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*J Immunol* 2009; 182:1912-1918; doi: 10.4049/jimmunol.0803777

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A Role of IgM Antibodies in Monosodium Urate Crystal Formation and Associated Adjuvant Activity

Uliana Kanevets,* Karan Sharma,* Karen Dresser,† and Yan Shi2*

Uric acid is released from injured cells and can act as an adjuvant signal to the immune system. Uric acid crystals invoke strong inflammatory responses in tissues. Although their biological effects are evident and the associated signaling mechanisms are becoming clear, it remains unexplained as to why uric acid precipitates rapidly in vivo, in sharp contrast to the minimal crystallization in vitro. We report in this study that a group of IgM Abs is able to bind to these crystals, which is interesting in light that B cell-deficient mice do not sense the proinflammatory adjuvant effect of uric acid. The titers of these Abs increase upon immunization with uric acid crystals. We have produced large quantities of such mAbs. The purified IgM Abs can significantly facilitate uric acid precipitation to form the inflammatory crystals in vitro. Infusion of these Abs into B cell-deficient mice significantly increases the basal level of inflammation in these recipients and restores the host’s ability to sense uric acid’s adjuvant activity. Therefore, we have identified a factor in determining uric acid precipitation and possibly its ability to function as an endogenous adjuvant. This finding suggests a new mechanism of the pathogenesis of gouty arthritis and uric acid-induced immune activation.


Received for publication November 11, 2008. Accepted for publication December 15, 2008.

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This work was supported by grants to Y.S. from Alberta Heritage Foundation for Medical Research and Canadian Institutes of Health Research.

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Abbreviations used in this paper: MSU, monosodium urate; DC, dendritic cell; MPO, myeloperoxidase; UBA, uric acid-binding Ab.

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www.jimmunol.org/cgi/doi/10.4049/jimmunol.0803777
Materials and Methods

Mice, cells, and reagents

All mouse strains were housed at the University of Calgary Animal Research Centre under protocols approved by the University of Calgary Animal Research Review Committee. m.mMT mice were purchased from The Jackson Laboratory. Control IgM Abs were either purchased from Sigma-Aldrich or produced from similar MSU negative-staining mAb hybridomas. The CD11b.1 hybridoma (FITC) Abs were from eBioscience. Secondary goat anti-mouse IgG plus IgM (H + L, 1:150, 0.05-0.068), IgM (μ-chain, 115-095-020), and F(ab')2 (115-096-146) Abs were purchased from Jackson ImmunoResearch Laboratories. IgM fragmentation kit was from Pierce. SIINFEKL peptide was a gift from K. Rock (University of Massachusetts Medical School, Worcester, MA). 51Cr label was freshly ordered for each experiment from MP Biomedical. C5 complete ELISA kit was purchased from BD Biosciences. All other cytokine measurement reagents were made in our laboratory from chemicals (all from Sigma-Aldrich). Dendritic cell (DC) cultures, cell lines, all other cytokine sources Centre under protocols approved by the University of Calgary Animal Research Review Committee. 

Immunization, mAb production, and HPLC Ig purification

C57BL/6 (B6) and BALB/c mice were immunized with 1 mg of suspended MSU crystal i.p. in PBS without any adjuvant, and were boosted with the same dose biweekly for 2 mo. Antiserum were collected at this point for the FACS analysis. Tail blood (≥100 μl) was mixed with 1 μl of heparin and spun in a microcentrifuge to remove blood cells. Splenocytes were then harvested and fused with A3A tumor cells using a standard hybridoma protocol (26), with 105 tumor partners fused with 4 × 106 splenocytes. Between weeks 2 and 4, 25 μl of cell supernatants from growing hybridomas were used to determine the IgM clones. All clones with moderate to high binding were kept, along with several negative-binding IgM controls.

Cytotoxic T cell lysis assay

C57BL/6 or m.mMT mice were s.c. immunized with 5 μg of OVA-coated 1-μm-diameter latex beads in 100 μl of PBS, as previously described (2). For those that received the Ab infusion, 500 μg of purified Abs was injected via tail vein on the same day before the immunization. The immunized mice were rested for 7 days and were then sacrificed, and 5 × 106 splenocytes were stimulated in 10 ml of culture medium in the presence of 10-7 M SIINFEKL peptide. Five days later, 5000 51Cr-labeled EL4 cells pulsed with 10-6 M SIINFEKL were used as target at the E:T ratios indicated. Unpulsed EL4 cells were used as a background control, which produced negligible reading.

Hybridoma culture, typing, fragmentation, and purification

B cell hybridomas were cultured in RPMI 1640 with 10% FBS plus 1 mM HEPES, 25 μM 2-ME, penicillin/streptomycin antibiotics, and hypoxanthine-aminopterin-thymidine medium. As a standard operation, the mAb from the supernatant was isolated from 800 ml of the supernatants collected. An equal volume of the saturated solution of ammonium sulfate was slowly added to the supernatant to precipitate the Ab. The mixture was centrifuged, and the precipitate was collected and dissolved using 30–50 ml of PBS. This solution was then dialyzed three times against PBS overnight. The solution was then analyzed using HPLC. The HPLC analysis was performed with a Shimadzu Prominance system with a CBM-20-A controller, a LC-20AB fluid pump, and a SPD-M20A diode array. A total of 500 μl of the Ab solution was injected and analyzed on an Alltech MacroSphere GPC 300A 7u column (Alltech 88181) with an isocratic buffer containing 10 mM phosphate and 120 mM NaCl (pH 7.2). The samples were run at a rate of 1 ml/min over 40 min. The fractions collected were tested for the ability to bind MSU crystals, and then the active fractions were collected. This mixture was then further subjected to concentration via centrifugation with a Macrosep 300 K filter (Pall Filtron). The leftover fraction was tested for protein concentration using the bicinchoninic acid protein assay kit (Pierce). The fractions were then further purified with a UNO 12 column (Bio-Rad) with a Bio-Rad FPLC, with 10 mM ethanalamine (pH 9.6) as buffer A, and buffer A plus 1.5 M NaCl as buffer B, resolved at a rate of 1 ml/min in 25 min from 0% buffer B to 80% buffer B. IgM fragmentation and the associated analysis were performed with a kit and instructions from Pierce (ImmunoPure IgM Fragmentation Kit).

Flow cytometric analysis of MSU crystals

The flow analysis for MSU crystals was developed in our laboratory by modifying standard cell-based FACS analysis. A total of 100 μl of 1 mg/ml MSU crystal suspension was mixed with 100 μl of mAb supernatant or 1 μg of purified mAb (or as indicated otherwise) and incubated at room temperature for 20 min. The crystals were then washed twice with PBS and incubated with 0.5 μl of 1 mg/ml second Ab (FITC) in 100 μl for 10 min. The crystals were then washed again before FACS analysis. On a BD FACScan (BD Biosciences), MSU crystals showed a smaller size (~5–10% on a linear scale) on the forward scatter, and higher granularity (1 to several folds higher on a linear scale) on the side scatter. Typically, 40,000–50,000 total events were collected with CellQuest, and data were analyzed with Flowjo (Tree Star). Xanthine crystals were produced by first incrementally adding 1 N NaOH into 5 mg/ml xanthine suspension to dissolve the powder at high pH. Once completely dissolved, 1 N HCL was added to bring pH back to 7.0. The crystals thus precipitated were washed in 95% ethanol twice, and then in acetone. The crystals were dried before use.

Uric acid precipitation assay

A supersaturated uric acid solution (1 mg/ml) was incubated with various concentrations of a control protein (OVA) and MSU (27) (both by sizing and analysis by exchange column, ~99% pure by HPLC analysis) Ab, either UBA 11, UBA E6, UBA fragment, or a control in 1 ml at the indicated concentrations for 6 h. The supernatants were removed and wells were flushed. A total of 1 ml of 0.01 N NaOH solution was added to the wells, and 20 μl of the solution was added to 1 ml of water and read at 292 nm with a spectrophotometer. The conversion to the quantity of uric acid was achieved using a standard curve for UV absorbance from a set of uric acid solutions with known concentrations, as follows: Y (uric acid in μg) = 0.0766 × (UV 292 absorption) + 0.0298; R2 for the curve is 0.9937.

Serum uric acid measurement

Mice were injected i.v. with 500 μg of indicated Abs or PBS for 3 days, with or without the co-injection of 1 mg of soluble uric acid each day. Twenty-four hours after the last injection, blood from mice was collected into vials with 1 μl of heparin, and serum was isolated via centrifugation. A total of 2 μl of serum was mixed with 198 μl of buffer A, which contained 10 mM ethanalamine and 120 mM NaCl adjusted to pH 9.6. This mixture was passed through a nanosep 30 K filter (Pall Filtron), which would allow the uric acid in the sample to pass through, but would prevent possible protein contaminants from the sample. A total of 100 μl of the sample was then injected into the HPLC. Uric acid was resolved on a PolyWAX LP column (The Nest Group) by a gradient of 0–60% B (10 mM ethanalamine and 11.2 mM NaCl adjusted to pH 9.6) at 0.5 ml/min over 15 min. The HPLC analyses with EZ-start software were done at UV 292 nm. Peak area at 292 nm as a percentage of total area of the sample was used for the calculation of uric acid levels in the serum to minimize run-to-run or injection volume variability, as reported previously (25).

MPO measurement

MPO measurements were performed by a method adapted from Bradley et al. (27) on lung tissue isolated on day 4 from mice injected with Ab and uric acid solution. The tissue was homogenized with a 50 mM potassium phosphate buffer that contained 0.5% hexadecyltrimethyl ammonium bromide (pH 6.0). The samples were then vortexed, and the supernatant was centrifuged for 4 min at 5000 rpm. The 7-μl supernatant samples were placed in a 96-well plate, and 200 μl of 0.167 mg/ml o-dianisidine hydrochloride solution containing 0.0005% (w/v) of hydrogen peroxide was added and the changes in absorbance at 450 nm were measured using a microplate reader. The absorbance measurements were then converted to MPO units. The MPO values were generated by multiplying the UV reading by a factor of 0.2528 per a standard protocol. One MPO unit of activity is defined as amount of MPO required to degrade 1 μmol of peroxide per minute at 25°C.

Complement C3 reading

The blood from mice i.v. injected with Ab and uric acid was isolated using the caccard puncture technique (Pierce). The mice were then decapitated, and blood was placed in a microfuge tube containing 1 ml of EDTA and centrifuged at 13,000 rpm to collect serum. The serum samples were then prepared, according to the manufacturer’s protocol (Mouse Complement C3 ELISA kit; Kamiya Biomedical). The absorbance at 450 nm was measured using a
microplate reader. The absorbance was then converted to C3 concentration in mg/ml using a standard curve fitting.

**Neutrophil activation**

Blood samples were collected from mice injected with UBA E6 or control Ab as in the serum uric acid reading, and were subjected to hemolysis treatment and then stained with CD11b (FITC) and CD62L (PE) Abs. The samples were then analyzed with flow cytometry.

**Statistical analysis**

All results were reproduced in at least three independent assays. All error bars are 1 SD of the test sample groups. Student’s t test (2 tail) was used to produce the p values shown in the graphs.

**Results**

**B cells are required for the uric acid adjuvant effect**

We hypothesized that some serum factor, most likely Abs, facilitated and enhanced the immune activation associated with MSU crystals. As such, the proposal predicts that the MSU-mediated immune activation should be substantially decreased in the absence of Igs in vivo due to the reduced MSU crystal precipitation. We tested the validity of this prediction using muMT (IgH) mice, which lack an important domain of the Ig μ-chain, and therefore lack mature B cells (28). We used a well-established protocol of cytolytic T cell (CTL) induction by s.c. immunization of OVA-coated latex beads (2, 29). Splenocytes from the immunized mice 7 days after the immunization were stimulated with an OVA CD8 epitope peptide (SIINFEKL). Five days later, the resulting T cells were counted, and the target EL4 (H-2B) cells were labeled with 51Cr and pulsed with the same peptide. A 5-h CTL assay was performed at the E:T ratio indicated, and as described in Materials and Methods. The two lines in each panel represent two independent mice. The data are representative of three independent repeats. Most of the error bars (1 SD) were covered by the symbols.

that other factors associated with B cells, most likely Abs, played a role in the sensing of uric acid by the host.

**The presence of MSU-binding Abs**

We immunized B6 and BALB/c mice with preformed crystals to generate MSU crystal-binding Abs. The mice were immunized with 1 mg of MSU crystals biweekly for 2 mo in the absence of any additional adjuvant. The sera collected from the tail blood from the immunized mice strongly stained MSU crystals as determined by FACS analysis (Fig. 2A). Unimmunized mice also had MSU-binding Abs, albeit at titers somewhere between half to 1 log lower than the immunized mice (Fig. 2A). As a control, the MSU immune sera failed to stain xanthine crystals (Fig. 2B). We produced over 300 mAbs from the immunized mice by fusing the splenocytes with a tumor partner A3A. Approximately 30% of total clones showed various degrees of staining of the MSU crystals, with 9% showing substantial binding (over a log shift compared with the second Ab alone). Of the mAbs that showed binding, we tested their subtypes by analyzing the supernatants with an Ab subtyping kit, and determined that 85% of them are IgM κ (Fig. 2D), whereas IgGs and one IgM A made up the rest. This indicates that MSU-binding B cells do not frequently undergo Ab class switching. This is a common feature of T-independent B cell responses, as expected for a nonprotein Ag (MSU crystals) that would not have a cognate T cell response. The lack of adjuvant in our immunization protocol could also explain the absence of class switch.

Additional controls showed that serum from muMT mice had very low staining of MSU even after a similar immunization schedule, suggesting that the substances bound to MSU are Abs, in line with Fig. 2 that they were detectable by anti-Ig Abs (Fig. 3A). The low staining of muMT serum of MSU seems to suggest that other serum factors can also interact with the crystal surface, albeit...
to a much lesser extent than the Abs. Whether such a low binding had any functional consequence was not further studied in this report. Likewise, the binding of two control Abs (against mouse and human MHC class I molecules) showed minimal staining as well (Fig. 3B). These results and the finding that some similarly produced IgM did not stain the MSU crystals suggest that the binding is likely to be specific. The strong affinity and binding of these Abs suggest a possible involvement of Ig-mediated cell activation in the biological functions of MSU crystals, especially in light of the possibility of IgM immune complex formation.

UBAs facilitate uric acid crystallization

In the process of generating UBAs, we fortuitously discovered that the presence of these Abs altered MSU precipitation (detail not shown). To investigate the issue, we incubated supersaturated uric acid (1 mg/ml, pH 8.0) solution with a control protein and purified UBAs (UBA 11 or UBA E6) at different concentrations (Fig. 4 and data not shown). After 6 h (Fig. 4) or overnight (data not shown), the MSU crystals settled at the bottom of the tissue culture plate. After carefully washing the plate to remove residual soluble uric acid, the crystals were resolubilized with 0.01 N NaOH, and the amount of precipitation was quantified with UV 292 absorbance. Fig. 4 shows that the rate of crystal precipitation was in fact a function of the concentration of UBAs, and was less affected by a control protein (OVA). This result indicates that uric acid precipitation is significantly facilitated by UBAs. This observation seems to suggest that nonspecific interaction with other proteins might alter uric acid precipitation to a certain extent, as implicated in an earlier report (9).

One outstanding issue in the UBA-mediated MSU precipitation is the mode of contact between the crystal surface and the Ab. To study the binding mechanism between UBAs and the crystal surface, we digested UBAs with an IgM fragmentation kit and produced various Ig domains. Using the same amount of UBA (E6) as the starting point, the resulting F(ab’)_2 completely retained the binding, nearly indistinguishable from the original IgM (Fig. 5A). Fc5u (the Fab-deleted pentameric H chain) demonstrated no staining at all (Fig. 5A). Therefore, MSU binding is F(ab’)_2 dependent.

However, the binding itself is not equivalent to the actual crystal precipitation. We mixed a supersaturated uric acid solution (1 mg/ml) with 50 μg/ml E6 or its fragments, and incubated the solution for 6 h. Surprisingly, despite the unabated binding to MSU crystals, the F(ab’)_2 fragment completely failed to precipitate the soluble uric acid (Fig. 5B). Other control structures, including Fc5 and a control IgM, MOPC that does not bind MSU, had no effect at all. It is clear from this assay that Ab affinity is necessary, but not sufficient for the uric acid precipitation. The pentameric IgM configuration appears to be a crucial structure for the nucleation and growth of the MSU crystals.

As with other Abs, one concern was the ability of these Abs to block MSU crystal interaction with immune cells. We reported early on that MSU crystals are potent stimulators of DC activation (2). However, it was possible that MSU-mediated activation of DCs might be blocked in the presence of these Abs. We tested the effect of UBAs on DC activation by coincubating DCs with MSU and either UBAs or control Abs, as indicated. In the presence of 200 μg/ml MSU crystals, 7-day B6 bone marrow DCs induced by GM-CSF and IL-4 were activated to express surface CD86, a characteristic DC activation marker. None of the individual UBA or their combination altered MSU-mediated DC activation, similar to that of irrelevant control Abs (Fig. 5C). These data argue that the presence of these Abs does not interfere with the strong adjuvant activities of MSU crystals, at least with regard to DCs.
UBAs increase the inflammatory indicators in muMT mice. A. B6 mice were injected i.v. with soluble uric acid (1 mg per day) and indicated Abs (500 μg/day). UA, uric acid; E6C7, UBA; cAB, control IgM. Serum uric acid levels were analyzed by HPLC, as described in Materials and Methods. B. The lung tissue from test C57BL/6 mice was homogenized and analyzed, as described in Materials and Methods. The MPO values were generated by multiplying the UV reading by a factor of 0.2528 per a standard protocol. C. The serum C3 levels in test muMT mice after 3 days of uric acid/Ab infusion were determined using an ELISA kit (see Materials and Methods). D. Blood CD11b+ cells (neutrophils) were gated and analyzed for their CD62L expression.

UBAs restore the sensing of uric acid as an immune adjuvant

The notion that UBA mediates uric acid precipitation predicts that blood uric acid levels should drop following the injection of the Ab. We therefore injected soluble uric acid i.v., followed by an infusion of UBA (see Materials and Methods) (25). Three days later, we collected the sera from tail blood. The sera were passed through a 30 K cutoff filter to reduce the UV absorption interference. The pass-through was then injected into a weak anion exchange HPLC column, and the uric acid peak absorption at 292 nm was recorded as a percentage of total UV absorption of each run (a method to minimize run-to-run human injection error). Fig. 6A shows that in the presence of UBA, the recipient mice demonstrated reduced serum uric acid levels, whereas a control IgM failed to do so. This result was consistent with our hypothesis that UBAs may indeed cause a phase change of uric acid in vivo. As an outcome, the soluble uric acid available in the serum was reduced.

Our work presented above led to the proposal that the presence of UBA determines the precipitation of uric acid and the associated inflammation and adjuvanticity. We first studied whether the presence of UBAs alters the overall inflammatory state in vivo, because it is expected that the precipitation of MSU crystals will cause certain inflammatory reactions. We measured two common indicators, MPO in the lung and total serum complement C3. MPO is a product of neutrophils, and its level indicates the accumulation of these cells and tissue inflammation (27). The serum C3 is produced in response to inflammation as well and can be regarded as a general indicator of inflammatory state (31). Fig. 6, B (MPO measurement) and C (complement C3 concentration), shows that, following the injection of uric acid and UBA, both of the factors were significantly elevated after 3 days. The control IgM Abs did not have such an effect. One interesting observation was that in both cases, injection of UBA (E6C7) alone led to higher readings, although the C3 elevation with UBA alone was not statistically significant in comparison with the control (Fig. 6C). It is possible that UBAs can cause endogenous uric acid precipitation without any additional infusion. It is also possible that injection of UBA causes stress in the recipient, and thus increases the uric acid levels in the animal in response to this sudden infusion of great amounts of Abs. Because the increases in MPO measurements suggested the activation/migration of neutrophils into the tissues, we analyzed their CD62L (L-selectin) level by FACS. Neutrophil activation in inflammation is usually accompanied by the loss of this surface adhesion molecule as the neutrophils migrate into the tissue. UBA injection substantially reduced the expression of CD62L on blood neutrophils, whereas the control IgM appeared to have a smaller, but noticeable effect (Fig. 6D).

The most critical prediction for our study was that the availability of UBA restores the sensing of uric acid as an adjuvant in B cell-deficient mice. We therefore injected UBA and a control IgM i.v. into muMT mice and repeated the assay of Fig. 1. If the presence of UBAs enables uric acid precipitation, then the defect in sensing uric acid shown in Fig. 1 should be restored. Fig. 7 shows that the addition of UBA significantly elevated the CTL activity to the levels similar to that of the wild-type mice (Fig. 1). Interestingly, the control Ab also occasionally provided an enhancing effect, although to a lesser extent (data not shown). This may suggest that the presence of IgM itself is an important means of local immune response, which may come in addition to uric acid precipitation. Overall, the results were in line with our hypothesis that MSU-binding Abs are a critical factor in determining the uric acid-mediated adjuvant effect and inflammation. These UBAs most likely achieve such an effect via their ability to cause MSU crystallization.

Discussion

We report in this study that IgM Abs enhance the precipitation of uric acid from solution at relatively low concentrations. Gout has been a recognized disease for centuries (32), and the underlying cause was discovered many years ago (33). To date, many mathematical models have been proposed to delineate the rate of crystalization in relationship with environmental factors, such as the temperature, pH, salt, vibration, and even the material of the container in which the experiments were conducted (7–10). Yet a formula for predicting the rate of crystal formation remains elusive. The robust onset of gouty arthritis therefore also remains a peculiar event.

Our discovery that uric acid is a danger signal released by dying cells is another example of uric acid’s biological effect (2). We reported earlier that a crystalization event is required for this adjuvant effect. We have made substantial progress in understanding the molecular nature of UBAs at the biological level.
how MSU crystals, once formed, interact with DCs by a nonreceptor-dependent, chemical response with the cellular membrane (23). But no underlying mechanisms with regard to the actual phase transition have been revealed, particularly in vivo. It has long been known that uric acid is one of the principal antioxidative systems in mammals, and higher species progressively lose the ability to process this substance, in a likely attempt to gain benefits from its protective functions (34, 35). Primates have complete lost any enzymatic activities to process uric acid to allantoin due to a mutational silencing (36, 37). This progression permits the increasing likelihood of uric acid accumulation and possible crystallization. Our work suggests that UBAs are an important facilitating factor that results in the actual crystal precipitation.

In gout, it is likely that sustained high uric acid levels lead to macroscopic MSU precipitation. There is some clinical evidence to support our hypothesis. In humans, Ig-coated MSU crystals are found in gouty patients (16–18, 38). Electron microscopy revealed that the Fab portion of the Ab is attached to the crystal surface with Fc pointing away from the clusters (14). This is the correct configuration of immune complexes, which frequently triggers activation and inflammatory phagocytosis of accessory cells and APCs. A clear exception is that in humans, IgGs are found to bind MSU (16–18, 38), and the role of MSU-binding IgM has not been studied. A potential explanation for this observation in humans is that gout is a recurring condition with robust cytokine production. Certain IgM responses may be driven to switch as a result of repeated stimulation and using cytokines as a class switch trigger (39). Another important distinction is that to date we have not been able to directly visualize gout-like MSU crystals in mouse tissues, because the intrinsic residual uricase activities prevent the formation of macroscopic crystals. In studying the underlying mechanism of uric acid-mediated immune activation, a uricase knockout mouse model would work best; however, these mice are not viable due to acute kidney failure (40). To us, it has also been proven futile to visualize the macroscopic MSU crystal formation in situ. Before our production of UBAs, there had been no immunological reagents to directly detect MSU crystals in the tissue. The preparation steps for histological or other microscopic analysis, such as Gomori’s methenamine-silver method, are incompatible with the microcrystals because they dissolve or dismount quickly in washing steps (our own observation). This report further extends the call for a suitable animal model for this important immunological topic.

The situation may be different in the uric acid immune adjuvant effect. The influx of IgM at the site of tissue stress (41, 42) may precipitate as well anchor the microcrystals in situ of uric acid release. Because the formation of the crystals is at the source of tissue destruction by injury or infection, this would establish a situation in which DCs and other APCs may come into the close proximity of Ag (self or nonself) and adjuvant (MSU crystals), efficiently enabling immune induction. On another note, it has long been reported that IgM is attracted to the site of injury and is a potent mediator of subsequent tissue damage (42). Whether MSU is one source of the attraction remains to be determined.

It is important to note that the source of the MSU-binding Abs is unknown. We can probably exclude the involvement of B2 B cells due to the lack of cognate protein Ag presentation. B1 and marginal zone B cells differ in the target locations that they protect (43). B1 B cells are mostly located in body cavities, whereas marginal zone B cells tend to engage pathogens that penetrate the bloodstream. In humans, MSU crystals appear most often in joints. Furthermore, from the substantial amounts of UBAs present in the uninmune mice, it seems to suggest that B1 cells are the source of UBAs, because B1 Abs are produced without antigenic stimulation. Signaling-wise, we have recently gained a comprehensive understanding of how MSU crystals can trigger immune cell activation by directly engaging cell surface lipids, particularly cholesterol. This leads to plasma membrane lipid sorting and nonspecific aggregration of ITAM-containing receptors and downstream activation essentially similar to a phagocytic event (23). However, we have no evidence to rule out immune complex-mediated activation. It is known that IgM complexes trigger complement fixation, which is ~1000-fold more efficient than IgG complexes (44). Small peptides released during complement fixation, such as C3a and C5a, are anaphylatoxins (45). Additionally, it has been suggested that there might exist a mouse µ-chain FcR (46, 47), which would presumably trigger immune activation akin to the better defined FcR-mediated inflammatory responses (48, 49).

It should be prudently noted that without direct visualization of crystals in vivo, this report lacks an important piece of evidence to support our central hypothesis. The interpretation of our data, particularly the possibility of Ab-induced MSU crystal formation in vivo, remains speculative. With advent of new imaging technologies, we remain hopeful that MSU crystal formation in vivo will be directly monitored in due time. Such a methodological development will either substantiate or nullify the main hypothesis of this report, although our proposal that Abs are a substantial component in uric acid-mediated inflammation would still stand.

Our work has added a new angle in the study of urate precipitation and the subsequent inflammatory responses. In recent years, the interest in solid structure-mediated immune activation has grown significantly. However, one question has remained unresolved: where do the endogenous crystals come from? Our work is therefore a timely finding. One interesting fact to consider is that in the original report by Addadi and colleagues (19, 20), the authors tested several immunization crystals and found that each immune serum produced in rabbit led only to the precipitation of the original immunizing crystal. This type of specificity and the common presence of the crystal-binding tissue factors among tested mammals may indicate that MSU-specific Abs are not an oddity. Other crystal depositions in humans could involve Abs as well, for precipitation and subsequent inflammatory responses.

In summary, our work implicates a role of IgM Abs in MSU crystal formation and potentially subsequent adjuvanticity/inflammation, and provides evidence that it is a biologically relevant event in vivo. It may add a line of investigation on other crystal-related arthropathies and pathogeneses, in addition to addressing a lingering question in MSU crystal-related immunological research in general.

Acknowledgments
We thank Dr. Kenneth Rock for the facility support; Drs. LianJun Shen, Rachel Gerstein, Paul Kubes, and Raymond Welsh for reagents and discussion; Drs. Yang Yang and Pere Santamaria for critical review of the manuscript; Shelly Galusha and Melanie Desrosiers for technical assistance; and Ashley D. Mucsi and Winston Pang for editorial assistance.

Disclosures
The authors have no financial conflict of interest.

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