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Decreased Alveolar Macrophage Apoptosis Is Associated with Increased Pulmonary Inflammation in a Murine Model of Pneumococcal Pneumonia

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Regulation of the inflammatory infiltrate is critical to the successful outcome of pneumonia. Alveolar macrophage apoptosis is a feature of pneumococcal infection and aids disease resolution. The host benefits of macrophage apoptosis during the innate response to bacterial infection are incompletely defined. Because NO is required for optimal macrophage apoptosis during pneumococcal infection, we have explored the role of macrophage apoptosis in regulating inflammatory responses during pneumococcal pneumonia, using inducible NO synthase (iNOS)-deficient mice. iNOS−/− mice demonstrated decreased numbers of apoptotic macrophages as compared with wild-type C57BL/6 mice following pneumococcal challenge, greater recruitment of neutrophils to the lung and enhanced expression of TNF-α. Pharmacologic inhibition of iNOS produced similar results. Greater pulmonary inflammation was associated with greater levels of early bacteremia, IL-6 production, lung inflammation, and mortality within the first 48 h in iNOS−/− mice. Labeled apoptotic alveolar macrophages were phagocytosed by resident macrophages in the lung and intratracheal instillation of exogenous apoptotic macrophages decreased neutrophil recruitment in iNOS−/− mice and decreased TNF-α mRNA in lungs and protein in bronchial alveolar lavage, as well as chemokines and cytokines including IL-6. These changes were associated with a lower probability of mice becoming bacteremic. This demonstrates the potential of apoptotic macrophages to down-regulate the inflammatory response and for the first time in vivo demonstrates that clearance of apoptotic macrophages decreases neutrophil recruitment and invasive bacterial disease during pneumonia. The Journal of Immunology, 2006, 177: 6480–6488.
prior in vitro findings, we predicted that these mice could have decreased AM apoptosis and this model could then be examined to determine the effects of decreased AM apoptosis, as opposed to NO deficiency, on the regulation of the inflammatory response, using a model of pneumococcal pneumonia.

We demonstrate that iNOS\(^{-/-}\) mice have decreased rates of AM apoptosis in vivo and that this is associated with a greater degree of inflammation in the lung. Furthermore, we demonstrate that instillation of apoptotic macrophages into mouse lungs decreases the greater lung inflammation observed in iNOS\(^{-/-}\) mice and decreases the likelihood of early invasive pneumococcal disease.

**Materials and Methods**

**Animals**

iNOS-deficient mice backcrossed onto a C57BL/6 background were obtained from The Jackson Laboratory and maintained as a homozygous colony. C57BL/6 mice (Harlan U.K.) were used as wild-type controls. Female mice were used throughout. For aminoguanidine treatment, C57BL/6 mice were treated with 2.5% aminoguanidine in their drinking water, with 5% glucose added to increase palatability, starting 7 days before pneumococcal infection (16). Control mice were treated with an equivalent dose of glucose. All animal experiments were conducted in accordance with the Home Office Animals (Scientific Procedures) Act of 1986 and received local ethical committee approval.

**In vivo pneumococcal infection model**

Infection of mice with 10\(^7\) CFU of type 2 pneumococci (strain D39) or mock infection with PBS was by direct tracheal instillation after anesthesia with ketamine (100 mg/kg i.p.) and acepromazine (5 mg/kg i.p.) as previously described (9). In survival studies, mice showing physical signs of severe illness were culled and their time of death recorded as the time at which they were culled.

**Collection of bronchial alveolar lavage, blood, and lungs**

Mice were killed by an overdose of sodium pentobarbitone and exsanguinated by cardiac puncture. Bronchial alveolar lavage (BAL) was performed as described (9). The cell differential was by review of cytospin preparations (9). Viable bacterial counts in lung and blood were obtained as described (9). The cell differential was by review of cytospin preparations.

Apoptosis detection was by Annexin V (BD Biosciences)/ToPro 3 (Molecular Probes) staining and flow cytometry or by nuclear morphology on cryosections (9). Viable bacterial counts in lung and blood were obtained as described (9).

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**Cytokine/chemokine mRNA detection**

Whole lung was harvested and immediately stored in RNAlater (Sigma-Aldrich). Total RNA was extracted from lung tissue by homogenization into TRIzol (Invitrogen Life Technologies) following the manufacturer’s protocol. RNA quality was assessed by comparison to 28S and 18S ribosomal RNA bands after electrophoresis and hybridization to Agilent Bioanalyzer 2100 LabChip using the RNA 6000 LabChip Kit (Agilent Technologies, Inc.). The RNA was not degraded, as judged by the ratio of 28S to 18S bands on the gel and by the ratio of peak areas from the RNA 6000 LabChip Kit analysis.

To perform TaqMan quantitative real-time PCR (qRT-PCR), RNA was DNA digested (Ambion) following the manufacturer’s protocol for routine DNase treatment. cDNAs were synthesized from RNA by reverse transcription using random hexamer primers. Relative expression of TNF-\(\alpha\) normalized to \(\beta\)-actin was by TaqMan gene expression assays (Applied Biosystems) for TNF-\(\alpha\) (Mm00443258_m1) and \(\beta\)-actin (Mm00607939_s1) following the manufacturer’s protocol.

**Cytokine production**

Cytokines in BAL were measured using Duoset ELISA development kits (R&D Systems) for mouse TNF-\(\alpha\), KC (CXCL1), MIP-2 (CXCL2), and IL-6 following the manufacturer’s protocols (19). Limits of detection were 15 pg/ml.

**Lung injury**

Protein levels in BAL were measured using a BCA protein assay (Pierce), following the manufacturer’s protocol (20).

**Histopathology**

Unlavaged lungs were fixed via the trachea with 10% buffered formalin at 20 cm H\(_2\)O, paraffin-embedded sections prepared, sectioned, stained with H&E and independently evaluated by two pathologists (P. G. Ince and S. S. Cross) using a BH2 Olympus microscope.

**Instillation of apoptotic bodies**

Apoptotic AM were generated from BAL of donor C57BL/6 mice. BAL was spun for 5 min at 1000 \(\times\) g and cells resuspended in RPMI 1640 (Invitrogen Life Technologies) 10% heat-inactivated FCS (Bioclear), 100 \(\mu\)g/ml streptomycin, and 100 U/ml penicillin (Invitrogen Life Technologies) and incubated at 37\(^\circ\)C for 4 h to allow macrophage adherence. AM were washed and labeled with 2 \(\mu\)M CellTracker Red (Molecular Probes) for 30 min at 37\(^\circ\)C. Apoptosis was induced by exposure to 120 mJ/cm\(^2\) irradiation (Stratalinker 1800; Stratagene). Twelve hours post-UV treatment, AM were gently scraped and washed in PBS. After this treatment, mean values for AM were 69% Annexin V\(^-\), 8% ToPro3\(^+\), and 65% showed loss of \(\Delta m\) (9, 11). In additional experiments, AM were made apoptotic by treatment with 10 \(\mu\)M staurosporine for 16 h. Cells were resuspended in PBS at 5 \(\times\) 10\(^5\) cells/ml. A total of 1 \(\times\) 10\(^4\) apoptotic AM were delivered to the lungs by direct intratracheal instillation immediately after pneumococcal infection. This number was chosen to provide approximately enough apoptotic cells to restore the numbers of apoptotic cells in the iNOS\(^{-/-}\) mice to the level observed in the C57BL/6 mice. Twelve to 48 hours postinfection, mice were killed and BAL performed. To confirm phagocytosis of apoptotic bodies, resident AM of some C57BL/6 mice were labeled with PKH2 green fluorescent phagocytic cell linker compound (Sigma-Aldrich) (18). PKH2 dye (stock 1 \(\times\) 10\(^{-3}\) M) was diluted 1/5 with diluted B and 100 \(\mu\)l was administered i.v. Forty-two hours, post-PKH2 treatment apoptotic AM were delivered to the lungs by direct intratracheal instillation. Lungs were lavaged 30 min postinstillation and cytokine preparations made from lung cells. Coverslips were mounted with VectaShield containing 4,6-diamidino-2-phenylindole (DAPI; Vector Laboratories), and the cells imaged with a DeltaVision Microscope.

**Ex vivo AM pneumococcal infection**

C57BL/6 and iNOS-deficient mice were killed with an overdose of sodium pentobarbitone and BAL was performed as described using RPMI 1640 (Invitrogen Life Technologies) plus 10% heat-inactivated FCS (Bioclear), 100 \(\mu\)g/ml streptomycin, and 100 U/ml penicillin (Invitrogen Life Technologies) (9). BAL was spun for 5 min at 1000 \(\times\) g and cells resuspended in RPMI 1640 plus 10% heat-inactivated FCS without antibiotics and incubated at 37\(^\circ\)C for at least 4 h to allow macrophage adherence. Infection was with type 1 pneumococci opsonized with serum from mice immunized with pneumovax vaccine at a multiplicity of infection of 10 (5). Killing was assessed by analyzing colony counts in supernatants at 4 and 20 h postinfection (21). Apoptosis was assessed at 20 h postinfection by nuclear morphology after DAPI staining as described (21).

**Statistics**

Results are recorded as mean and SEM. Survival was calculated by Kaplan-Meier followed by log-rank analysis. Parametric or nonparametric testing was performed with the indicated tests using Prism 4.0 software (GraphPad). Significance was defined as \(p < 0.05\).

**Results**

**AM apoptosis is reduced in iNOS\(^{-/-}\) mice during pneumococcal infection**

Because NO contributes to apoptosis of human macrophages during pneumococcal infection in vitro (11), but important differences exist between rodent and human macrophages with regard to the level of NO produced (22), we first confirmed that NO also played a role in apoptosis in murine AM when cultured ex vivo and challenged with pneumococci. As shown, murine AM from iNOS\(^{-/-}\) mice demonstrated lower levels of apoptosis (Fig. 1A) and decreased killing of bacteria (Fig. 1B) as compared with wild-type cells, thus confirming a similar phenotype to that previously found in human macrophages (11). We next examined levels of macrophage apoptosis in vivo in iNOS\(^{-/-}\) mice (15). Pneumococcal-infected C57BL/6 mice had significantly greater percentages of apoptotic cells in BAL in comparison to mock-infected mice (Fig.
Pneumococcal-infected iNOS−/− mice, however, had significantly fewer apoptotic cells in BAL than did pneumococcal-infected C57BL/6 mice (Fig. 2A). Because the BAL fluid from the pneumococcal-infected mice contained significant numbers of neutrophils at each time point, we determined the level of apoptosis in the macrophage population by flow cytometry (9). There were greater percentages of apoptotic macrophages in pneumococcal-infected C57BL/6 mice than in mock-infected mice, but the levels of apoptotic macrophages were significantly decreased in iNOS−/− mice as opposed to C57BL/6 mice (Fig. 2B). When apoptosis was measured in neutrophils, we also found evidence of decreased neutrophil apoptosis in iNOS−/− mice as opposed to C57BL/6 mice after pneumococcal infection (Fig. 2C).

Effect of iNOS deficiency on neutrophil numbers in BAL
There was a significant increase in the number of neutrophils in BAL from iNOS−/− mice as compared with C57BL/6 mice 24–48 h postinfection (Fig. 3A). Although there was evidence of increased neutrophil viability in iNOS−/− mice (reduced neutrophil apoptosis (Fig. 2C) and no increase in necrosis, data not shown), increased recruitment was also likely to contribute to increased neutrophil numbers. The iNOS−/− mice had no significant increase in bacteria in the lung at 12–48 h (Table I), excluding the possibility that a significantly greater bacterial load was the stimulus for neutrophil recruitment. Similarly, peripheral blood neutrophil counts were similar between C57BL/6 mice and iNOS−/− mice, suggesting an intrinsic difference in neutrophil numbers did not explain the differences in numbers of BAL neutrophils after infection, data not shown.

Neutrophil recruitment in pneumococcal infection demonstrates important differences as compared with other stimuli such as LPS with a prominent upstream role for TNF-α (23). We performed RNase protection assays on lungs 12 h after infection to identify cytokines and chemokines that were up-regulated in iNOS−/− mice and were able to demonstrate significant up-regulation of TNF-α message (Fig. 3, B and C). We also observed up-regulation of mRNA for the chemokines KC and MCP3 (Fig. 3, B and C), chemokines known to be associated with neutrophil recruitment in pneumococcal infection (23) and in iNOS deficiency (24). Measurement of cytokines in BAL revealed iNOS−/− mice had a significant increase in TNF-α production relative to C57BL/6 mice 12 h postinfection and a nonsignificant trend toward greater production at 24 h following pneumococcal infection (Fig. 3D). Levels of TNF-α in PBS-treated mice of both strains were below the level of detection. KC values in BAL fluid 12 h after infection from iNOS−/− vs C57BL/6 mice were 803 ± 28.6 vs 578.8 ± 96.58 pg/ml (p < 0.05, n = 4).

Pharmacologic inhibition of iNOS modulates macrophage apoptosis and neutrophil recruitment
To investigate whether these findings were related to decreased production of NO and/or one of its reaction products or whether they resulted from some nonspecific effect associated with iNOS deficiency, for example, increased levels of a cofactor or metabolite required for the functional activity of iNOS, we inhibited iNOS activity pharmacologically. Inhibition with aminoguanidine, a specific inhibitor of iNOS (25), replicated the findings in iNOS−/− mice with decreased total or macrophage apoptosis and increased neutrophil numbers or TNF-α levels in BAL all observed (Fig. 4).

iNOS−/− mice demonstrate enhanced levels of lung inflammation and bacteremia
The increased numbers of neutrophils and proinflammatory cytokines in the lung in iNOS−/− mice were associated with enhanced early mortality in the first 42 h after infection, 22.0% iNOS−/− vs 4.9% C57BL/6 mice, p < 0.05. Fisher’s exact test, but this was not associated with a significant decrease in clearance of bacteria from...
the lung (Table I). However, by 10 days there was no difference in overall mortality in the two groups of mice 54% iNOS<sup>/−</sup> vs 48% C57BL/6 mice (Fig. 5A). We have previously shown that decreased levels of macrophage apoptosis are associated with invasive pneumococcal disease (5, 9) and in keeping with this finding iNOS<sup>/−</sup> mice had >1 log higher bacterial colony counts in the blood at 12–48 h after infection (Fig. 5B). IL-6 production is a marker of sepsis-related mortality and poor outcomes in models of pneumococcal disease (26) and increased IL-6 production in the lungs of pneumococcal-infected mice was apparent 48 h after infection (Fig. 5B). The phagocytic capacity of AM for apoptotic cells in vitro is low in comparison to the capacity to ingest opsonized particles (18) or the capacity of peritoneal macrophages to phagocytose apoptotic cells (27). Nevertheless, macrophages with internalized apoptotic cells are observed in BAL from mice with pneumococcal infection (9). We estimated the percentage of extracellular apoptotic cells per cytospin, i.e., apoptotic cells that had not been phagocytosed by macrophages (Fig. 7A), and the percentage of intracellular apoptotic cells (Fig. 7B). Although, over all the time points studied, only a relatively low percentage of macrophages (C57BL/6 4.4 ± 0.3%; iNOS<sup>/−</sup> 5.7 ± 0.5%) contained apoptotic cells these apoptotic cells accounted for a significant percentage of the total apoptotic events (C57BL/6 44 ± 2.4%; iNOS<sup>/−</sup> 55.7 ± 3.5%), arguing in favor of a relatively efficient clearance mechanism for apoptotic cells in vivo during pneumococcal infection. As illustrated, there were a lower percentage of intracellular apoptotic bodies in iNOS<sup>/−</sup> mice at all time points (Fig. 7B).

**Instillation of apoptotic macrophages into the lung corrects markers of greater lung inflammation in iNOS<sup>/−</sup> mice**

Because ingestion of apoptotic cells can down-regulate macrophage production of proinflammatory cytokines (13), we tested whether exogenous apoptotic macrophages could decrease the pulmonary inflammation observed during pneumococcal infection. UV treatment induces high levels of macrophage apoptosis (5) and efficiently induced AM apoptosis. A suspension of apoptotic AM had no residual antimicrobial effect as there was no difference between bacterial colony counts in the lung 24 h after infection in the presence or absence of these cells: median and interquartile range for C57BL/6 mice in the presence of apoptotic cells 1.6 ± 0.8 × 10<sup>8</sup> vs 1.0 ± 0.3 × 10<sup>8</sup> CFU (8.3 ± 10<sup>5</sup>–8.3 × 10<sup>5</sup>) in the absence of apoptotic cells (n = 11, p = 0.81), Mann-Whitney. Instilled Cell Tracker Red-labeled apoptotic AM were phagocytosed by resident AM labeled with the green fluorescent dye, PKH2 (Fig. 8A). The greater numbers of recruited neutrophils in the lungs of iNOS<sup>/−</sup> mice could be the direct result of NO deficiency increasing production of proinflammatory cytokines (28, 29). However, instillation of apoptotic AM decreased inflammation in the lung (Fig. 8B). There were decreased numbers of recruited neutrophils in the lung 12–48 h after instillation of apoptotic macrophages in iNOS<sup>/−</sup> mice and to a lesser extent in C57BL/6 mice (Fig. 8B). In the iNOS<sup>/−</sup> mice, instillation of apoptotic AM reduced the numbers of neutrophils in the lung to a level comparable to the number that were recruited into C57BL/6 mice.

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**Table I. Microbiological outcomes**

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<td>C57BL/6</td>
<td>1.8 × 10&lt;sup&gt;5&lt;/sup&gt; (1.5 × 10&lt;sup&gt;5&lt;/sup&gt;–3.1 × 10&lt;sup&gt;5&lt;/sup&gt;)</td>
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<td>iNOS&lt;sup&gt;/−&lt;/sup&gt;</td>
<td>2.0 × 10&lt;sup&gt;6&lt;/sup&gt; (1.2 × 10&lt;sup&gt;5&lt;/sup&gt;–2.8 × 10&lt;sup&gt;6&lt;/sup&gt;)</td>
<td>1.7 × 10&lt;sup&gt;6&lt;/sup&gt; (8.3 × 10&lt;sup&gt;5&lt;/sup&gt;–2.5 × 10&lt;sup&gt;6&lt;/sup&gt;)</td>
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*Median (interquartile range).*

*Percent with detectable bacteria.*
mice not instilled with apoptotic AM. Results were identical regardless of whether AM were derived from iNOS\(^{-/-}\) or C57BL/6 mice and in preliminary data if AM were made apoptotic by staurosporine treatment as opposed to UV exposure (data not shown). Therefore, decreased numbers of apoptotic macrophages contribute to the increased lung inflammation during pneumococcal infection of iNOS\(^{-/-}\) mice and instillation of additional apoptotic macrophages reverses the proinflammatory phenotype associated with NO deficiency following pulmonary challenge with pneumococci.

**Effect of apoptotic cells on TNF-\(\alpha\) expression in iNOS\(^{-/-}\) mice**

In keeping with the pivotal role of TNF-\(\alpha\) expression in recruitment and activation of neutrophils during pneumococcal pneumonia (23), we found that TNF-\(\alpha\) mRNA expression in the lung was reduced by instillation of apoptotic cells in iNOS\(^{-/-}\) mice (Fig. 9, A and B). Similar findings were observed by qRT-PCR (Fig. 9C). This was confirmed by significant reduction in TNF-\(\alpha\) expression

[FIGURE 4. iNOS inhibition reduces apoptosis and increases pulmonary inflammation after pneumococcal infection. A, The percentage of apoptotic events (apoptotic cells and bodies) in cytospins of bronchial alveolar lavage (BAL) from C57BL/6 mice 24 h after instillation of 10^7 CFU of type 2 pneumococci (Spn), in the absence (H\(_2\)O) or presence of aminoguanidine (AG) treatment. B, Representative dot plots to show the percentage macrophage apoptosis (annexin V-PE/ToPro 3\(^{-}\) cells as determined by flow cytometry. C, Percentage of apoptotic macrophages. D, Number of recruited neutrophils (#PMN); E, TNF-\(\alpha\) concentration in BAL in the same experiments as A; n = 8. Mean + SEM, **, \(p < 0.01\), *** \(p < 0.001\), \(t\) test; ■, H\(_2\)O; dark gray box, AG.]

[FIGURE 5. Increased lung inflammation in iNOS\(^{-/-}\) mice after pneumococcal infection. A, Survival of wild-type iNOS-sufficient mice (C57BL/6) and iNOS-deficient (iNOS\(^{-/-}\)) mice after intratracheal instillation of 10^7 CFU of type 2 pneumococci, n = 23–24/group. B, Concentration of bacteria in blood of C57BL/6 and iNOS\(^{-/-}\) mice 12 and 48 h after intratracheal instillation of 10^7 CFU of type 2 pneumococci. The percentage of C57BL/6 vs iNOS\(^{-/-}\) mice with bacteremia at 12 h was 53 vs 82\% and at 48 h was 61 vs 81\%. C and D, Concentration of (C) IL-6 and (D) protein in bronchial alveolar lavage (BAL) 48 h after intratracheal instillation of 10^7 CFU of type 2 pneumococci, n = 8–16. Mean + SEM, *, \(p < 0.05\), **, \(p < 0.01\), *** \(p < 0.001\), Mann-Whitney; ■, C57BL/6; □, iNOS\(^{-/-}\).]

[FIGURE 6. Histologic appearance of increased lung inflammation in iNOS\(^{-/-}\) mice after pneumococcal infection. Representative appearances of lung sections stained with H&E from C57BL/6 and iNOS-deficient mice (iNOS\(^{-/-}\)) 48 h after intratracheal instillation of 10^7 CFU of type 2 pneumococci as viewed by \(\times 10\) and \(\times 40\) objective. Images were obtained from one of four mice in each group reviewed by two independent pathologists.
by ELISA, following instillation of apoptotic cells to iNOS−/− mice (Fig. 9D). These alterations were persistent for up to 48 h after infection (data not shown). Further screen of lung RNase protection assays and BAL ELISAs showed other changes in early cytokine/chemokine expression such as changes in IL-1 isoforms, MCP3 (data not shown), KC (Fig. 10A), and MIP-2 (Fig. 10B), but these differences were not as marked as for TNF-α expression. IL-6 levels were markedly reduced following instillation of apoptotic cells (Fig. 10C). Further support for the beneficial effect of phagocytosis of apoptotic macrophages in iNOS−/− mice infected with pneumococci comes from the observation that reduction in the number of neutrophils and cytokines such as TNF-α and IL-6 expression was associated with a decreased likelihood of mice developing invasive pneumococcal disease after instillation of apoptotic cells (Fig. 11).

**Discussion**

We demonstrate decreased rates of macrophage apoptosis in the lungs of iNOS−/− and aminoguanidine-treated mice during pneumococcal infection. The decrease in macrophage apoptosis in iNOS−/− mice is associated with development of earlier bacteremia and death, and also with increased markers of inflammation in the lung. Instillation of apoptotic macrophages into the lungs of iNOS−/− mice reversed the features of increased inflammation and reduced the development of invasive bacterial disease.

The potential benefits of host-mediated macrophage apoptosis in the innate response to infection are not fully understood. Apoptosis is a mechanism which removes unwanted cells, thus limiting inflammation and tissue injury (30). It is plausible that macrophage apoptosis during bacterial infection plays a role in down-regulating the inflammatory response in addition to its role in enhancing microbial killing (5). The innate response to pneumococci involves pulmonary inflammation (31), induction of host-mediated macrophage apoptosis (9) and a requirement for resolution of the inflammatory response for a successful outcome (12). Because iNOS−/− mice have decreased levels of macrophage apoptosis during pneumococcal infection, we anticipated this model would provide insights into the relationship between regulation of inflammation and induction of macrophage apoptosis.

The roles of NO in murine models of pneumococcal pneumonia have been conflicting reflecting the variety of strains of mice studied, varying doses and strains of bacteria, and the use of pharmacologic agents as opposed to genetically modified mice (32–34). Our findings with type 2 pneumococci have been reproduced with type 1 pneumococci, demonstrating that the findings of less macrophage apoptosis and greater neutrophil infiltration and TNF-α expression in the lung were not specific to the strain we used (data not shown). Although we have previously found that NO contributes to macrophage killing of pneumococci (11), we have also demonstrated that during low-dose infection other elements of the host response, including recruited neutrophils can compensate for decreased bacterial killing by AM (9). Clearly NO is only one factor involved in both microbicidal killing and AM apoptosis induction and other factors also contribute. Furthermore, NO also contributes to host defense by mechanisms independent of its effects on macrophage function (28, 35). On the basis of our findings of increased inflammation in iNOS−/− mice, we cannot exclude a role for NO produced in other cells for the phenotype we observed. However, we show that removal of one factor involved in host defense resets the AM response, and indeed the total innate host response in the lung, leading to an altered threshold at which the next element in the host response becomes critical to controlling infection. As Kerr et al. (34) have clearly illustrated at intermediate doses of pneumococci (albeit in mice of a different genetic background to those we studied), host defense is unable to compensate for NO deficiency and larger numbers of bacteria are recovered.
from the lungs of iNOS−/− mice. In the current study, we have used a high dose of pneumococci and have been unable to document any overall defect in bacterial killing in the lungs but have demonstrated greater lung inflammation, consistent with prior fulminating infection models using NO inhibitors (33). Our inability to demonstrate a clear microbiologic or survival difference in iNOS−/− mice suggests significant redundancy and compensation exists in this aspect of innate immunity in the lung and also that, at the high doses we used, the compensatory measures are overwhelmed so they are no longer the critical determinant of outcome in the setting of iNOS deficiency.

Nevertheless, decreased macrophage apoptosis, in the setting of decreased NO production, was associated with increased levels of bacteremia, in keeping with our previous observation that inhibition of macrophage apoptosis is associated in particular with increased levels of invasive pneumococcal disease (5, 11). Importantly, instillation of apoptotic AM in iNOS−/− mice reversed many of the features of increased inflammation, even though these mice had the same defects in NO production in macrophages and other cells as the iNOS−/− mice that did not receive apoptotic cells. This suggests that a relative deficiency in numbers of apoptotic cells and their clearance is a significant contributory factor to increased lung inflammation and bacterial tissue invasion. Nevertheless, the pathogenesis of bacteremia is multifactorial and many other factors play a role.

NO has a wide range of actions including cell signaling function (36). The proinflammatory effects of NO deficiency in the lung have been demonstrated both by pharmacologic inhibition of iNOS, and also by study of iNOS−/− mice (37). NO modifies
levels of the transcription factor NF-κB (38) and decreases NF-κB binding to the regulatory region of proinflammatory cytokine genes such as TNF-α and IL-6 (29). It remains possible the effects we observed are mediated by a reaction product of NO not by NO itself. However, pharmacologic inhibition of iNOS replicated the phenotype of iNOS−/− mice, arguing in favor of mediation by NO or a reactive nitrogen species rather than a nonspecific result of deletion of the iNOS gene (35).

In addition to its signaling role, NO also plays a role in the regulation of apoptosis (39). Ingestion of apoptotic bodies by macrophages modifies cytokine production following stimulation with microbial products (14). Although phagocytosis of apoptotic bodies in the unstimulated macrophage results in production of anti-inflammatory cytokines such as TGF-β (13), the effect of their ingestion in the context of stimulation of TLRs in vitro is to shorten the duration of proinflammatory cytokine production (14). We instilled apoptotic AM to address whether decreased levels of phagocytosis of apoptotic macrophages contributes to decreased pulmonary inflammation during pneumococcal infection and confirmed this was the case. We conclude that one mechanism by which NO deficiency contributes to lung inflammation is by decreasing macrophage apoptosis and one consequence of this is that macrophages are less likely to phagocytose apoptotic cells and therefore reset their cytokine expression profile. Interestingly, we previously demonstrated that AM depletion also increased neutrophil recruitment during pneumococcal infection in vivo (9). Although AM depletion would be anticipated to decrease total NO production it would also decrease the number of AM undergoing apoptosis during infection and the number of AM phagocytosing apoptotic cells. Although absolute NO deficiency or high level AM deficiency is unlikely during bacterial pneumonia, our findings raise the possibility that subtle manipulation of NO generation or levels of macrophage apoptosis could exert beneficial effects on the degree of pulmonary inflammation during bacterial pneumonia. Although the clearance of apoptotic cells alters cytokine expression profiles, these data have been generated in vitro using apoptotic neutrophils or lymphocytes (40, 41). This study provides, to our knowledge, the first data on the role of apoptotic macrophages in the regulation of the inflammatory phenotype during a model of pneumonia. Importantly, we have replaced the same cell type as our knowledge, the first data on the role of apoptotic macrophages in the regulation of the inflammatory phenotype during a model of pneumonia. Importantly, we have replaced the same cell type as in the regulation of the inflammatory phenotype during a model of pneumonia. Importantly, we have replaced the same cell type as in the regulation of the inflammatory phenotype during a model of pneumonia. Importantly, we have replaced the same cell type as in

In conclusion, we provide evidence, in a model of pneumonia, that phagocytosis of apoptotic AM is associated with reduced TNF-α expression, neutrophil recruitment, and invasive pneumococcal disease. These studies further define the role of host-mediated macrophage apoptosis during bacterial infection and highlight its impact on microbiologic and inflammatory outcomes.

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Disclosures
The authors have no financial conflict of interest.

References


6488 MACROPHAGE APOPTOSIS REDUCES INFLAMMATION IN PNEUMONIA