Neutrophils Are a Key Component of the Antitumor Efficacy of Topical Chemotherapy with Ingenol-3-Angelate

Jodie M. Challacombe, Andreas Suhrbier, Peter G. Parsons, Brad Jones, Peter Hampson, Dean Kavanagh, G. Ed Rainger, Melanie Morris, Janet M. Lord, Thuy T. T. Le, Diem Hoang-Le and Steven M. Ogbourne

*J Immunol* 2006; 177:8123-8132; doi: 10.4049/jimmunol.177.11.8123
http://www.jimmunol.org/content/177/11/8123

---

**References**

This article cites 52 articles, 23 of which you can access for free at: http://www.jimmunol.org/content/177/11/8123.full#ref-list-1

**Subscription**

Information about subscribing to *The Journal of Immunology* is online at: http://jimmunol.org/subscription

**Permissions**

Submit copyright permission requests at: http://www.aai.org/About/Publications/JI/copyright.html

**Email Alerts**

Receive free email-alerts when new articles cite this article. Sign up at: http://jimmunol.org/alerts
Neutrophils Are a Key Component of the Antitumor Efficacy of Topical Chemotherapy with Ingenol-3-Angelate¹

Jodie M. Challacombe,2* Andreas Suhrbier,2,3* Peter G. Parsons,*, Brad Jones,*, Peter Hampson,† Dean Kavanagh,† G. Ed Rainger,† Melanie Morris,* Janet M. Lord,† Thuy T. T. Le,* Diem Hoang-Le,* and Steven M. Ogbourne*

Harnessing neutrophils for the eradication of cancer cells remains an attractive but still controversial notion. In this study, we provide evidence that neutrophils are required to prevent relapse of skin tumors following topical treatment with a new anticancer agent, ingenol-3-angelate (PEP005). Topical PEP005 treatment induces primary necrosis of tumor cells, potently activates protein kinase C, and was associated with an acute T cell-independent inflammatory response characterized by a pronounced neutrophil infiltrate. In Foxn1nu mice depleted of neutrophils and in CD18-deficient mice (in which neutrophil extravasation is severely impaired) PEP005 treatment was associated with a >70% increase in tumor relapse rates. NK cell or monocyte/macrophage deficiency had no effect on relapse rates. Both in vitro and in mice, PEP005 induced MIP-2/IL-8, TNF-α, and IL-1β, all mediators of neutrophil recruitment and activation. In vitro, PEP005 activated human endothelial cells resulting in neutrophil adhesion and also induced human neutrophils to generate tumoricidal-reactive oxygen intermediates. Treatment of tumors with PEP005 significantly elevated the level of anticancer Abs, which were able to promote neutrophil-mediated Ab-dependent cellular cytotoxicity (ADCC) in vitro. PEP005 treatment of tumors grown in SCID mice was also associated with >70% increase in tumor relapse rates. Taken together, these data suggest a central role for neutrophil-mediated ADCC in preventing relapse. PEP005-mediated cure of tumors therefore appears to involve initial chemoablation followed by a neutrophil-dependent ADCC-mediated eradication of residual disease, illustrating that neutrophils can be induced to mediate important anticancer activity with specific chemotherapeutic agents. The Journal of Immunology, 2006, 177: 8123–8132.

The Journal of Immunology, 2006, 177: 8123–8132.

The compound ingenol-3-angelate (PEP005) is currently in clinical trials for skin cancer and solar keratoses. PEP005 is a diterpene ester extracted from the plant Euphorbia peplus, the crude sap of which has a long history of use for various ailments including self-treatment of skin cancers (11). We have shown previously that daily topical applications of PEP005 for 3 days to a number of mouse and human tumors grown s.c. in both C57BL/6 and Foxn1nu mice resulted in tumor cure without significant relapse (12), indicating that T cells are not required for effective anticancer activity. A notable feature of topical PEP005 treatment was induction of an acute erythema, which was associated with a significant infiltration of inflammatory cells, predominantly neutrophils. At high concentrations, PEP005 is acutely cytotoxic to tumor cells, causing cell death by induction of primary necrosis (12). At lower concentrations, PEP005 activates protein kinase C (PKC) (13) and PKC activators are known to stimulate a number of immune responses including induction of proinflammatory cytokines and neutrophil activation (14–16). In this study, we provide evidence that neutrophils are required to prevent relapse following topical treatment with PEP005. Relapse rates were high in two animal models where neutrophil activity was deficient, and both in vitro and in vivo PEP005 was shown to induce mediators associated with neutrophil recruitment and activation. The high relapse rates in SCID mice (B cell deficient), the induction of anticancer Abs by PEP005 treatment, and the ability of such Abs to mediated ADCC with neutrophils in vitro, support the view that neutrophil-mediated ADCC is involved in preventing relapse. Thus, PEP005 appears to have a two stage mechanism of action, initial chemoablation followed by a neutrophil-mediated ADCC-dependent eradication of residual disease.
Materials and Methods

Cells and cell culture

The B16 melanoma line (CRL-6322; American Type Culture Collection (ATCC)), the LK2 UV-induced mouse squamous cell carcinoma line (17), and the human melanoma lines MM96L (18) and Mel10538 (19) were cultured at 37°C and 5% CO₂ in RPMI 1640 medium (Invitrogen Life Technologies) supplemented with 10% FCS (CSL Biosciences), 100 μg/ml streptomycin, and 100 IU/ml penicillin (Invitrogen Life Technologies) (complete medium). Human epidermal keratinocytes were isolated from newborn foreskin and cultured in the presence of a mitomycin C-treated 3T3 feeder layer as described previously (20). Human skin fibroblasts were isolated from tissue digests taken at the time of joint replacement surgery by Percoll density centrifugation as described previously (22) and purity was assessed by Giemsa staining and was routinely >95%.

PEP005 treatment and histology

LK2 cells (10⁶) were injected s.c. (two tumor sites per mouse and two mice per group) into the flank of 6- to 10-week-old Fostam (BALB/c) mice (Animal Resource Centre, Perth, Australia). The tumors and two patches of normal skin on the opposite flanks were excised. The tissue was cryopreserved, sectioned, and processed for histology and standard H&E staining post-PEP005 treatment and treated sites were excised, formalin fixed, paraffin wax embedded, and processed for histology and standard H&E staining. Slides were examined using the ScanScopeT2 slide scanner (Aperio Technologies). Approval for all animal experiments was obtained from the Queensland Institute of Medical Research Animal Ethics Committee.

PEP005 treatment in neutrophil-depleted mice

LK2 cells (10⁶) were injected s.c. (four tumor sites per mouse) into the flanks of 6- to 10-week-old Fostam mice (n = 3 mice and n = 12 tumors/group). After 2 wk (when the tumors had reached ~14 mm³), six mice were given i.p. injections of anti-Ly-6G anti-granulocyte Ab (100 μg in PBS) (RB6-8C5; BD Biosciences). Another six mice were injected i.p. with an isotype-matched control Ab (100 μg in PBS) (A95-1; BD Biosciences). Ab was injected on days −2, 0, and 2, relative to initiation of PEP005 treatment and treated sites were excised, formalin fixed, paraffin wax embedded, and processed for histology and standard H&E staining. Slides were examined using the ScanScopeT2 slide scanner (Aperio Technologies). Approval for all animal experiments was obtained from the Queensland Institute of Medical Research Animal Ethics Committee.

PEP005 treatment in CD18-deficient mice

B6.129S7-Il6g2tm1hoxjj mice (The Jackson Laboratory) are CD18 hypomorphic mice on the B6 background and have 2–10% of normal CD18 expression on their neutrophils (23). B6 mice (5 x 10⁵) were injected s.c. (one tumor site per mouse) into the flanks of 6- to 10-week-old Fostam mice (n = 3 mice and n = 12 tumors/group) and after 2 wk the tumors had reached ~14 mm³, six mice were treated with PEP005 or placebo. Tumor and erythema size were measured as above.

PEP005 treatment in NK-depleted mice

LK2 cells (10⁶) were injected s.c. (four tumor sites per mouse) into the flanks of 6- to 10-week-old Fostam mice (n = 3 mice and n = 12 tumors/group). After 2 wk when the tumors had reached ~14 mm³, mice were treated with PEP005 or placebo (on days 0, 1, and 2) and tumor volumes were monitored as above. The mice received on days −2 and −1 (relative to initiation of PEP005 treatment) anti-asialo GM1 monoclonal antiserum (50 μl per Cediq Laboratories). NK depletion (~90%) was confirmed 2 days after Ab administration using splenocytes from a parallel group of animals and FACS analysis (using the pan NK Ab DX5; BD Biosciences/BD Pharmingen).

PEP005 treatment in op/op mice

B16 cells (5 x 10⁵) were injected s.c. (four tumor sites) into the flanks of 6- to 10-week-old op/op (csf1m/csf1m⁻) mice (Herston Medical Research Centre, Brisbane, Australia) (n = 3 mice and n = 12 tumors/group). Tumors were treated with PEP005 or placebo and monitored as above.

PEP005 treatment in SCID mice

LK2 cells (10⁶) were injected s.c. (four tumor sites per mouse) into the flanks of 6- to 10-week-old SCID mice (Animal Resource Centre) (n = 3 mice and n = 12 tumors/group). Tumors were treated with PEP005 or placebo and monitored as above.

Statistical analysis

SPSS 13.0 for Windows (release 13.0) was used for statistical analysis. Cox regression analysis was used to compare relapse rates between different groups and was adjusted for the mouse effect when four tumors per mouse were used. The analysis showed that the influence of mouse on time to relapse was not significant (p = 0.88 for Fig. 2C and p = 0.315 for Fig. 4D), illustrating that each tumor essentially behaves independently.

In vivo cytokine responses

LK2 cells were injected, as above, into the flanks of 6- to 10-week-old Fostam mice (two tumor sites per mouse). After 2 wk (when the tumors had reached ~14 mm³), the tumors and two skin sites on the opposite flanks of the animals were treated once with PEP005 as above. Parallel mice (n = 1/time point) were euthanized 0, 1, 2, and 6 h after treatment, and the tumor and skin sites were excised. Total RNA was extracted and purified from skin and tumor tissue as per the manufacturer’s instructions (RNeasy Protect-midi kit; Qiagen). Total RNA (4 μg) was reverse transcribed using (Oligo d(T)₃) and Superscript III (Invitrogen Life Technologies). PCR product intensity after gel electrophoresis and staining was determined to be linear for the number of cycles used, thus the analysis was deemed to be semiquantitative. The PCR was conducted using the Gene-Amp PCR System 9700 (PerkinElmer), Dynazyme II DNA polymerase (Finnzymes), and Hot Star TaqDNA polymerase (Qiagen) as described previously (24). The primers were IL-β, 5'-CAG GAT GAG ATG ATC ACC AGC ACC, 3'-CTC TGC AGA CTC AAA CTC CAC; TNF-α, 5'-CCA GAC CCT CAC ACT CAG AT; 3'-GGT AGA GAA TGG ATG AAC AC; GAPDH, 5'-TGA AGG TCG GTG TGA ACG GAT TTG GC, 3'-CAG GTA GGC CAT GAT GTC CAC GAC; macrophage inflammatory protein 2 (MIP-2) 5'-TCC AGA CTC CAG CCA CAC TCC AGC, 3'-TCT CAG ACA GCC AGG ACC ATC AGG.

Cytokine production by PEP005-treated human cells

Cells of Mur1538 cells, keratinocytes, fibroblasts (75% confluent at ~5 x 10⁵ cells/well of a 24-well plate) and neutrophils (10⁵ cells/well of a 96-well plate) were incubated for 6 h in the absence or presence of PEP005 (1–100 ng/ml). The supernatants were harvested and analyzed for the presence of the following cytokines: TNF-α, IL-1β, IL-2, IL-4, IL-6, IL-8, IL-10, IL-12, and GM-CSF using a multiplex detection kit (BioSource International).

PEP005-induced activation of vascular endothelium

Adherence of neutrophils to vascular endothelium was measured using a standard static adhesion assay (25). First passage HUVECs were grown on glass coverslips in the wells of 24-well plates using Medium 199 (Invitrogen Life Technologies) supplemented with 20% FBS (Sigma-Aldrich), 1 ng/ml epidermal growth factor (Sigma-Aldrich), 1 μg/ml hydrocortisone (Sigma-Aldrich), 28 μg/ml gentamicin (David Bull Laboratories), and 2.5 μg/ml amphotericin B (Sigma-Aldrich). PEP005 was added to culture wells at 1–100 ng/ml and TNF-α was used as a positive control at 100 U/ml. Endothelial cells were exposed to TNF-α or PEP005 for 4 h before thorough washing to remove PEP005 or TNF-α. Human neutrophils (7.5 x 10⁵/well) were then added to the endothelial cells and allowed to adhere for 5 min at 37°C. Cells were then washed, fixed in 2% glutaraldehyde, and examined by phase contrast microscopy to determine the number of neutrophils adhering to the endothelial cell monolayer.

MM96L cell killing by neutrophils in vitro

Human neutrophils were added to MM96L human melanoma cells (5000 cells/0.1 ml of a 24-well plate in triplicate), with and without 10 ng/ml PEP005. After 24 h, the cultures were washed with PBS to remove neutrophils and PEP005 and were then maintained for a further 6 days in complete medium. The cells were washed with PBS, fixed in methanol, and the total
protein of the adherent MM96L cells was determined using sulforhodamine B as described previously (26). Cell survival was expressed as a percentage of total protein measured from wells containing only melanoma cells.

**Lytic mediator release by neutrophils in vitro**

Neutrophils (10^5) were treated with 10 ng/ml PEP005 for 2 h and assayed for generation of 1) superoxides measured using a lucigenin-based assay (27), 2) release of soluble TRAIL measured by ELISA using a commercial kit (BioSource International), and 3) release of defensins 1–3, measured by ELISA using a commercial kit (Cell Sciences).

**Anti-cancer Ab measurements**

B16 cells (10^6) were injected s.c. (one tumor per mouse) into the flanks of 6- to 10-wk-old C57BL/6 mice (Animal Resource Centre) and when tumors had reached ~14 mm^3 they were treated with PEP005 as above. On days 11 and 135, sera was taken and analyzed by ELISA for Abs specific for B16. A group of B16-bearing animals, which were not treated with PEP005 and a naive group were included. B16 cells were sonicated in carbonate buffer (pH 9) and absorbed onto Immuno Maxisorp 96-well plates (Nunc) overnight and dried. The plates were blocked with 5% FBS, 0.01% Tween 20 in PBS for 1 h at 37°C. Test sera was serially diluted in duplicate and probed with rat anti-mouse biotinylated primary Ab (BD Biosciences/BD Pharmingen) and HRP-labeled streptavidin (BioSource International) followed by ABTS substrate (Sigma-Aldrich) and measurement of OD at 405 nm.

**In vitro neutrophil ADCC assay**

LK2 tumors (~14 mm^3) were established in Foxn1nu mice (four tumor sites per mouse) and were treated with PEP005 as described above. On day 11 after treatment initiation, antisera was collected by heart puncture and treated area (Fig. 1, C). Skin obtained 6 h after topical application of PEP005 showed decreased integrity of hair follicles and sebaceous glands, as well as increased dilation of local blood vessels. An increase in neutrophil numbers was observed in the treated area (Fig. 1, E and F). Similar results were also seen in the PEP005-treated tumor sites at this time (Fig. 1, G and H). Twenty-four hours after treatment with PEP005, a large number of neutrophils and some macrophages were found in treated skin (Fig. 1, I and J), and in and around the treated tumor site, with neutrophils abundant at the periphery of the tumor. Neutrophils can be seen as the small spherical and uniform cells with red/pink cytoplasm and irregular blue nuclei. Areas with a pronounced neutrophil infiltrate are encircled (Fig. 1 F, H, J, and L). Some hemorrhage of local blood vessels was also apparent at this time (Fig. 1, K and L). High magnification images of areas at the periphery of the tumor following placebo treatment show intact tumor cells, RBC, and few leukocytes (Fig. 1M). In contrast, high magnification images of areas at the periphery of the tumor following PEP005 treatment...

**Results**

**Neutrophil recruitment and tissue morphology after PEP005 treatment**

The UV-induced murine squamous cell carcinoma line LK2 was grown as s.c. tumors in Foxn1nu mice. H&E staining of tumor sites and normal skin 24 h after topical treatment with placebo (isopropanol gel) revealed normal morphology with small numbers of leukocytes present (Fig. 1, A–D). Skin obtained 6 h after topical application of PEP005 showed decreased integrity of hair follicles and sebaceous glands, as well as increased dilation of local blood vessels. An increase in neutrophil numbers was observed in the treated area (Fig. 1, E and F). Similar results were also seen in the PEP005-treated tumor sites at this time (Fig. 1, G and H). Twenty-four hours after treatment with PEP005, a large number of neutrophils and some macrophages were found in treated skin (Fig. 1, I and J), and in and around the treated tumor site, with neutrophils abundant at the periphery of the tumor. Neutrophils can be seen as the small spherical and uniform cells with red/pink cytoplasm and irregular blue nuclei. Areas with a pronounced neutrophil infiltrate are encircled (Fig. 1 F, H, J, and L). Some hemorrhage of local blood vessels was also apparent at this time (Fig. 1, K and L). High magnification images of areas at the periphery of the tumor following placebo treatment show intact tumor cells, RBC, and few leukocytes (Fig. 1M). In contrast, high magnification images of areas at the periphery of the tumor following PEP005 treatment...
show numerous neutrophils, which can be identified by their multilobed nuclei and red staining nongranular cytoplasm (Fig. 1, N and O). A similar pattern of neutrophil infiltration was also observed when this experiment was repeated using C57BL/6 mice and B16 melanomas (data not shown). The acute inflammatory response that follows PEP005 treatment was therefore characterized by a pronounced neutrophil infiltrate.

As reported previously, the cosmetic effect after PEP005 treatment was very favorable. Approximately 3 wk after PEP005 treatment, the skin at the treatment site was similar to untreated skin and had normal elasticity, and by 2–3 mo, few if any signs of scarring or erythema were apparent (12).

Tumor relapse in neutrophil-depleted mice after topical chemotherapy with PEP005

We have previously shown that PEP005 was able to cure, without significant relapse, a number of different tumor types grown in Foxn1nu mice, illustrating that T cells are not required for effective treatment (12). Neutrophils are fully active in Foxn1nu mice and their importance for the anticancer activity of PEP005 was investigated by using the anti-Ly-6G Ab (RB6-8C5) to deplete these cells in mice bearing LK2 tumors. The tumors grew only marginally slower in control animals (that had received no Ab) and animals receiving control Ab (A95-1), compared with animals given the anti-Ly-6G (RB6-8C5) Ab (Fig. 2A).

In a separate experiment, LK2 tumors were established in control animals, animals receiving control Ab and animals receiving anti-Ly-6G Ab. When the tumors had reached \( \approx 14 \text{ mm}^3 \) they were treated topically with PEP005 daily for 3 days. Initial chemoablation of the tumors was apparent in all groups; however, after day 25 in the anti-Ly-6G Ab-treated group tumors began to re-emerge (Fig. 2B). Data from this experiment are also presented as a percentage of tumors relapsing in each group over time (Fig. 2C). In control animals, only 1 of 12 (8.3%) tumors relapsed and none of the tumors relapsed in animals that had received the control Ab (Fig. 2C). However, in animals whose neutrophils had been depleted with anti-Ly-6G Ab (see below), a significant increase in the relapse rate to 83% was observed (Fig. 2C) (Cox regression analysis \( p = 0.005, \text{ Wald statistic} = 7.92, \text{ for anti-Ly-6G Ab vs control Ab}). These data suggest that neutrophils are required to prevent relapse following PEP005 treatment.

Together with the PEP005-treated animals described in Fig. 2, B and C, a parallel group of animals was established that was treated with placebo (instead of PEP005). Acute erythema was absent in all placebo-treated animals, but apparent in all PEP005-treated tumor sites (Fig. 2D). However, the peak erythema on day 2 was \( \approx 50\% \) lower in the mice that received the anti-Ly-6G Ab compared with control Ab-treated animals (\( p < 0.001, \text{ unpaired Student’s t test} \) (Fig. 2D). The neutrophil infiltrate thus appeared to contribute significantly to the PEP005-induced inflammatory response.

The neutrophil counts in blood were determined for each of the groups shown in Fig. 2. The white cell counts in placebo-treated mice (Fig. 3A), mice treated with PEP005 (Fig. 3B), and mice treated with the control Ab (data not shown) were essentially identical. In contrast, the percentage of neutrophils in the blood fell dramatically from \( \approx 80\% \) of total leukocytes to \( \approx 3\% \) following anti-Ly-6G Ab treatment in both placebo-treated and PEP005-treated animals (Fig. 3, C and D). Neutrophil counts were restored to normal levels \( \approx 10–12 \text{ days} \) after the last Ab injection (in Fig. 3, C and D, the increase in the percentage of lymphocytes over days 0–10 did not reflect an increase in the number of lymphocytes per microliter of blood; data not shown).

**FIGURE 2.** The effect of granulocyte depletion on PEP005 treatment of LK2 tumors grown on Foxn1nu mice. A. Tumor growth in control mice and animals given anti-Ly-6G Ab. LK2 tumors were injected s.c. (\( n = 12 \text{ tumors/group}, 4 \text{ tumors/mouse, } 3 \text{ mice/group} \)) on day -14. Mice were injected with anti-Ly-6G Ab (100 \( \mu \text{g i.p., on days } -2, 0, \text{ and } 2 \)) ( ), or with control Ab (A95–1; 100 \( \mu \text{g i.p., on days } -2, 0, \text{ and } 2 \)) ( ) or with nothing ( ). Tumors were treated with placebo on days 0, 1, and 2. The tumor volumes represent the mean volume of individual tumors. Hash symbols indicate data points where one or more of the mice were euthanized (due to excessive tumor burden), with the means including the tumor volume(s) observed at euthanasia. B. Tumor growth in control mice and animals given anti-Ly-6G Ab after PEP005 treatment. As for A except LK2 tumors (\( \approx 14 \text{ mm}^3 \)) were treated with 10 \( \mu \text{g of PEP005 in isopropyl-based gel daily for } 3 \text{ days on days } 0, 1, \text{ and } 2 \) (symbol legend and hash symbols as for A). C. Percentage of tumor sites relapsing in control mice and animals given anti-Ly-6G Ab after PEP005 treatment. Data from the experiment described in B showing the percentage of tumor sites relapsing over time after PEP005 treatment (100% = 12 tumor sites) (symbol legend as for A). The tumors were deemed to have relapsed when they had reached 8 mm\(^3\) (the first tumor to relapse was on mouse 1, the next on mouse 3, then mouse 2, 2, 3, 3, 1, 2, and 1, illustrating that relapse rates were not different for individual mice and that each tumor could be considered as an independent entity). D. Erythema of tumor sites in control mice and animals given anti-Ly-6G Ab after PEP005 or placebo treatment. The erythema area represents the mean of the areas of erythema at the treatment sites (\( n = 12 \text{ tumors/group} \)). Symbol legend as in A plus placebo-treated mice after 1) anti-Ly-6G Ab administration ( ), 2) after control Ab administration ( ), and 3) after no Ab administration ( ). The experiments described in this figure were repeated twice and one representative experiment is shown.
Rapid relapse after PEP005 treatment of B16 tumors in CD18-deficient mice

Neutrophil extravasation into inflamed sites is severely impaired in CD18-deficient mice (23, 30, 31), whereas Ab and some T cell responses remain intact (32). B16 tumors grew faster in CD18-deficient mice compared with C57BL/6 mice (Fig. 4A), so PEP005 treatment was initiated earlier post-tumor cell implantation in the former so that tumor sizes were comparable in CD18-deficient and C57BL/6 mice. Although 100% of B16 tumors with an average volume of $14 \text{ mm}^3$ were cured with three topical applications of PEP005 in C57BL/6 mice, all tumors of a similar size treated in CD18-deficient mice rapidly relapsed after the same PEP005 treatment (Fig. 4A). An acute erythema was apparent after PEP005 treatment and the area was similar in CD18-deficient and wild-type mice, although the intensity was considerably less in the former animals. The erythema in CD18-deficient mice was mainly a discoloration with slight reddening seen on some mice, whereas wild-type mice showed pronounced reddening in all treated sites (data not shown).

Thus, in a second neutrophil-defective model, PEP005-treated tumors relapsed and the intensity of the PEP005-induced inflammation was reduced. Although multiple leukocyte interactions can be affected by CD18 deficiency, these data support the view that an acute inflammatory response with a dominant neutrophil infiltrate is required for the antitumor efficacy of PEP005.

**FIGURE 3.** Blood leukocyte counts in mice given anti-Ly-6G and control Abs. Tail vein blood was taken from the mice described in Fig. 2 and smeared onto glass slides. White cell counts were performed by microscopy after Quick Dip staining (cells on two slides were counted per time point). The number of individual cell types is expressed as a percentage of total white cells ($\pm$ SE); neutrophils (■), lymphocytes (▲), monocytes (□), and eosinophils (▲). A, Control mice treated with placebo (as in Fig. 2A). B, Control mice treated with PEP005 (as in Fig. 2B). C, Mice injected with anti-Ly-6G Ab and treated with placebo (as in Fig. 2A). D, Mice injected with anti-Ly-6G Ab and treated with PEP005 (as in Fig. 2B).

**FIGURE 4.** Relapse rates following PEP005 treatment in CD18-deficient, NK-depleted, op/op, and SCID mice. A, Rapid relapse of B16 melanoma in CD18-deficient mice. B16 cells were injected s.c. on the back of B6.129S7-Itgb2tm1Bay/J CD18 hypomorphic mice. Six days later when tumors had reached $14 \text{ mm}^3$, the animals were treated daily for 3 days (days 0, 1, and 2) with placebo (□) or 10 μg of PEP005 (△) (n = 9 tumors/group, 1 tumor/mouse). B16 cells were implanted s.c. on the backs of C57BL/6 mice and were treated, 7 days after inoculation (when the tumors had reached $14 \text{ mm}^3$), daily for 3 days with placebo (□) or 10 μg of PEP005 (△) (n = 8 mice/group, 1 tumor/mouse). The indicated tumor volumes represent the mean volume of individual tumors ($\pm$ SE). B, The role of NK cells. Percentage of LK2 tumor sites relapsing in control Foxn1nu mice (□) and Foxn1nu mice given anti-asialo GM1 Ab (△) after PEP005-treatment of $14 \text{ mm}^3$ tumors (n = 12 tumors/group, 4 tumors/mouse). C, The role of macrophages. B16 tumor growth in op/op mice following treatment with placebo (□) or 10 μg of PEP005 (△) (n = 12 tumors/group, 4 tumors/mouse; 100% = 12 tumor sites). D, Relapse of LK2 tumors following PEP005 treatment in SCID mice. LK2 tumors were injected s.c. into SCID and Foxn1nu mice (n = 12 tumors/group, 4 tumors/mouse; 100% = 12 tumor sites). After 2 wk when tumors had reached $14 \text{ mm}^3$, the tumors were treated with PEP005 as above and relapse rates monitored. Individual placebo-treated tumors reached an average size of $270 \text{ mm}^3$ by day 28 in both SCID and Foxn1nu mice (data not shown). The experiments in this figure were performed once.
The role of NK cells and macrophages

NK cells and macrophages are present in Foxn1nu mice and these cell types can be activated by PKC-activating agents (33, 34). To determine their potential contribution to the antitumor effects of PEP005, the experiment shown in Fig. 2 was repeated using a polyclonal anti-asialo GM1 polyclonal Ab to deplete NK cells (tumor growth rates in SCID and Foxn1nu mice were analyzed in triplicate for the indicated cytokines. ND, Not detectable.

Induction of MIP-2, TNF-α, and IL-1β after PEP005 treatment in vivo.

To determine whether the rapid inflammation induced by PEP005 requires B cells and Ab production, LK2 tumors were grown in Csfm−/− (op/op) mice, which lack functional macrophage-CSF, and are therefore severely monocytopenic (35). PEP005 treatment was able to cure, without relapse, 100% of tumors (Fig. 4C), indicating that macrophages do not have an important role in preventing relapse following PEP005 treatment.

Relapse of PEP005-treated tumors in SCID mice

To determine whether successful relapse free PEP005 treatment requires B cells and Ab production, LK2 tumors were grown in SCID mice and were treated with PEP005 and monitored for relapse. The relapse rate of PEP005-treated LK2 tumors grown in SCID mice (Fig. 4D) was very similar to that seen for PEP005-treated LK2 tumors grown in granulocyte-depleted Foxn1nu mice (Fig. 2C), with >80% of tumors relapsing by day 40. In contrast, nearly all the tumors were cured when similar sized LK2 tumors grown in Foxn1nu mice were treated with PEP005 (Cox regression analysis, p = 0.006, Wald statistic = 7.68), for relapse rates in SCID compared with Foxn1nu mice. LK2 tumors grew at similar rates in SCID and Foxn1nu mice and the PEP005-induced erythema was similar in the two mouse strains (data not shown). The high relapse rates in SCID mice indicates that anti-cancer Ab production following PEP005 treatment is also required to prevent relapse.

Prolinflammatory cytokine induction in human keratinocytes, fibroblasts, neutrophils, and melanoma cells after PEP005 treatment in vitro

To determine whether the inflammatory mediator profile induced in mice following PEP005 treatment also applies to human cells, the induction of cytokines and chemokines following PEP005 treatment of human keratinocytes, neutrophils, and a human melanoma cell line (Me10538) was assessed in vitro. Culture medium was analyzed for cytokines and chemokines 6 h after treatment with PEP005. IL-8, the human counterpart of MIP-2, was induced in all cells tested, with keratinocytes and neutrophils producing maximal levels at 5 ng/ml PEP005 (Table I). The

Table I. Induction of proinflammatory cytokines in human cells in vitro

<table>
<thead>
<tr>
<th>PEP005 (ng/ml)</th>
<th>Keratinocytes</th>
<th>Fibroblasts</th>
<th>Melanoma</th>
<th>Neutrophils</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>IL-8</td>
<td>TNF-α</td>
<td>IL-6</td>
<td>IL-8</td>
</tr>
<tr>
<td>0</td>
<td>995 ± 48</td>
<td>8 ± 1</td>
<td>ND</td>
<td>20 ± 1</td>
</tr>
<tr>
<td>1</td>
<td>3910 ± 148</td>
<td>510 ± 26</td>
<td>ND</td>
<td>79 ± 3</td>
</tr>
<tr>
<td>5</td>
<td>4775 ± 178</td>
<td>847 ± 37</td>
<td>ND</td>
<td>160 ± 14</td>
</tr>
<tr>
<td>10</td>
<td>3895 ± 198</td>
<td>498 ± 29</td>
<td>ND</td>
<td>215 ± 12</td>
</tr>
<tr>
<td>100</td>
<td>2950 ± 108</td>
<td>335 ± 21</td>
<td>ND</td>
<td>239 ± 9</td>
</tr>
</tbody>
</table>

* Cells (10⁶ cells per well for neutrophils and 5 × 10⁴ cells per well for the other cells) were incubated with the indicated concentration of PEP005 for 6 h and the supernatants were analyzed in triplicate for the indicated cytokines. ND, Not detectable.
reduced response of neutrophils at 100 ng/ml PEP005 (compared with 3 ng/ml) was probably due to apoptosis, as PEP005 was a potent inducer of neutrophil activation (see below), which leads to activation-induced cell death. TNF-α levels were induced ~60-fold in keratinocytes in the presence of 1 ng/ml PEP005, and IL-6 was marginally induced in fibroblasts after treatment with >10 ng/ml PEP005 (Table I). No IL-2, IL-4, IL-10, IL-12, or GM-CSF was detected in any cell type tested after PEP005 treatment (data not shown). Thus, PEP005 induced production of the neutrophil chemoattractant IL-8 in keratinocytes, fibroblasts, neutrophils, and tumor cells, and potently induced TNF-α production in keratinocytes.

**Activation of human endothelial cells by PEP005**

Recruitment of neutrophils to the site of inflammation requires activation of the vascular endothelium to promote neutrophil binding, which is a prerequisite for extravasation and tissue infiltration (37). PEP005 was able to activate vascular endothelial cells in a dose-dependent manner with significant neutrophil binding occurring at 10 ng/ml PEP005 (Fig. 6A). TNF-α is known to activate vascular endothelium; neutrophil binding induced by TNF-α was shown as a positive control (Fig. 6A).

**Induction of tumoricidal reactive oxygen species by PEP005-activated human neutrophils**

The production of potential antitumor agents by PEP005-stimulated human neutrophils was investigated. PEP005 concentrations >10 ng/ml were able to induce marked superoxide production by human neutrophils (Fig. 6B). At 100 ng/ml (but not 10 ng/ml), PEP005 induced a modest release of defensins (data not shown) which are neutrophil granule proteins that have been reported to have some anticancer activity (38). Soluble TRAIL (Apo-2 ligand)
Neutrophils have long been recognized as key players in innate defenses against microbial infections, but recently insights into their potential roles in anticancer activity have begun to emerge. Residual tumor cells by ADCC. This mechanism has recently been shown to be involved in the anti-cancer activity of adoptively transferred anti-cancer Abs (5–7). The high relapse rates of PEP005-treated tumors grown in (B cell defective) SCID mice, the increase in anti-cancer Abs following PEP005 treatment, and the ability of antisera from mice, whose tumors had been treated with PEP005, to mediate neutrophil recruitment and activation of neutrophils (2, 15, 46). PEP005 was also able to activate endothelial cells directly, perhaps providing a second rapid and direct boost to neutrophil recruitment. MIP-2/IL-8 by tumor and stromal cells may therefore represent a key immunotherapeutic aspect of PEP005 action, allowing it to harness the anticancer potential of neutrophils. The initial cytotoxic activity of PEP005 involves rapid disruption of the plasma membrane, rapid swelling of mitochondria and death by primary necrosis. The process does not appear to require PKC activation and occurs at PEP005 concentrations of ~100 μg/ml (12). At lower concentrations (10–100 ng/ml), PEP005 is a potent activator of PKC (13). This level of PEP005 is probably reached within a day or two around the tumor site due to drug dispersal and conversion of PEP005 to inactive products, which in tissue culture occurs with a half life of ~5–10 h (data not shown). Application of the PKC activator 12-O-tetradecanoyl-phorbol-13-acetate (TPA) to mouse skin also induces a pronounced T cell-independent influx of neutrophils (14, 15, 43). TPA is also able to stimulate neutrophils to produce reactive oxygen intermediates (44). The activation of PKC by PEP005 is therefore likely to be responsible for the PEP005-mediated recruitment and activation of neutrophils. PEP005 treatment induced MIP-2/IL-8, TNF-α, and IL-1β, all mediators that would contribute to the recruitment and activation of neutrophils (2, 15, 36). In vivo neutrophil extravasation requires activation of the endothelium (37). This process is induced by TNF-α and IL-8, which increase the affinity of integrins on neutrophils and their adhesion partners on the endothelial cells (45, 46). PEP005 was also able to activate endothelial cells directly, perhaps providing a second rapid and direct boost to neutrophil recruitment. MIP-2/IL-8 showed the greatest increase in response to PEP005 and previous studies have demonstrated that Foxn1nu mice challenged with human tumor cells expressing IL-8 showed extensive neutrophil influx to the tumor site resulting in dramatically reduced tumor growth (47). The release of MIP-2/IL-8 by tumor and stromal cells may therefore represent a key immunotherapeutic aspect of PEP005 action, allowing it to harness the anticancer potential of neutrophils. The high relapse rates of PEP005-treated tumors grown in (B cell defective) SCID mice, the increase in anti-cancer Abs following PEP005 treatment, and the ability of antisera from mice, whose tumors had been treated with PEP005, to mediate neutrophil ADCC in vitro, indicate that once recruited to the tumor site, killing of residual cancer cells by neutrophils involves ADCC. This mechanism has recently been shown to be involved in the anti-cancer activity of adoptively transferred anti-cancer Abs (5–7). How PEP005 treatment might lead to increased anti-cancer Ab
production remains unclear. However, the induction of primary necrosis of the tumor cells by PEP005 treatment (12) may be involved because primary necrosis has been shown to be immunostimulatory (48, 49). PKC activation (13) may also promote dendritic cell (50) and/or B cell maturation (51). In vitro, PEP005 was able to induce release of tumoricidal reactive oxygen intermediates. Furthermore, like TPA (52), PEP005 was unable to stimulate efficient degranulation of human neutrophils. However, PKC stimulation and FcR engagement have been shown to act synergistically on neutrophils to stimulate degranulation and killing of tumor cells (52), a synergy that also appears to apply to PKC stimulation and complement receptor engagement (Fig. 6E, ±10 ng/ml PEP005). PEP005 thus appears to provide an environment that is highly conducive to neutrophil-mediated anticancer activity. Neutrophils are recruited and activated by PEP005-induced inflammatory mediators and are also activated by the direct action of PEP005. The PEP005-induced anti-cancer Abs then provide tumor cell targeting and Fc and/or complement receptor-mediated triggering.

In summary, our data show that neutrophil-mediated killing of residual tumor cells is important for preventing relapse following PEP005 treatment of skin cancers. This study also provides additional evidence that in specific settings neutrophils can provide important anticancer activity.

Acknowledgments

We thank Prof. David Weedon (Bond University, Gold Coast, Australia) and Clay Winterford (Histotechnology Facility, University of Queensland, Queensland, Australia) for their help with histology. We also thank Sharon Mason for assistance with counting blood smears, David Radford and Andrew Filer for help provided with culture of normal keratinocytes and fibroblasts, and Hema Chahal for help with multiplex analysis.

Disclosures

S. M. Ogborne is now an employee of Peplin Ltd, but was not when the work being performed. A. Suhbir and P. G. Parsons are consultants for Peplin Ltd.

References