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*J Immunol* 2007; 178:520-529; doi: 10.4049/jimmunol.178.1.520
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The Journal of Immunology is published twice each month by The American Association of Immunologists, Inc., 1451 Rockville Pike, Suite 650, Rockville, MD 20852
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Print ISSN: 0022-1767 Online ISSN: 1550-6606.
Dendritic Cell Transmigration through Brain Microvessel Endothelium Is Regulated by MIP-1α Chemokine and Matrix Metalloproteinases

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Dendritic cells (DCs) accumulate in the CNS during inflammatory diseases, but the exact mechanism regulating their traffic into the CNS remains to be defined. We now report that MIP-1α increases the transmigration of bone marrow-derived, GFP-labeled DCs across brain microvessel endothelial cell monolayers. Furthermore, occludin, an important element of endothelial tight junctions, is reorganized when DCs migrate across brain capillary endothelial cell monolayers without causing significant changes in the barrier integrity as measured by transendothelial electrical resistance. We show that DCs produce matrix metalloproteinases (MMP) -2 and -9 and GM6001, an MMP inhibitor, decreases both baseline and MIP-1α-induced DC transmigration. These observations suggest that DC transmigration across brain endothelial cell monolayers is partly MMP dependent. The migrated DCs express higher levels of CD40, CD80, and CD86 costimulatory molecules and induce T cell proliferation, indicating that the transmigration of DCs across brain endothelial cell monolayers contributes to the maintenance of DC Ag-presenting function. The MMP dependence of DC migration across brain endothelial cell monolayers raises the possibility that MMP blockers may decrease the initiation of T cell recruitment and neuroinflammation in the CNS. The Journal of Immunology, 2007, 178: 520–529.

Dendritic cells (DCs) are in the CNS are sufficient to present Ags to naive T cells during autoimmune inflammatory diseases (1). Although DCs could develop from microglia in the brain (2, 3) or be found as resident cells in the CNS (4, 5), they also infiltrate into the CNS from the blood (1). The mechanism of DC infiltration into the CNS is yet to be determined.

During CNS inflammation, blood-borne immune cells access the CNS via the blood-brain barrier (BBB) (6, 7). Major components of the BBB, the brain microvessel endothelial cells, are closely interconnected with special cell-cell contacts, such as tight junctions (8), and along with astrocytes, pericytes, and neurons, they form the neurovascular unit (9). The neurovascular unit is important in the maintenance of the immune-privileged nature of the CNS and regulating cellular transmigration (10, 11). In inflammation, the BBB becomes compromised and allows cellular traffic into the CNS. Although factors that regulate lymphocyte and monocyte migration into the CNS have been extensively studied (12–19), the mechanism of DC interaction with brain microvessel endothelium and DC transmigration across the BBB remains to be defined. The concordance of DC accumulation in cerebrospinal fluid throughout experimental autoimmune encephalomyelitis (16, 20–24), DC localization to the proximity of inflamed microvessels in multiple sclerosis (MS) lesions (25), and the production of astrocyte-derived chemokines that promote DC recruitment into the CNS (26) make it likely that brain microvessel endothelial cells regulate DC recruitment into the CNS. Although DC transmigration across HUVEC and epithelial cell monolayers has been studied (27–31), the mechanism of DC migration across brain microvascular endothelium has not been investigated.

To explore the mechanism of DC transmigration across the brain endothelium, primary cultures of brain microvascular endothelial cells were used (32–34). Transmigration of DCs across brain microvascular cell monolayers was augmented in the presence of the MIP-1α chemokine and partly mediated by matrix metalloproteinases (MMP). DCs developed numerous filopodia and induced the rearrangement of occludin, an endothelial tight junction protein, without causing a detectable decrease of transendothelial electrical resistance (TEER) values, indicating that endothelial monolayers stayed intact during DC migration. DC transmigration across brain microvessel endothelial cell monolayers led to the up-regulation of CD40, CD80, and CD86 costimulatory molecules on DCs. Following their transmigration across brain microvessel endothelial cells, DCs induced Ag-specific T cell activation. Our findings reveal that DC transmigration across brain microvessel endothelium is regulated by MIP-1α chemokine, MMP, and occludin perturbation. Because the transmigration of DCs across brain microvessel endothelial cells contributes to the activation of Ag-specific T cells, the BBB may be a key element in the local restimulation of T cell responses in the proximity of brain microvessels and in the amplification of T cell-mediated immunity in the CNS.

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Received for publication May 4, 2006. Accepted for publication October 6, 2006.

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1 This work was supported by National Institutes of Health Grant RO1-NS 37570-01A2 (to Z.F.).

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4 Abbreviations used in this paper: DC, dendritic cell; BBB, blood-brain barrier; MS, multiple sclerosis; MMP, matrix metalloproteinase; TEER, transendothelial electrical resistance; BCEC, brain capillary endothelial cell; SEM, scanning electron microscopy; ZO-1, zona occludens-1; Tg, transgenic; NGAL, neutrophil gelatinase B-associated lipocalin; CLSM, confocal laser scanning microscopy.

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Materials and Methods

Mice

Four- to 6-wk-old female C57BL/6 mice were obtained from The Jackson Laboratory. GFP-transgenic (Tg) mice were a gift from Dr. P. Marrack (National Jewish Center for Immunology and Respiratory Medicine, Denver, CO) (35). OVA323–334 (SINFEKL) peptide-specific TCR Tg mice (OT-I) (36, 37) were provided by Dr. K. Hogquist (University of Minnesota, Minneapolis, MN). Both Tg strains were bred on the C57BL/6 background. The animals were housed according to the guidelines of the National Institutes of Health and University of Wisconsin-Madison Research Animals Resource Center.

Abs and secondary reagents

Abs anti-CD205 (clone NLDC-145) and anti-CD11c (clone N418) were purified from hybridoma cell supernatants (American Type Culture Collection) and conjugated with the Cy5 fluorochrome (Amersham Biosciences) and biotin (Sigma-Aldrich). Anti-CD11a (clone M17/4), anti-CD8 (clone 53.6-7), anti-Vo2 (B2.0.1), anti-1-A3 (clone AF6-120.1), anti-CD40 (H440-3), anti-CD80 (16-10A1), anti-CD86 (GL1), and anti-CD195 (CCR5) (clone C34-3448) Abs, streptavidin-Cy3-Chrome, and streptavidin-allophycocyanin conjugates were purchased from BD Pharmingen. Streptavidin-Alexa 568 conjugate was purchased from Molecular Probes and monoclonal goat anti-mouse MPM9 (AF9059) was purchased from R&D Systems.

Brain capillary endothelial cells (BCEC)

Mouse BCEC were isolated and cultured according to Deli et al. (38, 39). In brief, cerebra of 4- to 6-wk-old mice were mechanically homogenized and microvessels were isolated by two rounds of collagenase digestion and density centrifugation. Capillary fragments were directly seeded onto Falcon inserts (3-μm pore; Fisher Scientific) coated with collagen type IV/fibronectin (10 μg/ml; Sigma-Aldrich) in DMEM with 10% FBS (Fisher Scientific), 2 mM l-glutamine, 100 μg/ml penicillin, 100 μg/ml streptomycin, and 50 μg/ml gentamicin (Sigma-Aldrich). Isolated capillaries were cultured for 2 days in puromycin-supplemented medium (4 μg/ml; Sigma-Aldrich) to eliminate contaminating perivascular smooth muscle cells, followed by culture in medium supplemented with basic fibroblast growth factor (2 ng/ml; Roche) (40, 41). After reaching confluence, cells were cultured in serum-free DMEM/Ham’s F12 (Sigma-Aldrich) containing 2 mM l-glutamine, 100 U/ml penicillin/streptomycin, and 50 μg/ml gentamicin (Sigma-Aldrich). Isolated capillaries were cocultured with DCs for 3 h, fixed with 1% glutaraldehyde, and the phenotype of adhering DCs was analyzed according to Materials and Methods. Scanning electron microscopy shows that in the presence of chemokine MIP-1α there is a close contact between DCs and the BCEC monolayer, and DCs develop numerous filopodia (arrows) and protrusions (arrowheads; original magnification, ×1000). Inset, Higher magnification of DC morphology (original magnification, ×5000). The images shown are representative of three independent assays with similar observations.

Dendritic cells

DCs were generated as previously described (47, 48). Briefly, bone marrow obtained from femurs and tibias was washed and plated in 24-well plates in RPMI 1640 with 5% FBS supplemented with 100 U/ml penicillin/streptomycin and 20 ng/ml GM-CSF. GM-CSF was titrated from supernatants of the GM-CSF secreting X63 cell line (gift from Dr. A. Erdei, Eotvos University, Budapest, Hungary). On day 7, the nonadherent and loosely adherent cells were removed and replenished in the absence of GM-CSF. Following overnight incubation, the nonadherent cells were collected for use. Purity of DC cultures was >80% as determined by flow cytometry demonstrating high expression of CD11c, CD205, and MHC class II proteins as previously described (48).

Flow cytometry

Cells (0.5–1 × 10⁷) were processed for flow cytometric analysis as previously described (48). Single-cell suspensions were incubated for 30 min on ice with saturating concentrations of Abs in the presence of unlabelled Fc-RII/FCγRII-specific Ab (clone 2.4G2) to block FcR-mediated, non-specific binding. Cell surface staining was acquired on a four-color FACSCalibur flow cytometer and analyzed with CellQuest software version 3.1 (BD Immunocytometry Systems) and FlowJo (Tree Star) software version 5.4.5.

TEER

Endothelial tissue resistance measurement chamber (World Precision Instruments) was used to measure the electrical resistance of endothelial cell monolayers. TEER values (Ω × cm²) were calculated, following the manufacturer’s instructions, by subtracting the contribution of bare filters and medium and multiplying by the surface area of the membrane.

DC transendothelial migration assay

DC migration across in vitro BCEC monolayers was analyzed using QCM 24-well invasion assay. Migration index was calculated as the percentage of migrated DC number from the total cell population number (2.5 × 10⁷) added to the upper compartment of the migrating chambers for 24 h. Data are presented as the mean ± SD of five independent experiments. Each data point was run in triplicate and data were analyzed using two-tailed t tests. B, BCEC monolayers were cocultured with DCs for 3 h, fixed with 1% glutaraldehyde, and the phenotype of adhering DCs was analyzed according to Materials and Methods. Scanning electron microscopy shows that in the presence of chemokine MIP-1α there is a close contact between DCs and the BCEC monolayer, and DCs develop numerous filopodia (arrows) and protrusions (arrowheads; original magnification, ×1000). Inset, Higher magnification of DC morphology (original magnification, ×5000). The images shown are representative of three independent assays with similar observations.

Confocal laser scanning microscopy (CLSM)

Expression of endothelial occludin and DC distribution and localization on monolayers of BCEC were studied using CLSM. Confluent monolayers of endothelial cells were cocultured with GFP-labeled DCs for 3 h, rinsed with PBS supplemented with Ca²⁺ and Mg²⁺, and fixed in 1% paraformaldehyde in PBS. Samples were permeabilized in 0.1% Triton X-100 and 1% BSA in PBS and labeled with rabbit anti-occludin (4 μg/ml; Zymed Laboratories) pAb, followed by Alexa 568-conjugated goat anti-rabbit IgG (Chemicon International). The samples were analyzed by CLSM (MRC-1024; Bio-Rad), equipped with argon (488 nm) and helium-neon (568 nm) lasers. Serial sections were collected with a step increment of 0.1 μm in the z-axis starting at the apical surface. Cross-sections of xz and yz images were rendered using Visbio (version 2.31) software (University of Wisconsin, Madison, WI).
Scanning electron microscopy

BCEC monolayers cocultured with DCs for 3 h were fixed with 1% glutaraldehyde (Sigma-Aldrich) and 1% tannic acid (Mallinckrodt) in 0.1 M phosphate buffer (pH 7.4) for 10 min at room temperature and incubated in the same buffer overnight at 4°C. The monolayers were then washed with phosphate buffer and gradually dehydrated with 30, 50, 70, 80, 90, 95, and 100% (three times) of ethanol at room temperature. The insert membranes were carefully cut out and placed into the critical point dryer to exchange the ethanol content of the cells against CO2. The membranes then were coated with Au/Pd by TM200 (VCR Group) ion-beam sputter-coating device and analyzed by an Hitachi S-570 scanning electron microscope.

Zymography

The amount of secreted MMP in the cell culture supernatants was estimated by zymography. Gelatin (1 mg/ml; Sigma-Aldrich) was incorporated into polyacrylamide gel (10%). Concentrated cell samples (YM-10; Centricon) were mixed with an equal volume of 2× sample buffer (126 mM Tris-HCl (pH 6.8), 20% (v/v) glycerol, 0.01% (w/v) bromphenol blue; 4% (w/v) SDS), and electrophoresed for 3 h through the substrate gel. The gel was renatured in 2.7% (v/v) Triton X-100 for 1 h at room temperature and incubated overnight at 37°C in developing buffer (50 mM Tris-HCl (pH 7.5), 200 mM NaCl, 5 mM CaCl2, and 0.02% (v/v) Brij-35). The gel was stained with 0.25% (w/v) Coomassie Brilliant Blue in 25% isopropanol and 10% acetic acid and destained in 50% methanol and 10% acetic acid until bands with diminished staining appeared on a uniformly stained background. The molecular mass (kDa) of the gelatinases was estimated against markers of known molecular mass (Chemicon International). Gels were scanned (Hewlett-Packard Scanjet 5400c) and analyzed with a Scion Image program (a version of NIH Image; Scion Corporation).

Ag presentation assays

Splenocytes were isolated from OT-1 mice and labeled with CFSE as described previously (49) with minor modifications (50). In brief, OT-1 spleen cells were depleted of RBC, washed, CFSE (1 μM; Molecular Probes) labeled for 5 min at 37°C, and quenched with 20% FBS. DCs were pulsed with OVA (DC-OVA, 50 μg/ml; Sigma-Aldrich) for 16 h at 37°C, washed three times with PBS, and applied (2.5 × 10⁵) to the luminal side of BBB endothelial cells, or the membrane inserts alone. Migrated DC-OVA were collected after 24 h and combined in a ratio of 1:10 (DCs:T cells) with OVA-specific CFSE-labeled OT-1 TCR Tg splenocytes in culture. After 3 days, OT-1 cells were collected and analyzed by flow cytometry. The proliferation and activation of OT-1-specific TCR Tg T cells was measured by analyzing CFSE staining intensity and the level of CD11a/CD18 (LFA-1) expression. Fluorescent intensity was analyzed on gated Tg CD8⁺Va2⁺ T cells as was described previously (48, 51).

Statistical data analysis

Each data point was run in triplicate in three to five independent experiments as indicated and data were analyzed using two-tailed t tests. All values are expressed as mean ± SD. FACS measurements were also run in triplicates. Bivariate dot plots or probability contour plots were generated upon data reanalysis to display the frequencies of, and patterns by which, individual cells coexpressing certain levels of cell surface Ags. Histograms were generated based on these contour plots with CellQuest software version 3.1 (BD Immunocytometry Systems) and FlowJo (Tree Star) software version 5.4.5. Descriptive statistics were applied by calculating raw means and percentages.

Results

MIP-1α increases DC transmigration across brain microvascular endothelial cell monolayers

Because MIP-1α (also known as CCL3) has been suggested to play a critical role in the migration of DCs into peripheral tissues (52–55), and this chemokine is up-regulated under inflammatory conditions in the CNS (54, 56–60), we first sought to define the role of MIP-1α in DC migration across BCEC monolayers. As shown in Fig. 1A, GM-CSF-matured DCs were mobile and migrated across BCEC monolayers with high efficiency, even in the
absence of MIP-1α. However, there was a linear increase in DC transmigration across BCEC monolayers when increasing amounts of MIP-1α were applied (Fig. 1A). In the presence of MIP-1α, DCs strongly adhered to microvascular endothelial cells within 3 h and developed numerous filopodia (Fig. 1B, arrow), some of which reached across the endothelial cell monolayers (Fig. 1B, arrowheads). In the absence of MIP-1α, DCs adherence to brain microvessel endothelial cells was significantly compromised during the same time period (data not shown). These data indicate that GM-CSF-maturated DCs can migrate through brain microvessel endothelial cell monolayers and this migration is augmented in the presence of MIP-1α.

Increased MMP2/MMP9 secretion and occludin tight junction molecule reorganization correlate with DC migration across BCEC monolayers

During inflammatory diseases of the CNS, MMP have been implicated in the disruption of the BBB and regulation of cellular infiltration (61–64). Therefore, we studied whether DC-secreted MMP could contribute to the migration of DCs across brain endothelial cell monolayers. To evaluate the level of MMP secretion by DCs, supernatants from GM-CSF matured DCs were analyzed at 4, 6, 7, and 9 days of culture using gelatin zymography. As shown in Fig. 2, A and B, this method revealed a significant production of the latent or proform of MMP2 and MMP9 by DCs that accumulated in increasing amounts in the supernatants in a time-dependent manner. At day 4 of this in vitro culture period, DCs also expressed neutrophil gelatinase B-associated lipocalin (NGAL, 130-kDa band), which is known to covalently attach to MMP9 (65). Although NGAL production is predominantly associated with neutrophils, DC expression of this enzyme has also been suggested (66). In parallel with increased MMP production, NGAL expression declined in these cultures by time and was undetectable by day 6 of culture (Fig. 2A).

To determine MMP intracellular distribution in DCs, we performed immunocytochemistry and demonstrated punctate (vesicular) (Fig. 2C, arrowheads) as well as cytoplasmic localization of MMP9 (Fig. 2C, arrows). This is consistent with observations by Nguyen et al. (67) suggesting an accumulation of active MMP9 molecules in secretory vesicles that become effective during cell migration. We further demonstrated that MMP2 and MMP9 secretion by DCs could be blocked using an MMP-specific broad inhibitor, GM6001 (Fig. 2D). Zymography assays performed using DC culture supernatants indicated that GM6001 inhibited both MMP2 and MMP9 expression in a concentration-dependent manner (Fig. 2D).

It has been suggested that tight junction proteins can be degraded by MMP (68). To determine whether tight junction proteins were disrupted during DC transmigration across BCEC monolayers, we analyzed occludin expression and distribution on BCEC during DC migration. To measure the intercellular junctional integrity of the BCEC, the paracellular permeability of the monolayers was analyzed by measuring the TEER. After culture for 3 days in serum-supplemented medium, BCEC formed confluent monolayers that were well suited for studying the transcellular migration in vitro. The effect of DC coculture with BCEC on endothelial cell monolayer integrity was monitored from day 3 until day 5, after which experiments were performed (Fig. 3A). We also followed the effect of MIP-1α on electrical resistance of endothelial cell monolayers in the absence of DCs from day 3 until day 7 of in vitro culture (Fig. 3B). Both MIP-1α and DC interaction with BCEC caused no detectable changes in TEER values, suggesting that DCs were capable of diapedesis across the BCEC without disturbing the functional integrity of the barrier (Fig. 3).

To further analyze the mechanism of DC diapedesis across BCEC monolayers, we studied the distribution of tight junction molecules in the presence and absence of DCs. As shown in Fig. 4A, brain microvessel endothelial cells formed well-organized monolayers on basement membrane-coated filter inserts, presenting a uniform belt-like distribution of occludin tight junction molecules between endothelial cells. This appearance did not change in the presence of MIP-1α (Fig. 4B). Testing of MIP-1α treatment on zonula occludens-1 (ZO-1) molecule immunostaining also indicated that similarly to the distribution of occludin. MIP-1α had no effect on ZO-1 tight junction molecule distribution (data not shown). However, within 3 h after DCs were placed on the top of brain microvessel endothelial cell monolayers, discontinuous and poor organization of endothelial occludin was observed in the areas of DC interaction with brain microvessel endothelial cells (Fig. 4, C and D, arrows). In the absence of MIP-1α, there were fewer DCs adhering to microvascular endothelium (Fig. 4, C, arrowheads, and E). When DCs adhered to brain microvascular endothelial cell monolayers in the presence of MIP-1α, occludin distribution was different than that observed when endothelial cells were grown without DCs (Fig. 4D, arrows and z/y/z sections). Consistent with previous observations, DCs (42) also expressed occludin (yellow) in the presence of MIP-1α (69) (Fig. 4D, arrowhead). We applied the MMP inhibitor GM6001 to further understand the effect of MMP on occludin tight junction molecule distribution. We demonstrated that in the presence of GM6001, DCs (Fig. 4F, green and arrowheads) did not invade the endothelial tight junctions, leaving occludin distribution intact (Fig. 4F, right panel, arrows). These data indicate that MIP-1α-induced DC transmigration across brain microvessel endothelial cell monolayers is partly regulated by MMP and leads to occludin tight junction molecule redistribution. Because fewer DCs attached to the BCEC monolayer in the
absence of MIP-1α, DC-mediated occludin perturbation could not be investigated under these conditions. However, the finding that endothelial occludin distribution did not change significantly in the presence of MIP-1α without DCs (Fig. 4B) makes it likely that DCs and not MIP-1α chemokine play a critical role in modulating occludin distribution during DC diapedesis.

DC migration across BCEC monolayers can be partly blocked by MMP-specific inhibitor

Having established that GM-CSF-matured DCs produce MMP2 and MMP9 (Fig. 2), we next analyzed whether DCs would express MMP or induce MMP production by endothelial cells during their migration across BCEC monolayers. We have evaluated MMP production from BCEC supernatants cultured alone (BCEC) or with DCs (BCEC/DC) using gelatin zymography. As shown in Fig. 5, A and B, there was a significant increase of pro-MMP9 production measured in supernatants from BCEC/DC cocultures as compared with DCs cultured alone. This might indicate that MMP9 production by endothelial cells is induced by DCs. Three hours following medium exchange, endothelial cells grown alone did not produce pro-MMP9 but expressed low levels of active MMP9 and detectable amounts of pro-MMP2.

To determine whether secreted MMP facilitate DC transendothelial migration in addition to disruption of occludin, we analyzed DC traffic across the BCEC monolayers in the presence of control and active GM6001 peptide before and after MIP-1α treatment. Fig. 5C demonstrates that baseline DC migration across BCEC monolayers is significantly impaired in the presence of GM6001. MIP-1α-induced DC migration across BCEC monolayers can also be inhibited by GM6001 peptide, resulting in a return to nearly normal levels of migration.
baseline level of DC migration. This suggests that both baseline and MIP-1α-induced migration of DCs across brain microvessel endothelial cell monolayers is partly MMP dependent. However, additional mechanisms should also be considered, since both baseline and MIP-1α-induced migration of DCs across BCEC monolayers could not be entirely inhibited by MMP inhibitor (Fig. 5).

DCs activate naive CD8+ T cells upon their migration across brain endothelial cell monolayers

Previous studies have shown that DCs might be sufficient to present Ags to naive T cells and promote epitope spreading locally in the CNS (1). Whether DCs that migrate across brain microvessel endothelial cell monolayers would be capable of stimulating naive T cell responses was uncertain. To answer this question, we analyzed the Ag-presenting capability of OVA-pulsed DCs following their transendothelial migration. OVA-pulsed DCs were allowed to migrate across brain endothelial cell monolayers in the presence of MIP-1α for 24 h. As shown in Fig. 6A, DCs upon their migration expressed elevated levels of costimulatory molecules CD40, CD80, and CD86. Increased expressions of CD40, CD80, and CD86 costimulatory molecules on migrating DCs were not directly the result of MIP-1α treatment, because the expression of these molecules on DCs did not increase in the absence of brain microvessel endothelial cells (Fig. 6B, □). The CD80 (B7-1) molecule was up-regulated by MIP-1α on DCs in the presence of BCEC (Fig. 6B, □); however, the expression of CD80 remained unchanged after DC transmigration across brain microvessel monolayers (Fig. 6B, □). These data suggest that CD40, CD80, and CD86 costimulatory molecules are up-regulated upon the migration of DCs across brain microvessel cells and that this up-regulation is not dependent on the presence of MIP-1α. To further address the question of whether MIP-1α directly influences DC maturation, we measured the expression of the CCR5 chemokine receptor on DCs before and after migration in the presence and absence of MIP-1α treatment. As we show in Fig. 6C, MIP-1α treatment did not up-regulate CCR5 expression on nonmigrating DCs (or on DCs in in vitro cultures without brain endothelial cells). Some up-regulation of CCR5 on migrating DCs after MIP-1α treatment was shown; however, this up-regulation was not statistically significant when compared to nontreated cells. In conclusion, our data demonstrate that MIP-1α treatment alone is not sufficient for inducing DC maturation.

We further investigated whether DCs that migrated across brain microvessel endothelial cells were capable of presenting Ags to naive T cells. To accomplish this, we cocultured OVA-pulsed DCs collected from the lower chamber of migration wells with CFSE-labeled TCR Tg OT-1 T cells for 3 days in vitro. Cell division rapidly dilutes the CFSE signal from OT-1 progeny cells, and cell expansion could be measured by defining CSFE content using cytometry. As shown in Fig. 7 (right panel), migrated and OVA-pulsed DCs induced OT-1 T cell proliferation (black solid line) more efficiently than cultured, OVA-pulsed DCs (gray solid line), whereas CFSE-labeled OT-1 T cells cultured in the absence of DCs did not proliferate (dashed line, gray filled area). Up-regulation of...
LFA-1 activation marker on Ag-specific OT-1 T cells also indicated that DCs, upon their migration across brain microvessel endothelial cell monolayers, could induce T cell activation (Fig. 7, left panel). Taken together, these results indicate that DC transmigration across brain microvessel endothelium contributes to costimulatory molecule up-regulation and efficient Ag presentation by DCs.

**Discussion**

DCs are critical during the initiation, development, and maintenance of CNS autoimmunity and inflammation (1, 22, 25, 70). Previously, it was shown that DCs could be recruited into the CNS in multiple infectious and autoimmune diseases (1, 71–73); however, the pathway of their recruitment has not yet been elucidated. In this study, we have defined a role for MMP and MIP-1α in DC transmigration across brain microvessel endothelial cell monolayers. More importantly, we show that once DCs transmigrated across brain microvessel endothelial cell monolayers, they induced the activation of naive Ag-specific T cells.

Transmigration of immune cells from the blood into the CNS is critical in CNS inflammatory responses, and it is largely regulated by the BBB. Previous studies showed that multiple factors, including chemokines and adhesion molecules, play a role in leukocyte migration into the CNS (reviewed in Ref. 6 and Ransohoff et al. (11)), but factors regulating DC migration across the BBB are poorly understood. The mechanisms involved in DC interaction with nonbrain endothelial or epithelial cells have been examined by several groups (27, 30, 69, 74, 75). DC transmigration across mouse tracheal epithelial cells has been demonstrated to be severely impaired in MMP9-deficient mice (27). In these studies, similar to our findings, MIP-1α and MIP-3β elicited DC migration across tracheal epithelial cells. We show that the migration of DCs across the BCEC monolayers is very substantial in the absence of...
MIP-1α and increases in the presence of this chemokine. It was already shown that MIP-1α could have an effect on both endothelial cells and DCs. For example, Solanilla et al. (76) have demonstrated that MIP-1α and TNF-α stimulate endothelial production of membrane-bound and soluble Flt-3 ligand (76). A number of adhesion molecules on endothelial cells also have been shown to be up-regulated in the presence of MIP-1α and other chemokines (17). It is also known that immature DCs undergo chemotaxis in response to MIP-1α and up-regulate the CCR5 receptor that binds to this chemokine (77–80). In our work, we demonstrated that MIP-1α increases DC adhesion to BCEC and accelerates DC migration across the BBB.

MMPs were also shown to play a critical role in human DC migration across the Matrigel extracellular matrix, and the dominant MMP involved in this process was MMP9 (28). A physiologic function for p-glycoprotein (MDR-1) during the migration of DCs from skin via afferent vessels was also demonstrated (29). Because MDR-1 is a well-known transporter that mediates efflux of chemotherapeutic agents from the cytoplasm in brain endothelial cells, it is possible that this molecule also participates in DC transmigration across the BBB. Interestingly, it has also been reported that in DC migration across the BBB, both baseline and MIP-1α-induced DC migration across in vitro brain microvessel monolayers is further supported by the blocking effect of MMP inhibitor on DC migration. Because both baseline and MIP-1α-induced DC migration could be partly impaired using the MMP inhibitor GM6001, we suggest that other than MMP-dependent regulation of DC migration should be considered (Fig. 5C). This suggests that although MMP play a significant role in DC migration across BCEC, other mechanisms might also contribute to this process.

Recently, it has been shown that during inflammatory pain, disruption of BBB permeability is accompanied by alteration of occludin expression (85), which also suggests that loss of occludin integrity is related to inflammatory cell migration into the CNS in vivo. Although it is important to note that MIP-1α, among the other chemokines expressed by glial cells and macrophages around inflammatory plaques in MS and experimental autoimmune encephalomyelitis (86), is known to alter the integrity of the BBB, it has been previously shown that MIP-1α cannot be transported across the BBB (87) and does not affect occludin expression between epithelial cells (27). Our observations indicate that MIP-1α treatment alone does not result in perturbation of endothelial occludin (Fig. 4B) and ZO-1 (data not shown) expression in brain microvessel endothelial monolayers. Thus, discontinuous occludin expression during DC diapedesis is likely to be the result of DC transmigration and not MIP-1α chemokine treatment of BCEC in vitro.

Finally, we show that transmigration of DCs across brain microvessel endothelial cell monolayers allows Ag presentation to naive Ag-specific T cells. It has been shown previously that monocytes cultured with HUVEC differentiated into DCs within 2 days of culture, particularly after phagocytosing particles in the subendothelial collagen (29). In this study, we show that transmigration of GM-CSF-differentiated immature DCs through brain microvessel endothelial cell monolayers led to increased expression of costimulatory molecules CD40, CD80, and CD86 on the migrated DCs and facilitated their Ag presentation capability (Figs. 6 and 7). Because the bone marrow is the best known source for DC isolation, we used GM-CSF to differentiate DCs from the bone marrow to generate a sufficient number of cells to perform in vitro studies. It has been shown previously that GM-CSF-differentiated DCs resemble the phenotype of immature DCs circulating in the blood (88).

Recently, it was proposed that perivascular DCs in the CNS are critical in the initiation and maintenance of CNS inflammatory reactions (1, 89–91). Our data suggest that within the perivascular
space of the CNS, Ag-specific T cells could re-encounter Ags presented by DCs. Additionally, if a small frequency of naive T cells can access the perivascular space, activation of these cells could also happen. Although in our current model for T cell migration into the CNS the major prerequisite is that only activated T cells home to the nervous tissue, it has also been suggested that nonstimulated naive T cells are also capable of migrating into the brain (92). Our data show that after transmigration, DCs exhibit a mature phenotype with increased expression of costimulatory molecules (CD80, CD86, and CD40) and these cells retain their functional ability to activate naive CD8⁺ T cells in vitro.

In conclusion, we have demonstrated that DC transmigration across brain endothelial cell monolayers is increased in the presence of MIP-1α and partly dependent on MMP. Transmigrated DCs expressed high levels of costimulatory molecules and activated naive Ag-specific CD8⁺ T cells. Understanding the mechanism of DC transmigration across brain microvessel endothelial cells might lead to the design of targeted therapies to modify DC traffic in the CNS and thus inhibit immune-mediated inflammatory diseases of the CNS, such as MS.

Acknowledgments

We thank Dr. C. Weidenfeller for technical help with primary endothelial cells. We are also grateful to Philip Oshel (Biological and Biomaterials Preparation, Imaging and Characterization Laboratory, University of Wisconsin-Madison) for help with scanning electron microscopy and Khen MacVilay for flow cytometry.

Disclosures

The authors have no financial conflict of interest.

References
