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CD40-CD40 Ligand Interaction between Dendritic Cells and CD8^{+} T Cells Is Needed to Stimulate Maximal T Cell Responses in the Absence of CD4^{+} T Cell Help^{1}

Maria Genevive H. Hernandez,*† Lianjun Shen,* and Kenneth L. Rock^{2*}

Stimulation of CD40 on APCs through CD40L expressed on helper CD4^{+} T cells activates and “licenses” the APCs to prime CD8^{+} T cell responses. Although other stimuli, such as TLR agonists, can also activate APCs, it is unclear to what extent they can replace the signals provided by CD40-CD40L interactions. In this study, we used an adoptive transfer system to re-examine the role of CD40 in the priming of naive CD8^{+} T cells. We find an ~50% reduction in expansion and cytokine production in TCR-transgenic T cells in the absence of CD40 on all APCs, and on dendritic cells in particular. Moreover, CD40-deficient and CD40L-deficient mice fail to develop endogenous CTL responses after immunization. Surprisingly, the role for CD40 and CD40L are observed even in the absence of CD4^{+} T cells; in this situation, the CD8^{+} T cell itself provides CD40L. Furthermore, we show that although TLR stimulation improves T cell responses, it cannot fully substitute for CD40. Altogether, these results reveal a direct and unique role for CD40L on CD8^{+} T cells interacting with CD40 on APCs that affects the magnitude and quality of CD8^{+} T cell responses. The Journal of Immunology, 2007, 178: 2844 –2852.

 naïve T cells require contact with appropriately activated APCs in order to be primed (1–3). CD40-CD40L (CD154) interactions mediate one of the most effective APC-activating signals. CD40 is a TNFR family member that is constitutively expressed on all APCs and is up-regulated upon infection or inflammation (4–6). It binds to CD40L, a member of the TNF family, which is expressed mainly on activated CD4^{+} T cells (4–6). Stimulation of CD40 on dendritic cells (DCs), through either activated CD4^{+} T cells, soluble CD40L, or activating anti-CD40 Ab, up-regulates expression of costimulatory molecules CD80 and CD86, enhances production of cytokines (most notably IL-12), and promotes cross-priming to exogenous Ags (2, 7, 8). In vivo, CD40 stimulation increases the magnitude of CD4^{+} and CD8^{+} T cell expansion, leading to enhanced tumor protection and conversion of steady-state tolerance into immunity (9–15). Conversely, CD40-signaling blockade, mainly through anti-CD40L Ab, inhibits T cell activation and results in tolerance, e.g., to transplants, and control of some autoimmune diseases (16, 17).

The pivotal role of CD40-CD40L interactions in the generation of productive immune responses is highlighted by the phenotype of CD40-deficient and CD40L-deficient mice, which exhibit defects in both humoral and cellular immunity (4, 18–21). The reduced CD8^{+} T cell responses are thought to be largely due to impaired “licensing” of APCs. This process is considered to be the mechanism through which CD4^{+} T cells provide help for the generation of primary CD8^{+} T cell responses, especially to noninflammatory Ags (22). There are two models for how CD4^{+} T cell help occurs. One model involves a sequential two-cell interaction, first, between CD40L-expressing CD4^{+} T cells and CD40-expressing DCs leading to DC activation, and then between the activated DCs and CD8^{+} T cells (8, 23, 24). An alternative model involves a direct interaction between CD40L-expressing CD4^{+} T cells and CD40-expressing CD8^{+} T cells (25). In either case, CD4^{+} T cell help and CD40 activity are often considered to be identical because CD40 stimulation can restore CD8^{+} T cell responses primed in the absence of CD4^{+} T cell help (8, 23, 24).

Nonetheless, primary CD8^{+} T cell responses against some pathogens including lymphocytic choriomeningitis virus (LCMV), vesicular stomatitis virus, and Listeria monocytogenes are impaired in the absence of CD4^{+} T cells or CD40 (15, 26–28). These pathogens are thought to bypass the need for CD4^{+} T cell help in part because of their ability to directly activate DCs. Recent studies show that DCs and other APCs express TLRs that are able to bind microbial components such as LPS, CpG, dsRNA, and some viral proteins, e.g., respiratory syncytial virus fusion protein (29–31). Ligation of TLRs with these pathogen-associated molecular patterns induces similar effects as that of CD40 stimulation, e.g., activation of NF-kB, up-regulation of costimulatory molecules, production of cytokines, and promotion of cross-priming (29, 32, 33). It has even been shown recently that TLR agonists can abrogate tolerance induced by CD40L blockade (34). However, it is still not clear whether CD40-stimulated or TLR-stimulated DCs have identical CD8^{+} T cell priming capability in vivo.

Given the different pathways by which DCs can be activated, we re-examined the requirement for CD40-CD40L interaction in vivo. We asked what is its natural role in the priming of naive CD8^{+} T cell responses in the absence of presence of microbial pathogen-associated molecular patterns. Using an adoptive transfer system of TCR-transgenic (Tg) CD8^{+} T cells into wild-type (WT) or CD40^{−/−} hosts, we find that CD40 signaling on DCs, as well as other APCs, has an important function in inducing maximal T cell

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3. Abbreviations used in this paper: DC, dendritic cell; LCMV, lymphocytic choriomeningitis virus; Tg, transgenic; WT, wild type; BMDC, bone marrow-derived DC.

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proliferation and effector function. Surprisingly, this effect is observed even in the absence of CD4+ T cells. In this situation, CD40L expression by the responding CD8+ T cells contributes to the maximum response. Finally, we find that TLR stimulation cannot fully compensate for CD40 activity. Therefore, CD40 provides a unique and nonredundant signal for APC activation that impacts the ensuing naive CD8+ T cell responses.

Materials and Methods

Mice and cell lines

C57BL/6J, B6.129P2-Cd40+/−/Kb/J (CD40−/−), and B6.129S2– Cd40−/+Kb/J (CD40+/−) mice were purchased from The Jackson Laboratory and used from 5 to 10 wk of age. P-14 TCR-Tg and OT-I TCR-Tg breeders were originally obtained from Dr. R. Welsh (University of Massachusetts Medical School, Worcester, MA) and Dr. S. Jameson (University of Minnesota, Minneapolis, MN), respectively. These were bred with C57BL6/1g-thy1aspKpil+ mice (The Jackson Laboratory) to yield Thy1.1+ T cells. Additionally, P-14 mice were bred with B6.129P2-Tp−prcPep3/Boy mice to yield CD45.1+ T cells. Lastly, P-14/Thy1.1+ mice were bred with CD40−/− mice to yield CD40−/−Thy1.1+ P-14 T cells. All mice were bred and housed in specific pathogen-free conditions at the University of Massachusetts Medical School animal facility.

The fibroblast cell line L cell (DAP) transfected with full-length OVA and TIR-OVA fusion constructs has been described previously (35). B16 tumor cells expressing Flt3 ligand (B16-Flt3L) (36) were obtained from Dr. U. von Andrian (Center for Blood Research, Harvard Medical School, Boston, MA).

Dendritic cells

Bone marrow-derived DCs (BMDC) were generated by flushing cells from femurs and tibias of mice and culturing them in complete medium containing 10 ng/ml GM-CSF and 5 ng/ml IL-4 (Corixa). Fresh cytokines were added on day 4, and on day 7 both adherent and nonadherent cells were harvested. Splenic DCs were collected from mice that were injected s.c. with B16-Flt3L tumor cells 10–14 days previously. At the time of harvest, the spleen cellularity had increased up to 7-fold and contained 25–50% CD11c+ cells (clone N418; BD Pharmingen) by flow cytometry, with all DC subsets having expanded. DCs were pulsed with the minimal MHC class I epitope from LCMV glycoprotein (KAVYNFATC, gp33 peptide) or chicken OVA (SIINFEKL; OVA peptide) at a concentration of 1 μg per 5 × 10^6 cells for 2–4 h at 37°C. When WT and CD40-deficient DCs were pulsed with SIINFEKL under these conditions, they had the same levels of T cell responses.

In vitro restimulation of spleen cells. In some of the experiments, CD4+ T cell-depleted hosts were used.

In vivo CTL assay

The in vivo CTL assay was performed as described previously (37). Briefly, splenocytes targets were pulsed with relevant or irrelevant peptide and labeled with different concentrations of CFSE. The targets were then mixed at a 1:1 ratio and injected i.v. into immunized and unimmunized control mice. After 2–20 h, spleen or blood was collected and the percentage of target cell killing was calculated using the formula: 100 − (t% relevant peptide-pulsed in immunized/95% irrelevant peptide-pulsed in immunized) (% relevant peptide-peptide pulse in control/95% irrelevant peptide-pulse in control) × 100).

Statistical analysis

Data were analyzed for statistical significance with a two-tailed Student’s t test using Microsoft Excel software. Differences in T cell responses were considered significant when a probability value of p < 0.05 was obtained.

Results

Reduced CD8+ T cell expansion and effector function in CD40−/− hosts

Most of the previous studies showing a role for CD40 in APC activation and CD8+ T cell responses in vivo relied on exogenous stimulation of the receptor with agonistic Ab (9–12, 38). Two major limitations of these studies are the nonphysiologic nature of Ab-mediated stimulation and the possibility of nonspecific effects because of the numerous cell types that can express CD40. We therefore took the opposite approach and examined the priming of CD8+ T cells upon adoptive transfer into CD40-deficient animals. In this system, all APCs lack CD40 while the responding T cells. In this system, all APCs lack CD40 while the responding

Assessment of endogenous CD8+ T cell responses

In one set of experiments, WT, CD40−/−, and CD40−/− mice were immunized s.c. on one flank with 4–5 × 10^6 Tir-OVA cells. One week later, the mice were given peptide-pulsed target cells for an in vivo CTL assay. In a second set of experiments, WT mice were immunized i.v. with 1 × 10^6 WT or CD40−/− DCs pulsed with OVA peptide. On day 7, some mice received targets for an in vivo CTL assay while some mice were sacrificed for in vitro restimulation of spleen cells. In some of the experiments, CD4+ T cell-depleted hosts were used.

In vitro CTL assay

The in vivo CTL assay was performed as described previously (37). Briefly, splenocytes targets were pulsed with relevant or irrelevant peptide and labeled with different concentrations of CFSE. The targets were then mixed at a 1:1 ratio and injected i.v. into immunized and unimmunized control mice. After 2–20 h, spleen or blood was collected and the percentage of target cell killing was calculated using the formula: 100 − (t% relevant peptide-pulsed in immunized/95% irrelevant peptide-pulsed in immunized) (% relevant peptide-peptide pulse in control/95% irrelevant peptide-pulse in control) × 100).

In vitro restimulation

Splenocytes from DC-immunized mice were depleted of RBC and plated at 5 × 10^6 cells/well in a 24-well plate and stimulated with 1 μg of OVA peptide. After 4 to 6 days, OVA-specific CD8+ T cell responses were evaluated by intracellular IFN-γ staining.

Intracellular cytokine staining

Spleen and lymph node cells were incubated with the indicated peptide in the presence of brefeldin A (Golgi Plug; BD Pharmingen) and rIL-2 for 2 h at 37°C. The samples were stained with anti-CD8 and anti-Thy1.1 or anti-CD45.1 Ab, fixed, and permeabilized using Cytofix/Cytoperm buffer (BD Pharmingen), and stained with anti-cytokine Ab diluted in Perm/Wash buffer (BD Pharmingen) according to manufacturer’s instructions. Anti-IFN-γ (XMG1.2), anti-TNF-α (MP6-XT22), and anti-IL-2 (JES6-5H4) were purchased from BD Pharmingen or eBioscience. The samples were washed twice with Perm/Wash buffer and analyzed by flow cytometry.

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We next examined whether the Tg T cells became functional effectors by assaying for cytokine secretion. The number of P-14 T cells making IFN-γ in the spleen and lymph nodes of CD40−/− hosts is ~10-fold less than in the WT hosts (Fig. 1C). The same difference was observed for TNF-α and IL-2 production (data not shown).

We also tested the response of adoptively transferred OT-I TCR-Tg T cells, which are specific for the OVA peptide SIINFEKL bound to H-2Kb, to make sure that the observed effects were not confined to the P-14 TCR-Tg T cells. Instead of peptide, we immunized the hosts i.v. with a stable OVA-transfected cell line that also gets cross-presented by host DCs (35). On day 4 of the response, almost all of the OT-I T cells in both the WT and CD40−/− hosts had divided more than eight times (data not shown). However, the OT-I T cells accumulated to a lesser extent in the spleens and lymph nodes of CD40−/− compared with the WT hosts (Fig. 2A). The proportion, as well as the absolute number of IFN-γ-secreting cells, was also reduced by as much as 50% in the CD40−/− hosts (Fig. 2B). Furthermore, the killing of SIINFEKL-pulsed target cells was reduced in the CD40−/− hosts compared with the WT hosts (94% vs 56%; Fig. 2C). We also obtained similar results after s.c. immunization of the OVA transfectants (data not shown).

Altogether, these results demonstrate that although CD40 on APCs is not absolutely required to initiate naive CD8+ T cell priming, it is important in inducing T cells to undergo maximum expansion and differentiation into effectors.

**CD40−/− DC induce suboptimal T cell responses**

The preceding experiments examined T cell responses in which all APCs in the host are either expressing or not expressing CD40. Because DCs are considered to be the most potent APC (1), we analyzed T cell responses stimulated by CD40-deficient DCs. To do this, we immunized WT mice containing adoptively transferred P-14 T cells with WT or CD40−/− BMDCs pulsed with LCMV gp33 peptide. In this situation, the only cell that lacks CD40 is the DCs. The preceding experiments also showed that the lack of CD40 activity did not simply compromise presentation but rather interfered with the activation of the T cells. The data also showed that the killing of target cells by CD40−/− DCs was also reduced. However, the T cells accumulated to a lesser extent in the spleens and lymph nodes of CD40−/− compared with the WT hosts (Fig. 2A). The proportion, as well as the absolute number of IFN-γ-secreting cells, was also reduced by as much as 50% in the CD40−/− hosts (Fig. 2B). Furthermore, the killing of SIINFEKL-pulsed target cells was reduced in the CD40−/− hosts compared with the WT hosts (94% vs 56%; Fig. 2C). We also obtained similar results after s.c. immunization of the OVA transfectants (data not shown).

Altogether, these results demonstrate that although CD40 on APCs is not absolutely required to initiate naive CD8+ T cell priming, it is important in inducing T cells to undergo maximum expansion and differentiation into effectors.
contrast, CD40−/− DCs induced significantly weaker T cell expansion as well as IFN-γ production at all time points (Fig. 3, A and B). The reduced T cell response induced by the CD40−/− DCs led to a corresponding decrease in the ability of the hosts to eliminate peptide-pulsed targets during the peak of the response (Fig. 3C). By day 12, however, there is very little CTL activity left regardless of the immunizing DC. This is most probably due to the low numbers of effector cells that remain at this point.

We also analyzed CD8+ T cell responses induced by DCs generated in vivo. Because there is only a small population of DCs in the spleen and they are difficult to isolate in large numbers, we first injected WT and CD40−/− mice with B16 tumor cells that secrete Fli-3 ligand. Fli-3L has previously been shown to induce the differentiation and expansion of functionally mature DC subsets in vivo (36, 40). In our case, injection of B16-Fli3L cells resulted in up to 50-fold expansion of CD11c+ cells in the spleen, with no difference in DC subsets between WT and CD40−/− mice (data not shown). We pulsed the cells with LCMV gp33 peptide and used them to stimulate P-14 T cells in vivo. Similar to in vitro-derived BMDCs, CD40−/− splenic DCs induced much less T cell proliferation and cytokine production on day 4 of the response (Fig. 3D and data not shown).

The above results are concordant with the data obtained using CD40-deficient hosts. Moreover, they directly demonstrate a key role for CD40 on DCs in inducing maximal naive CD8+ T cell responses because they are the only cells lacking this receptor in these experiments.

**Peptide-pulsed DC induce CD4+ T cell-independent CD8+ T cell responses**

CD8+ T cell responses to noninflammatory Ags such as peptides, soluble proteins, particulate Ags, and cell-associated Ags, including peptide-pulsed DCs are largely dependent on CD4+ T cells (15, 41). It is believed that CD40L on activated CD4+ T cells is needed to stimulate CD40 on APCs and trigger licensing, and that this is the mechanism by which CD4+ T cells provide help for CD8+ T cell responses. In our system, CD4+ T cell responses might be generated to bovine serum proteins presented by cultured BMDCs or if the LCMV 13mer peptide contained a MHC class II epitope. To determine whether CD4+ T cells are responsible for activating CD40 on the DCs in our experiments, we examined P-14 T cell responses induced by peptide-pulsed DCs in CD4+ T cell-deficient hosts. We chose to acutely deplete CD4+ T cells with GK1.5 Ab because this gives the most complete elimination of these cells. Surprisingly, we found that WT DCs stimulated an equivalent P-14 T cell response with or without CD4+ T cells (Fig. 4). This indicates that the CD8+ T cell response to peptide-pulsed DCs can occur independently of CD4+ T cell help. Depletion of CD4+ T cells resulted in a slight increase in the absolute number of IFN-γ-producing cells, which perhaps suggests a possible effect of regulatory T cells. However, this difference was not statistically significant. Interestingly, CD40−/− DCs still induced weaker P-14 T cell priming in the absence of CD4+ T cells. This implies that CD4+ T cell and CD40 activity are not always equivalent. Moreover, it suggests that CD4+ T cells are not the only cells capable of activating CD40 on APCs during an immune response. To investigate this further, we did most of our subsequent experiments in CD4+ T cell-depleted hosts.

**Pivotal role of CD40-CD40L interaction in endogenous CD8+ T cell response**

We next sought to extend our analysis of naive CD8+ T cell priming to a non-Tg system, in which host mice have normal T cell frequencies. In the first set of experiments, we analyzed in vivo...
FIGURE 4. Helper-independent CD8⁺ T cell responses. CFSE-labeled Thy1.1⁺ P-14 T cells were adoptively transferred into WT mice, which were depleted of CD4⁺ T cells or left untreated. One day later, the mice were immunized i.v. with 1 x 10⁶ gp33 peptide-pulsed or unpulsed DCs from the indicated mice. Spleens were harvested on day 4 and analyzed as in Fig. 1. A, Percentage of P-14 T cells in the total CD8⁺ T cell population. B, Absolute number of IFN-γ-producing P-14 T cells. Each circle represents an individual mouse; the bars represent the means. The numbers in parentheses below indicate the percentage of P-14 T cells secreting IFN-γ. The results shown are representative of three independent experiments with three mice per group. *, p < 0.01.

CTL activity in WT, CD40⁻/⁻, and CD40L⁻/⁻ mice 1 wk after immunization with OVA-transfected cells. As expected, unimmunized WT, CD40⁻/⁻, and CD40L⁻/⁻ mice were unable to lyse OVA peptide-pulsed targets (data not shown). Immunized WT mice exhibited strong CTL responses, being able to lyse as much as 80% of targets (Fig. 5A). In contrast, both the immunized CD40⁻/⁻ and CD40L⁻/⁻ mice showed much weaker CD8⁺ T cell priming as target cell killing in these mice was reduced to ~20% (Fig. 5A). Depletion of CD4⁺ T cells had no effect on the ability of WT animals to mount a primary CTL response (Fig. 5B).

In the second set of experiments, we examined endogenous CD8⁺ T cell responses in WT hosts upon immunization with WT or CD40⁻/⁻ DCs pulsed with OVA peptide. WT DCs induced a very strong CTL response, resulting in almost complete elimination of target cells (Fig. 5C). In contrast, CD40⁻/⁻ DCs induced more modest CTL activity (Fig. 5C) and this weaker response was observed whether or not CD4⁺ T cells were present. We could not detect CD8⁺ T cell cytokine production directly ex vivo in either WT DC or CD40⁻/⁻ DC immunized mice (data not shown). Upon in vitro restimulation, we found that only CD8⁺ T cells from mice immunized with WT DCs were able to secrete IFN-γ (Fig. 5D).

Interestingly, we observed a reduction in the number of IFN-γ-producing cells in the absence of CD4⁺ T cells. This appears to be a difference between the TCR-Tg vs the endogenous (polyclonal) T cell response and could be due to differences in T cell affinity and/or precursor frequency. However, it is important to note that despite the reduced number of endogenous IFN-γ-producing CD8⁺ T cells in CD4⁺-depleted hosts, the response remains CD4⁺-dependent.

Overall, these results indicate that the CD40-CD40L interaction is required to generate a highly effective primary response from an endogenous polyclonal CD8⁺ T cell population and this requirement is observed even in the absence of CD4⁺ T cell help.

FIGURE 5. Endogenous CD8⁺ T cell responses are compromised in the absence of CD40 or CD40L. A, WT, CD40⁻/⁻, and CD40L⁻/⁻ mice were immunized s.c. with 5 x 10⁶ OVA-transfected cells or left unimmunized. One week later, OVA peptide-specific CTL responses were assessed using an in vivo CTL assay. Specific target cell killing from the different mice are presented as mean ± SD. The results shown are representative of three independent experiments with three to five mice per group. B, WT mice depleted of CD4⁺ T cells or left untreated were immunized as in A. In vivo CTL activity against OVA peptide-pulsed targets was assessed 1 wk later and the data are presented as mean ± SD. The results shown are representative of three independent experiments. C, CD4⁺-depleted or undepleted WT mice were immunized i.v. with 1 x 10⁶ OVA peptide-pulsed or unpulsed DCs from the indicated mice. In vivo CTL activity against OVA peptide-pulsed targets was determined 1 wk later and presented as mean ± SD of specific target killing. D, CD4⁺-depleted or undepleted mice were immunized as in C. On day 7, spleenocytes were harvested and restimulated in vitro with 1 µg/ml OVA peptide. After 6 days, intracellular IFN-γ production was assessed following an additional 5-h incubation with OVA peptide. Representative plots are shown, with the numbers indicating the percentage of CD8⁺ T cells producing IFN-γ. The frequency of IFN-γ⁺ cells in mice immunized with unpulsed DCs was ~0.5%. The results shown are representative of two independent experiments with three mice per group.

CD40L expression by CD8⁺ T cells contributes to maximal response

We hypothesized that in the absence of CD4⁺ T cells, CD40L might be provided to DCs by the CD8⁺ T cells themselves. To test this hypothesis, we examined the response of CD40L-deficient P-14 T cells in WT hosts after immunization with the LCMV 13-mer peptide. In this system all the host APCs express CD40 and all the host T cells express CD40L; only the adoptively transferred T cells are unable to express CD40L. P-14/CD40L⁻/⁻ T cells proliferated 2- to 3-fold less compared with WT P-14 T cells in the presence or absence of CD4⁺ T cells (Fig. 6A). The proportion and absolute number of P-14/CD40L⁻/⁻ T cells secreting IFN-γ was also significantly decreased (Fig. 6B). The reduced P-14/ CD40L⁻/⁻ T cell response parallels that of WT P-14 T cells stimulated by CD40L⁺ DCs in CD4⁺ T cell-deficient hosts.

We next investigated whether the reduced CD8⁺ T cell response was due to deficient activation of the CD40-positive host APCs and not to an inherent defect of the CD40L-deficient T cells. Injection of agonistic anti-CD40 Ab along with the LCMV 13-mer increased the P-14/CD40L⁻/⁻ T cell response to WT levels (Fig. 6C). This increased response was not observed when CD4₀⁻/₀ hosts were used (data not shown) ruling out the possibility of the Ab directly activating the transferred T cells, which can express CD40. Furthermore, adoptive transfer of a 1:1 mixture of WT and CD40L-deficient P-14 T cells resulted in equal expansion and
preliminary experiments, we also obtained similar data using CFSE-labeled Thy1.1 WT or CD40L-deficient P-14 T cells. One day later, the mice were immunized i.v. with 1 μg of LCMV 13-mer or left unimmunized. On day 4, P-14 T cell expansion (A) and IFN-γ production (B) were analyzed as in Fig. 1. Each circle represents an individual mouse; the bars represent the means. The numbers in parentheses below indicate the percentage of P-14 T cells secreting IFN-γ. The results shown are representative of three independent experiments with three mice per group. C, CD4-depleted mice containing WT or CD40L-deficient P-14 T cells were immunized i.v. with 1 μg of LCMV 13-mer and injected i.p. with 50 μg of the agonistic anti-CD40 Ab FGK45. Spleens were harvested on day 4 and the percentage of P-14 T cells in the total CD8+ T cell population as well as absolute numbers of IFN-γ-producing P-14 T cells were determined. Each circle represents an individual mouse; the bars represent the means. The numbers in parentheses below indicate the percentage of P-14 T cells secreting IFN-γ. The results shown are representative of three independent experiments with three mice per group.

IFN-γ production of both T cell populations (Fig. 6D and data not shown). Injection of the anti-CD40L blocking Ab MR1 inhibited the ability of the WT CD8+ T cells to rescue the response of the CD40L-deficient T cells (Fig. 6D). Therefore, the CD40L-deficient T cells were fully functional when the WT CD8+ T cells provided the CD40L signal, presumably to the CD40-expressing APCs.

Our data so far do not indicate whether Ag-specific CD40L-expressing CD8+ T cells from the host might be providing “help” to the adoptively transferred T cells. To address this issue, we used CD40L+/− mice, which are completely unable to express CD40L, as hosts. The magnitude of T cell expansion was very similar in WT and CD40L+/− hosts, but more importantly, the CD40L-deficient P-14 T cells still exhibited ~2-fold reduction in proliferation compared with the WT P-14 T cells (data not shown). In preliminary experiments, we also obtained similar data using RAG−/− hosts, which lack both endogenous CD4+ and CD8+ T cells (data not shown).

All of these results show that there is no inherent defect in the ability of the CD40L-deficient CD8+ T cells to be primed and their reduced response can be attributed to their inability to activate APCs. These data provide functional evidence that CD8+ T cells are expressing CD40L. We verified that this is the case by stimulating the TCR-Tg CD8+ T cells with peptide or PMA and ionomycin and detecting low levels of CD40L on the surface of WT cells but not CD40L−/− cells (data not shown); this is consistent with some other reports that CD8+ T cells can express CD40L (42–45). Therefore, CD40L expression by the responding CD8+ T cells contributes to the generation of a maximal primary response.

**TLR stimulation does not compensate for CD40 or CD40L deficiency**

TLR ligands stimulate APCs to mature and enhance their ability to induce T cell activation and differentiation (29, 32). To determine whether TLR stimulation can substitute for CD40 signaling, we

![Graph A](image1.png)

**FIGURE 6.** Reduced responses by CD40L-deficient CD8+ T cells. CD4-depleted or undepleted WT mice were adoptively transferred with CFSE-labeled Thy1.1 WT or CD40L-deficient P-14 T cells. One day later, the mice were immunized i.v. with 1 μg of LCMV 13-mer or left unimmunized. On day 4, P-14 T cell expansion (A) and IFN-γ production (B) were analyzed as in Fig. 1. Each circle represents an individual mouse; the bars represent the means. The numbers in parentheses below indicate the percentage of P-14 T cells secreting IFN-γ. The results shown are representative of three independent experiments with three mice per group. C, CD4-depleted mice containing WT or CD40L-deficient P-14 T cells were immunized i.v. with 1 μg of LCMV 13-mer and injected i.p. with 50 μg of the agonistic anti-CD40 Ab FGK45. Spleens were harvested on day 4 and the percentage of P-14 T cells in the total CD8+ T cell population as well as absolute numbers of IFN-γ-producing P-14 T cells were determined. Each circle represents an individual mouse; the bars represent the means. The numbers in parentheses below indicate the percentage of P-14 T cells secreting IFN-γ. The results shown are representative of three independent experiments with three mice per group.

![Graph B](image2.png)

**FIGURE 7.** CD40−/− DC stimulated with TLR agonists induce reduced CD8+ T cell responses. WT mice were depleted of CD4+ T cells and adoptively transferred with CFSE-labeled Thy1.1+ P-14 T cells. One day later, the mice were immunized i.v. with 1 × 10^6 gp33 peptide-pulsed or unpulsed DCs from the indicated mice that were stimulated with representative TLR agonists in vitro or left untreated. Spleens were harvested on day 4 and analyzed as in Fig. 1. A, Percentage of P-14 T cells in the total CD8+ T cell population. B, Absolute number of IFN-γ-producing P-14 T cells in the spleen. Each circle represents an individual mouse; the bars represent the means. Mice immunized with unpulsed DC had a background of 2.5 × 10^3 IFN-γ+ cells. C, Mean percentage of P-14 T cells secreting IFN-γ. The results shown are representative of three independent experiments with two to three mice per group.
CD40L expression is not only by professional APC but also by hematopoietic precursors, epithelial cells, endothelial cells, and even activated T cells (4–6). Because CD40 plays a role in the generation of CD8⁺ T cells responses, it was important to determine on which cell CD40 was acting. This issue has not been resolved in many previous studies that have explored CD40 function using agonistic anti-CD40 Ab, which bind to all CD40-expressing cells. Our adoptive transfer experiments map the key role of CD40 to APCs in the host. Moreover, we demonstrate that when DCs are the only APC lacking CD40, the reduction in T cell responses is similar to that observed when all host APCs were CD40 deficient. Therefore, CD40 is working at least in part on DCs. Our findings do not rule out the possibility that CD40 might also have the same stringent requirement for CD40 in the priming of naive CD4⁺ T cells (20, 46).

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Another important question was what cell was the source of CD40L that was needed to stimulate APCs in vivo. It has generally been thought that Th cells are the principal source of CD40L in CD40-dependent responses. However, our finding that the absence of helper CD4⁺ T cells did not affect the CD40 dependence of the CD8⁺ T cell responses indicated that some other cells provided CD40L for APC activation. Because CD8⁺ T cells can express CD40L (42–46) and they are able to directly interact with APCs, we therefore reasoned that they might be able to provide their own help. This possibility has been hinted at by some earlier studies (26, 50, 51); however, the experiments in which we used CD40L-deficient CD8⁺ TCR-Tg T cells provided the first direct test of this hypothesis. These mutant cells exhibited the same defective responses that were observed when WT T cells were stimulated with CD40L-deficient APCs in the presence or absence of CD40⁺ T cells. Moreover, WT P-14 T cells were able to rescue the P-14/
CD40L−/− T cell response, formally showing that CD8+ T cells can provide help in “trans.” NK cells, NKT cells, and platelets also express CD40L (5). Moreover, NKT cells have been shown to directly activate DCs in a CD40-CD40L-dependent manner (52–54). However, we find that depletion of NK cells, NKT cells, and platelets had no effect on the generation of primary CD8+ T cell responses in our experimental systems (data not shown). We conclude that Ag-specific CD8+ T cells can directly activate APCs through CD40L and thereby provide their own help in the absence of CD4+ T cells.

It has been shown in other systems that CD4+ T cells provide help through CD40L (8, 23, 24). Given our results, it is likely that CD40L expressed by both CD4+ and CD8+ T cells can activate APCs and contribute to the amplification of a normal immune response. It is remarkable that we observed CD8+ T cell responses in the absence of help in light of the general requirement for CD4+ T cells in CD8+ T cell responses against noninflammatory Ags. The precursor frequency and affinity of responding CD8+ T cells have been shown to affect helper dependence (55, 56). However, titrating the number of adoptively transferred T cells still resulted in detectable CD40-dependent responses in the presence or absence of CD4+ T cells (data not shown). Moreover, we were able to detect endogenous primary CTL responses; aside from having a low frequency, the responders in this case also consisted of a spectrum of affinities. In the APC-licensing model, CD40 signaling is usually equated with CD4+ T cell help. The fact that we observed CD40-dependent responses despite the absence of helper CD4+ T cells indicates that the two are not always equivalent.

The final issue that our studies address is how CD40 stimulation compares to microbial (TLR) stimulation for licensing APCs to support CD8+ T cell responses. Although it is known that APCs must be activated to stimulate naïve T cell responses, it has been unclear whether all activating stimuli are similarly effective in this process, particularly for responses in vivo. By incubating WT and CD40−/− DCs with TLR ligands before immunization, we were able to directly show that the two signals have nonredundant effects on the stimulatory property of DCs. Although TLR ligands were able to amplify CD8+ T cell responses in the absence of CD40 signaling, these responses never reached the levels that were induced when CD40 signaling was present. In other words, maximum CD8+ T cell proliferation could only be achieved when both CD40 and TLR are stimulated. This result is consistent with reports that TLR stimulation can influence CD40 responses for cytokine production and amplification of CD8+ T cell responses (57, 58).

CD40 stimulation of DCs has been shown to be important for production of IL-12, which promotes CD8+ T cell expansion and differentiation (7, 59). However, we found that WT and CD40-deficient DCs made similar levels of IL-12 upon incubation with either WT or CD40L-deficient CD8+ T cells (data not shown). Moreover, we found that with or without TLR ligation, there was no difference in MHC–peptide levels as well as costimulatory molecule expression (CD80 and CD86) and IL-12 production between WT and CD40−/− DCs (data not shown). Therefore, the different responses induced by CD40- and TLR-stimulated DCs are not due to differences in conventional “costimulatory repertoire” and the underlying molecular mechanism(s) remain to be determined. This is especially important in light of recent efforts that establish the need to distinguish between phenotypically and functionally mature DCs (3). CD40-matured DCs have been reported to be more phenotypically stable compared with TLR-matured DCs (60). In addition, CD40 stimulation has been shown to increase the lifespan of DCs (61). It has also been shown that CD40 induces higher levels of CD70 (CD27L) on DCs compared with TLRs and this correlates with increased immunogenicity even in the absence of helper CD4+ T cells (62, 63). Whether any of these previously reported mechanisms and/or other ones account for the effects we observed will require further studies.

In conclusion, our data support a new model of CD8+ T cell-mediated APC “licensing,” in which CD40L expressed by Ag-specific CD8+ T cells interacts with CD40 on APCs, leading to maximal CD8+ T cell responses that can be primed in the absence of CD4+ T cell help. Our findings also reveal a unique role for CD40 signaling on APC activation that cannot be fully replaced by TLR stimulation. It will be interesting to examine further whether the CD40 dependence of the CD8+ T cell response extends to noninflammatory Ags considering that some pathogen-specific responses occur in the absence of CD40 ligation. It is also important to study whether CD40-CD40L interactions are needed not just during primary but also during other phases of the immune response. One of the major issues that still needs to be resolved is determining what is the ultimate fate of CD8+ T cells primed in the absence of CD40. One study has shown that systemic administration of Ag-loaded CD40−/− DCs failed to sustain the activation and led to deletional tolerance of CD4+ T cells (64). There is also evidence that CD40L is involved in the generation but not maintenance of LCMV-specific memory CD8+ T cells (65). In light of these findings, we are currently investigating whether CD40-CD40L interaction also plays a role in the generation, maintenance, and response of memory CD8+ T cells.

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Disclosures
The authors have no financial conflict of interest.

References
CD40-CD40L INTERACTION IN HELP-INDEPENDENT CD8+ RESPONSES


