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*J Immunol* 2009; 182:6834-6843; doi: 10.4049/jimmunol.0803860
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Costimulation Modulation Uncouples Protection from Immunopathology in Memory T Cell Responses to Influenza Virus\textsuperscript{1}

John R. Teijaro,* Modesta N. Njau,* David Verhoeven,* Smita Chandran,* Steven G. Nadler,‡ Jeffrey Hasday,† and Donna L. Farber\textsuperscript{2,*}

The rapid effector functions and tissue heterogeneity of memory T cells facilitate protective immunity, but they can also promote immunopathology in antiviral immunity, autoimmunity, and transplantation. Modulation of memory T cells is a promising but not yet achieved strategy for inhibiting these deleterious effects. Using an influenza infection model, we demonstrate that memory CD4 T cell-driven secondary responses to influenza challenge result in improved viral clearance yet do not prevent the morbidity associated with viral infection, and they exacerbate cellular recruitment into the lung, compared with primary responses. Inhibiting CD28 costimulation with the approved immunomodulator CTLA4Ig suppressed primary responses in naive mice infected with influenza, but was remarkably curative for memory CD4 T cell-mediated secondary responses to influenza, with reduced immunopathology and enhanced recovery. We demonstrate that CTLA4Ig differentially affects lymphoid and nonlymphoid responses to influenza challenge, inhibiting proliferation and egress of lymphoid naive and memory T cells, while leaving lung-resident memory CD4 T cell responses intact. Our findings reveal the dual nature of memory T cell-mediated secondary responses and suggest costimulation modulation as a novel strategy to optimize antiviral immunity by limiting the memory T cell response to its protective capacities. The Journal of Immunology, 2009, 182: 6834 – 6843.

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Received for publication November 17, 2008. Accepted for publication March 23, 2009.

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\textsuperscript{1}This project was supported by National Institutes of Health Grants AI50632 and AI077029 and by a grant from Bristol-Myers Squibb awarded to D.L.F.

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www.jimmunol.org/cgi/doi/10.4049/jimmunol.0803860
context of extensive lung immunopathology and morbidity. Strikingly, protection and immunopathology of this memory CD4 T cell-driven secondary response can be uncoupled by inhibiting the CD28 pathway with CTLA4Ig. We show that in primary responses to influenza infection, CTLA4Ig suppresses the CD4 T cell response, resulting in reduced viral clearance and recovery. In contrast, CTLA4Ig treatment of mice with influenza-specific memory CD4 T cells resulted in improved clinical outcome and reduced morbidity to sublethal influenza infection, as well as increased survival to lethal influenza challenge. We demonstrate that CTLA4Ig treatment maintains enhanced and rapid lung viral clearance mediated by memory CD4 T cells, yet reduces lung immunopathology. In vivo, CTLA4Ig inhibits naive and memory CD4 T cell lymphoid responses and T cell recruitment to the lung, while not affecting in situ lung-specific memory T cell responses, accounting for differential effects on primary vs secondary responses. These results suggest a new strategy to optimize antiviral immunity to influenza and other ubiquitous pathogens where memory T cells readily develop and persist, and they further emphasize the importance of the host immune status in determining the outcome of immunotherapies.

Materials and Methods

Mice

BALB/c mice (8–16 wk of age) were obtained from the National Cancer Institute Biological Testing Branch, and congenic BALB/c (Thy1.1) mice were bred as homozygotes. Influenza hemagglutinin (HA)-TCR transgenic mice expressing a transgene-encoded TCR (clonotype 6.5) specific for HA peptide (110–119) and I-E\(^{b}\) (23) were bred as heterozygotes onto BALB/c (Thy1.2) or BALB/c (Thy1.1) hosts. RAG2\(^{-/-}\) mice on BALB/c genetic backgrounds were obtained from Taconic and maintained under specific pathogen-free conditions. Mice were maintained in the Animal Facility at the University of Maryland School of Medicine (Baltimore, MD), and animal protocols were approved by the Institutional Animal Care and Use Committee.

Reagents

The following purified Abs were purchased from Bio X Cell: anti-CD8 (TIB 105), anti-CD4 (GK1.5), anti-I-A\(^{d}\) (212.A1), and anti-Thy-1 (TIB 238). The 6.5 anti-clonotype Ab directed against the HA-TCR (23) was purified and conjugated to biotin (Pierce). Allophycocyanin- or PE-conjugated CD62L, PE-conjugated CD90.1 and CD90.2, FITC-conjugated CD90.1 and CD90.2, and PerCP-conjugated anti-CD4 were purchased from BD Pharmingen. PE-conjugated FoxP3 Ab was purchased from eBioscience. Murine CTLA4Ig was obtained from Bristol Myers-Squibb, and murine IgG2a isotype control was obtained from Bio X Cell. Influenza HA peptide (110–120, SFERFEIFPKF) was synthesized by the Biopolymer Laboratory at the University of Maryland School of Medicine.

Influenza virus infection

Influenza virus (A/PB/8/34) was generously provided by Dr. Walter Gerhard (Wistar Institute) and grown in the alantoic fluid of 10-day-old embryonated chicken eggs as described (24). Determination of influenza viral titers in viral stocks, lung homogenates, or bronchoalveolar lavage (BAL) fluid was accomplished by the tissue culture infectious dose 50% assay (TCID\(_{50}\)) as described (25), with titers expressed as the reciprocal of the dilution of lung extract that corresponds to 50% virus growth in Madin-Darby canine kidney (MDCK) cells, calculated by the Reed-Muench method.

For in vivo infection using sublethal doses of influenza, mice were anesthetized with isofluorane, and 20 \(\mu\)l of PR8 influenza virus containing 500 TCID\(_{50}\) was administered intranasally. For lethal influenza infection, mice were infected as above with 5000 TCID\(_{50}\) PR8 influenza virus (2LD\(_{50}\)), and weight loss and mortality were monitored daily. All infected mice were housed in the biocontainment suite, the University of Maryland at Baltimore animal facility, where tissue harvest from infected mice was also performed. Isolation of BAL fluid was obtained from anesthetized mice by flushing the alveolar space with PBS followed by withdrawal of lavage liquid. BAL fluid samples were centrifuged to pellet cells, and the supernatant was analyzed for viral content by the TCID\(_{50}\) assay described above.

Hemagglutination inhibition assay

The concentration of neutralizing anti-influenza Abs was measured in serum from 10-day-infected animals using the HA inhibition assay as described (26). Briefly, serum was heat inactivated for 30 min at 56°C, diluted 1/5 in PBS, and preabsorbed with 1% chicken RBC for 30 min. Serial 2-fold dilutions of serum were subsequently incubated in duplicate wells with 4 agglutinating units of virus for 1 h at room temperature, then 50 \(\mu\)l of a 1% chicken RBC solution was then added to each well and incubated for 45 min at room temperature. The HA inhibition titer was expressed as the reciprocal of the serum dilution where agglutination was inhibited in duplicate wells.

Generation of influenza-specific memory CD4 T cells

Generation of HA-specific memory CD4 T cells in congenic BALB/c (Thy1.1) hosts was accomplished as previously described (27, 28). Briefly, naive CD4 T cells were purified from spleens of HA-TCR mice and primed in vitro by culture with 5.0 \(\mu\)g/ml HA peptide and mitomycin C-treated, T-depleted BALB/c splenocytes as APCs in complete Click’s media (Irvine Scientific) for 3 days at 37°C. The resultant activated HA-specific effector cells were transferred into congenic BALB/c (Thy1.1) hosts (5 \(\times\) 10\(^6\) memory) to yield “HA-memory” mice with a stable population of HA-specific memory CD4 T cells (27–29). HA-specific memory CD4 T cells were also generated by transfer of 5 \(\times\) 10\(^6\) primed, HA-specific effector cells into RAG2\(^{-/-}\) recipient mice and harvested 2–3 mo posttransfusion as previously described (12, 27, 30, 31). HA-specific memory CD4 T cells isolated from these RAG2\(^{-/-}\) recipients were labeled with 5 \(\mu\)M CFSE (Invitrogen) and adoptively transferred into secondary BALB/c (Thy1.1) hosts, which were subsequently infected with influenza.

Polyclonal memory CD4 T cells specific for influenza were generated by infecting BALB/c mice intranasally with 500 TCID\(_{50}\) PR8 influenza. Total splenic CD4 T cells containing influenza virus-specific memory CD4 T cells were harvested 12–16 wk postinfection. The relative frequencies of influenza-specific IFN-\(\gamma\) and IL-2-secreting memory CD4 T cells in response to stimulation with HA peptide or whole influenza virus particles were determined using ELISpot as previously described (27, 31), and spots were enumerated using the ImmunoSpot ELISPOT reader (CTL; BD Biosciences).

Flow cytometry and intracellular cytokine staining

Cells were stained with fluorochrome-conjugated Abs as described (12), fixed, and acquired using an LSR II flow cytometer (BD Biosciences) with a minimum acquisition of 100,000 events and analyzed using FACSDiva software (BD Biosciences). Intracellular cytokine staining was performed as described previously (27). Briefly, lymphocytes from the spleen and lungs of influenza infected mice treated with CTLA4Ig or IgG2a were isolated 6 days postinfection, cultured in vitro for 4 h in the presence of PMA (25 ng/ml), ionomycin (1 \(\mu\)g/ml), and monensin (1 \(\mu\)l/ml) (GolgiStop; BD Pharmingen), surface stained, fixed in Cytoperm/Cytofix solution (BD Pharmingen), and stained intracellularly with IFN-\(\gamma\) or isotype control IgG1 Ab in Perm/Wash solution (BD Pharmingen). Stained cells were analyzed using an LSR II flow cytometer and FACSDiva software (BD Biosciences).

Histopathology of lung samples

For preparation and isolation of lung tissue for histological examination, mice were euthanized by isofluorane inhalation, trachea were exposed, and lungs were inflated with 4% paraformaldehyde at constant pressure. Lungs were then removed from the chest cavity, fixed in paraformaldehyde, embedded in paraffin wax, sectioned and stained with H&E by the Pathology Core Facility (University of Maryland at Baltimore), and analyzed by light microscopy.

In vivo BrdU labeling

Influenza virus-infected mice treated with CTLA4Ig or IgG2a were administered BrdU (1 mg, i.p.) for 3 consecutive days starting at day 3 postinfection. Spleen and lung lymphocytes were harvested at day 6 postinfection and resuspended in stain buffer. Cells were surface stained, fixed and permeabilized (Cytofix/Cytoperm, Perm/Wash; BD Biosciences), incubated with DNase (Sigma-Aldrich), and stained intracellularly with fluorescently labeled anti-BrdU Abs at 4°C. Cells were subsequently analyzed on the LSR II (BD Biosciences).
Statistics
Results are expressed as the mean value from individual groups ± SD indicated by error bars. Significance between experimental groups was determined by the two-tailed Student’s t test, assuming a normal distribution for all groups.

Results
Model for analyzing memory CD4 T cell-mediated secondary responses to influenza virus challenge

To analyze secondary responses to influenza virus infection directed exclusively by memory CD4 T cells, we established complementary models using TCR-transgenic and polyclonal influenza-specific T cells. In the TCR-transgenic model, naive TCR-transgenic CD4 T cells specific for influenza HA were obtained from HA-TCR transgenic mice (23), primed in vitro with HA peptide and APCs, and the resultant HA-specific effector cells were transferred into unmanipulated, congenic BALB/c hosts where they develop into long-lived, resting memory T cells (27, 28). The resultant “HA-memory” mice contain a stable population of HA-specific memory CD4 T cells, which comprise 0.5–5% of total endogenous CD4 T cells (Ref. 27 and data not shown) and exhibit the phenotype, function, and heterogeneous tissue distribution of in vivo primed polyclonal memory CD4 T cells, as we previously showed (12, 27, 29–31).

For generating polyclonal influenza-specific memory CD4 T cells, we infected BALB/c mice intranasally with a sublethal dose of PR8 influenza, isolated CD4 T cells 2–4 mo postinfection, and determined the frequency of influenza-specific memory CD4 T cells by ELISPOT (12). Equal numbers of CD4 T cells from previously primed mice were transferred into BALB/c hosts to generate “polyclonal flu-memory” recipients with a full complement of endogenous T cells. The total numbers of flu-specific memory CD4 T cells in these flu-memory hosts were back-calculated based on the ELISPOT results.

We assessed whether influenza-specific memory CD4 T cells could coordinate a protective immune response to influenza challenge, initially by comparing responses in BALB/c naive and HA-memory hosts infected with a sublethal dose of PR8 influenza (500 TCID50) with mock-infected mice as controls. We assessed the progression of disease by monitoring daily weight loss, and analyzed viral clearance by determining lung viral titers as in Fig. 1.

CTLA4Ig treatment improves the clinical outcome of memory CD4 T cell responses to influenza challenge while maintaining viral clearance

We compared the effects of inhibiting CD28 costimulation using CTLA4Ig, on the physiological outcomes of primary and memory CD4 T cell responses to influenza challenge. For costimulation modulation in vivo, we treated naive, HA-memory, or polyclonal-memory mice with murine IgG2a or CTLA4Ig at the 10 mg/kg clinical dose (12) before and following influenza challenge (Fig. 2A) and measured weight loss and viral titers as in Fig. 1A. In naive mice, both control- and CTLA4Ig-treated animals lost extensive weight following influenza challenge (Fig. 2B, left), with CTLA4Ig-treated naive infected mice having higher lung viral loads and mortality at 6 days postinfection compared with infected IgG2a control-treated naive mice (Fig. 2B and data not shown). This suppression of antiviral primary responses is consistent with a previous report (35) and the known CD28 requirement for naive T cell activation.

In contrast to the undesirable effects of CTLA4Ig on primary immune responses to influenza, CTLA4Ig treatment of mice with influenza-specific memory CD4 T cells improved the clinical outcome to influenza challenge. Whereas IgG2a-treated HA-memory mice exhibited progressive weight loss from 1 to 6 days postinfection comparable to infected naive mice, CTLA4Ig-treated HA-memory mice lost weight initially and then began to recover weight by day 4, with a steady weight gain until necropsy at day 6 postinfection.
FIGURE 2. CTLA4Ig optimizes secondary responses to influenza, while suppressing primary responses. A. Protocol for CTLA4Ig treatment of naive and HA-memory mice. Lower arrows denote time points for administration of CTLA4Ig or IgG2a, and upper arrows indicate time points for infection and mouse harvest. B. CTLA4Ig effects on the primary response to influenza in naive BALB/c mice treated and infected as in Fig. 2A. Left, Daily weight loss. Right, Lung viral titers 6 days postinfection. Virus challenge determined as in Fig. 1. Results are from four to five mice per group and are representative of three independent experiments. C, CTLA4Ig effects on the memory CD4 T cell-mediated secondary response in HA-memory mice treated and infected as in A. Left, Daily weight loss following infection in control IgG2a- or CTLA4Ig-treated HA-memory mice compared with naive or mock-infected mice (*, p = 0.016 for weights of CTLA4Ig- vs IgG2a-treated mice at day 6; n = 4 mice/group). Right, Viral titers from lung homogenates harvested 6 days postinfection as in Fig. 1. (p = 0.02 between naive and IgG2a- or CTLA4Ig-treated HA-memory mice; n = 4 mice/group). Results are representative of six independent experiments. D, Cumulative weight loss at day 6 postinfection of naive, HA-memory, or polyclonal-memory mice treated and infected as in A. Weight loss data are compiled from three independent experiments for naive mice (n = 9), three experiments using polyclonal memory mice (n = 8), and four experiments with HA-memory mice (n = 10). Each experiment contained three to five mice per experimental group. Value is p = 0.0001 when comparing HA-memory mice treated with IgG2a vs CTLA4Ig and p = 0.01 comparing recipients of polyclonal memory CD4 T cells treated with IgG2a and CTLA4Ig.

6 (Fig. 2C, left). Importantly, CTLA4Ig treatment did not appreciably affect the ability of HA-specific memory T cells to clear virus as seen by the comparable low viral titers in the lungs of IgG2a- and CTLA4Ig-treated HA-memory mice 6 days after influenza challenge (Fig. 2C, right). In polyclonal flu-memory mice, CTLA4Ig treatment also resulted in reduced weight-loss morbidity (Fig. 2D) and maintenance of lung viral clearance (data not shown). Comparing morbidity data from multiple experiments (Fig. 2D) reveals that CTLA4Ig treatment did not affect morbidity of naive mice infected with influenza, while it significantly reduced morbidity of HA- and polyclonal-memory mice, with CTLA4Ig-treated mice losing only 10–15% of their body weight compared with 25–30% weight loss of IgG2a-treated memory mice. HA- and polyclonal-memory mice treated with CTLA4Ig also exhibited fewer clinical signs of influenza-induced morbidity, including ruffled fur and hunched posture, compared with IgG2a-treated mice (data not shown). These results indicate that CTLA4Ig administration appears to optimize memory CD4 T cell-mediated antiviral responses by reducing morbidity while maintaining viral clearance, contrasting its suppressive effect on primary anti-influenza responses.

The reduced morbidity in response to influenza challenge observed in CTLA4Ig-treated HA-memory mice prompted us to ask whether CTLA4Ig treatment would provide protection from a lethal influenza virus challenge. We challenged CTLA4Ig or IgG2a-treated naive or HA-memory mice with a lethal dose (2LD50) of influenza virus and monitored morbidity and mortality daily. Mortality from this lethal dose began at days 7–8 postinfection, with all mice within IgG2a- and CTLA4Ig-treated naive groups succumbing to lethal challenge at 8–10 days postinfection (Fig. 3). The presence of memory CD4 T cells in HA-memory mice results in partial protection from lethal influenza infection, with 50% of IgG2a-treated mice succumbing to infection (Fig. 3). CTLA4Ig treatment of HA-memory mice resulted in improved survival from lethal challenge, with surviving mice experiencing less weight loss overall (Fig. 3 and data not shown). These results show that CTLA4Ig treatment can also improve protective immunity to lethal challenge in the presence of influenza-specific memory CD4 T cells.

Because CTLA4Ig inhibits primary T cell and Ab responses (7) and Abs are considered essential for complete viral clearance in naive mice (36), we asked whether the improved clinical outcome...
and viral clearance in CTLA4Ig-treated memory mice persisted at later times postinfection. We assessed influenza responses of differentially treated naive and memory mice up to day 10 postinfection, which corresponds to the peak Ab response and complete viral clearance in naive and HA-memory mice treated with CTLA4Ig or IgG2a as determined in Fig. 1. The designation “un.” (undetectable) indicates viral titers below the detection limit of the assay. D, Anti-influenza virus Ab titers in serum determined by HAI assay (see Materials and Methods) 10 days postinfection in naive and HA-memory mice. Titers are expressed as the reciprocal dilution equal to 1 HAI. Value is $p = 0.02$ when comparing IgG2a- and CTLA4Ig-treated naive mouse groups, $p = 0.006$ when comparing IgG2a- vs CTLA4Ig-treated HA-memory groups. Results are representative of two independent experiments with three to five mice per experimental group.

CTLA4Ig treatment of memory mice reduces lung immunopathology

The comparable viral clearance, yet disparate clinical outcomes in CTLA4Ig vs IgG2a-treated, flu-infected memory mice, prompted examination of lung pathology in these differently treated groups following influenza challenge. We examined H&E-stained sections from influenza-infected naive, IgG2a- or CTLA4Ig-treated HA-memory and polyclonal flu-memory mice. As compared with uninfected mice, lungs from infected naive mice contained mononuclear infiltrates within the interstitial tissue and near the large airways along with moderate airway damage characterized by hypertrophy in the alveolar epithelium. Additionally, these mice had moderate epithelial hypertrophy with dispersed consolidation surrounding the bronchial airways (Fig. 5A). In contrast, lungs from influenza-challenged control mice with either HA-specific or polyclonal flu-specific memory CD4 T cells had extensive diffuse mononuclear infiltrates around the airways and throughout the interstitium, leading to disruption of normal alveolar architecture and...
severe consolidation near most of the bronchial airways. In tandem, we observed acute damage to the airway epithelium as evidenced by desquamation throughout the alveoli and sloughing within the bronchial airways (Fig. 5A), connoting extensive lung immunopathology. Importantly, this lung immunopathology in flu-infected memory mice was dramatically reduced by CTLA4Ig treatment, as exemplified by reduced mononuclear cell infiltration and alveoli hypertrophy, and an increased number of alveoli with normal architecture in CTLA4Ig-treated compared with IgG2a-treated polyclonal- and HA-memory mice (Fig. 5A).

Consistent with the extensive infiltration in memory mice observed by histopathology, we also found increased numbers of endogenous CD4 T cells in the lungs of influenza-challenged HA-memory (Fig. 5B) and polyclonal flu-memory mice (Fig. 5C) compared with flu-infected naive mice. This enhanced accumulation of CD4 T cells in the lungs of memory mice was reduced by CTLA4Ig treatment in both HA and polyclonal memory groups (Fig. 5, C and D). We also investigated whether there were increased numbers of CD8 T cells in the lungs of flu-memory mice and whether CD8 T cell recruitment to the lungs was affected by CTLA4Ig. Interestingly, we found a decreased number of CD8 T cells in the lungs of flu-infected memory mice compared with naive mice (Fig. 5D), possibly due to reduced CD8 T cell priming due to early lung viral clearance in HA-memory mice (Fig. 1B). These results indicate that increased CD8 T cell recruitment to the lung does not occur in the presence of flu-specific memory CD4 T cells. Moreover, CTLA4Ig treatment did not significantly decrease or alter the number of CD8 T cells in the lungs of influenza-infected naive or memory hosts (Fig. 5D). These results show that CTLA4Ig has more profound inhibitory effects on the endogenous CD4 compared with the CD8 T cell compartment during influenza virus infection.

**CTLA4Ig reduces the accumulation and expansion of memory CD4 T cells in spleen and lung following influenza challenge**

To determine mechanisms for the improved antiviral response and clinical outcome mediated by memory CD4 T cells in the presence of CTLA4Ig, we used the HA-memory model to analyze the effects of costimulation inhibition on the responding memory CD4 T cell population. In control-treated HA-memory hosts, influenza infection resulted in extensive expansion and accumulation of HA-specific memory T cells in both the spleen and lungs, with HA-specific memory T cells comprising 25–50% of total lung CD4 T cells at 6 days postinfection (Fig. 6A, left). However, in flu-challenged CTLA4Ig-treated mice, there was a marked reduction in the frequency and absolute numbers of HA-specific memory T cells in the spleen and lungs (Fig. 6A). Comparing the absolute numbers of HA-specific memory cells in lung and spleen from IgG2a- and CTLA4Ig-treated infected mice (Fig. 6A, right) reveals that CTLA4Ig treatment inhibited the accumulation of memory T cells in the lung (5-fold inhibition) to a greater extent than in spleen (2-fold inhibition).

We asked whether the reduced numbers of memory T cells in the spleen of CTLA4Ig-treated mice resulted from reduced proliferation of memory T cells in vivo, by analysis of CFSE-labeled HA-specific memory CD4 T cells. Memory CD4 T cells isolated from RAG2−/− adoptive hosts were CFSE labeled and transferred to mice treated and infected as in Fig. 2A. We found extensive in vivo proliferation of HA-specific memory T cells in both IgG2a- and CTLA4Ig-treated groups; however, the proportion and absolute numbers of minimally divided (CFSE<sup>lo</sup>) CD4 T cells was higher in CTLA4Ig-compared with control-treated mice (Fig. 6B). These results show that CTLA4Ig reduces the proliferative expansion of splenic memory CD4 T cells in response to influenza infection.

The reduced accumulation of flu-specific memory CD4 T cells in the lung could be due to diminished T cell expansion and/or altered homing and recruitment to the lung. To address potential differences in homing capacity, we examined expression of the lymph node homing receptor molecule CD62L on memory CD4 T cells in influenza-specific memory CD4 T cells. CFSE-labeled HA-specific memory CD4 T cells were transferred (1 × 10<sup>6</sup>/mouse) into congenic BALB/c hosts, which were subsequently infected with 500 TCID<sub>50</sub> PR8 influenza virus. Left, CFSE dilution of HA-specific memory CD4 T cells 5 days postinfection, with the marker indicating percentage of undivided memory cells. Right, Absolute number of undivided HA-specific memory CD4 T cells expressed as an average of four mice per group. Value is p = 0.004 when comparing the absolute numbers of undivided Thy 1.2<sup>+</sup> memory CD4 T cells in IgG2a- and CTLA4Ig-treated mice.

**FIGURE 6.** CTLA4Ig treatment inhibits proliferation and expansion of influenza-specific memory CD4 T cells to viral challenge. A. Reduced frequency and absolute numbers of HA-specific memory CD4 T cells in flu-infected HA-memory mice treated with CTLA4Ig. Left, Flow cytometry plots show the frequency of CD4<sup>+</sup> Thy1.2<sup>+</sup> HA-specific memory CD4 T cells in the spleen and lung 6 days following influenza challenge of IgG2a- and CTLA4Ig-treated HA-memory mice, with the percentage of HA-specific memory CD4 T cells from total CD4 T cells indicated in each plot. The absolute number of HA-specific memory CD4 T cells in spleen and lung tissue was calculated from microscopic cell count by trypan blue exclusion of dead cells. Values are p = 0.03 and 0.002 when comparing the absolute numbers of Thy1.2<sup>+</sup> memory CD4 T cells in the spleen and lung tissues, respectively, from mice treated with IgG2a and CTLA4Ig. Results are representative of six independent experiments with four to five mice per group. B, CTLA4Ig treatment reduces in vivo proliferation of HA-specific memory CD4 T cells. CFSE-labeled HA-specific memory CD4 T cells were transferred (1 × 10<sup>6</sup>/mouse) into congenic BALB/c hosts, which were subsequently infected with 500 TCID<sub>50</sub> PR8 influenza virus. Left, CFSE dilution of HA-specific memory CD4 T cells 5 days postinfection, with the marker indicating percentage of undivided memory cells. Right, Absolute number of undivided HA-specific memory CD4 T cells expressed as an average of four mice per group. Value is p = 0.004 when comparing the absolute numbers of undivided Thy 1.2<sup>+</sup> memory CD4 T cells in IgG2a- and CTLA4Ig-treated mice.
CTLA4Ig treatment alters homing receptor expression of activated HA-specific memory CD4 T cells. A. Increased CD62L expression on HA-specific memory CD4 T cells in CTLA4Ig- vs IgG2a-treated mice following influenza challenge. Upper, CD62L expression by CD4<sup>+</sup> Thy1.2<sup>+</sup> spleen and lung HA-specific memory CD4 T cells isolated from IgG2a- and CTLA4Ig-treated, flu-challenged HA-memory mice, with percentage CD62L<sup>high</sup> indicated in each histogram. Lower, The frequency of CD62L<sup>high</sup> memory CD4<sup>+</sup> T cells in spleen and lung tissues compiled from five independent experiments (n = 22). Value is p = 2 x 10<sup>-5</sup> for the frequency of CD62L expression HA-specific memory T cells between IgG2a- and CTLA4Ig-treated mice. B. Maintenance of CD62L<sup>high</sup> expression on proliferating memory CD4<sup>+</sup> T cells in the presence of CTLA4Ig. CFSE-labeled memory CD4<sup>+</sup> T cells were transferred into congenic hosts and analyzed after infection and treatment as in Fig. 6B. Plots show CD62L expression vs CFSE dilution on gated CD4<sup>+</sup> Thy 1.2<sup>+</sup> memory T cells 5 days postinfection of IgG2a- and CTLA4Ig-treated mice. (Fig. 7A), indicating that CTLA4Ig did not affect the CD62L profile of lung memory CD4<sup>T</sup> cells and rather had biased effects on CD62L expression by spleen memory CD4<sup>T</sup> cells.

The predominant CD62L<sup>high</sup> phenotype of splenic memory CD4<sup>T</sup> cells in CTLA4Ig-treated flu-infected mice could result from impaired memory CD4<sup>T</sup> cell activation or from reduced CD62L<sup>low</sup> in control-treated mice and were equally divided between CD62L<sup>high</sup> and CD62L<sup>low</sup> phenotypes in CTLA4Ig-treated mice (Fig. 7B). These results indicate that CTLA4Ig partially inhibits CD62L<sup>low</sup> down-regulation on memory CD4<sup>T</sup> cells responding to influenza virus, suggesting that the capacity of lymphoid memory CD4<sup>T</sup> cells to home to nonlymphoid sites, such as the lung, is curtailed.

CTLA4Ig treatment has biased effects on lymphoid memory CD4<sup>T</sup> cells

To evaluate the cellular mechanism for the differential effects of CTLA4Ig treatment on primary and secondary immune responses to influenza infection, we analyzed in vivo responses of naive and memory CD4<sup>T</sup> cells in both lymphoid and nonlymphoid tissues by BrDU incorporation. We administered BrDU to naive or HA-memory mice infected with influenza, harvested spleen and lung tissue 6 days postinfection, and measured the extent of BrDU incorporation in each tissue from the differentially treated groups. In naive mice infected with influenza, BrDU incorporation of endogenous CD4<sup>T</sup> cells in both the spleen and lung of control-treated mice was substantially inhibited by CTLA4Ig treatment (Fig. 7A), with mock-infected controls having minimal BrDU incorporation in both tissues (0.5–1% and 1–3% in spleen and lung, respectively). In flu-infected HA-memory mice, both spleen and lung-resident memory CD4<sup>T</sup> cells in control-treated mice exhibited extensive BrDU incorporation following influenza infection (Fig. 8B, left) that exceeded BrDU incorporation in the primary CD4<sup>T</sup> cell response (Fig. 8A). In the presence of CTLA4Ig, BrDU incorporation by spleen-memory CD4<sup>T</sup> cells was markedly reduced (5-fold reduction), whereas BrDU incorporation by lung-memory CD4<sup>T</sup> cells was not affected (Fig. 8B, top left and bottom left). BrDU incorporation of endogenous CD4<sup>T</sup> and CD8<sup>T</sup> cells in...
spleen and lung of infected memory mice was inhibited by CTLA4Ig treatment, similar to that seen in naive mice (data not shown). These results strongly suggest that CTLA4Ig preferentially inhibits spleen or lymphoid-derived naive and memory CD4 T cells, while leaving intact in situ lung memory CD4 T cells; however, we cannot rule out that BrdU incorporation of lung memory CD4 T cells in the lung may have migrated from lymphoid sites.

A hallmark of memory CD4 T cell recall is their rapid effector function. We therefore measured the capacity of HA-memory CD4 T cells recovered from the spleen and lung of CTLA4Ig- or IgG2a-treated mice to produce IFN-γ 6 days after influenza virus challenge. We observed a biased reduction in early IFN-γ production from spleen memory CD4 T cells (2-fold) of CTLA4Ig-treated mice; however, we cannot rule out that BrdU incorporation of lung memory CD4 T cells in the lung may have migrated from lymphoid sites.

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Discussion

We demonstrate herein that memory CD4 T cells mediate secondary responses to influenza infection characterized by efficient viral clearance in the context of extensive immunopathology and morbidity. Strikingly, the physiological outcome of a memory CD4 T cell-mediated secondary response to influenza can be significantly improved by targeting the CD28 pathway with the costimulation modulator CTLA4Ig. While CTLA4Ig is suppressive for primary immune responses to influenza, leading to increased viral loads, reduced lung function, and increased morbidity, CTLA4Ig treatment of naive mice infected with influenza suppresses the initiation of T cell and Ab responses in lymphoid tissues, impairing the antiviral response. In contrast, memory CD4 T cells are present in both lymphoid and lung tissue, and they require CD28 costimulation mainly for Ag-driven IL-2 production and proliferation (12). While CTLA4Ig inhibited lymphoid memory CD4 T cell expansion, it did not affect in situ lung memory CD4 T cell expansion and effector cytokine production, and therefore viral clearance was maintained. Our results further reveal a specific role for CD28 costimulation in homing to nonlymphoid sites during a viral infection, and they are consistent with earlier findings that CD28 controls T cell migration to peripheral sites in the absence of infection (40). These effects of CTLA4Ig treatment on T cell homing may be a mechanism for the clinical efficacy of abatacept in reducing immunopathology in rheumatoid arthritis, known to be perpetuated by memory CD4 T cells (14).

In addition to its differential effects on lymphoid and nonlymphoid responses, CTLA4Ig treatment had disparate effects on cytokine production by memory CD4 T cells. We show herein that CTLA4Ig preferentially inhibits IL-2 production from lung memory...
CD4 T cells, while leaving intact IFN-γ production. We propose that the ability of CTLA4Ig to differentially inhibit IL-2 vs IFN-γ responses may be directly related to the uncoupling of immunomodulation and protection in secondary influenza responses. IFN-γ production has been shown to be crucial for protection in secondary responses to influenza and other viral infections (41, 42), although it can be dispensable for clearance of influenza virus during primary responses (43, 44). The ability of lung memory CD4 T cells to rapidly produce IFN-γ in the presence of CTLA4Ig despite a suppressed Ab and endogenous CD4 and CD8 T cell responses suggests that IFN-γ production in situ may mediate rapid viral clearance by memory CD4 T cells, a possibility we are presently investigating. Conversely, IL-2 production by memory CD4 T cells, which is important for their expansion (12), can contribute to increased infiltration into lung tissue and the resultant immunopathology. Thus, highly expansive memory T cells may be detrimental when site-specific immunity is required in respiratory virus infections. We propose that for protective immunity to influenza, the quality and location of memory T cells is more important than their absolute frequency, also a key issue for vaccine design (45).

We demonstrate that targeting CD28 costimulation can optimize influenza-specific antiviral secondary responses, suggesting a new clinical strategy for ameliorating influenza morbidity. Morbidity and mortality from influenza infection have been attributed to pathological immune responses characterized by excessive cytokine secretion and inflammatory infiltration into the lung (21, 46); however, a cellular mechanism for influenza-induced immunopathology has not been identified. We show herein that memory CD4 T cells can exacerbate inflammation and infiltration in the lung in secondary responses to influenza, similar to findings of memory CD4 T cell-mediated immunopathology in other viral systems, including respiratory syncytial virus (47, 48), dengue virus (49), and hepatitis (50). Additionally, previous studies have identified a role for CD8 T cells in lung immunopathology during primary influenza infection (51, 52). As memory CD8 T cells have also been shown to require CD28 costimulation for optimal proliferation in vivo (10, 11), CTLA4Ig treatment may also show efficacy in preventing CD8 T cell-mediated immunopathology. Thus far, strategies for reducing immunopathology through inhibition of inflammatory cytokines (53) or global T cell immunosuppression (54) have been ineffective or have blocked protective immune responses, impairing viral clearance. Here, we show that CTLA4Ig may provide the appropriate type of immunosuppression to differentially curtail pathological immune reactions while maintaining site-specific antiviral responses mediated by memory T cells.

Memory T cell responses to influenza are clinically relevant given their presence in healthy individuals (17, 18), as well as recent identification of memory CD4 T cells that cross-react with avian influenza (H5N1) epitopes in the peripheral blood of healthy humans exposed to seasonal influenza variants (19, 20). These findings emphasize the clinical importance of understanding memory T cell responses to influenza and other viruses, and the clinical applicability of immunotherapies that enhance a memory T cell response. We propose that an illness resulting from influenza infection in an immune-experienced individual may mask the underlying memory T cell-mediated viral clearance, and that immunomodulation may be an effective way to manifest the protective features of T cell memory.

Our findings strongly suggest that considering both the mode of immunomodulation together with the host immune status are critical parameters for evaluating the efficacy of immunotherapies. Previous studies in transplantation have found that the presence of memory T cells interferes with or prevents the effectiveness of tolerance induction strategies or immunosuppression (55, 56), indicating that memory T cells may represent a barrier to effective treatment. We demonstrate herein that immunomodulation of a memory response can result in a positive clinical outcome to a respiratory virus infection. These studies, together with our results, suggest that considering memory T cells when designing and testing immunotherapies is important for evaluating their efficacy and potential utility in antiviral immunity, autoimmunity, and transplantation.

Acknowledgments

The authors extend their gratitude to Wendy Lai for mouse colony maintenance, Dr. Mark Cowan for help in analyzing the histology slides, and Daniel Perez and Haichen Song for help with growing influenza virus.

Disclosures

D.L.F. received a research grant from Bristol-Myers Squibb in partial support of this work. S.G.N. is an employee of Bristol-Myers Squibb and owns stock in the company.

References


23. Kirberg, J., A. Baron, S. Jakob, A. Rolink, K. Karjalainen, and H. von Boehmer. 1994. Thymic selection of CD8+ single positive cells with a class II major his-


neutralizing antibodies of immunoglobulin G (IgG) but not of IgM or IgA iso-

munol. 35: 3173–3186.


nol. 6: 414–419.


42. Selin, L. K., S. M. Varga, I. C. Wong, and R. M. Welsh. 1998. Protective het-
erologous antiviral immunity and enhanced immunopathogenesis mediated by memory T cell populations. J. Exp. Med. 188: 1705–1715.


istration of anti-IFN-γ antibody to β2-microglobulin-deficient mice delays influ-
enza virus clearance but does not switch the response to a T helper cell 2 phe-

