma cell line R331 (Fig. 2A), the mammary carcinoma cell line 4T1 (Fig. 2B), and a colon carcinoma cell line CT26L5 (10^5) (C). On days 0, 3, 7, 11, and 15 (A) or 3, 7, and 11 (B) after tumor inoculation, mice were treated i.p. with anti-CD1d mAbs (200 μg) or clg. Additionally, some groups of mice were also injected i.p. on day 0 and then twice weekly with depleting anti-CD4, anti-CD8, and/or anti-ASGM1 Abs. Tumor sizes are represented as the mean ± SEM, representative of two independent experiments. Statistical differences in tumor sizes between mice injected with clg and anti-CD1d mAbs and mAb-treated mice injected with anti-CD1d mAbs were determined by Mann-Whitney Rank Sum test (*, p < 0.05).
FIGURE 6. Role of IFN-γ and IL-12 in tumour suppression by anti-CD1d mAb. Groups of BALB/c WT mice (n = 5) were inoculated s.c. with the renal carcinoma cell line R331 (5 × 10^5) (A), the mammary carcinoma cell line, 4T1 (2 × 10^5) (B), and the colon carcinoma cell line CT26L5 (1 × 10^5) (C). On days 0, 3, 7, 11, and 15 (A) or 3, 7, and 11 (B and C) after tumour inoculation, mice were treated i.p. with anti-CD1d mAbs (200 µg) or cIg. Additionally, some groups of mice were also injected i.p. on day 0 and then twice weekly with either neutralizing anti-IL-12 and/or anti-IFN-γ Abs or cIg. D, Groups of BALB/c WT mice (n = 5) were inoculated s.c. with the mammary carcinoma cell line 4T1 (2 × 10^5). Commencing on day 3 or 11 after tumour inoculation, mice received either PBS or 500 ng IL-12 for 5 or 4 successive days, respectively. Tumor sizes are represented as the mean ± SEM, representative of two independent experiments. Statistical differences in tumor sizes between mice injected with cIg and anti-CD1d mAbs and mAb-treated mice injected with anti-CD1d mAbs were determined by Mann-Whitney Rank Sum test (p < 0.05).

Anti-CD1d mAb-induced tumor suppression is dependent on IL-12 and IFN-γ

To determine the cells and cytokines involved in anti-CD1d mAb-mediated tumor suppression, we inoculated groups of BALB/c WT mice, BALB/c SCID mice, or BALB/c Jot18 mice, with R331 or 4T1 or CT26L5 tumors and treated with anti-CD1d mAb on days 0, 3, 7, 11, and 15 (for R331) or days 3, 7, and 11 (for 4T1 and CT26L5) (Figs. 4, 5, and 6). Additionally, some groups of mice were depleted of CD8^+ T cells and/or NK cells or neutralized with anti-IFN-γ and/or IL-12. Interestingly, depletion of NK cells, but not CD8^+ T cells, almost completely abrogated the antitumor effect of anti-CD1d mab against R331 tumor growth (p = 0.0119, 0.0079) (Fig. 4A) and depletion of both NK cells and CD8^+ T cells was completely inhibitory (p = 0.0119) (Fig. 4A). Similar R331 tumor growth suppression by anti-CD1d mAb was observed in Jot18^−/− and SCID mice, indicating that T cells, including type I NKT cells, were not required (Fig. 5A). Neutralization of either IL-12 and/or IFN-γ completely inhibited the ability of anti-CD1d mAb to suppress R331 tumor growth compared with similar groups of mice treated with cIg and anti-CD1d (p < 0.05) (Fig. 6A). Overall, these data suggested that anti-CD1d mAb stimulated IL-12 and subsequent NK cell activation and IFN-γ secretion was largely responsible for activity against s.c. R331. By contrast, while either IL-12 or IFN-γ neutralization also completely inhibited the antitumor activity of anti-CD1d mAb against 4T1 and CT26L5 tumors (p < 0.05) (Fig. 6, B and C), NK cells, type I NKT cells, and CD4^+ T cells were dispensable (p > 0.4) (Fig. 4, B and C, and Fig. 5B). Interestingly, depletion of CD8^+ T cells resulted in a markedly reduced ability of anti-CD1d mAb to suppress 4T1 and CT26L5 tumor growth (p = 0.0119, 0.0079) (Fig.
4, B and C). Thus, in models where type II NKT cells are postu-
lated to suppress effector tumor-specific CD4+ and CD8+ T cells, anti-CD1d mAb predominantly mediates its antitumor ef-
fect via CD8+ T cells in an IFN-γ- and IL-12-dependent man-
ner. Despite the importance of host IL-12, recombinant mIL-12
given at day 0 or 8 after tumor inoculation did not provide
tumor protection equivalent to anti-CD1d mAb, indicating that
the role of IL-12 was context dependent, rather than systemic
(Fig. 6D).

Discussion
This is the first report to document the potential antitumor activity
and mechanism of action of anti-CD1d mAb. Anti-CD1d mAbs
systematically activated CD1d+ APCs, as measured by in vitro and in
vivo production of IL-12 and IFN-γ. Tumor growth inhibition was
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anti-CD1d mAbs to coincidently activate CD1d

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References
D1-restricted natural killer T cells mature dendritic cells, leading to strong adaptive immunity.
8. Lynch, D. H., A. Andreassen, E. Maraskovsky, J. Whitmore, R. E. Miller, and
J. C. Schuh. 1997. FLIC ligand induces tumor regression and antigen-specific immune
11. Vonderheide, R. H., K. T. Flaherty, M. Khalil, M. S. Stumacher, D. L. Bajor,
12. Chiiodoni, C., M. Iezzi, C. Guiducci, S. Sangarelli, I. Alessandrini, C. Ratti,
and M. A. Exley. 2006. Loss of IFN-γ production by in vivo activated NK T
16. Vonderheide, R. H., K. T. Flaherty, M. Khalil, M. S. Stumacher, D. L. Bajor,
ligation on human monocytes directly signals rapid NF-κB activation and pro-
18. Terabe, M., J. Swann, E. Ambrosino, P. Sinha, S. Takaku, Y. Hayakawa,