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The Tyrosine Kinase Pyk2 Mediates Lipopolysaccharide-Induced IL-8 Expression in Human Endothelial Cells

Appakkudal R. Anand,* Magali Cucchiarini,† Ernest F. Terwilliger,† and Ramesh K. Ganju2*

Secretion of proinflammatory cytokines by LPS activated endothelial cells contributes substantially to the pathogenesis of sepsis. However, the mechanism involved in this process is not well understood. In the present study, we determined the role of a nonreceptor proline-rich tyrosine kinase, Pyk2, in LPS-induced IL-8 (CXCL8) production in endothelial cells. First, we observed a marked activation of Pyk2 in response to LPS. Furthermore, inhibition of Pyk2 activity in these cells by transduction with the catalytically inactive Pyk2 mutant, transfection with Pyk2-specific small interfering RNA, or treatment with Tyrphostin A9 significantly blocked LPS-induced IL-8 production. The supernatants of LPS-stimulated cells exhibiting attenuated Pyk2 activity blocked transendothelial neutrophil migration in comparison to the supernatants of LPS-treated controls, thus confirming the inhibition of functional IL-8 production. Investigations into the molecular mechanism of this pathway revealed that LPS activates Pyk2 leading to IL-8 production through the TLR4. In addition, we identified the p38 MAPK pathway to be a critical step downstream of Pyk2 during LPS-induced IL-8 production. Taken together, these results demonstrate a novel role for Pyk2 in LPS-induced IL-8 production in endothelial cells. The Journal of Immunology, 2008, 180: 5636–5644.

The endothelium lines the inner surface of blood vessels and functions as an interactive barrier between blood and tissue. Exposed to blood flow, it is the primary target for inflammatory agents during local and systemic inflammation (1, 2). Endothelial cells express chemokines that initiate the activation and recruitment of circulating leukocytes at sites of tissue inflammation (3, 4). The bacterial endotoxin, LPS, an essential component of the surface of Gram-negative bacteria (5), has potent proinflammatory properties by acting on many cell types including endothelial cells (3, 6). High levels of LPS are a major cause of Gram-negative septic shock, in which LPS induces numerous changes, including up-regulation of adhesion molecules as well as procoagulant activity, enhanced endothelial permeability, and secretion of proinflammatory mediators by the endothelium (3, 4). Among these activities, secretion of IL-8 (CXCL8) by LPS-activated endothelial cells contributes substantially to inflammatory responses (8). IL-8 displays a chemotactic activity for neutrophils, which are the first line of immune cells to be recruited to infected areas (9, 10). IL-8 also activates neutrophils to generate several toxic products, such as arachidonic acid metabolites. Thus, during septicemia, IL-8 participates in a series of cellular events that severely damage the endothelium and surrounding tissues. However, the precise mechanism of LPS-induced signaling that leads to IL-8 secretion in endothelial cells is poorly understood. Elucidation of this pathway may be of immense significance in the design of anti-inflammatory therapies that act by regulating chemokine responses leading to septic shock-related events.

The proline-rich kinase 2, Pyk2, also known as RAFTK or CAKg, is a cytoplasmic tyrosine kinase related to focal adhesion kinase (FAK) (11). However, unlike FAK, Pyk2 exhibits a more restricted tissue expression pattern primarily in epithelial cells, neuronal cells, fibroblasts, hematopoietic cells, and endothelial cells (11–13). Pyk2 has been shown to be activated in response to a broad range of stimuli, including extracellular signals that elevate intracellular Ca2+ concentration, agonists of G protein-coupled receptors, and engagement of Ag receptors on T cells, B cells, and mast cells (12, 14–16). Pyk2 is also activated in response to inflammatory cytokines, stress signals, and integrin-mediated cell adhesion (12, 17, 18). It is fast emerging as a critical “platform” kinase that couples several receptors (including integrin and chemokine receptors) with a variety of downstream effectors, thus regulating various functions such as cell adhesion, migration, proliferation, and survival (11). Although the expression of Pyk2 has been identified in endothelial cells, very few reports have focused on its role in these cells. In view of recent studies that link the nonreceptor tyrosine kinase Pyk2 with inflammation (19, 20), our present study evaluated its role in mediating IL-8 secretion in LPS-stimulated endothelial cells.

Materials and Methods

Reagents, cells, and culture conditions

LPS and cyclohexamide were obtained from Sigma-Aldrich. The Pyk2 inhibitor (Tyrphostin A9), p38 MAPK inhibitor (SB203580), ERK kinase inhibitor (PD080593), Pam.Cys (TLR2 agonist), and polymyxin B were obtained from Calbiochem. Py99, Pyk2, phospho-Pyk2, and phospho-FAK Abs were obtained from BioSource International, whereas TLR2, TLR4, IL-8, phospho-ERK, ERK, phospho-p38, and p38 Abs were obtained from Santa Cruz Biotechnology. Isotype controls were purchased from BD Transduction Laboratories. HUVEC were purchased from Clonetics. Cells were grown at 37°C in 5% CO2 in endothelial growth medium EGM2-MV

Keywords: LPS, Pyk2, IL-8, TLR4, HUVEC

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containing 2% FBS, 12 μg/ml bovine brain extract, 10 ng/ml human recombinant epidermal growth factor, 1 μg/ml hydrocortisone, and GA-1000 (1 μg/ml gentamicin and amphotericin B), according to the recommendations of the supplier. The human monocytic THP-1 cell line obtained from the American Type Culture Collection was cultured in RPMI 1640 medium supplemented with 10% FBS at 37°C in 5% CO2 atmosphere.

**Stimulation**

In all experiments, HUVEC were grown to 80% confluency in 6-well assay plates. The cells were stimulated with LPS in the presence of 0.5% FBS. In the case of inhibitor treatments (Tyrophostin A9, SB203580, PD98059), HUVEC were pretreated with inhibitor for 1 h after which they were stimulated with LPS for various time periods. The supernatant was used for the Western blotting and immunoprecipitation analyses. THP-1 cells were differentiated into macrophage-like cells by treating with 100 ng/ml cycloheximide. Supernatants were collected, centrifuged, and processed for IL-8 quantification by commercially available ELISA kits (Endogen), per the manufacturer’s instructions.

**Recombinant adeno-associated virus (AAV) transduction**

High-efficiency gene delivery of the dominant negative Pyk2 mutant, Pyk2K457A (Pyk2MT), or a control gene, β-galactosidase (β-Gal), was accomplished using a recombinant AAV-based method. The AAV vectors were prepared as previously described (21). Briefly, the kinase-inactive Pyk2 mutant or the same vector encoding β-Gal cDNA (as a control for nonspecific effects of viral infection) were inserted into a CMV-immediate early promoter and an SV40 fragment (providing an intron as well as a polyadenylation function) in one of our lab’s standard AAV vector plasmids, pACP. Each gene cassette was framed between AAV2 inverted terminal repeats. The vectors were then packaged in AAV2 virions using a three-plasmid transient cotransfection system consisting of the vector plasmids pXX2 and pXX6. Cell lysates were collected 2 days after transfection, and the virions were isolated. The vector preps were then dialyzed before use. Genomic titers of the preparations, as determined by real-time PCR were \(-10^{11}\) copies/ml. Before being exposed to the virus, the HUVEC were cultured overnight in complete medium. HUVEC were transduced by application of the AAV in a minimal amount of serum-free medium for 90 min at 37°C in a cell culture incubator. Equal volumes of complete endothelial basal medium (EBM) containing 4% serum were added to the cells to achieve a final serum concentration of 2%. The cells were then cultured overnight in complete medium. HUVEC were transduced by application of the AAV in a minimal amount of serum-free medium for 90 min at 37°C in a cell culture incubator. Equal volumes of complete endothelial basal medium (EBM) containing 4% serum were added to the cells to achieve a final serum concentration of 2%. The cells were then cultured overnight in complete medium. HUVEC were transduced by application of the AAV in a minimal amount of serum-free medium for 90 min at 37°C in a cell culture incubator. Equal volumes of complete endothelial basal medium (EBM) containing 4% serum were added to the cells to achieve a final serum concentration of 2%. The cells were then cultured overnight in complete medium.

**Cycloheximide treatment**

HUVEC were either unstimulated or stimulated for 24 h with LPS (100 ng/ml) in the presence or absence of 10 μg/ml cycloheximide. Supernatants were harvested and the IL-8 production measured by ELISA.

**IL-8 ELISA**

After stimulation, the culture supernatants were collected, centrifuged, and processed for IL-8 quantification by commercially available ELISA kits (Endogen), per the manufacturer’s instructions.

**FIGURE 1.** LPS induces tyrosine phosphorylation and enzymatic activity of Pyk2 in endothelial cells. HUVEC were stimulated with LPS (100 ng/ml) for the indicated periods of time (A) or with various concentrations of LPS for 15 min (B). The lysates were then immunoprecipitated with Abs to Pyk2. The immunoprecipitates were analyzed by Western blotting with Abs to phosphotyrosine (Py99). The same blot was then probed with anti-Pyk2 Ab. C and D, Lysates obtained from HUVEC stimulated with LPS (100 ng/ml) for various periods of time were also analyzed by Western blotting with Abs to phospho-FAK (p-FAK) (C) or with Abs to phospho-Pyk2 (Tyr 402) and phospho-Pyk2 (Tyr 580) (D). E, The kinase activity in the immunoprecipitates was assessed as the ability of Pyk2 to phosphorylate the synthetic substrate poly(Glu-Tyr)4:1. A typical autoradiograph is shown (117 kDa). (top), NRS, Normal rabbit serum; PC, positive control. The total abundance of Pyk2 protein was analyzed by Western blot analysis of the lysates, as indicated (bottom). Data show one representative experiment of three independent experiments performed.
Isolation of neutrophils

Human neutrophils were purified from normal donors by dextran sedimentation and Ficoll gradient centrifugation followed by hypotonic lysis of erythrocytes. The purity of the prepared neutrophils was 95% (as judged by the morphology of stained cytocentrifuged preparations) and the viability was 98% (as judged by the trypan blue dye exclusion method).

TEM assay of neutrophils

Briefly, 100,000 HUVEC were added to fibronectin-coated 24-well Transwell culture chambers with a pore size of 3 μm (Costar; Corning) and grown for 3 days in 5% CO₂ at 37°C. A total of 0.6 ml of medium from the untreated or LPS/Tyrphostin A9 or AAV/Pyk2MT-treated HUVEC was added to the lower compartment. In the upper compartment, 1 x 10⁶ neutrophils in 0.1 ml of the EBM containing 0.5% FBS were added onto the HUVEC monolayer. Supernatants pretreated with 1000 U/ml polymyxin B and 20 μg/ml IL-8 Ab served as controls. The chambers were incubated for 4 h at 37°C in 5% CO₂. The cells in the lower compartment were counted on a hemocytometer. The results are presented as mean ± SD of three separate experiments and are expressed as the increase in the number of cells migrating toward the lower compartment.

Western blotting and immunoprecipitation

Total cellular extracts from the LPS-treated cells were prepared by lysing the cells in radioimmunoprecipitation assay buffer (50 mM Tris-HCl (pH 7.4), 1% Nonidet P-40, 0.25% sodium deoxycholate, 150 mM NaCl, 1 mM PMSF, 10 μg/ml aprotinin, leupeptin, and pepstatin, 10 mM sodium vanadate, 10 mM sodium fluoride, and 10 mM sodium pyrophosphate). Proteins (50 μg) were size-fractionated by 8% SDS-PAGE and transferred onto nitrocellulose membranes. The membranes were blocked for 2–3 h in 5% nonfat milk and then incubated with the respective primary and secondary Abs for 2–3 h each. The membranes were washed three to four times for 15 min each with TBS and 0.05% Tween 20, and later developed by chemiluminescence (ECL System; GE Healthcare).

For the immunoprecipitation, equal amounts of protein from the stimulated time points were clarified by incubation with protein A-Sepharose CL-4B or GammaBind Sepharose beads (Amersham Biosciences) for 1 h

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**FIGURE 2.** Inhibition of Pyk2 activation blocks LPS-induced IL-8 production in endothelial cells. A, HUVEC were transduced with recombinant AAV-expressing Pyk2 mutant (Pyk2MT) or AAV β-Gal vector control (VC). Overexpression of the mutant was demonstrated by Western blot analysis with anti-Pyk2 Abs 48 h after transduction (top left). Anti-actin Ab was used as an internal control (bottom left). Transduced cells were tested for their ability to produce IL-8 upon LPS stimulation (100 ng/ml) using a commercial ELISA kit (Endogen) (right). *, p < 0.05 vs the AAV vector control (AAV-VC). Data represent the mean ± SD of three independent experiments. B, HUVEC were pretreated with vehicle or Tyrphostin A9 (5 μM) for 1 h at 37°C. The same blots were probed with anti-Pyk2 Abs. HUVEC preincubated with vehicle or various concentrations of Tyrphostin A9 for 1 h were cultured without or with LPS (100 ng/ml) and the concentration of IL-8 in the culture supernatants was determined 24 h after stimulation (right). *, p < 0.05 compared with the vehicle control. Data represent the mean ± SD of three independent experiments. C, HUVEC were transfected with Pyk2-specific siRNA and nontargeting siRNA (NT siRNA) using Lipofectamine in the absence or presence of LPS (100 ng/ml). The knockdown of Pyk2 expression was analyzed by Western blotting with anti-Pyk2 Abs (left). Anti-actin Ab was used as an internal control. The supernatants from the siRNA transfected cells were analyzed for IL-8 expression after stimulation with LPS (100 ng/ml) (right). *, p < 0.05 compared with the nontargeting siRNA control. D, HUVEC were treated with LPS (100 ng/ml) in the absence or presence of 10 μg/ml cyclohexamide (CX). After 24 h, the supernatants were collected and analyzed for IL-8 expression. *, p < 0.05 compared with the LPS control. Data represent the mean ± SD of three independent experiments.
At 4°C. The Sepharose beads were removed by brief centrifugation, and the supernatants were incubated with different primary Abs for 2 h at 4°C. Immunoprecipitation of the antigen–antibody complexes was performed by incubation at 4°C overnight with 50 μl of protein A-Sepharose or Gammabind Sepharose (50% suspension). Nonspecific interacting proteins were removed by washing the beads thrice with radioimmunoprecipitation RIPA buffer and once with PBS. Immunoprecipitated complexes were solubilized in 50 μl of 2X Laemmli buffer, and further analyzed by Western blotting, as described.

Immune complex kinase assay
HUVEC were lysed in modified RIPA buffer and equal amounts of cell lysates were immunoprecipitated with anti-Pyk2 Ab. The immunoprecipitates were incubated for 12 min at 23°C with 50 μl of protein A-Sepharose or Gammabind Sepharose (50% suspensions). Nonspecific interacting proteins were removed by washing the beads thrice with radioimmunoprecipitation RIPA buffer and once with PBS. Immunoprecipitated complexes were solubilized in 50 μl of 2X Laemmli buffer, and further analyzed by Western blotting, as described.

Cell surface staining and flow cytometry analysis
To detect TLR2 and TLR4 expression, HUVEC and THP-1 cells were stained with the monoclonal primary Abs, respectively, followed by staining with FITC- or PE-conjugated secondary Abs. Flow cytometry was conducted using a FACScalibur cytometer and these data were analyzed by CellQuest software (BD Biosciences).

Small interfering RNA (siRNA)-mediated knockdown of Pyk2
RNA interference-mediated knockdown of Pyk2 was performed using SMARTpool Pyk2 duplex RNA oligonucleotides obtained from Dharmacon. A nontargeting siRNA (Qiagen) was used as the control. HUVEC were transfected with the siRNA using Lipofectamine 2000 reagent (Invitrogen Life Technologies), according to the manufacturer’s instructions. Pyk2 siRNA-mediated knockdown was estimated by detection of Pyk2 expression by Western blot analysis, 48 h after the initial transfection.

Statistical analysis
Reported data are mean ± SD of at least three independent experiments performed in duplicate or triplicate. The statistical significance was determined by the Student’s t test.

Results

LPS induces the tyrosine phosphorylation of Pyk2, but not FAK, in endothelial cells
To investigate the signaling processes involved in LPS-induced cytokine production, we examined the ability of LPS to activate Pyk2. Pyk2 is a nonreceptor tyrosine kinase that is expressed abundantly in endothelial cells (22). Stimulation of endothelial cells with LPS (100 ng/ml) was accompanied by increased tyrosine phosphorylation of Pyk2, as demonstrated by immunoprecipitation experiments. We observed that LPS induces both a concentration and time-dependent tyrosine phosphorylation of Pyk2 (Fig. 1, A and B). The increase in Pyk2 phosphorylation was observed with as low as 10 ng/ml LPS and was most evident at 100 ng/ml LPS, 5–30 min after stimulation. Similar amounts of Pyk2 were immunoprecipitated in the samples, as demonstrated by blotting with anti-Pyk2 Ab (Fig. 1, A and B, bottom). In contrast, LPS did not induce an increase in FAK phosphorylation (Fig. 1C). We, therefore, focused the rest of our studies on Pyk2.

Pyk2 has a structure similar to FAK as it contains an autophosphorylation site (Tyr881), sites involved in kinase activation (Tyr405), and a site homologous to the Grb2-binding site (Tyr579/580), in FAK (11). Endothelial cells treated with LPS were analyzed for the phosphorylation and activation of Pyk2 using Abs specific for these sites of tyrosine phosphorylation. We observed that LPS enhanced the phosphorylation of residues 402 and 580 (Fig. 1D), indicating an important function for these residues in LPS-induced Pyk2 activation. However, LPS did not markedly induce phosphorylation of residue 881 (data not shown).

LPS induces Pyk2 tyrosine kinase activity
To assess whether increased Pyk2 tyrosine phosphorylation leads to the induction of Pyk2 enzyme activity, Pyk2 immunoprecipitates from control or LPS-stimulated HUVEC were assayed for...
Pyk2 activity based on its ability to phosphorylate a synthetic substrate poly(Glu-Tyr)₄:₁ in an in vitro kinase assay (Fig. 1E). As a positive control, Pyk2 activity was stimulated with 100 ng/ml vascular endothelial growth factor for 2 min. LPS significantly increased the activation of Pyk2 and its tyrosine kinase activity peaked at 15 min.

Inhibition of Pyk2 activation attenuates LPS-induced IL-8 production

Because Pyk2 is rapidly being recognized as a tyrosine kinase of central importance in diverse cell types, we hypothesized that Pyk2 could be involved in the LPS signaling that mediates acute inflammatory responses in endothelium. Thus, we tested the possible role of Pyk2 in LPS-induced cytokine production. Tyrphostin A9 (AG17) is the most selective of tyrosine kinase inhibitors that block the Pyk2 signaling pathway (23), thus providing an effective tool for investigating the role of this tyrosine kinase in cellular signaling. Our initial studies testing the possible involvement of Pyk2 in LPS-induced cytokine expression using Ab arrays demonstrated that certain cytokines that are known to be up-regulated on stimulation with LPS, such as GROα, IL-1ra, IL-6, IL-8, IP-10, and migration inhibitory factor, were inhibited by the specific Pyk2 inhibitor, Tyrphostin A9 (data not shown). Because IL-8 is the prototypic CXC chemokine (9) and it contributes most significantly to the inflammatory process in endothelial cells (7), we further investigated the molecular mechanisms of Pyk2 in LPS-induced IL-8 production.

We used three approaches to study the role of Pyk2 in regulating IL-8 production: 1) expression of a dominant negative form of Pyk2 that removes a key lysine involved in ATP binding, 2) use of a specific chemical inhibitor, and 3) knockdown of Pyk2 protein expression using siRNA.

As a first approach, we constructed AAV vectors expressing a kinase-inactive Pyk2 mutant (Pyk2K457A) and then HUVEC were transduced with this dominant negative Pyk2. In this mutant, mutation of Lys⁴⁵⁷ to alanine in the tyrosine kinase domain of Pyk2 creates an inactive Pyk2 that inhibits wild-type Pyk2 activity. We observed that endothelial cells transduced with the catalytically inactive Pyk2 mutant (AAV-Pyk2MT) exhibited significantly attenuated LPS-induced IL-8 production as compared with the
vector control (β-Gal) transduced cells (Fig. 2A, right). The inhibitory effect was present over a range of LPS stimulations (10–100 ng/ml) (data not shown) with maximal inhibition occurring at 100 ng/ml LPS. It should be noted that a basal production of IL-8 was always present in the supernatant. Overexpression of Pyk2MT in HUVEC was confirmed by Western blot analysis of the cell lysates (Fig. 2A, left). This result provides direct evidence for the role of Pyk2 in LPS-induced IL-8 production.

We also demonstrated that Tyrphostin A9, a specific inhibitor of Pyk2, inhibited LPS-induced Pyk2 phosphorylation in a time-dependent manner (Fig. 2B, left). Furthermore, the inhibitor blocked LPS-induced IL-8 expression in the endothelial cells in a concentration-dependent manner (Fig. 2B, right).

As a third approach, we decreased Pyk2 expression in HUVEC using siRNA Pyk2. Our optimization studies indicated that the expression of Pyk2 protein was specifically decreased by transfection with the siRNA Pyk2 duplex (100 nM), but not by transfection with a nontargeting siRNA (Fig. 2C, left). At this siRNA concentration (100 nM), there was minimal loss of cell viability (<10%) (data not shown). There was no difference in the Pyk2 knock down in the absence or presence of LPS (100 ng/ml). The Pyk2 siRNA-treated cells showed reduced IL-8 expression compared with cells treated with nontargeting siRNA control (Fig. 2C, right). Taken together, these data suggest that Pyk2-dependent activation is an important part of the LPS-IL-8 signaling pathway.

Cycloheximide treatment at 10 μg/ml completely blocked LPS-induced IL-8 production, confirming that the up-regulation of IL-8 expression was due to de novo synthesis (Fig. 2D). To exclude a possible toxic effect of cycloheximide, cells that had been cultured in its presence were washed and restimulated with LPS in the absence of cycloheximide, and IL-8 production was measured after 24 h. HUVEC were still able to respond to LPS (data not shown).

Inhibition of Pyk2 activation reduces the LPS-induced TEM of neutrophils

IL-8 has been shown to be a potent mediator of the extravasation and accumulation of neutrophils at the sites of injury by their adhesion to and migration through the endothelial lining (24). Hence, the TEM of neutrophils toward the supernatants of LPS-stimulated endothelial cells transduced with AAV-Pyk2MT or pretreated with Tyrphostin A9 was assessed to determine whether the IL-8 produced was functionally active. As shown, transduction of LPS-stimulated endothelial cells with Pyk2MT and pretreatment with Tyrphostin A9 significantly inhibited neutrophil migration across a HUVEC monolayer compared with the supernatants of cells treated with LPS only (Fig. 3A). To confirm that the carryover of Tyrphostin A9 did not influence IL-8-mediated neutrophil migration, rIL-8 (100 ng/ml) was added to the supernatants with or without Tyrphostin A9. There was no difference in IL-8-mediated transmigration of neutrophils toward these supernatants with or without Tyrphostin A9 (data not shown). In addition, we also observed that supernatants from the Pyk2-specific siRNA-treated cells showed attenuated neutrophil transmigration, in comparison

FIGURE 5. Pyk2 regulates LPS-induced IL-8 expression through the p38 MAPK pathway. A, HUVEC was pretreated with vehicle or Tyrphostin A9 (5 μM) for 1 h at 37°C. The cells were then stimulated with LPS for various periods of time in EBM with 0.5% FBS. The cells were lysed and analyzed by Western blotting with anti-phospho-ERK1/2 (p-ERK1/2) Abs (upper left blots) or anti-phospho-p38 (p-38) Abs (lower left blots). The same blots were probed with anti-ERK1/2 or anti-p38 Abs, respectively. Data show one representative experiment of three independent experiments. For quantitative analysis of protein phosphorylation, the ratio of phosphorylation vs total protein in each lane was obtained by densitometry. The phosphorylation index of ERK and p38 was determined by calculating the value of this ratio in each lane and presenting the ratio as the fold increase over the control value (unstimulated sample; 0), which was designated as 1 (right). B, HUVEC pretreated with vehicle, the p38 MAPK inhibitor (SB203580, SB) (left) or the ERK kinase inhibitor (PD98059, PD) (right) for 1 h were stimulated with 100 ng/ml LPS and then the concentration of IL-8 in the supernatants was measured 24 h after stimulation. *, p < 0.05 compared with the vehicle control. Data represent mean ± SD of three independent experiments.
to the supernatants of cells treated with nontargeting siRNA (Fig. 3B). The TEM of neutrophils toward any residual LPS present in the supernatant was eliminated by pretreatment with polymyxin B. Polymyxin B was used at a concentration of 1000 U because almost complete inhibition of LPS-induced IL-8 expression was seen at this concentration (Fig. 3C).

To specifically test the role of IL-8 in migration, 20 μg/ml human IL-8-specific goat Ab was added to the supernatants for 1 h before the assay was initiated. The results demonstrated that anti-IL-8 Ab blocked the neutrophil migration across the HUVEC monolayer, which was induced by the LPS-treated supernatants (Fig. 3D). These results confirmed that the secreted IL-8 in the supernatants was functionally active and hence indicated a physiological function of Pyk2 activity in regulating IL-8 production.

**LPS-induced Pyk2 activation leading to IL-8 expression is mediated by TLR4**

Previous studies have indicated that TLR4, and to a certain extent TLR2, are the main receptors for LPS in endothelial cells (25–27). To investigate the involvement of TLR2 and TLR4 in LPS-responsive HUVEC, we first analyzed the expression pattern of TRIs in HUVEC by FACS analysis. TLR4 was detected at moderate levels on the surface of HUVEC (Fig. 4A, top left). However, TLR2 expression was minimal (Fig. 4A, bottom left). THP-1, a monocytic cell line was used as a positive control for expression of TLR2 and TLR4 (Fig. 4A, right).

To further determine the functional role of TLR4 and TLR2 in LPS-induced Pyk2 activation and IL-8 expression, the ability of anti-TLR4- and anti-TLR2-neutralizing Abs to block these LPS-induced effects was examined. At concentrations of 10 μg/ml, anti-TLR4 Ab significantly inhibited both the Pyk2 activation (Fig. 4B) and IL-8 production (Fig. 4C) induced by LPS. However, anti-TLR2 Ab did not have any effect. The functional ability of the TLR2 and TLR4 blocking Ab was confirmed by inhibition of PamCys-induced IL-8 production and LPS-induced IL-8 production, respectively, in vitamin D3-differentiated THP-1 cells (Fig. 4D). In THP-1 cells, PamCys, an analog of bacterial lipoprotein has been shown to induce IL-8 production via TLR2 (28), whereas LPS has been shown to induce IL-8 production through TLR4 (29). Taken together, these findings suggest that LPS requires cell surface TLR2 to induce Pyk2 activation leading to IL-8 production in endothelial cells.

Pyk2 regulates IL-8 production through the p38 MAPK pathway

Pyk2 has been shown to act as an essential intermediate providing a link between extracellular stimuli and MAPK signaling pathways in various cell lines (30). Of the signaling pathways activated by LPS in endothelial cells, the p44/42 MAPK and p38 MAPK pathways have been shown to be directly involved in the production of cytokines (31–34). Hence, we determined whether inhibition of Pyk2 with Tyrophostin A9 blocked MAPK activation. Treatment of HUVEC with LPS resulted in the phosphorylation of both ERK and p38 MAPK, as reported previously (31–34). However, pre-treatment with Tyrophostin A9 significantly inhibited the LPS-induced p38 phosphorylation (Fig. 5A, left), suggesting that p38 MAPK may be an important downstream molecule of Pyk2 in LPS-induced IL-8 production. The phosphorylation of p38 MAPK was shown in Fig. 5A (right). The involvement of the p38 MAPK pathway was confirmed by the dose-dependent blocking of LPS-induced IL-8 with a specific inhibitor of p38 MAPK, SB203580 (Fig. 5B, left). Though the use of a specific ERK kinase inhibitor, PD98059, reduced the LPS-induced expression of IL-8 slightly, the difference was not statistically significant (Fig. 5B, right).

**Discussion**

Pyk2 is rapidly becoming recognized as a tyrosine kinase of central importance in diverse cell types such as neuronal cells, hematopoietic cells, liver epithelial cells, and vascular smooth muscle cells (11, 12, 14, 35). However, little is known about Pyk2 regulation in vascular endothelial cells. In this study, we present data indicating that activation of Pyk2 plays a key role in modulating LPS-induced IL-8 production in endothelial cells. We have demonstrated Pyk2 activation by an increase in both tyrosine phosphorylation and enzyme activity of Pyk2. Though previous studies on endothelial cells have shown that Pyk2 is tyrosine-phosphorylated in response to stimulation with G protein-coupled receptor agonists, vascular endothelial growth factor, and the cytokine IL-1α (13, 36), this study is the first to our knowledge to demonstrate the activation of Pyk2 in response to LPS in endothelial cells.

The proline-rich kinase-2 Pyk2 is a cytoplasmic tyrosine kinase showing considerable sequence homology and structural similarity to FAK, including consensus motifs in the catalytic domain. Despite the high sequence homology, structural similarities, and common downstream effector molecules between FAK and Pyk2, recent studies provide evidence of their different cellular localization and regulatory mechanisms (37, 38). In the present study, LPS did not induce the tyrosine phosphorylation of FAK, and therefore the activities of Pyk2 and FAK are likely to be regulated by different upstream molecules in LPS-stimulated endothelial cells.

Several lines of recent evidence suggest that Pyk2 tyrosine phosphorylation could be closely linked to inflammatory processes in multiple cell types (12, 13, 20). In endothelial cells, the LPS-induced production of inflammatory cytokines has been reported to contribute significantly to the inflammatory process (7). Preliminary cytokine array studies in our laboratory indicated IL-8 to be among the most prominent chemokines attenuated in LPS-stimulated endothelial cells treated with Tyrophostin A9 (data not shown). Hence, we further investigated the molecular mechanism by which Pyk2 regulates LPS-induced IL-8 production using AAV-expressing a Pyk2 kinase-inactive mutant, Tyrophostin A9, and Pyk2-specific siRNA. Using all three approaches, we were able to demonstrate decreased LPS-induced IL-8 production, in comparison to the controls. Overexpression of a catalytically inactive form of Pyk2, Pyk2-K457A in pulmonary endothelial cells has been demonstrated to result in defects in cell adhesion, spreading and migration (39, 40). Although Pyk2-specific siRNA blocked IL-8 expression compared with the nontargeted siRNA control, the basal levels of IL-8 in the controls treated only with siRNA were marginally higher than the levels in untreated control. The higher basal levels of IL-8 produced on treatment with only siRNA are in agreement with a recent study that shows that siRNA may nonspecifically trigger the production of IL-8 (41). The physiological relevance of these experiments was confirmed by the inhibition of human neutrophil transmigration toward the supernatants of the LPS-treated HUVEC that were transduced with AAV-Pyk2MT, pretreated with Tyrophostin A9 or transfected with Pyk2-specific siRNA, in comparison to the LPS-treated controls, suggesting that Pyk2 modulates neutrophil infiltration across the endothelial cells via regulation of IL-8 production.

Increasing evidence suggests that central to the recognition of LPS expression in endothelial cells is a family of transmembrane proteins known as TLRs that have leucine-rich repeats in their
extracellular domains (42, 43). Most effector cells of the innate immune system, such as monocytes and endothelial cells express TLR2 and TLR4 (26). Our studies on the expression of these two receptors in HUVEC confirmed a previous report (26) that indicated that HUVEC predominantly express TLR4 and weakly express TLR2. Using anti-TLR4 blocking Abs, we were able to demonstrate the inhibition of both LPS-induced Pyk2 activation and IL-8 production. From this demonstration, it seems reasonable to suggest that TLR4 is an important part of the Pyk2 signaling pathway, acting to transmit LPS-dependent signals that lead to IL-8 production.

Pyk2 has been shown to function as an essential intermediate providing a link between extracellular stimuli and signaling pathways involving MAPKs (30, 44). The central role of p38 MAPK-dependent signaling for LPS-induced endothelial cell activation has been highlighted in several recent studies (32, 33, 45). We were able to demonstrate that p38 MAPK is an important part of the signaling cascade downstream of Pyk2 in LPS-IL-8 pathway in endothelial cells. Based on our data, we therefore propose a model that suggests that, as a result of the interaction of LPS with TLR4 on the cell surface, a signaling route is initiated via Pyk2 and the p38 MAPK pathway leading to the production of IL-8 (Fig. 6), possibly through NF-κB activation. Activation of NF-κB, downstream to p38 MAPK, is known to be an essential prerequisite for LPS-induced IL-8 expression in endothelial cells (33, 46).

In summary, we show that LPS induces Pyk2 activation in endothelial cells. The specific inhibition of Pyk2 activity was paralleled by a strong reduction in both LPS-induced IL-8 production and neutrophil chemotaxis, pinpointing a key role of Pyk2 in the LPS-induced production of the inflammatory cytokine. Because LPS-induced IL-8 production is known to be involved in the pathogenesis of sepsis and several inflammatory diseases, these results indicate that Pyk2 may represent a novel target for the development of innovative therapeutic strategies against inflammatory conditions.

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Disclosures

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