Helicobacter pylori CagA-Dependent Macrophage Migration Inhibitory Factor Produced by Gastric Epithelial Cells Binds to CD74 and Stimulates Procarcinogenic Events

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Macrophage migration inhibitory factor (MIF) is a proinflammatory cytokine that has recently been implicated in carcinogenesis. *Helicobacter pylori*, which is closely linked to gastric cancer, induces the gastric epithelium to produce proinflammatory cytokines, including MIF. MIF can bind to CD74, which we have previously shown to be highly expressed on the surface of gastric epithelial cells (GEC) during *H. pylori* infection. In this study, we sought to investigate the role of the *H. pylori*-induced MIF on epithelial proliferation and procarcinogenic events. Upon establishing a role for the *H. pylori* CagA virulence factor in MIF production, MIF binding to CD74 on GEC was confirmed. rMIF and *H. pylori* were shown to increase GEC proliferation, which was decreased when cagA− strains were used and when CD74 was blocked by mAbs. Apoptosis was also decreased by MIF, but increased by cagA+ strains that induced much lower amounts of MIF than the wild-type bacteria. Furthermore, MIF binding to CD74 was also shown to decrease p53 phosphorylation and up-regulate Bcl-2 expression. This data describes a novel system in which an *H. pylori* virulence factor contributes to the production of a host factor that in turn up-regulates procarcinogenic events by the gastric epithelium.


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**Helicobacter pylori** is a common human pathogen that infects over 50% of the world’s population. Infection with this organism is a major cause of chronic gastritis, gastric and duodenal ulcers, and gastric carcinomas throughout the world (1, 2). Pathogenesis of infection often includes inflammation, mucosal damage, or gastric atrophy, and requires close association of the bacteria with the gastric epithelium, leading to chronic epithelial responses. There is now considerable evidence linking this pathogen to changes in the gastric mucosa that contribute to gastric cancer (3, 4). Gastric cancer is the second leading cause of mortalities due to cancer in the world, is often undetectable in the early stages, and has an overall survival rate of only 10–20% (5). Links between chronic inflammation and carcinogenesis may play an important role in gastric cancer, but are not well understood.

The main virulence factor of *H. pylori* that has been suggested to induce procarcinogenic events in gastric epithelial cells (GECs) is the cytotoxin-associated gene A product, CagA protein (6, 7). CagA is injected into cells by *H. pylori*’s type IV secretion system. Once CagA is injected into the cell, it specifically binds the Src homology 2-containing protein tyrosine phosphatase, SHP-2, in a tyrosine phosphorylation-dependent manner and stimulates phosphatase activity (8, 9). This activity induces various cell responses that result in cytoskeleton rearrangement leading to a cell scattering/hummingbird phenotype, and cell-to-cell disassociation that may result in disruption of the epithelial barrier integrity. ERK1/2 activation is also suggested to be induced after CagA is phosphorylated (10), and may also lead to increased inflammatory responses associated with *H. pylori* infection (11, 12). Another important observation about CagA+ *H. pylori* infections is an increase in gastric cell proliferation (13, 14), which has been observed both in vivo and in vitro. The mechanisms leading to this increase in proliferation remain unknown.

*H. pylori* has recently been shown to induce production of macrophage migration inhibitory factor (MIF) by several cell types (15). MIF is a versatile cytokine that mediates both innate and adaptive immunity and plays a role in chronic inflammation. It has been shown to up-regulate NF-κB, ERK1/2, AP-1, and protein kinase C pathways leading to increased expression of IL-1β and IL-8 mRNA (16, 17). Also, it is important to consider the role that MIF plays in cell growth and procarcinogenic signaling. It is thought to promote tumor growth and viability by modulating immune responses and supporting tumor-associated angiogenesis (18, 19). The central mechanisms MIF affects to increase cell proliferation and decrease apoptosis are p53 inhibition and reduction of oxidative stress-induced apoptosis (20, 21). The inhibition of p53 appears to result in a more robust inflammatory response, which may also explain its importance in chronic inflammation.

CD74, or the class II MHC-associated invariant chain, can act as a receptor for MIF (22). CD74 is known for its role in the regulation of intracellular transport and Ag-presenting functions of class II MHC molecules. Recent studies have shown that CD74 is not only expressed intracellularly, but also on a variety of cell surfaces, including GECs (23, 24). We and others have shown that bound CD74 can induce signaling events leading to NF-κB activation (25, 26). CD74 may also play a role in cancer. Cell surface
expression of CD74 was shown to be up-regulated during gastric cancer when tumor cells were examined by immunohistochemical staining (27). Patients with low CD74 expression were shown to have better outcomes than those with high CD74 expression. In more general cancer immunotherapy studies, antisense suppression of CD74 has been demonstrated to be an effective therapeutic treatment in several studies (28, 29).

This study investigated GEC production of MIF in response to *H. pylori*. This response was shown to be CagA dependent. MIF was shown to bind to CD74 in this system and increase cell proliferation that could be blocked by anti-CD74 mAbs. Cell proliferation was not increased by CagA knockout strains, but apoptosis rates of GECs were increased to levels that were higher than those of cells treated with wild-type *H. pylori*. Phosphorylated p53 was increased by CagA-deficient strains, but not by wild-type strains, and this increase was abrogated when rMIF was added to cell cultures suggesting that MIF induced by *H. pylori* infection decreases phosphorylation of p53, and increases GEC proliferation. These observations strongly suggest a link between MIF, a proinflammatory cytokine, and precarcinogenic events.

### Materials and Methods

#### Cell lines

The GEC line N87 (CRL-5822) was obtained from the American Type Culture Collection (ATCC) and maintained in RPMI 1640 medium supplemented with 10% FCS and 1 mM glutamine. The nontransformed fetal gastric cell line HS-738 (CRL-7860) was also obtained from ATCC and maintained in DMEM with high glucose, 10% FCS, and 1 mM glutamine.

#### Bacterial cultures

*H. pylori* strains 26695, a strain isolated from a patient with gastritis in the United Kingdom, and 51B, a strain isolated from a patient with gastritis in Japan, and the cagA isogenic mutants were obtained from Dr. Y. Yamaoka (Baylor College of Medicine, Houston, TX). These autonomous knockout strains were prepared as previously described (30). All strains were grown on blood agar plates (BD Biosciences) at 37°C under microaerophilic conditions, as previously described (31). Bacteria were transferred after 48 h into Brucella broth containing 10% FBS for 24 h. After centrifugation at 2500 rpm, bacteria were resuspended in sterile PBS. The concentration of bacteria per milliliter was determined by measuring the OD<sub>600</sub> with a spectrophotometer (DU-65; BD Biosciences) and comparing the value to a standard curve generated by quantifying viable organisms from aliquots of bacteria at varying concentrations that were also assessed by OD.

#### MIF ELISA

*H. pylori* strains were incubated with GECs in a 100:1 bacteria:cell ratio in 24-well plates for 8-h time intervals up to 32 h. Supernatants from wells were used to quantify the production of MIF by ELISA. The MIF ELISA kit was obtained from R&D Systems, and assays were performed according to the manufacturer’s instructions.

#### MIF binding to CD74

rMIF (R&D Systems), 1 ng, was added to GEC lysates, and rotated for 2 h at 4°C. Lysate mixtures were precleared with 10 μl of protein A/G beads (Santa Cruz Biotechology) for 2 h at 4°C. MIF was immunoprecipitated using protein A/G beads that were preincubated with anti-MIF mAb (R&D Systems) for 2 h at room temperature. After washing, beads were washed with the lysate mixture of MIF and GEC lysates. Beads were then washed four times and the bound material was eluted, samples were run on SDS PAGE, and transferred to a nitrocellulose membrane for Western blotting. The membrane was incubated with the anti-CD74 Ab clone Pin.1 (StressGen Bioreagents at a 1/3,000 dilution for 2 h at room temperature). After washing four times for 15 min with TBS Tween 20, donkey anti-mouse HRP-conjugated secondary Ab (Santa Cruz Biotechnology) was used diluted 1/1,000 and incubated with the membrane for 1 h at room temperature. The membrane was again washed four times for 15 min. Immunoreactive proteins were detected using Super Signal West Pico Chemiluminescent Substrate (Pierce Biotechnology).

#### Flow cytometry

For flow cytometry analysis of Bel-2 and activated caspase-3, cells were lysed using BD Cytofix/Cytoperm (BD Biosciences) according to the manufacturer’s instructions. Anti-Bel-2 (BD Biosciences) or isotype control Abs before the addition of MIF. Strain-specific ELISPOTs were performed on 96-well plates (BD Biosciences) at 37°C under microaerophilic conditions, as previously described (31). Bacteria were transferred after 48 h into Brucella broth containing 10% FBS for 24 h. After centrifugation at 2500 rpm, bacteria were resuspended in sterile PBS. The concentration of bacteria per milliliter was determined by measuring the OD<sub>600</sub> with a spectrophotometer (DU-65; BD Biosciences) and comparing the value to a standard curve generated by quantifying viable organisms from aliquots of bacteria at varying concentrations that were also assessed by OD.

#### Proliferation assay

CellTiter 96 Non-Radioactive Cell Proliferation kit (Promega) was used to measure cell proliferation. Cells were cultured in a 96-well plate overnight in serum-free medium at 1 × 10<sup>6</sup> cells/well. A standard curve was established using wells seeded with 1/2 dilutions of cells starting from 5 × 10<sup>3</sup> to 6.4 × 10<sup>6</sup>. Cells were treated with 0.1, 1.0, or 10 ng/ml MIF or 1:1, 10:1, or 100:1 bacteria:cell ratio of wild-type and cagA PAI or cagA knockout strains for 24 h. Some cells were pretreated with anti-CD74 clone MB-748 (BD Biosciences) or isotype control Abs before the addition of MIF. For a positive control, 1 ng/ml PMA was added to wells for 24 h. Cells were then washed with medium to remove bacteria, and the dye solution added according to manufacturer’s instructions for 1 h at 37°C. After addition of stop solution, OD was read at 490 nm using an ELISA plate reader. Cell numbers were calculated using the standard curve.

#### Apoptosis

Cell Death Detection ELISA (Roche) kits were used to measure cell death. Cells were treated with wild-type and cagA-deficient knockout strains using the same concentrations and time intervals that were used for proliferation assays. rMIF was added to some wells before cagA knockouts. For wild-type strains, anti-MIF neutralizing Abs were added to some wells and isotype controls were used as well. Cells were lysed according to the manufacturer’s instructions, lysates were used for ELISA, and OD was measured at 405 nm.

#### p53 phosphorylation

Total and phosphorylated p53 levels were measured by intracellular bead arrays (BioSource International). Beads conjugated with mAbs for p53 were prepared according to manufacturer’s instructions and incubated with lysates from GECs treated with *H. pylori* strains 26695, 51B, or their CagA-deficient mutants for 24 h. Some samples were pretreated with rMIF (10 ng/ml) before the addition of *H. pylori*. After washing, samples were run on a Bio-Plex System and analyzed using Bio-Plex Manager software (Bio-Rad).

Phosphorylated p53 was also detected by Western blotting. *H. pylori* strains or rMIF was incubated with GECs for 24 h. Cells were lysed as described previously (32). Lysates were centrifuged at 14,000 × g for 10 min at 4°C. The resulting protein (5 μg) from each sample were loaded into wells of 10% SDS gels for electrophoresis, and transferred to nitrocellulose membranes for Western blotting. The membranes were treated with incubated with anti-phospho (S15) p53 (R&D Systems), and anti-β-actin (Sigma-Aldrich) as a loading control. After washing four times for 15 min with TBS Tween, donkey anti-mouse HRP-conjugated secondary Ab (Santa Cruz Biotechnology) was used diluted 1/10,000 and incubated with the membrane for 1 h at room temperature. The membrane was again washed four times for 15 min. Immunoreactive proteins were detected using Super Signal West Pico Chemiluminescent Substrate (Pierce Biotechnology).

#### Statistical analysis

Results are expressed as mean ± SEM. MIF production and cell turnover rates were compared using an ANOVA analysis of the variance and considered significant if p < 0.05.

#### Results

**H. pylori induces MIF production by GECs in a CagA-dependent manner**

MIF has recently been suggested to play an important role in both innate and adaptive immunity and the dysregulation of cell cycle...
events in carcinogenesis. Because these factors are important during \textit{H. pylori} infection, we sought to determine MIF production by GECs in response to \textit{H. pylori}. Furthermore, because the cag pathogenicity island injection of CagA into cells plays such an important role in many GEC responses induced by \textit{H. pylori}, cagA-deficient strains were compared with wild-type strains of \textit{H. pylori} at a moderate infection of 10:1 bacteria:cell ratio. Fig. 1, A and B, indicate that \textit{H. pylori} strains 26695 and 51B induced MIF production by the HS-738 nontransformed fetal gastric cell line and the N87 gastric carcinoma cell line, respectively. Increases in MIF production were detected as early as 8 h postinfection and continued to increase up to 32 h after infection. When the CagA-deficient strains were used at 24 h, the induction of MIF was just slightly above basal levels produced by both cell lines (Fig. 1, C and D), whereas the wild-type (CagA\textsuperscript{+}) induced a robust MIF response, suggesting that MIF production by GECs is dependent on CagA injection into the cells.

\textbf{MIF binds to CD74 on GECs}

CD74 has been suggested to act as a receptor for MIF in several studies (22, 33). We have shown that GECs express large amounts of CD74 (24, 25), which is up-regulated during inflammatory conditions and \textit{H. pylori} infection (34). Consequently, we examined the role of CD74 as a receptor for MIF on GECs by immunoprecipitation and Western blot analysis. rMIF was incubated with N87 cell lysates. MIF was immunoprecipitated along with GEC proteins bound to it. Western blot analysis using anti-CD74 mAbs revealed that CD74 was coprecipitated with MIF in samples and in control samples immunoprecipitated with anti-CD74 as indicated by the presence of a 33-kDa band (Fig. 2A).

Attachment of MIF to CD74 was also analyzed by flow cytometry. Biotinylated MIF was incubated with GECs and stained with streptavidin-PE for analysis. As shown in Fig. 2B, MIF attached to cells, but this attachment was dramatically decreased upon incubating cells with anti-CD74 before the addition of MIF. These studies suggest that CD74 is the primary receptor for MIF on GEC surfaces because blocking CD74 led to very little attachment of MIF to the cells.

\textbf{H. pylori-induced MIF increases GEC proliferation}

Various reports have shown that MIF increases proliferation of some cell types. In this study, we investigated the ability of MIF to induce proliferation of GECs. rMIF or \textit{H. pylori} strains and their respective CagA-deficient knockout strains were incubated with cells for 24 h. Proliferation was measured by nonradioactive cell proliferation colorimetric assay, as used in multiple recent studies (14, 33). Standard curves of known amounts of cells were run with each assay to extrapolate cell number from treated samples. As seen in Fig. 3A, GEC proliferation was increased starting at 0.1 ng/ml rMIF, which was in the range of that produced by GECs in response to \textit{H. pylori}, and increased further with each 10-fold increase in MIF concentration. To investigate the role of CD74 in the

\textbf{FIGURE 1.} GECs produce MIF in response to \textit{H. pylori}. MIF production by (A) HS-738 and (B) N87 GECs after exposure to \textit{H. pylori} strains 51B or 26695 for various time points assayed by ELISA. MIF levels were decreased when (C) HS-738 and (D) N87 cells where treated with CagA-deficient mutants compared with wild-type \textit{H. pylori} at 24 h. The means are shown as the results of duplicates in four experiments, \textit{n} = 8.

\textbf{FIGURE 2.} MIF binds to CD74 on GECs. A, rMIF was mixed with N87 cell lysates and immunoprecipitated with anti-MIF Abs with bound cell proteins. Western blot analysis with anti-CD74 revealed the presence of CD74. N87 lysates were run as a control in the \textit{center lane}, and N87 cell lysates immunoprecipitated with isotype control Abs were run in the \textit{left lane}. B, Biotinylated rMIF was incubated with HS-738 cells and streptavidin-PE for flow cytometry. Anti-CD74 Abs blocked attachment to cells compared with the solid peak control.
treated with (FIGURE 3. MIF induces GEC proliferation. HS-738 and N87 cells were
liferation (where all conditions showed significant increases in cell number (compared with unblocked cells (increased cell numbers compared with wild types (as the results of duplicates in four experiments, compared with a standard curve of known amounts of cells. The means are shown was measured by CellTiter 96 Non-Radioactive Cell Proliferation kit and com-
pared with conditions in the presence of , proliferation was decreased (Fig. 3D), further suggesting the role of MIF in 
H. pylori-induced cell proliferation.

H. pylori-induced MIF decreases GEC apoptosis

Because H. pylori is known to induce both cell proliferation and apoptosis (31, 35), apoptosis of GECs with wild-type H. pylori and CagA-deficient strains, which induce very little MIF, were investigated for their ability to induce apoptosis of GECs. Apoptosis was measured by Cell Death Detection ELISA for histone-associated DNA fragmentation (nucleosome fragment formation) at 24 and 48 h. Fig. 4A indicates the apoptosis index of HS-738 cells in the presence of H. pylori 26695 in 1:1, 10:1, and 100:1 bacteria:cell ratios. Although apoptosis rates with the 10:1 bacteria:cell ratio was only slightly increased over that of 1:1, a much larger increase in apoptosis was observed with 100:1 bacteria:cell ratio, which parallels the decrease in proliferation seen with 100:1 in Fig. 3B. When the 26695 CagA-deficient mutant strain was examined for its ability to induce GEC apoptosis, cell death was double of that induced by the wild-type, parental strain. Similar results were seen with strain 51B and its corresponding CagA-deficient mutant (data not shown here). To determine a role for MIF in the decreased apoptosis rates seen with wild-type H. pylori, rMIF (10 ng/ml) was added to cultures incubated with CagA-deficient strains in amounts equivalent to those produced during H. pylori 100:1 bacteria:cells (Fig. 1). Addition of MIF to cultures with CagA-deficient mutants resulted in a decrease in apoptosis (Fig. 4B), similar to that seen with wild-type 26695 in Fig. 4A, demonstrating an important role for MIF in decreasing H. pylori-induced apoptosis.

To further investigate the effect of MIF on apoptosis, expression of the activated form of caspase-3 was measured in GECs after exposure to 100:1 bacteria:cell ratio. Some cells were pretreated with 1 ng/ml rMIF for 24 h before exposure to H. pylori strains. As seen in Fig. 4C, H. pylori induced cells to produce some active caspase-3, which was produced at a higher rate with the CagA-deficient bacteria than the wild type. When cells were pretreated with MIF before CagA− bacteria, the levels of active caspase-3 were decreased (Fig. 4D), suggesting that MIF can down-regulate apoptotic events.
H. pylori-induced MIF decreases phosphorylation of p53

MIF has been shown to decrease p53 expression and phosphorylation in several studies (36, 37). Because CagA has been shown here to induce MIF production by GECs, the role of rMIF and CagA-induced MIF in the expression and phosphorylation of p53 was investigated. The levels of total p53 and phosphorylated p53 (S15) were also measured by intracellular bead arrays. The units of phosphorylation per nanogram of total p53 were calculated as recommended by the manufacturer. Fig. 5A illustrates that the level of phosphorylated p53 detected in cell treated with the CagA-deficient bacteria was more than double that of cells treated with the wild type, which was consistent with both strains tested. When rMIF was added to cells in similar amounts to that produced when cells were incubated with wild-type H. pylori before CagA strains, the level of phospho-p53 was decreased to similar levels seen in cells treated with wild-type bacteria. To further verify these findings, Western blot analysis was performed with cell samples treated under the same conditions. Similarly, treating cells with CagA-deficient 26695 resulted in higher levels of phospho-p53 than those induced in cells treated with wild-type 26695 (Fig. 5B). Similar results were seen with N87 cells and H. pylori strain 51B, but are not shown here. When rMIF was added, the levels of phosphorylated p53 decreased. These results, along with other studies suggesting a role for CagA in gastric cancer, and studies suggesting that MIF inhibits p53 function (36), indicate that MIF may play an important role in regulating p53 during H. pylori infections.

H. pylori-induced MIF induces Bcl-2 expression

Because Bcl-2 expression is up-regulated during H. pylori infection and gastric atrophy and cancer (38, 39), and down-regulated in

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**FIGURE 4.** MIF decreases GEC apoptosis. HS-738 cells incubated with H. pylori 24 and 48 h were examined for apoptosis by histone-associated DNA fragmentation where (A) strain 26695 incubated with cells resulted in less DNA fragmentation than the CagA-deficient knockout (p < 0.05) and (B) CagA knockout DNA fragmentation was decreased with addition of rMIF (p < 0.05). The means are shown as the results of duplicates in four experiments. n = 8. Active caspase-3 was measured by flow cytometry as shown in example figures where (C) H. pylori strain 26695 induces some expression, which is increased with the CagA- strain and (D) reduced when rMIF is incubated with cells before infection in example figures compared with the isotype control shown with a solid peak when compared with solid peak isotype control.

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**FIGURE 5.** p53 phosphorylation induction by H. pylori. Phosphorylation of p53 is shown with HS-738 cells by (A) intracellular bead array and (B) Western blotting where p53 phosphorylation is increased by H. pylori strain 26695 and 51B compared with untreated control (p < 0.05), but increased more with the CagA- strain compared with 26695 (***, p < 0.05), but decreased in the presence of rMIF (***, p < 0.05).
apoptotic cells (40), the expression of Bcl-2 was investigated during infection with \( H. pylori \) and upon pretreatment with rMIF. Flow cytometric analysis revealed that Bcl-2 expression is induced in GECs upon exposure to \( H. pylori \), but is decreased when cells were exposed to CagA-deficient strains in comparison to wild-type strains (Fig. 6A). Incubation of samples with rMIF alone also induced expression of Bcl-2 (Fig. 6B). These results indicate that MIF may also play an important role in the induction of Bcl-2 expression seen in GECs during \( H. pylori \) infection.

Discussion

Proinflammatory cytokines such as MIF and IL-8 have been attributed to increased proliferative signaling in a variety of systems (20, 33, 41, 42). MIF has been shown to play an important role in the regulation of proinflammatory immune responses in inflammatory disorders, and proliferative responses in multiple cancer types. MIF has recently been shown to be produced in response to \( H. pylori \) by multiple cell types (15). Because CagA has been shown to induce inflammatory responses (11, 12), and it has been associated with gastric cancer (43, 44), the role of CagA in MIF production by GECs was investigated in this study. By using CagA-deficient knockout strains, we established that CagA is responsible for MIF production by GECs because while wild-type strains induced MIF production, very little MIF was present in cultures treated with CagA knockout strains.

GECs express CD74 and this expression increases during infection with \( H. pylori \) both in vivo and in vitro (24, 34). CD74 was recently shown to be used by MIF as a receptor (22). Consequently, MIF binding to CD74 on GECs was examined here, and found to be a major receptor for MIF in this system as well. Similar to MIF, CD74 has been shown to be up-regulated in cancer studies, including gastric cancer (27). In a previous study, we also found that CD74 surface expression is increased during inflammatory conditions, and during \( H. pylori \) infection (34). This is especially significant in this system because \( H. pylori \) can also use CD74 as a point of attachment to GECs (25). The dramatic increase in CD74 expression during infection and the high turnover rate of CD74 (22) suggests that both MIF and \( H. pylori \) can use CD74 as a receptor in this system. MIF binding to CD74 has been shown to increase proliferation of different cell types (22, 45, 46). Similarly, MIF induced proliferation of GECs here in a dose-dependent fashion, and at comparable amounts to what is produced during \( H. pylori \) infection. Concurrently, \( H. pylori \) infection also led to increased proliferation, which was decreased when CagA knockout strains were used. Proliferation was also decreased when CD74 was blocked with Abs or MIF was neutralized by Abs in the medium. Under the same conditions, apoptosis of cells was decreased in the presence of MIF, and cagA knockout strains, where MIF production is much lower, induced greater levels of apoptosis than wild-type strains. rMIF added to cultures with the cagA knockout strains brought apoptosis levels back down suggesting that the effects MIF has on the cells may down-regulate apoptosis. The results of our studies agree with several other independent studies, including those by Moss et al. and Rokkas et al. (13, 14) using biopsies which showed that cagA strains increase GEC proliferation without a parallel increase in apoptosis. However, a separate study by Moss et al. (47) disagreed with these and suggested that there is not a deficiency in apoptosis with cagA strains, but the authors of this study also proposed that host factors may be more important than bacterial factors in determining the outcome of infection. Our studies agree that a host factor, MIF, may be crucial in shifting the balance of proliferation/apoptosis to one that is more positive for proliferation.

Concurrent with the decrease in apoptosis attributed to MIF, MIF may inhibit p53 function (36, 37), which is considered an important tumor suppressor because it is an important factor in apoptotic pathways. As for the effect of \( H. pylori \) on p53, there is conflicting information available. Several studies suggest there are no changes in p53 levels of the gastric mucosa during infection (48, 49), while others suggest that p53 levels are increased during infection (50, 51). However, these studies only looked at total p53 present, but failed to investigate levels of phosphorylated p53. In this study, we measured the amount of phosphorylated p53 per nanogram of total p53. We found that the total amount of p53 did not fluctuate much during infection, but there were changes in phosphorylated p53. Although wild-type \( H. pylori \) did induce some phosphorylation of p53, CagA-deficient strains induced more than double the levels. The addition of rMIF decreased these levels to levels similar to those induced by the wild-type bacteria. Based on these studies, MIF appears to have the ability to decrease p53 phosphorylation, which may elucidate how it decreases apoptosis. There are certainly bacterial factors present that induce some p53 phosphorylation, as seen with wild-type \( H. pylori \), but MIF appears to decrease the overall amount of phosphorylation, and apoptosis seen. This is especially evident with cells infected with CagA-deficient strains along with rMIF. Because \( H. pylori \) has also been proposed to mutate p53, the possibility that MIF may be involved should also be investigated.

To further examine the role for MIF in the regulation of apoptosis and increased proliferation, Bcl-2 expression by infected GECs was investigated. Bcl-2 regulates apoptotic processes, and can lead to apoptotic resistance when highly expressed. This protein has been shown to be up-regulated during \( H. pylori \) infection with cagA strains (52) and \( H. pylori \)-associated gastric cancers (38, 39). In this study, Bcl-2 levels were expressed at much higher levels in GECs with wild-type strains than with CagA-deficient...
strains. rMIF was also able to increase expression further suggesting the role for MIF in driving cells toward a carcinogenic phenotype.

The data presented here are the first to suggest a crucial role for MIF in the procarcinogenic signaling induced by H. pylori infection of GECs. Production of MIF by these cells in response to H. pylori depends upon the presence of CagA, which is also the bacterial factor suggested to play a role in procarcinogenic epithelial responses. rMIF induced the same proliferative responses that CagA+ H. pylori strains, and could therefore be an important piece to the gastric cancer puzzle.

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Disclosures

The authors have no potential conflict of interest.

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