Helicobacter pylori-Induced Invasion and Angiogenesis of Gastric Cells Is Mediated by Cyclooxygenase-2 Induction through TLR2/TLR9 and Promoter Regulation

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Helicobacter pylori-Induced Invasion and Angiogenesis of Gastric Cells Is Mediated by Cyclooxygenase-2 Induction through TLR2/TLR9 and Promoter Regulation

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Cyclooxygenase-2 (COX-2) plays a crucial role in Helicobacter pylori-associated gastric cancer. In this study, we report that H. pylori-induced COX-2 expression enhances the cancer cell invasion and angiogenesis via TLR2 and TLR9, which can be attenuated by the specific COX-2 inhibitor NS398 or celecoxib. The cAMP response element (CRE) and AP1 sites, but not κB on the COX-2 promoter, are involved in MAPKs-regulated COX-2 expression. Differential bindings of the CREB-1, ATF-2, c-jun to the CRE site, and the c-fos, c-jun, ATF-2 to the AP1 site are demonstrated by DNA affinity protein-binding, supershift, and chromatin immunoprecipitation assays. Activations of these transcription factors were attenuated by different MAPKs inhibitors. The mutants of TLR2, TLR9, or MAPKs inhibited H. pylori-induced COX-2 promoter, CRE, and AP-1 activities. MAPKs inhibitors attenuated the H. pylori-induced COX-2 mRNA and protein expressions. These results indicate that H. pylori acts through TLR2 and TLR9 to activate MAPKs, especially p38, and their downstream transcription factors (CREB-1, ATF-2, c-jun, and c-fos), resulting in the activations of CRE and AP-1 on the COX-2 promoter. These intracellular networks drive the COX-2-dependent PGE$_2$ release and contribute to cell invasion and angiogenesis. The Journal of Immunology, 2005, 175: 8242–8252.

Helicobacter pylori is a Gram-negative bacterium that colonizes the human stomach (1). It has been identified as a major pathogen leading to the development of a wide range of gastroduodenal diseases, including gastritis, duodenal ulcer, gastric ulcer, gastric adenocarcinoma, and gastric mucosa-associated lymphoid tissue lymphoma (2). Eradication of H. pylori prevents the development of new cancers or continued growth of occult cancers in patients with early carcinoma (3), suggesting that H. pylori infection may play a role not only in the initiation, but also in the promotion of gastric carcinoma. Cyclooxygenase-2 (COX-2) is the rate-limiting enzyme for the production of prostanooids (prostaglandins and thromboxanes) from arachidonic acid. Up-regulation of COX-2 plays a central role in the inflammatory changes and tissue damages associated with the chronic H. pylori infection, and is also involved in the gastric tumorigenesis (4, 5). The expression of COX-2 protein is significantly higher in patients with gastric cancer than those with nonulcer dyspepsia (6). Therefore, COX-2 expression is assumed to play a key role in H. pylori-associated gastric cancer in addition to the propagation of gastric inflammation (7). Recent studies have demonstrated that COX-2-mediated PG biosynthesis affects carcinogenesis via several different mechanisms. Angiogenesis, the formation of new blood vessels from an existing vascular bed, is a key step in tumor growth and metastasis (8). Up-regulations of IL-8, VEGF, angiogenin, urokinase-type plasminogen activator, and matrix metalloproteinase 9 genes by H. pylori have been reported to regulate the angiogenesis and invasion of human gastric carcinoma (9, 10). The implication of COX-2 in tumor angiogenesis has also been documented in the colon cancer cells (11). However, the role of H. pylori-induced COX-2 in the cell invasion and angiogenesis of gastric cancer cells remains elusive.

Epithelial cells, like macrophages, recognize microbial infections through TLRs, which recognize conserved motifs on pathogens that are not found in higher eukaryotes (12). All TLRs activate a common signaling pathway that culminates into the activation of NF-κB and MAPKs (13). The involvement of TLR2 and TLR9 in H. pylori-induced COX-2 expression via the PKC/ε-Src/NF-κB activation pathway has been reported from this lab (14). Activations of MAPKs by H. pylori have been reported and found to regulate the expressions of many genes such as matrix metalloproteinase 7 and IL-8 (15, 16). Therefore, the role of TLR2 and TLR9 in MAPKs activation induced by H. pylori leading to COX-2 expression is further investigated in this study.

Our recent study has demonstrated that H. pylori strains isolated from gastric cancer patient (HC) induced a higher COX-2 expression than those from gastritis and peptic ulcer. However, the virulence genes of icaA, vacA, babA2, caga 3′ repeat region and hrgA among various H. pylori strains failed to show any association with COX-2 expression despite a higher COX-2 induction capability of HC (14). Involvement of cAMP response element (CRE) site in the H. pylori-induced COX-2 expression was demonstrated (14). The importance of CRE and/or AP-1 sites in mediating COX-2 transcription, particularly on the murine and human promoter has been reported (17–20). The AP-1 site (−67/−61) on the human COX-2 promoter adjacent to the downstream CRE site (−59/−53) and to the upstream pNFAT site (−76/−68) was first mentioned by
Fresno and colleagues (17) and Flamand and colleagues (18). By probing consensus sequences containing both CRE and AP-1 sites, Dunnemann and colleagues (21, 22) demonstrated that AP-1 components bind to the CRE site. However, the individual role of CRE and AP-1 could not be clearly differentiated because AP-1 components probably bind to the AP-1 and/or CRE sites. In the present study, DNA affinity protein-binding and gel shift assays were performed using probes containing CRE and AP-1 site consensus sequences, respectively. Thus, the components binding to the CRE or AP-1 site could be clearly distinguished.

The investigation of the relationship between \( H. \) pylori infection, COX-2 overexpression, and gastric cancer development is still lacking. In this study, we explored a novel intracellular network involving TLRs, MAPKs, and transcription factors binding to the CRE and AP-1 sites on the human COX-2 promoter, leading to COX-2 expression and \( \text{PGE}_2 \) release, which in turn promote gastric cancer cell invasion and angiogenesis.

**Materials and Methods**

**Bacterial strains and growth conditions**

\( H. \) pylori strains were isolated from patients undergoing gastroscopy for the evaluation of upper gastrointestinal symptoms. At the time of endoscopy, two biopsy specimens were taken from the antrum for bacterial culture. Different strains of clinical \( H. \) pylori isolates from patients with nonulcer dyspepsia with gastritis, duodenal ulcer, and gastric cancer were termed as HS, HD, and HC, respectively. Columbia agar with 5% sheep blood was used for \( H. \) pylori culture. The bacterial cells were cultured at 37°C in a microaerophilic chamber (Don Whitley) containing 10% CO\(_2\), 5% O\(_2\), and 85% N\(_2\).

**Cell culture and \( H. \) pylori infection experiments**

The human gastric cancer epithelial cell lines AGS (American Type Culture Collection (ATCC) no. CRL-1739) and MKN45 (Japanese Cancer Research Bank no. 0254) were obtained from the ATCC and RIKEN, respectively. Both of them were cultured in RPMI 1640 supplemented with 10% FBS, 100 U/ml penicillin, and 100 \( \mu \)g/ml streptomycin. AGS and MKN45 cells were cocultured with \( H. \) pylori in the antibiotics-free RPMI 1640 supplemented with 10% FCS. Bacterial cells were resuspended in PBS buffer (pH 7.4) and used for the infection experiment (14).

**Preparation of nuclear extracts and the EMSA**

AGS cells cocultured with \( H. \) pylori for the indicated time were harvested. The cell suspensions were collected and washed with PBS buffer (pH 7.4). The bacterial cells were pelleted. Cell pellets were then resuspended in PBS buffer (pH 7.4) and used for the infection experiment (14).

**In vitro invasion assay**

The invasion assay was conducted using the Transwell cell culture chambers (Corning Costar; No. 3422) according to the manufacturer’s recommendation with some modifications. Briefly, polyvinylpyrrolidone-free polycarbonate filters (8.0 \( \mu \)m pore size; Nucleopore) were precoated with 5 \( \mu \)g of Matrigel (BD Biosciences) on the upper surface. AGS or MKN45 cells were harvested with 1 mM EDTA and resuspended in the antibiotics-free RPMI 1640 supplemented with 0.1% FBS. The cell suspension (2.5 \( \times \) 10\(^5\) cells) was added to the upper compartment of the chamber. Six hours later, cells were cocultured with \( H. \) pylori after the pretreatment with either NS-398, celecoxib, or \( \text{PGE}_2 \) for 30 min. After 48 h incubation, the top side of the insert membrane was swabbed free of cells with a cotton swab. The bottom side was fixed with 3.7% formaldehyde and stained with 0.5% crystal violet in 20% methanol. The crystal violet dye retained on the filters was extracted with DMSO and colorimetrically assessed by measuring its absorbance at 590 nm on an ELISA reader (Bio-Tek).

**DNA affinity protein-binding assay (DAPA)**

Binding of transcriptional factors to the COX-2 promoter DNA sequences was assessed. After coculture with \( H. \) pylori for the indicated time, nuclear extracts were prepared. The bioin-labeling double-stranded oligonucleotides were synthesized based on the human COX-2 promoter sequence (25). “CRE + AP1 wt” (sense, 5’-biotin-AAGAACATCTGGTTTCTACATG-3’; antisense, 5’-CCATGCGAGAATGCTGGTCTTT-3’) contains CRE and AP1 binding sites; “CRE mut” (sense, 5’-biotin-AAGAACATCTGGTTTCTACATG-3’; antisense, 5’-CCATGCGAGAATGCTGGTCTTT-3’) contains wild-type (wt) CRE and AP1 and wt CRE. The sequence underlined means the “core region” for the transcription factor binding sites, and the sequence with italic type means the “mutated base pair.” The binding assay was performed by mixing 200 \( \mu \)g of nuclear extract proteins, 2 \( \mu \)g of biotin-labeled DNA oligonucleotides, and 20 \( \mu \)l of streptavidin agarose beads (4%) with 70% slurry. The mixture was incubated on a rotator for 1 h with shaking. The beads were pelleted and washed with cold PBS three times. The bound proteins were separated by SDS-PAGE, followed by Western blot analysis, and finally probed with specific Abs.

**PGE\(_2\) release ELISA**

\( \text{PGE}_2 \) was quantified from the collected culture media using ELISA by a commercial kit from Amersham Biosciences according to the manufacturer’s instructions.

**ChIP analysis**

ChIP analysis was performed as described (26). Immunoprecipitated DNA was purified and amplified using the COX-2 promoter region using the 5’ primers 5’-TACCGGGAATTCAGTCTGTCCACATG-3’ and 3’ primer 5’-CATTCGCTGCACTTGCACATGG-3’, which contains the API binding site; “API” 5’-AGAACATCTGGTTTCTACATG-3’, which contains the CRE binding site; “CRE” 5’-CATTCGCTGCACTTGCACATGG-3’, which contains the CRE binding site, were synthesized, annealed, and endlabeled with [γ-\( ^{32} \)P]ATP using T4 polynucleotide kinase. EMSA was performed as described previously (25). Four different antisera were performed. The polyclonal Abs specific to c-fos, c-jun, CREB-1, ATF-2, and p65 were added to the nuclear extracts 30 min prior to the binding reaction, and the DNA/nuclear protein complexes were separated on a 4.5% polyacrylamide gel.
2 µg of total RNA, which was reverse transcribed into cDNA using the oligo dT primer, then the cDNA was amplified for 30 cycles using two oligonucleotide primers derived from a published COX-2 sequence (5'-CACGACCCTACGCTACAGT-3' and 5'-CTCTGTCATGGAAGCCTG-3'), c-fos sequence (5'-GAATACATGGTCTGAGCAAAACTGCGCAA-3' and 5'-CCTGATGCAAGAAGGAGCAGAATT-3'), c-jun sequence (5'-GGAGGACCTTCTAGATCAGGCCTCAAC-3' and 5'-GAACACCTGCTCGATCTGACGTCCTTT-3'), and then oligonucleotide primers from a β-actin sequence (5'-GAGGAAGGTTCACCCCCACTGTGCCCATCTA-3' and 5'-CTAGAAGCATTTGGGCGGAGGATGGG-3'). For COX-2, a PCR cycle consisted of a denaturation step (94°C, 1 min), an annealing step (60°C, 1 min), and an elongation step (72°C, 1.5 min). The total reaction took 35 cycles, which was followed by an additional extension step (72°C, 7 min). For β-actin, PCR cycle was conducted for 30 s at 94°C, 30 s at 65°C, and 1 min at 70°C. The PCR products were subjected to electrophoresis in a 1.5% agarose gel. Quantitative data was obtained using a computing densitometer and ImageQuant software (Molecular Dynamics).

**Plasmids**

The COX-2 promoter construct pGS459 (−459/+9) was a generous gift from H. H. Wang (University of Texas, Houston, TX). CRE (CRE-Luc), API (API-Luc), and NF-κB (κB-Luc) luciferase reporters were from Stratagene. The dominant-negative mutant of ERK2 was from Dr. M. Cobb (South-Western Medical Center, Dallas, TX). p38 (T180A/Y185F) was from Dr. L. H. Wang (University of Texas, Houston, TX). The CRE site (53) on the human COX-2 promoter is added for exogenous PGE2 to induce COX-2 expression in malignant gastric cell invasion, a Matrigel assay was performed. The invasion of AGS cells was inhibited by two COX-2 inhibitors, NS-398 and celecoxib (Fig. 1E). To investigate the mechanism of COX-2-dependent gastric tumor cell invasion, exogenous PGE2 was added. PGE2 (50 nM) enhanced cell invasion and reversed the inhibition of NS-398 (Fig. 1E). Similar results were seen in MKN45 cells cocultured with HC (Fig. 1F). HD, which induced COX-2 expression and PGE2 release in the MKN45 cells, also elicited MKN45 cell invasion (Fig. 1F).

To investigate the in vitro angiogenesis caused by H. pylori, Matrigel tube formation assay was performed in HUVECs. The tube structures were significantly formulated when HUVECs were treated with the condition media collected from AGS or MKN45 cells cocultured with HC (Fig. 1G). Condition media collected from HS also induced a smaller extent of tube formation. When pretreated by NS-398, the tube formation induced by HC was incomplete and the number was reduced (Fig. 1G). These results demonstrated that PGE-release to the media due to COX-2 expression elicited tube formation in HUVECs.

**H. pylori acts through TLR2/TLR9-mediated COX-2 to induce tumor cell invasion and tube formation**

Because TLR2 and TLR9 were demonstrated to be involved in the H. pylori-induced COX-2 expression (14), their role in the H. pylori-induced cell invasion was examined. The AGS cell line overexpressing COX-2, AGS/COX-2, was established and used to examine the role of COX-2 in cell invasion and angiogenesis. As shown in Fig. 2A, this stable clone had an abundant expression of COX-2 and showed high invasiveness comparing to the mock, AGS/pCDNA3. Neutralizing Abs to TLR2 and TLR9 attenuated the H. pylori-induced cell invasion, but they did not affect the invasiveness of AGS/COX-2 cells (Fig. 2A). These results demonstrated that H. pylori induced cell invasion via TLR2/TLR9-mediated COX-2 expression in AGS cells.

Condition media collected from AGS/COX-2 induced a significant tube formation (Fig. 2B). Neutralizing Abs to TLR2 and TLR9 attenuated the H. pylori-induced tubular formation. However, they did not affect the tube formation induced by AGS/COX-2 cells (Fig. 2B). These results demonstrated that TLR2/TLR9-dependent COX-2 expression and PGE2 release mediated by H. pylori-induced tube formation in vitro.

**Statistical analysis**

Data were analyzed using Student’s t test. Values of p < 0.05 were considered significant.

**Results**

**COX-2-dependent cell invasion and angiogenesis induced by H. pylori from cancer patient**

Different strains of H. pylori from patients with gastric cancer (HC), gastritis (HS), and duodenal ulcer (HD) were analyzed for their capability to induce COX-2 expression and PGE2 release in the gastric epithelial cells, AGS and MKN45. As in our previous report (14) and as shown in Fig. 1, A and C, HC induced a time-dependent increase of COX-2 expression and PGE2 release in AGS cells. It was evident at 6 h after H. pylori infection and reached maximum at 24 h. A similar result was also observed in MKN45 cells after coculture with HC (Fig. 1, B and D). COX-2 expression and PGE2 release were seen in MKN45 cells induced by HD. However, the extent was smaller than that of HC. To evaluate the role of H. pylori-induced COX-2 expression in malignant gastric cell invasion, a Matrigel assay was performed. The invasion of AGS cells was inhibited by HC, but not HS and HD, and HC-induced AGS cell invasion was inhibited by the two COX-2 inhibitors, NS-398 and celecoxib (Fig. 1E).
FIGURE 1. COX-2-dependent tumor cell invasion and angiogenesis induced by *H. pylori* isolates from patients. AGS (A) or MKN45 (B) cells were cocultured with HC, HS, or HD at a bacterium to cell ratio of 150:1 for the indicated time. Whole cell lysates were prepared and subjected to Western blot using Ab specific to COX-2 or actin. PGE₂ releases in the media collected from AGS (C) or MKN45 (D) cells were measured using ELISA. The PGE₂ production was normalized to the total protein level. AGS (E) or MKN45 (F) cells were treated with 50 nM PGE₂ or cocultured with HS, HD, or HC for 48 h followed by pretreatment with 10 μM NS-398, 40 μM celecoxib, 10 nM PGE₂, or 50 nM PGE₂ on Transwell inserts coated with Matrigel. The percent invasion was measured by comparing with the basal (*, p < 0.05 as compared with the basal; **, p < 0.05 as compared with HC; #, p < 0.05 as compared with HC in the presence of NS-398). The images of invasive cells stained with crystal violet were shown in the lower panels. G, Photomicrographs depicted a random field of view and showed the alignment of HUVECs treated with condition media collected from gastric epithelial cells rendered with different treatments. "Basal" means the media collected from cells without any treatment. "HS" means the media collected from cells cocultured with HS for 16 h. "HC" means the media collected from cells cocultured with HC for 16 h. "HC + NS398" means the media collected from cells cocultured with HC for 16 h followed by pretreatment with NS398 for 30 min (right panel). Quantification of the extent of tube formation was shown as relative to the control (left panel). Data are presented as means ± SE of three independent experiments with six replicates per condition (*, p < 0.05 vs basal; **, p < 0.05 as compared with HC alone).
To further confirm the results from DAPA, supershift experiments were performed. Anti-ATF-2, c-jun, CREB1, but not c-fos and p65, Abs blocked the CRE-specific DNA-protein complex formation using probe with CRE consensus sequences (Fig. 3D, lanes 3–7). Anti-ATF-2, c-fos, c-jun, not CREB1 and p65, Abs blocked the AP1-specific DNA-protein complex formation using probe with AP1 consensus sequences (Fig. 3E, lanes 3–7). These data confirmed the bindings of CREB-1, ATF-2, and c-jun to the CRE site, and c-fos, c-jun, and ATF-2 to the AP1 site.

The in vivo recruitments of CREB1, ATF-2, c-jun, and c-fos to the COX-2 promoter induced by H. pylori were demonstrated by ChIP assay. As shown in Fig. 3F, time-dependent increases in the binding of these transcription factors to the promoter region -109/+7 containing CRE and AP1 sites were found.
FIGURE 3. Time-dependent increase of H. pylori-induced CREB-1, ATF-2, c-Fos, and c-Jun bindings to the COX-2 promoter in gastric epithelial cells. A, Schematic illustration of the consensus sequences of CRE plus AP-1 sites on the human COX-2 promoter labeled with biotin. The underlined shows the region corresponding to CRE and AP1 consensus sequences. Bases mutated in the CRE or AP1 site are indicated by bold letters. B and C, H. pylori-induced in vitro bindings of CREB-1, ATF-2, c-Fos, and c-Jun to the COX-2 promoter were examined. Time-dependent increases in the components binding to the CRE and AP1 sites, their transcriptional fos and/or c-Jun bindings to the COX-2 promoter in gastric epithelial cells. H. pylori, whereas SB203580 and SP600125 inhibited the c-Jun bindings to the COX-2 promoter in gas.

Regulation of H. pylori-induced c-fos and c-jun mRNA expressions, and CREB-1 and ATF-2 activations by MAPKs

Because c-fos and/or c-Jun were demonstrated to be the components binding to the CRE and AP1 sites, their transcriptional regulations were examined. Time-dependent increases in the c-fos and c-jun mRNA expressions in AGS cells infected by H. pylori were seen (Fig. 4A). Because the expressions of c-fos and c-Jun were regulated by MAPKs (28), the roles of ERK1/2, p38, and JNK in this event were examined. Similar to our previous findings (28), PD98059 (50 μM) blocked TNF-α-induced ERK1/2 activation without having any effect on the p38 and JNK activations, and SB 203580 (30 μM) caused complete inhibition on p38 activation without affecting ERK1/2 and JNK activations. SP600125 inhibited JNK1/2 activation in a dose-dependent manner without any effect on ERK1/2 and p38 activations (data not shown). PD98059 and SB203580 were shown to inhibit the H. pylori-induced c-fos mRNA expression, whereas SB203580 and SP600125 inhibited the c-jun mRNA expression (Fig. 4B, lanes 3 and 4; lanes 4–6). Taken together, these results indicated the involvement of ERK and p38, not JNK, in the regulation of c-fos mRNA expression and p38 and JNK, not ERK, in the c-jun mRNA expression.

Activations of CREB-1 and ATF-2 were examined by their phosphorylation. H. pylori-induced phosphorylation of CREB-1 in the nuclear extracts was seen after coculture with H. pylori for 10 min and sustaining for 120 min (Fig. 4C, lanes 2–6). Phosphorylation of ATF-2 in the nuclear extracts reached maximum at 10 min and declined at 60 min (Fig. 4E, lanes 2–4). The roles of MAPKs in the phosphorylations of CREB-1 and ATF-2 were examined. ERK and p38, not JNK, were found to regulate the phosphorylation of CREB-1. However, all three MAPKs were shown to regulate the ATF-2 phosphorylation (Fig. 4, D and F).

Involvements of TLR2/9, ERK, p38, and JNK in H. pylori-mediated COX-2 promoter activity via activations of CRE and AP-1

Because TLR2/9 and CRE and AP1 sites on the COX-2 promoter were demonstrated to be involved in the H. pylori-induced COX-2 expression, H. pylori-induced CRE and AP-1 activation through TLR2/9 was examined. As shown in Fig. 5, A
and B, H. pylori-induced CRE and AP-1 luciferase activities were inhibited by the TLR2 and TLR9 mutants, suggesting the involvements of these two TLRs. Because the binding components of CRE and AP-1 sites and their regulations by MAPKs were demonstrated, the roles of MAPKs in regulating the H. pylori-induced CRE and AP-1 luciferase activities were further confirmed using the dominant-negative p38, ERK2, and JNK mutants (Fig. 5, C and D).

**Discussion**

An increase in COX-2 expression has been implicated in the genesis of a variety of human cancers, including colorectal, gastric, pancreatic, esophageal, brain, and lung cancers (29). COX-2 may play a role as survival factor in protecting cells from apoptosis (30). Furthermore, functions like the encouragement of cell proliferation (31), the induction of tumor cell invasion (32, 33), the stimulation of angiogenesis (11), and the inhibition of immunosurveillance (34) have also been described. SC-236, a selective COX-2 inhibitor, was effective in reducing angiogenesis driven by the basic fibroblast growth factor in a Matrigel model (35). The expression of COX-2 in the newly formed blood vessels within tumors grown in animals was observed, whereas the quiescent vascula expressed only COX-1, indicating the involvement of COX-2-derived PGs in tumor angiogenesis (35). We demonstrated that H. pylori isolated from gastric cancer patients induced the COX-2 expression and PGE2 release, and they also increased the angiogenesis and cell invasion in the AGS and MKN45 cells. The highly specific COX-2 inhibitors, NS-398 and celecoxib, were shown to...
inhibit these phenomena. The introduction of exogenous PGE$_2$ reversed the inhibitory effect of NS-398. Similar positive correlation between PGs productions and tumor invasiveness was observed in the lung cells (32). Cell line transfected with COX-2 (AGS/COX-2) also exhibited high invasiveness and angiogenesis. Our previous study has shown that neutralizing Abs for TLR2 and TLR9 blocked H. pylori–induced COX-2 expression (14). In this study, they also attenuated the H. pylori–induced but not the AGS/COX-2 cells-induced invasion and angiogenesis, confirming the involvement of TLR2 and TLR9 in the effect of H. pylori. These
results were in line with the recent studies of *H. pylori* activating innate immunity and NF-κB signaling via TLR2 but not TLR4 (36–38), and finding of TLR9 expression on the gastric epithelial cell surface (39). Moreover, TLR2 and TLR9 agonists were found to promote an angiogenic switch (40). These findings together revealed a novel function of TLR2 and TLR9 in *H. pylori*-induced COX-2 expression, cell invasion, and angiogenesis. However, the addition of both anti-TLR2 and TLR9 Abs to the culture could not completely block cell invasion and tube formation (data not shown), indicating that other TLR receptors or alternative mechanisms may be functional.

The effectiveness of NS-398 or celecoxib on inhibiting cell invasion and angiogenesis of AGS and MKN45 gastric cancer cell lines points to a promising therapeutic target for gastric cancer prevention and treatment, as exemplified by the great success of COX-2 inhibitor in the colon cancer prevention and treatment (41). Therefore, COX-2 and its metabolites are the necessary “angiogenic” and “invasive/permissive” factors and are sufficient to induce tumor invasiveness and angiogenesis. Recently, the use of nonsteroidal anti-inflammatory drugs is also reported to be associated with a decreased risk of gastric cancer in a systematic review and metaanalysis of epidemiological evidence (42). Intriguingly, the protective effect of nonsteroidal anti-inflammatory drugs was observed only in individuals who were *H. pylori* IgG-positive but not in noninfected subjects (43).

The mechanism that *H. pylori* induced COX-2 expression at transcriptional level was further examined. Our recent data demonstrated the involvements of CRE (−59/−53), NF-IL6, and NF-κB sites in the *H. pylori*-induced COX-2 expression (14). Juttner et al. (44) also reported the involvement of CRE/E box (−56/−48) in *H. pylori*-induced COX-2 expression. However, we further verified and investigated the AP-1 element (−67/−61). The transcription factors of AP-1 family (Fos, Jun, and ATF) and CREB/ATF family (ATF and CREB) can form either homo- or heterodimer. Jun-Jun and Jun-Fos dimers preferentially bind to the AP-1 site, whereas Jun-ATF prefers binding to the CRE sequence (45). Using DAPA and supershift assays, the respective bindings of CREB-1, ATF-2, and c-jun to the CRE site, and c-fos, c-jun, and ATF-2 to the AP1 site after *H. pylori* coculture were demonstrated. In addition, ChIP assays demonstrated the in vivo recruitments of CREB-1, ATF-2, c-jun, and c-fos to the proximal sites of COX-2 promoter (−109/+7). We are the first to clearly verify the respective binding components of CRE and AP-1 sites on the COX-2 promoter both in vivo and in vitro. Dannenberg and colleagues and others claimed the binding of AP-1 components to the CRE site using oligonucleotide consensus sequences containing both CRE and AP-1 sites (21, 22, 46). In their experiments, the individual role of CRE and AP-1 could not be clearly differentiated because they claimed the bindings of c-fos, c-jun, and ATF-2 (AP-1 components) to the CRE site (21). However, Bowden and colleagues (47) found the bindings of CREB and ATF-1 (CREB components) to the CRE site using similar probes. We and Flamm and colleagues (18) used oligonucleotides containing respective AP-1 site- and CRE site-specific consensus binding sequences. Flamm and colleagues also demonstrated the involvements of CRE and AP-1 sites in the COX-2 promoter activation in human herpes virus 6-infected monocytes. They found the binding of CREB-1 to the CRE site and c-jun to the AP-1 site (18). Therefore, respective binding components of CRE and AP-1 sites on the COX-2 promoter were explored. Our unpublished data demonstrated the specific binding of CREB/ATF-2 to the CRE site (−53/−45), and c-fos/c-jun to the AP-1 site (−936/−930) on the cyclin D1 promoter after *H. pylori* infection. The additional bindings of c-jun to the CRE site and ATF-2 to the AP1 site in the present study might be due to the adjacency of these two elements on the COX-2 promoter, as they are far apart on the cyclin D1 promoter.

Present studies explored that *H. pylori* induced activations of CRE and AP-1 through MAPKs pathways. As lung A549 cells expressing ICAM-1 and releasing IL-8 through MAPKs-induced AP-1 activation (28, 48), ERK and p38 regulate c-fos mRNA expression, and JNK and p38 regulate c-jun mRNA expression in the present AGS cells. Activations of ERK and c-fos expression by *H. pylori* have also been noted (49). CREB-dependent transcription is regulated through the phosphorylation of serine 133, which was first found to be mediated by PKA (50). Our finding provided evidence for an alternative mechanism whereby CREB was phosphorylated by MAPKs as found by Yu et al. (51). ATF-2 has been reported to be activated by JNK and p38 (52, 53). In this study, we showed that ATF-2 was activated by all three MAPKs. Therefore, the binding components of CRE and AP-1 sites are differentially regulated by the MAPKs. Intriguingly, p38 could regulate all these transcription factors.

In summary, we provided a mechanistic view of COX-2 in the *H. pylori*-mediated carcinogenesis. *H. pylori* was shown to act through TLR2 and TLR9 to activate the MAPKs (ERK1/2, p38, JNK) and their downstream transcription factors (CREB-1, ATF-2, c-fos, and c-jun), resulting in the activations of CRE and AP-1 on the COX-2 promoter. These intracellular networks drive the COX-2-dependent PGE2 release, and they may contribute to the carcinogenesis and metastasis in patients colonized with these strains through the enhancement of tumor cell invasion and angiogenesis. A schematic diagram exhibiting the
comprehensive intracellular networks in gastric cancer cells is displayed in Fig. 6, and the application of the selective COX-2 inhibitor may provide an alternative therapy to reduce the development of gastric cancer.

Disclosures

The authors have no financial conflict of interest.

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